



Stochastic processes in gravitropism

Citation

Meroz, Yasmine, and Renaud Bastien. 2014. "Stochastic processes in gravitropism." *Frontiers in Plant Science* 5 (1): 674. doi:10.3389/fpls.2014.00674. <http://dx.doi.org/10.3389/fpls.2014.00674>.

Published Version

doi:10.3389/fpls.2014.00674

Permanent link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:13581122>

Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA>

Share Your Story

The Harvard community has made this article openly available.
Please share how this access benefits you. [Submit a story](#).

[Accessibility](#)



Stochastic processes in gravitropism

Yasmine Meroz* and Renaud Bastien*

Applied Mathematics, School of Engineering and Applied Science, Harvard University, Cambridge, MA, USA

Edited by:

Karen Alim, Harvard University, USA

Reviewed by:

Jeanne Marie Harris, University of Vermont, USA

Jacques Dumais, Universidad Adolfo Ibáñez, Chile

***Correspondence:**

Yasmine Meroz and Renaud Bastien, Applied Mathematics, School of Engineering and Applied Science, Harvard University, 29 Oxford St., Cambridge, MA 02138, USA
e-mail: ymeroz@seas.harvard.edu; rbastien@seas.harvard.edu

In this short review we focus on the role of noise in gravitropism of plants – the reorientation of plants according to the direction of gravity. We briefly introduce the conventional picture of static gravisensing in cells specialized in sensing. This model hinges on the sedimentation of statoliths (high in density and mass relative to other organelles) to the lowest part of the sensing cell. We then present experimental observations that cannot currently be understood within this framework. Lastly we introduce some current alternative models and directions that attempt to incorporate and interpret these experimental observations, including: (i) *dynamic sensing*, where gravisensing is suggested to be enhanced by stochastic events due to thermal and mechanical noise. These events both effectively lower the threshold of response, and lead to small-distance sedimentation, allowing amplification, and integration of the signal. (ii) The role of the cytoskeleton in signal-to-noise modulation and (iii) in signal transduction. In closing, we discuss directions that seem to either not have been explored, or that are still poorly understood.

Keywords: stochasticity, gravitropism, signal amplification, gradual response, stochastic resonance, noise, cytoskeleton

1. INTRODUCTION

Tropism is the reorientation of a plant organ according to the direction of an external stimulus, one example of which is gravitropism (reorientation relative to the direction of gravity). While the first scientific investigations took place roughly 300 years ago (Dodart, 1703; Astruc, 1709), fundamental aspects of underlying mechanisms still remain elusive. Generally, the main steps of tropisms include the sensing of a stimulus, transduction of a signal, and lastly the response of the organ. Any sensory system is confronted with three main challenges: (i) sensitivity to weak stimuli, (ii) robustness to noise (be it internal or external), and (iii) a gradual response. Sensorimotor systems in other living organisms are known to be affected by noise at any one of these stages, and a large body of work investigates these phenomena (Mortensen and Suhl, 1991; Douglass et al., 1993a; Patel and Kosko, 2005; Faisal et al., 2008; Li et al., 2012). Only in the recent past has the role of noise been known to play a crucial role in tropisms. We note that all types of tropism rely on exquisitely sensitive and robust sensors. In light of this, the principal aim of this review is to establish the role of noise in gravitropism. We shall present the conventional model of gravitropism, and then point out a number of experimental observations that do not comply with it, while identifying various sources of noise at different stages of signal perception and transduction. Finally we shall indicate current alternative models and directions that attempt to incorporate these experimental observations, focusing on the role of noise.

