



Post-activation muscle potentiation and its relevance to cyclical behaviours

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1 **Post-activation muscle potentiation and its relevance to cyclical behaviors**

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7 Keywords: post-activation potentiation, staircase potentiation, muscle, work, frequency

8 **Abstract**

9 Muscle can experience post-activation potentiation (PAP), a temporary increase in
10 force and rate of force development, when contractions are closely timed; therefore,
11 cyclical behaviors are likely affected by PAP, as succeeding contraction cycles can lead to
12 potentiation over several subsequent cycles. Here, we examined PAP during *in situ* cyclical
13 contractions of the mallard lateral gastrocnemius (LG). Surface swimming, a cyclical
14 behavior, was mimicked with work-loops utilizing *in vivo* LG length change and stimulation
15 parameters. Tests were performed at mallards' preferred cycle frequency as well as at
16 lower and higher frequencies. Like muscles from mammals, anurans, and arthropods, the
17 mallard LG exhibited PAP with increases in peak force, average force rate, and net work.
18 Staircase potentiation occurred over two or more work-loop cycles, resulting in gradual
19 increases in PAP. The number of cycles needed to reach maximum work varied with cycle
20 frequency, requiring more cycles at higher cycle frequencies. PAP occurred under *in vivo*-
21 like stimulation parameters, suggesting a potentially important role of PAP in animal
22 locomotion, especially in cyclical behaviors.

23

24 **Introduction**

25 The contraction history of a muscle can influence its performance. Post-activation
26 potentiation (PAP) is one such history-dependent phenomenon where muscle force is
27 higher for a given submaximal stimulation, if the muscle has been recently stimulated. The
28 potentiated state may be the predominate condition for muscles *in vivo* [1]. PAP is caused
29 by phosphorylation of the myosin regulatory light chain (MRLC), which increases the rate
30 of cross-bridge formation [2], thereby increasing both the magnitude of force and rate of
31 force development for submaximal activation [3,4]. PAP has been studied for more than a
32 century (for review [5]). Much of the foundational work on PAP focused on isolated muscle
33 fibers [4] or whole muscles [1,2,3,6] *in vitro*. PAP was also measured in humans as an
34 increase in twitch force following a maximal voluntary contraction [7,8], demonstrating
35 that voluntary activation could elicit PAP. More recently, increases in human performance
36 during high-power behaviors following a tetanic contraction have been studied [5,9,10];
37 however, differences in timing of MRLC phosphorylation relative to improved exercise
38 performance led to concerns that these studies measured a different phenomenon [5]. Thus,

39 a gap in understanding remains between measurements of PAP and naturally-occurring
40 contractile behaviors.

41 Cyclical behaviors are likely affected by PAP since conditioning by one cycle can lead
42 to potentiation in subsequent cycles. MRLC phosphorylation levels remain high for up to a
43 minute following a conditioning stimulus [2,11], and PAP can affect muscle forces for
44 several minutes [1,7,8] or longer under non-fatiguing conditions [6]. Observations of PAP
45 during low-frequency isometric contractions uncovered a phenomenon known as staircase
46 potentiation – a gradual rise in muscle force over several contractions [3,6,12,13]. Staircase
47 potentiation has been observed during twitch [3,6,13] and incompletely-fused tetanic
48 contractions [12]. We, therefore, hypothesize that staircase potentiation may also affect
49 force and work production during dynamic contractions.

50 Relatively few studies have examined PAP during dynamic contractions [2,11,14].
51 The work-loop technique is a powerful tool for mimicking dynamic muscle contractions
52 and is particularly useful for studying cyclical behaviors [15]. Work is the area inside of a
53 force versus length loop, so both increased force and force rate should together increase
54 work output. An experiment examining a single work-loop in the potentiated state suggests
55 PAP increases work output [2], and work has been observed to increase over several work-
56 loops [15, 16, 17]. Although the term post-activation potentiation has been restricted to
57 certain conditioning stimuli and responses [5,13], we define PAP as any increases in muscle
58 force, rate of force development, or work following prior activation.

59 Here, we used the work-loop technique *in situ* to examine how PAP affects work
60 output over repeated cycles in mallard lateral gastrocnemius (LG). The LG is an important
61 ankle extensor during walking and surface swimming, and *in vivo* LG function during these
62 behaviors has been described [18]. The current study differs from previous studies of PAP
63 by measuring PAP while mimicking the *in vivo* LG length change and activation patterns
64 observed during surface swimming, a cyclical behavior. Mallards use a cycle frequency of
65 2.6 Hz during surface swimming [18,19], but use a range of cycle frequencies during
66 terrestrial locomotion [20]. To explore the effect of cycle frequency, we measured work
67 across successive cycles at 1.0, 2.6, and 4.0 Hz. We tested three hypotheses: (H1) PAP
68 increases muscle work output under *in vivo*-like conditions; (H2) because of the staircase

69 effect [13], PAP builds over cycles, resulting in increased work with each subsequent cycle
70 until it reaches some maximum; and (H3) cycle frequency affects the degree of potentiation.

71

72 **Methods**

73 All animal procedures were approved by Harvard University's Institutional Animal
74 Care and Use Committee (protocol 20-09). Mallard ducks (*Anas platyrhynchos*, n=5) were
75 anesthetized with 1-2% isoflurane:oxygen gas mixture using mask induction. To measure
76 fascicle length, sonomicrometry crystals (2 mm, Sonometrics Corporation) were implanted
77 at each end of a mid-belly LG fascicle (Fig. S1) and secured (5-0 silk suture, Ethicon). The
78 muscle's origin remained intact, but all other connective tissue attachments were removed
79 to allow free movement of the LG. To measure muscle force and impose length changes,
80 kevlar thread was tied proximal to the tendon's fibrocartilage pad and connected to a
81 muscle ergometer lever arm (10 N, Aurora Scientific Inc.) modified by drilling a hole at half
82 its length to increase its upper force limit (18.3 N). The animal was mounted on a moveable
83 stage by clamping the femur mid-shaft and securing the foot to a flat plate (Fig. S1); this
84 setup ensured that the limb and muscle origin remained stationary and the muscle line of
85 action was aligned with the ergometer. The muscle was stimulated (pulse duration: 0.2 ms,
86 Grass S48, Grass Instruments, Warrick, RI) intramuscularly through medial and lateral
87 pairs of 24 gauge, stainless steel wire electrodes separated proximally and distally as far
88 possible (~3 cm) and sutured in place (Fig. S1).

89 The voltage that induced maximal twitch force was determined, with voltage
90 reduced (~2-3 V) to elicit submaximal force ($35 \pm 5\%$ maximum), resulting in dynamic
91 forces likely similar to forces produced during slow swimming [18]. Using stimulation
92 frequencies of 125 (n=3) or 250 Hz (n=2), we determined the submaximal isometric L_0
93 (length at which submaximal tetanic force is produced). We then used the work-loop
94 technique [15] to measure muscle work while mimicking strain and activation patterns
95 observed during surface swimming *in vivo* [18]. Specifically, ramp changes in muscle length
96 were applied over 5-15 cycles with stimulation onset at -1.6% phase, stimulation duration
97 of 26%, and lever arm excursion of 9 mm (Fig. 1), which corresponded to fascicle length
98 changes of 5.4 ± 0.9 mm and strains (maximum: 0.03 ± 0.04 , minimum: -0.21 ± 0.04)
99 consistent with *in vivo* strains observed during surface swimming [18, KTB personal

100 observations]. Keeping the length change, activation phase, and duty factor constant, we
101 tested three cycle frequencies (1.0, 2.6, and 4.0 Hz) in a random order with a 5 min rest
102 between series, consistent with previous PAP work-loop experiments [2]. Because of in-
103 series compliance, minimum strains decreased over cycles with increased force production,
104 but these differences were small (change in strain from first to last cycle ~ 0.02 for 2.6 & 4
105 Hz and ~ 0.001 at 1 Hz). Isometric force production at L_0 was monitored throughout the
106 experiment to ensure that force decreases were $< 10\%$.

107 Post-mortem dissections confirmed sonomicrometry crystal alignment. Resting LG
108 fascicle and inter-crystal lengths were measured and the ratio of these was used to
109 calculate time-varying fascicle lengths (time-varying segment lengths * resting fascicle
110 length/inter-crystal distance). Force and length data were filtered (low-pass, 1st order, dual
111 pass Butterworth, 20 Hz cutoff; Fig. 1C,D) before plotting length versus force to calculate
112 resulting loop areas (i.e., net work, Fig. 1E). Peak force and the average rate of force
113 development ($F_{\text{rate}} = \text{change in force} / \text{duration from stimulation onset to maximum force}$)
114 were measured. 2nd order polynomials were fitted to net work, peak force, and F_{rate} versus
115 cycle number for each frequency to calculate (1) the maximum net work, peak force, or F_{rate}
116 and (2) the cycle when the maximum occurred. Percent change in net work, peak force, and
117 F_{rate} were calculated as: $((\text{maximum} - \text{cycle 1 value}) / \text{cycle 1 value}) * 100$. Percent change
118 data are available on Dryad (doi:10.5061/dryad.69p8cz8z4).