2. GRAVITROPISM

Gravitropism occurs both in roots and shoots. Roots manifest positive gravitropism, reorienting themselves toward the

direction of gravity, while shoots manifest negative gravitropism, reorienting in the opposite direction (Sack, 1991). Cells, called statocytes, have been identified as being specialized in sensing gravity. In roots these are located in the root cap, and in shoots they form a cylindrical sheath surrounding the stem along the shoot (Morita and Tasaka, 2004). Recognizing that gravity acts on mass, the perceiving elements of these sensory cells have generally been identified as organelles with high mass and density (relative to other organelles). These organelles are termed *statoliths*. Starch-filled organelles called *amyloplasts* have been determined to play this role, though not exclusively as shall be seen later (Blancaflor and Masson, 2003; Perbal and Driss-Ecole, 2003; Morita and Tasaka, 2004; Moulija and Fournier, 2009). Upon displacement of the organ from its vertical orientation, the statoliths sediment to the new low point of the sensing cell. The cell is then able to recognize this change in orientation and transmits an associated signal. However, we shall see in the next Section that this picture does not account for a number of experimental observations.

Before proceeding it is interesting to note that stochastic processes dictate the bounds of both the sensitivity of gravity sensors (through thermal noise) and their selectivity (through the activation energy) (Björkman, 1988; Hasenstein, 2009). The sedimentation of statoliths is a mechanical process, and as such is affected by both mechanical and thermal noise (Björkman, 1988, 1992). Both gravity and thermal energy cause the gravity sensor to move, but thermal energy depends on temperature alone, and not on mass. Therefore, thermal energy acts as background noise that the gravity sensor must overcome, constituting a lower bound on sensitivity (Perbal et al., 2002). As for selectivity, the sensor should be selectively activated through a reaction by the stimulus

rather than thermal motion. Higher selectivity can be obtained by raising the activation energy of the reaction high enough so that it is rarely stimulated spontaneously, but this also translates to a higher threshold stimulus, meaning that there is a trade-off between the sensor's sensitivity and its selectivity (Björkman, 1988, 1992).

3. CAVEATS OF THE CLASSIC MODEL OF AMYLOPLAST SEDIMENTATION: EXPERIMENTAL OBSERVATIONS

We present a few experimental observations that do not agree with the classical model of static sedimentation of amyloplasts. Firstly, presentation time (the shortest stimulus needed for a response) can be as short as 7 s (Larsen, 1969). The average amyloplast sedimentation rate is estimated to be of the order of $0.154 \mu\text{m}/\text{min}$ (Perbal, 2009). Given that the width of a statocyte cell is of the order of $10 \mu\text{m}$, it should take the amyloplast several minutes to sediment to the bottom – a couple of orders magnitude longer than the mentioned presentation time.

Gravitropic response has been found to be highly sensitive; experiments carried out in microgravity or clinostatting, have revealed a threshold of $10^{-4} g$ (Shen-Miller et al., 1968; Merkis et al., 1985). Such high sensitivity requires large responding masses such as amyloplasts, and signal averaging. Indeed oat coleoptiles respond equally to 1 s stimuli administered every 5 s, and to 0.5 s stimuli administered every 1 s (Raven and Rubery, 1982), also suggesting signal averaging or integration.

Mutants with 60% of the starch of the wild type have been found to exhibit almost the same degree of sensitivity as the wild type (Kiss et al., 1996). In starchless mutants statoliths are lighter and therefore do not sediment. However, these mutants are still able to sense gravity, though with increased variability in the shoots orientation (Caspar and Pickard, 1989; Kiss et al., 1989; Vitha et al., 2000). Starchless plants therefore seem to rely on smaller masses, leading to an increased relative noise, as exhibited by the increased variability. Indeed apart from amyloplasts, prominent in higher plants, other statoliths have been identified: Chara relies on BaSO₄-enriched vesicles (Hemmersbach et al., 1999), and fungi such as Phycomyces rely on protein crystals (Schimek et al., 1999). These statoliths have a much lower mass compared to amyloplasts, inconsistent with the requirement of large responding masses, and making them considerably weaker sensors.