119 Statistical tests were performed in R (v. 3.6.0), with mixed-effects linear models
120 used to predict the percent change in net work, peak force, and F_{rate} and the maximum
121 work cycle number. Individual was a random effect and cycle frequency, stimulation
122 frequency, and strain at peak force (taken from cycle 2) were added iteratively as fixed
123 effects. Models were compared using ANOVA with $\alpha = 0.05$. Stimulation frequency and strain
124 at peak force did not significantly improve the models ($p > 0.05$), so were not included in
125 further analysis. Frequency significantly improved the models ($p < 0.05$) for all variables.
126 Diagnostic plots for the selected model (frequency as a fixed effect and individual as a
127 random effect) revealed assumptions of homoscedasticity and normal distribution of
128 residuals were reasonable. We performed pairwise comparisons using the emmeans
129 package and Tukey's adjustment. All reported values are mean \pm s.e.m.

130

131 **Results**

132 We performed work-loop experiments to determine the degree of work, force, and
133 F_{rate} PAP over 5-15 work-loop cycles at 1.0, 2.6, and 4.0 Hz. The number of work-loop cycles
134 required to reach maximum work varied by frequency ($p < 0.001$, Fig. 2A,B, Table 1). The
135 maximum work cycle number significantly differed between 1 Hz and 2.6 Hz and 1 Hz and
136 4 Hz, but not between 2.6 and 4 Hz (Table 2).

137 Cycle frequency explained a significant amount of the variation in percent change
138 (i.e., PAP) for net work (Fig. 2A,C; $p = 0.001$), peak force (Fig. D; $p < 0.001$), and F_{rate} (Fig. E;
139 $p = 0.001$). The percent change in net work was significantly different from zero at 4 Hz, but
140 not 2.6 and 1 Hz (Table 1). Peak force and F_{rate} both significantly increased over work-loop
141 cycles at 2.6 and 4 Hz, but not at 1 Hz (Table 1). Pairwise comparisons showed PAP in net
142 work, peak force, and F_{rate} was significantly different between 2.6 and 4 Hz and 1 and 4 Hz,
143 but not between 1 and 2.6 Hz (Fig. 2; Table 2).

144

145 **Discussion**

146 We examined the effect of PAP on mallard LG cyclical force and work performance,
147 by imposing *in vivo* length change and activation data to mimic the contractile behavior of
148 this muscle during surface swimming [18]. Under these conditions, PAP resulted in a 37%
149 increase in peak force, a 64% increase in F_{rate} , and a 52% increase (n.s.) in net work (Fig. 2,
150 Table 1), suggesting PAP is a relevant and substantial contractile phenomenon at the
151 mallard's preferred cycle frequency.

152 Past work provided evidence of PAP in mammals [1,2,3,4,7,8,17], anurans [6,21],
153 fish [16], and insects [15]; that birds also experience PAP is not surprising. The mallard LG
154 is a mixed fibered muscle, with mostly type II fibers (74-100%, depending on the sample
155 location, [22]). Previous work has shown that muscles with large populations of type II fast
156 fibers demonstrate larger increases in force and F_{rate} than type I slow-fibered muscles [8].

157 We observed increases in both force and F_{rate} (Fig. 2, Table 1), consistent with
158 previous findings [2,3,4]. Unlike isometric contractions where force and F_{rate} can be
159 considered separately, force and F_{rate} are interrelated during dynamic contractions. Force
160 drops when muscles shorten to lengths below L_o , so a greater F_{rate} will permit more force

161 development prior to substantial shortening. Thus, we cannot draw independent
162 conclusions about these factors here.

163 During a dynamic contraction, changes in force and F_{rate} should also correspond to a
164 change in work (H1). We observed a significant increase in peak force and F_{rate} for both 2.6
165 and 4 Hz, but a significant increase in net work only at 4 Hz (although there was a tendency
166 toward significance at 2.6 Hz). Work potentiation has previously been observed during a
167 single work-loop of a mouse fast-fibered muscle (extensor digitorum longus) following a
168 conditioning stimulus [2]. The current study builds on this finding by examining work
169 potentiation over several cycles of the mallard LG. Our observation of gradual increases in
170 net work, peak force, and F_{rate} over 4-10 cycles (Figs. 1E,2A) indicates that PAP is
171 cumulative during dynamic contractions (H2). Our findings are consistent with (1)
172 staircase potentiation, where force increases over several isometric stimulations
173 [3,6,12,13], and (2) observations of work increasing over several cycles [15,16,17]. PAP can
174 build and possibly be maintained over a large number of cycles for sustained locomotion.

175 At higher cycle frequencies, (1) net work, peak force, and F_{rate} potentiation was
176 larger and (2) a larger number of cycles were needed to achieve this potentiation,
177 supporting our hypothesis that cycle frequency would affect PAP (H3). Krarup [3] similarly
178 observed that higher contraction frequencies elicited larger twitch force potentiation but,
179 in contrast to our findings, found that peak potentiation was reached with fewer stimuli at
180 higher contraction frequencies. The lack of a significant effect of PAP on LG work at lower
181 frequencies could be due to longer inter-cycle times, allowing the effects of PAP to diminish
182 before the next cycle commences. Alternatively, the effects of fatigue due to longer
183 contraction durations at lower frequencies could counteract the effects of PAP, causing
184 work potentiation to be lower and to peak with fewer contractions. Because we maintained
185 a constant duty factor of activation across frequencies, the total stimulation duration and
186 number of spikes was higher at lower frequencies. PAP is thought to offset the effects of
187 fatigue [7,23], possibly helping to maintain force production over a series of cycles. Thus,
188 PAP may very well affect work output by mitigating the effects of fatigue and reducing the
189 drop in force that might otherwise be observed.

190 The present study demonstrates that PAP occurs under conditions that mimic a
191 cyclical swimming behavior and could elicit meaningful increases in muscle work, force,

192 and F_{rate} under certain cycle frequencies. Further exploration is needed of how changes in
193 force and work production resulting from PAP relate to locomotor performance in
194 naturally-behaving animals.

195

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203 Vertebrate Locomotion, and NIH AR055648.

204

205 **Tables & Figure Captions**

206 **Figure 1.** Work-loop experiments. (A) Ramp length change pattern applied via the lever
207 arm and stimulation timing. Stimulation offset (1.6%) and duration (26%) are expressed
208 as % of cycle and scaled to cycle frequency. Example (B) lever position and stimulation, (C)
209 fascicle length change from sonomicrometry, and (D) force from the ergometer during a
210 work-loop series at 2.6 Hz. (E) Force versus length work (area inside the loop) loops at 2.6
211 Hz. Length is expressed as an absolute measure (mm) and as strain (relative to $L_{o, \text{submax}}$).
212 All loops are counterclockwise (positive work). Loops progress from light to dark with each
213 subsequent loop. Thicker lines indicate the period of activation.

214

215 **Figure 2.** Post-activation potentiation occurred during work-loops in the mallard LG. (A)
216 Percent change in net work versus cycle number at three frequencies: 1 Hz (gray, filled
217 circles), 2.6 Hz (black, filled circles; mallards' preferred surface swimming cycle frequency),
218 and 4 Hz (open circles). Solid lines are 2nd order polynomial fits. (B) Maximum work cycle
219 number was significantly higher at 2.6 Hz and 4 Hz than at 1 Hz, but not between 2.6 and 4
220 Hz (Table 2). Percent change in (C) net work, (D) peak force, and (E) F_{rate} were significantly
221 greater at 4 Hz than 1 Hz and 2.6 Hz, but not between 1 Hz and 2.6 Hz (Table 2). All points
222 are averages \pm s.e.m. Lower case letters (a,b) indicate groups for Tukey's pairwise

223 comparison, with different letters assigned to groups that were significantly different
 224 ($p < 0.05$). Sample size is indicated by the number on each point.

225

226 **Table 1.** Maximum work cycle number and mean percent change for net work, peak force,
 227 and F_{rate} by frequency. P-values < 0.05 (bolded) indicate percent change is significantly
 228 different from zero.

Frequency		Max Work Cycle	Work (%)	Force (%)	F_{rate} (%)
1 Hz	mean \pm s.e.m.	4 \pm 1	16 \pm 9%	10 \pm 4%	29 \pm 8%
	p-value		0.577	0.535	0.570
2.6 Hz	mean \pm s.e.m.	8 \pm 1	52 \pm 14%	37 \pm 11%	64 \pm 20%
	p-value		0.051	0.016	0.026
4 Hz	mean \pm s.e.m.	10 \pm 1	115 \pm 35%	69 \pm 17%	117 \pm 27%
	p-value		<0.001	<0.001	0.002

229

230

231 **Table 2.** P-values for a Tukey-adjusted pairwise-comparison for maximum work cycle
 232 number and percent change in net work, peak force, and F_{rate} by frequency. Bolded p-values
 233 indicate a significant difference (< 0.05).