Ma and Hasenstein (2007) investigated the possible dependency of gravisensing on thermal or mechanical noise, by studying the effect of temperature and external mechanical oscillations on the rate of gravitropic curvature on flax. They found that vertical oscillations prior to reorientation enhance gravisensing. The most effective enhancement occurred for oscillations of around 5 Hz and 0.5 mm amplitude (10 or 20 Hz did not promote curvature), reducing the time to reach half-maximal curvature by 18 min compared to non-oscillated roots (almost half the time). Similarly to mechanostimulation, temperature was also found to enhance gravitropic response. For vibration and temperature studies, the presentation times decreased almost 6-fold. Together with other work where mechanoperception was found to be enhanced by temperature (Douglass et al., 1993a,b), this may

suggest that thermal noise plays a role in the process. The extent of this role needs to be better quantified since temperature also affects other biological processes, some of which, as shall be seen later, affect sensitivity.

The word statolith is derived from the Greek, meaning “stationary stone” – however saltatory movement of statoliths have been recorded (Morita, 2010): for example amyloplasts in corn coleoptiles (Sack et al., 1984), in maize root columella cells (Sack et al., 1986), and in the stem endodermis (Saito et al., 2005). In some cases the origin of the saltatory movements has been identified: actin-based lifting of statoliths in Chara rhizoids (Braun et al., 2002), in Arabidopsis endodermal cells amyloplasts perform F-actin-dependent saltatory movements (Saito et al., 2005). Indeed some of these saltations are non-Brownian movements constituting a simple type of cytoplasmic streaming (Sack and Leopold, 1985).

Recent work has identified the actin cytoskeleton as a crucial component in plant gravitropism (Blancaflor, 2002, 2013), both at the level of signal perception and transduction. Disrupting F-actin enhances sensitivity and causes an over-response in curvature for roots (Hou et al., 2003, 2004) and shoots (Yamamoto and Kiss, 2002). Studies also indicate that disrupting F-actins with Lat-B enhances the movement of some amyloplasts (Hou et al., 2004), but reduces the movement of the vast majority of amyloplasts (Palmieri and Kiss, 2005; Saito et al., 2005). Instead, inhibition of myosin ATPase activity with BDM increases amyloplast movement in vertically oriented hypocotyls. After reorientation, amyloplasts exhibit less displacement in response to gravity, continuing to saltate, and the organ curves less in response to gravity (Palmieri et al., 2007).

In summary, the classical model of static sensing of amyloplasts sedimenting to the lowest point in the sensing cell does not generally comply with the following observations: (i) the sedimentation speed of amyloplasts is orders of magnitude slower than the speed required to explain recorded presentation times, (ii) highly sensitive gravitropic response and signal integration (iii) gravity can also be sensed via much weaker sensors than amyloplasts (although with greater variance of response curves), (iv) applying mechanical or thermal noise results in enhanced sensing, (v) saltatory movements of statoliths. Moreover, the role of the cytoskeleton in gravitropism, both at the level of signal perception and transduction, needs to be understood.

4. CURRENT ALTERNATIVE MODELS AND DIRECTIONS

We now present some current alternative models and directions that attempt to incorporate and interpret these experimental observations. We focus on the role of the cytoskeleton in both signal-to-noise modulation and signal transduction, and the dynamic sensing model which suggests stochastic events enhance gravisensing via different approaches. The underlying notion is that noise has a central role in regulating and enhancing the gravitropic responses in plants.

4.1. THE ROLE OF THE CYTOSKELETON IN SIGNAL-TO-NOISE MODULATION

Several studies have proposed that amyloplasts and the actomyosin system are part of a feedback mechanism that fine-tunes

the gravitropic response (Hou et al., 2004; Palmieri and Kiss, 2005; Saito et al., 2005). Indeed the cytoskeleton has been found to affect saltatory movements of amyloplasts, as mentioned in Section 3. One can interpret these results also in terms of changes in the signal-to-noise ratio (Palmieri et al., 2007): As mentioned earlier, the movement of the vast majority of amyloplasts is inhibited when the actin cytoskeleton is disrupted (including saltatory movements, i.e., reduction of noise). The few mobile amyloplasts therefore impart a gravity signal, which, in the absence of noise, may be perceived as amplified. Therefore, amyloplasts may be seen as indicators of noise and the direction of gravity signal. In this context the cytoskeleton can be seen as modulating the signal-to-noise ratio.

4.2. DYNAMIC SENSING: STOCHASTIC EVENTS ENHANCE GRAVISENSING

Hasenstein (2009) attempts to propose an alternative model to the classic picture of amyloplast sedimentation. Following the caveats presented in Section 3, the gravisensing system must respond to mechanical and thermodynamic noise, be compatible with the undisputed function of statoliths but must also be able to function in the absence of amyloplasts. Rather than perceiving gravity statically at the lowermost site of sedimentation, which produces one stimulus per settled statolith, it is suggested (Hasenstein, 2009) that stochastic events due to thermal and mechanical noise (for example due to cytoplasmic streaming (Kato et al., 2002; Morita et al., 2002) and cellular activities such as cytoskeletal tread milling), affect any susceptible particle in two ways: (i) effective lowering of mechano-stimulation, allowing smaller masses to act as statoliths, and (ii) gravisensing occurs via continuous small-distance sedimentation, allowing the amplification, and integration of the signal.

Let us expand on these two points. Stochastic events cause smaller masses to exceed the minimum energy needed to activate the ensuing signaling process. This is in agreement with *stochastic resonance*, a stochastic process where noise effectively lowers the threshold of mechano-stimulation, thus allowing to discern weak signals (Wiesenfeld and Moss, 1995). Stochastic resonance has been found to play a role in many mechano-transduction systems, ranging from crayfish (Douglass et al., 1993a) to hair cells in animal and human auditory system (Jaramillo and Wiesenfeld, 1998, 2000; Indresano et al., 2003). Secondly, it is suggested (Hasenstein, 2009) that gravisensing occurs via short distance sedimentation, rather than static perception at the lowermost site of sedimentation which produces one stimulus per settled statolith. This is in accordance with the saltatory motion described in Section 3. In this case sedimentation events take place continuously, constantly stimulating the system, allowing the amplification, and integration of the signal. The resulting signal is therefore continuously updated and dynamic.

In this context, heavier particles may lead to more interactions (and more energetic ones) between statoliths and membranes, cytoskeletal elements, or other sensitive structures, including vacuoles (Kato et al., 2002). Gravity affects the distribution of the combined statoliths and detectable events, which increase in the direction of the gravity vector.

4.3. THE ROLE OF THE CYTOSKELETON IN SIGNAL TRANSDUCTION

The interaction between amyloplasts and F-actin has already been mentioned earlier in relation to gravity perception and noise. In the context of signal transduction, it has been proposed that the sedimentation of amyloplasts through the F-actin network inside the statocyte may possibly impact a mechano-sensitive surface at the cell periphery (for reviews, see Perbal and Driss-Ecole, 2003). Alternatively, amyloplasts may disrupt the tensional integrity of the F-actin network, mechanically stimulating ion channels in the plasma membrane Palmieri et al., 2007.

Hasenstein (2009) also suggests that some correcting activity is associated with the (F-actin) cytoskeleton, i.e., regulation of the signal transduction: Short-distance sedimentation of statoliths and other particle lead to interactions between statoliths and membranes, cytoskeletal elements, or other sensitive structures, including vacuoles (Kato et al., 2002). These events may trigger a signal associated with the membrane, such as the opening or activation of stretch-activated channels (Perbal et al., 2004) or auxin efflux carriers (PIN3 proteins). Related investigations on mechano-transduction in human leukemia cells show that F-actin disassembly reduces the amplitude of stretch-activated currents but not the probability of channel opening (Staruschenko et al., 2005), suggesting that some correcting activity associated with the (F-actin) cytoskeleton is necessary to regulate the channels. As long as gravistimulation occurs, basal channels either on the endoplasmic reticulum or plasma membrane may open and provide discrete but frequent signals that indicate the direction of the gravitational vector. Disruption of the actin filaments may prevent closure and cause persistent channel conductivity, compatible with experimental observations where disrupting F-actin enhances the sensitivity and cause an over-response in curvature for roots (Hou et al., 2003, 2004) and shoots (Yamamoto and Kiss, 2002).