Comparison	Max Work Cycle	Work	Force	F_{rate}
1 Hz vs. 2.6 Hz	0.011	0.234	0.065	0.072
2.6 Hz vs. 4 Hz	0.288	0.045	0.048	0.046
1 Hz vs. 4 Hz	0.001	0.004	0.001	0.002

234

235 **References**

236

237 1. Brown IE, Loeb GE. 1998. Post-activation potentiation – A clue for simplifying models of
238 muscle dynamics. *Amer. Zool.* **38**, 743-754. (doi:10.1093/icb/38.4.743)

239

240 2. Grange RW, Vandenboom R, Xenj J, Houston ME. 1998. Potentiation of in vitro concentric
241 work in mouse fast muscle. *J. Appl. Physiol.* **84**, 236-243. (doi:10.1152/jappl.1998.84.1.236)

242

243 3. Krarup C. 1981. Enhancement and diminution of mechanical tension evoked by staircase
244 and by tetanus in rat muscle. *J. Physiol.* **311**, 355-372. (doi:

245 [10.1113/jphysiol.1981.sp013589](https://doi.org/10.1113/jphysiol.1981.sp013589))

246

247 4. Sweeney HL, Stull JT. 1990. Alteration of cross-bridge kinetics by myosin light chain
248 phosphorylation in rabbit skeletal muscle: implications for regulation of actin-myosin
249 interactions. *Proc. Natl. Acad. Sci. USA.* **87**, 414-418. (doi: [10.1073/pnas.87.1.414](https://doi.org/10.1073/pnas.87.1.414))

250

251 5. Blazeovich AJ, Babault N. 2019. Post-activation potentiation versus post-activation
252 performance enhancement in humans: historical perspective, underlying mechanisms, and
253 current issues. *Front. Physiol.* **10**, 1359. (doi: [10.3389/fphys.2019.01359](https://doi.org/10.3389/fphys.2019.01359))

254

255 6. Isaacson A. 1969. Post-staircase potentiation, a long-lasting twitch potentiation of
256 muscles induced by previous activity. *Life Sci.* **8**, 337-342. (doi: [10.1016/0024-
257 3205\(69\)90225-2](https://doi.org/10.1016/0024-3205(69)90225-2))

258

259 7. Vandervoort AA, Quinlan J, McComas AJ. 1983. Twitch potentiation after voluntary
260 contraction. *Exp. Neurol.* **81**, 141-152. (doi: [10.1016/0014-4886\(83\)90163-2](https://doi.org/10.1016/0014-4886(83)90163-2))

261

262 8. Hamada T, Sale DG, MacDougall JD, Tarnopolsky MA. 2000. Postactivation potentiation,
263 fiber type, and twitch contraction time in human knee extensor muscles. *J. Appl. Physiol.* **88**,
264 2131-2137. (doi:[10.1152/jappl.2000.88.6.2131](https://doi.org/10.1152/jappl.2000.88.6.2131))

265

266 9. Hodgson M, Dockerty D, Robbins D. 2005. Post-activation potentiation: underlying
267 physiology and implications for motor performance. *Sports Medicine.* **35**, 585-595. (doi:
268 [10.2165/00007256-200535070-00004](https://doi.org/10.2165/00007256-200535070-00004))

269

270 10. McCann MR, Flanagan SP. 2010. The effects of exercise selection and rest interval on
271 postactivation potentiation of vertical jump performance. *J. Strength Cond. Res.* **24**, 1285-
272 1291. (doi: [10.1519/JSC.0b013e3181d6867c](https://doi.org/10.1519/JSC.0b013e3181d6867c))