5. DISCUSSION

In this short review we focused on the role of noise in the gravitropic response of plants. We described the conventional model based on the sedimentation of high-mass statoliths to the lowest part of the sensing cell, and continued to present experimental observations that cannot currently be understood within this framework. We then pointed out some current alternative models and directions, that attempt to incorporate and interpret these experimental observations. These include the role of the cytoskeleton in signal-to-noise modulation and signal transduction, and the dynamic sensing model, where stochastic events enhance gravisensing. The common thread to all of these is the idea that intrinsic noise has a central role in regulating and enhancing the gravitropic responses in plants. Having said that, much is still not fully understood in the sensing and signal transduction mechanisms, and more so the role of noise in these. We propose here a number of directions that seem to have either not been explored, or that are still poorly understood.

- (i) Accommodation – gravitropic sensitivity is found to depend on the strength of the stimulus (more sensitive for weaker stimuli), reminiscent of the Weber-Fechner law of perception (Moullia et al., 2006; Moullia and Fournier, 2009).

Indeed, root statocytes have been shown to be more sensitive in microgravity compared to those grown on a 1 g centrifuge in space (Perbal and Driss-Ecole, 2003).

- (ii) Teasing apart sources of noise at different stages of perception: the initial sensing, signal transduction, and finally organ response. An example in gravitropism is that any external mechanical stimulus can only affect the initial sensory system, not the down-stream biochemical signal transduction. This sort of argument is more elusive when differing between the signal transduction and the organ response.
- (iii) Mathematical modeling – in recent years there has been a rise in mathematical modeling at the molecular, cellular and organ level, and has proven to offer valuable insights in the tropism mechanisms (e.g., Shafir and Forgacs, 2002; Moulia and Fournier, 2009; Bastien et al., 2013 and many others). Current mathematical models lack a stochastic framework that would address the addition of noise to the system. One way of taking this into consideration is to add noise into dynamic equations of, e.g., the curvature of the shoot (Bastien et al., 2013), reminiscent of a Langevin equation. In line with the previous point, it is important to distinguish between different sources of noise, and thus incorporate them in a meaningful way. By doing so one may attribute the noise to different terms: the response, proprioception, graviception or even the stimulus itself. Secondly, the stochastic mechanisms mentioned here require modeling and may be incorporated within an adequate integrative mathematical framework. Such an understanding may help pose a new understanding of the problem.

ACKNOWLEDGMENT

We wish to thank L. Mahadevan for fruitful discussions and financial support, and Mattia Gazzola for helpful comments on the manuscript. Yasmine Meroz is an Awardee of the Weizmann Institute of Science National Postdoctoral Award Program for Advancing Women in Science.

REFERENCES

- Astruc, J. (1709). Conjecture sur le redressement des plantes inclinés à l'horizon. *Mem. Acad. R. Sci. Paris* 1708, 463–470.
- Bastien, R., Bohr, T., Moulia, B., and Douady, S. (2013). Unifying model of shoot gravitropism reveals proprioception as a central feature of posture control in plants. *Proc. Natl. Acad. Sci. U.S.A.* 110, 755–760. doi: 10.1073/pnas.1214301109
- Björkman, T. (1988). "Perception of gravity by plants," in *Advances in Botanical Research*, Vol. 15, ed J. A. Callow (San Diego, CA: Academic Press), 1–41
- Björkman, T. (1992). Perception of gravity by plants. *Adv. Space Res.* 12, 195–201. doi: 10.1016/0273-1177(92)90283-4
- Blancaflor, E. B. (2002). The cytoskeleton and gravitropism in higher plants. *J. Plant Growth Regul.* 21, 120–136. doi: 10.1007/s003440010041
- Blancaflor, E. B. (2013). Regulation of plant gravity sensing and signaling by the actin cytoskeleton. *Am. J. Bot.* 100, 143–152. doi: 10.3732/ajb.1200283
- Blancaflor, E. B. and Masson, P. H. (2003). Plant gravitropism. Unraveling the ups and downs of a complex process. *Plant Physiol.* 133, 1677–1690. doi: 10.1104/pp.103.032169
- Braun, M., Buchen, B., and Sievers, A. (2002). Actomyosin-mediated statolith positioning in gravisensing plant cells studied in microgravity. *J. Plant Growth Regul.* 21, 137–145. doi: 10.1007/s003440010052
- Caspar, T. and Pickard, B. (1989). Gravitropism in a starchless mutant of *Arabidopsis* - implications for the starch-statolith theory of gravity sensing. *Planta* 177, 185–197. doi: 10.1007/BF00392807
- Dodart, D. (1703). Sur l'affectation de la perpendiculaire remarquable dans toutes les tiges, dans plusieurs racines, et autant qu'il est possible sur toutes les branches des plantes. *Mem. Acad. R. Sci. Paris* 1700, 47–63.
- Dougllass, J. K., Wilkens, L., Pantazelou, E., and Moss, F. (1993a). Noise enhancement of information transfer in crayfish mechanoreceptors by stochastic resonance. *Nature* 365, 337–340. doi: 10.1038/365337a0
- Dougllass, J. K., Wilkens, L. A., and Moss, F. (1993b). "Noise-assisted information transfer in crayfish mechanoreceptors: stochastic resonance in a neuronal receptor," in *Proceedings of the SPIE 2036, Chaos in Biology and Medicine*, Vol. 2036 (San Diego, CA: SPIE), 152–161. doi: 10.1117/12.162707
- Faisal, A. A., Selen, L. P. J., and Wolpert, D. M. (2008). Noise in the nervous system. *Nat. Rev. Neurosci.* 9, 292303. doi: 10.1038/nrn2258
- Hasenstein, K. H. (2009). Plant responses to gravity - insights and extrapolations from ground studies. *Gravit. Space Biol.* 22, 21–32.
- Hemmersbach, R., Volkman, D., and Hader, D. P. (1999). Graviorientation in protists and plants. *J. Plant Physiol.* 154, 1–15. doi: 10.1016/S0176-1617(99)80311-3
- Hou, G., Kramer, V., Wang, Y., Chen, R., Perbal, G., Gilroy, S., et al. (2004). The promotion of gravitropism in *Arabidopsis* roots upon actin disruption is coupled with the extended alkalization of the columella cytoplasm and a persistent lateral auxin gradient. *Plant J.* 39, 113–125. doi: 10.1111/j.1365-313X.2004.02114.x