273

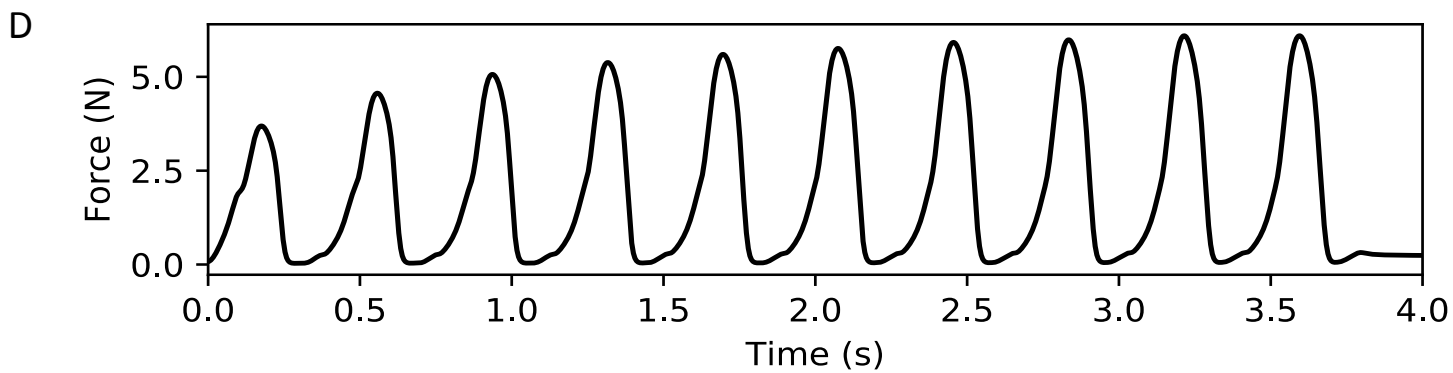
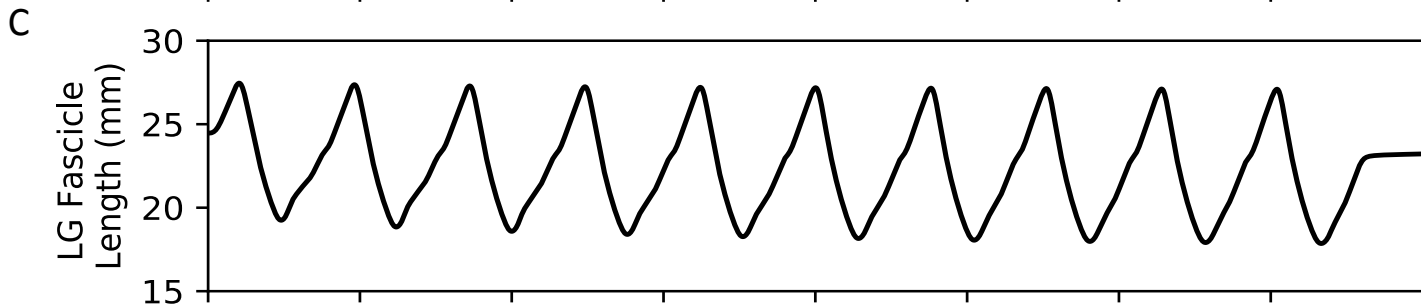
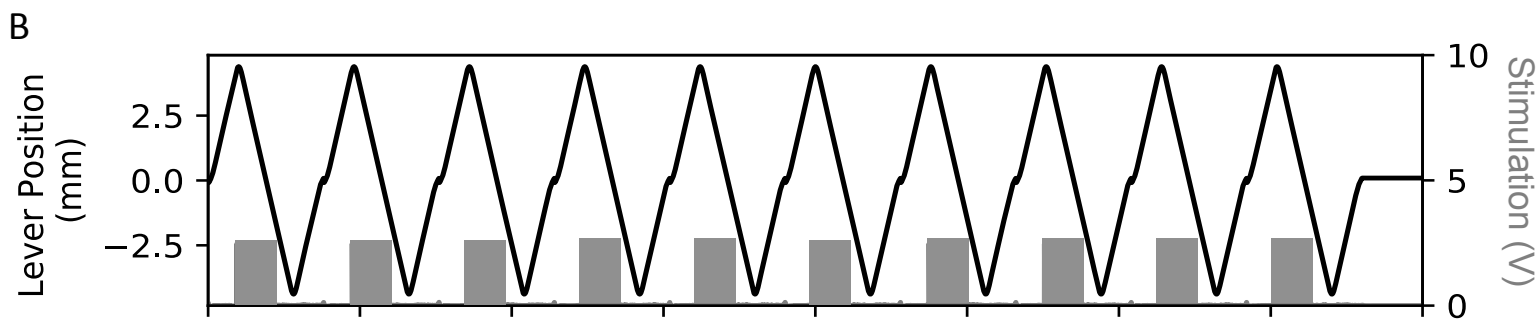
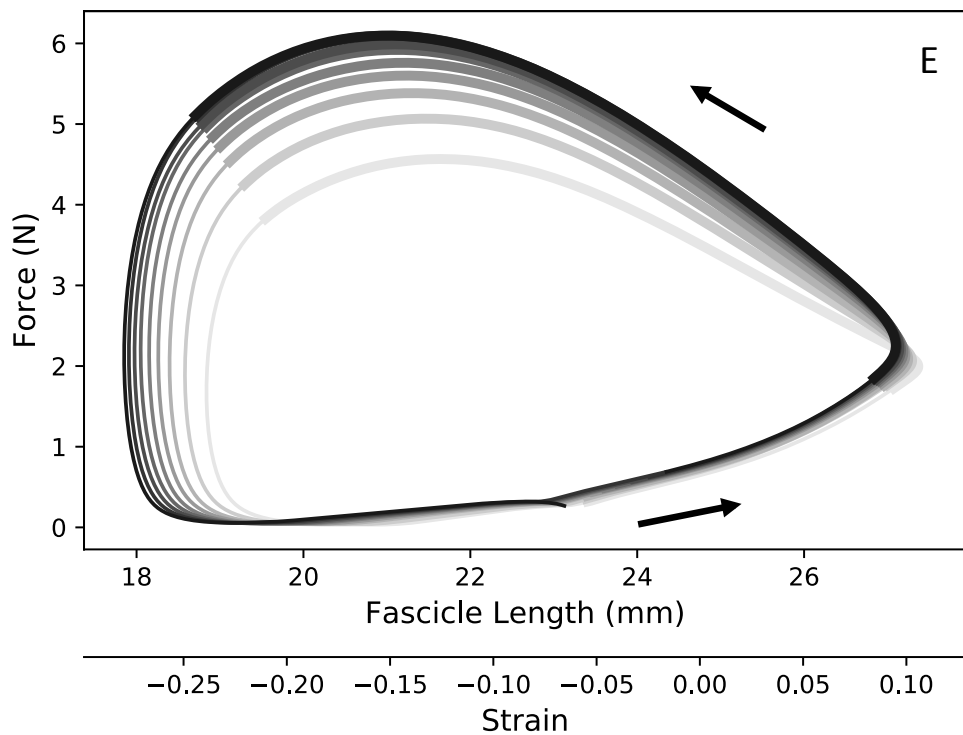
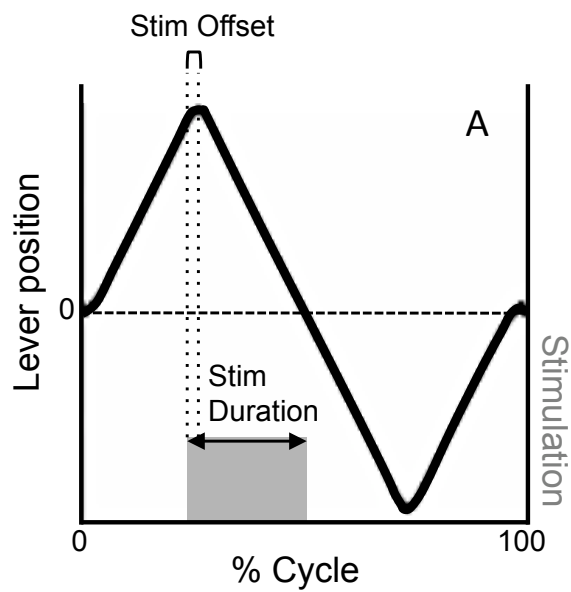
274 11. Grange RW, Cory CR, Vandenboom R, Houston ME. 1995. Myosin phosphorylation
275 augments force-displacement and force-velocity relationships in mouse fast muscle. *Am. J.*
276 *Physiol.* **269**, C713-C724. (doi: [10.1152/ajpcell.1995.269.3.C713](https://doi.org/10.1152/ajpcell.1995.269.3.C713))