- Hou, G., Mohamalawari, D., and Blancaflor, E. (2003). Enhanced gravitropism of roots with a disrupted cap actin cytoskeleton. *Plant Physiol.* 131, 1360–1373. doi: 10.1104/pp.014423
- Indresano, A., Frank, J., Middleton, P., and Jaramillo, F. (2003). Mechanical noise enhances signal transmission in the bullfrog sacculus. *J. Assoc. Res. Otolaryngol.* 11, 363–370. doi: 10.1007/s10162-002-3044-4
- Jaramillo, F. and Wiesenfeld, K. (1998). Mechano-electrical transduction assisted by Brownian motion: a role for noise in the auditory system. *Nat. Neurosci.* 1, 384–388. doi: 10.1038/1597
- Jaramillo, F. and Wiesenfeld, K. (2000). Physiological noise level enhances mechano-electrical transduction in hair cells. *Chaos Soliton. Fract.* 11, 1869–1874. doi: 10.1016/S0960-0779(99)00123-X
- Kato, T., Morita, M., and Tasaka, M. (2002). Role of endodermal cell vacuoles in shoot gravitropism. *J. Plant Growth Regul.* 21, 113–119. doi: 10.1007/s003440010047
- Kiss, J., Hertel, R., and Sack, F. (1989). Amyloplasts are necessary for full gravitropic sensitivity in roots of *Arabidopsis thaliana*. *Planta* 177, 198–206. doi: 10.1007/BF00392808
- Kiss, J., Wright, J., and Caspar, T. (1996). Gravitropism in roots of intermediate-starch mutants of *Arabidopsis*. *Physiol. Plant.* 97, 237–244. doi: 10.1034/j.1399-3054.1996.970205.x
- Larsen, P. (1969). The optimum angle of geotropic stimulation and its relation to the starch statolith hypothesis. *Physiol. Plant.* 22, 469–488. doi: 10.1111/j.1399-3054.1969.tb07401.x
- Li, Z., Li, Y., Yi, Y., Huang, W., Yang, S., Niu, W., et al. (2012). Dissecting a central flip-flop circuit that integrates contradictory sensory cues in *C. elegans* feeding regulation. *Nat. Commun.* 3, 776. doi: 10.1038/ncomms1780
- Ma, Z. and Hasenstein, K. H. (2007). Noise amplification of plant gravisensing. *Adv. Space Res.* 39, 1119–1126. doi: 10.1016/j.asr.2006.12.005
- Merkis, A. I., Laurinavichyus, R. S., and Shvyagzhedne, D. V. (1985). Gravitational sensitivity and growth of plants in weightlessness. *Izvest. Akad. Nauk SSSR: Ser. Fizich.* 49, 715–723.
- Morita, M., Kato, T., Nagafusa, K., Saito, C., Ueda, T., Nakano, A., et al. (2002). Involvement of the vacuoles of the endodermis in the early process of shoot gravitropism in *Arabidopsis*. *Plant Cell* 14, 47–56. doi: 10.1105/tpc.010216
- Morita, M. T. (2010). Directional gravity sensing in gravitropism. *Annu. Rev. Plant Biol.* 61, 705–720. doi: 10.1146/annurev.arplant.043008.092042
- Morita, M. T. and Tasaka, M. (2004). Gravity sensing and signaling. *Curr. Opin. Plant Biol.* 7, 712–718. doi: 10.1016/j.pbi.2004.09.001
- Mortensen, U. and Suhl, U. (1991). An evaluation of sensory noise in the human visual system. *Biol. Cybern.* 66, 37–47. doi: 10.1007/BF00196451

- Mouliat, B., Coutand, C., and Lenne, C. (2006). Posture control and skeletal mechanical acclimation in terrestrial plants: implications for mechanical modeling of plant architecture. *Am. J. Bot.* 93, 1477–1489. doi: 10.3732/ajb.93.10.1477
- Mouliat, B. and Fournier, M. (2009). The power and control of gravitropic movements in plants: a biomechanical and systems biology view. *J. Exp. Bot.* 60, 461–486. doi: 10.1093/jxb/ern341
- Palmieri, M. and Kiss, J. Z. (2005). Disruption of the F-actin cytoskeleton limits statolith movement in Arabidopsis hypocotyls. *J. Exp. Bot.* 56, 2539–2550. doi: 10.1093/jxb/eri248
- Palmieri, M., Schwind, M., Stevens, M., Edelmann, R., and Kiss, J. (2007). Effects of the myosin ATPase inhibitor 2,3-butanedione monoxime on amyloplast kinetics and gravitropism of Arabidopsis hypocotyls. *Physiol. Plant.* 130, 613–626. doi: 10.1111/j.1399-3054.2007.00926.x
- Patel, A. and Kosko, B. (2005). Stochastic resonance in noisy spiking retinal and sensory neuron models. *Neural Netw.* 18, 467478. doi: 10.1016/j.neunet.2005.06.031
- Perbal, G. (2009). From ROOTS to GRAVI-1: twenty five years for understanding how plants sense gravity. *Microgravity Sci. Technol.* 21, 3–10. doi: 10.1007/s12217-008-9064-x
- Perbal, G. and Driss-Ecole, D. (2003). Mechanotransduction in gravisensing cells. *Trends Plant Sci.* 8, 498–504. doi: 10.1016/j.tplants.2003.09.005
- Perbal, G., Jeune, B., Lefranc, A., Carnero-Diaz, E., and Driss-Ecole, D. (2002). The dose–response curve of the gravitropic reaction: a re-analysis. *Physiol. Plant.* 114, 336–342. doi: 10.1034/j.1399-3054.2002.1140302.x
- Perbal, G., Lefranc, A., and Driss-Ecole, D. (2004). Mechanotransduction in root gravity sensing cells. *Physiol. Plant.* 120, 303–311. doi: 10.1111/j.0031-9317.2004.0233.x
- Raven, J. A. and Rubery, P. H. (1982). Coordination of development Hormone receptors, hormone action, and hormone transport. in *The Molecular Biology of Plant Development*, eds H. Smith and D. Grierson (Oxford: Blackwell), 28–48.
- Sack, F. and Leopold, A. (1985). Cytoplasmic streaming affects gravity-induced amyloplast sedimentation in maize coleoptiles. *Planta* 164, 56–62. doi: 10.1007/BF00391025
- Sack, F., Suyemoto, M., and Leopold, A. (1984). Kinetics of amyloplast sedimentation in gravistimulated maize coleoptiles. *Planta* 161, 459–464. doi: 10.1007/BF00394578
- Sack, F., Suyemoto, M., and Leopold, A. (1986). Amyloplast sedimentation and organelle saltation in living corn columella cells. *Am. J. Bot.* 73, 1692–1698. doi: 10.2307/2444235
- Sack, F. D. (1991). Plant gravity sensing. *Int. Rev. Cytol.* 127, 193–252. doi: 10.1016/S0074-7696(08)60695-6
- Saito, C., Morita, M. T., Kato, T., and Tasaka, M. (2005). Amyloplasts and vacuolar membrane dynamics in the living graviperceptive cell of the Arabidopsis inflorescence stem. *Plant Cell* 17, 548–558. doi: 10.1105/tpc.104.026138
- Schimek, C., Eibel, P., Horie, T., Galland, P., and Ootaki, T. (1999). Protein crystals in phycomyces sporangiophores are involved in graviperception. *Adv. Space Res.* 24, 687–696. doi: 10.1016/S0273-1177(99)00400-7
- Shafir, Y. and Forgacs, G. (2002). Mechanotransduction through the cytoskeleton. *Am. J. Physiol. Cell Physiol.* 282, C479–C486. doi: 10.1152/ajpcell.00394.2001
- Shen-Miller, J., Hinchman, R., and Gordon, S. (1968). Thresholds for georesponse to acceleration in gravity-compensated a vena seedlings. *Plant Physiol.* 43, 338–344. doi: 10.1104/pp.43.3.338
- Statuschenko, A., Negulyaev, Y., and Morachevskaya, E. (2005). Actin cytoskeleton disassembly affects conductive properties of stretch-activated cation channels in leukaemia cells. *Biochim. Biophys. Acta* 1669, 53–60. doi: 10.1016/j.bbmem.2005.02.013
- Vitha, S., Zhao, L., and Sack, F. (2000). Interaction of root gravitropism and phototropism in Arabidopsis wild-type and starchless mutants. *Plant Physiol.* 122, 453–461. doi: 10.1104/pp.122.2.453
- Wiesenfeld, K. and Moss, F. (1995). Stochastic resonance and the benefits of noise - from ice ages to crayfish and squids. *Nature* 373, 33–36. doi: 10.1038/373033a0
- Yamamoto, K. and Kiss, J. (2002). Disruption of the actin cytoskeleton results in the promotion of gravitropism in inflorescence stems and hypocotyls of Arabidopsis. *Plant Physiol.* 128, 669681. doi: 10.1104/pp.010804

Conflict of Interest Statement: The Guest Associate Editor Karen Alim declares that, despite being affiliated to the same institution as the authors, the review process was handled objectively and no conflict of interest exists. The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 29 August 2014; accepted: 12 November 2014; published online: 26 November 2014.

Citation: Meroz Y and Bastien R (2014) Stochastic processes in gravitropism. *Front. Plant Sci.* 5:674. doi: 10.3389/fpls.2014.00674

This article was submitted to *Plant Evolution and Development*, a section of the journal *Frontiers in Plant Science*.

Copyright © 2014 Meroz and Bastien. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.