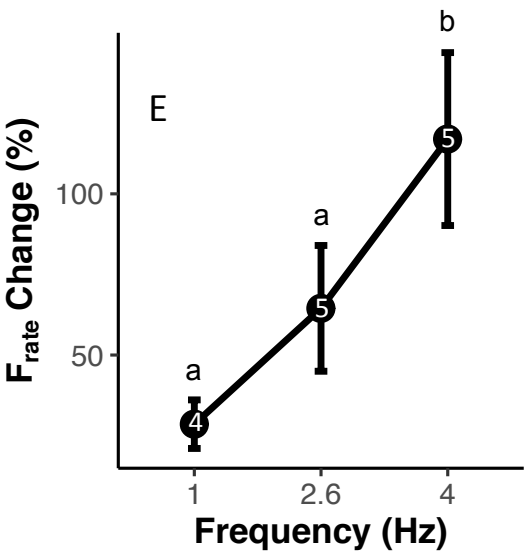
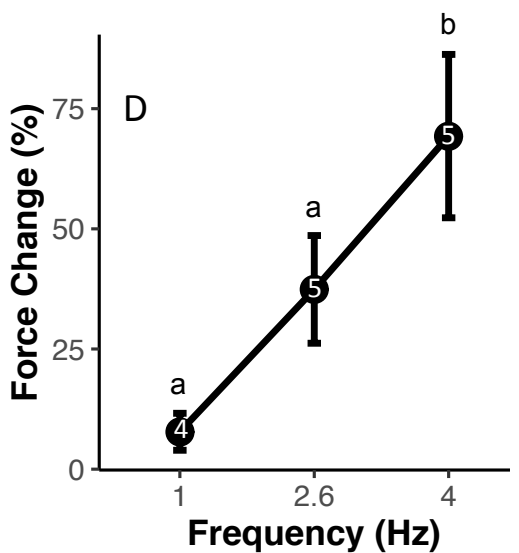
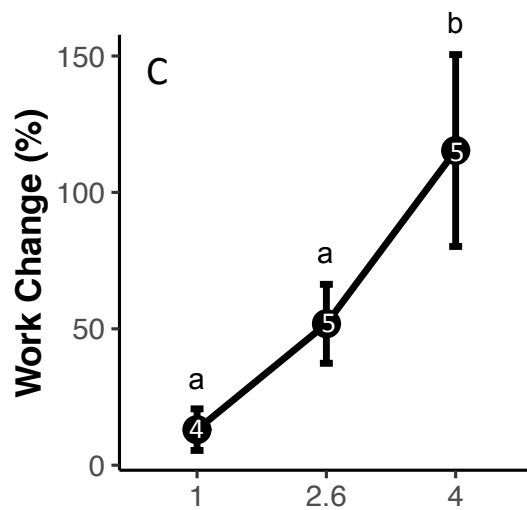
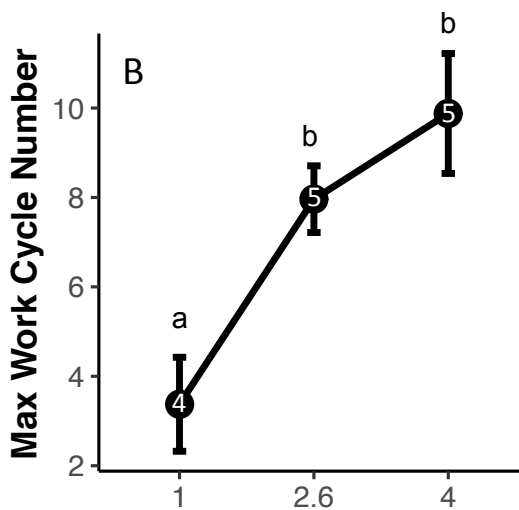
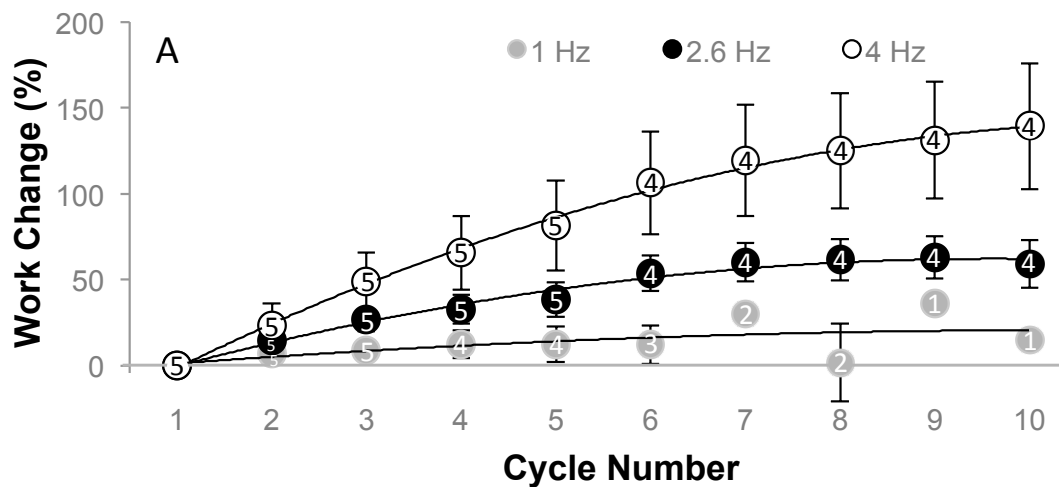
277

278 12. MacIntosh BR, Willis JC. 2000. Force-frequency relationship and potentiation in
279 mammalian skeletal muscle. *J. Appl. Physiol.* **88**: 2088-2096. (doi:

280 [10.1152/jappl.2000.88.6.2088](https://doi.org/10.1152/jappl.2000.88.6.2088))

- 281
282 13. MacIntosh BR. 2010. Cellular and whole muscle studies of activity dependent
283 potentiation. In *Adv. Exp. Med. Bio.* vol. **682**. Springer New York, NY. 315-342.
284 (doi:[10.1007/978-1-4419-6366-6_18](https://doi.org/10.1007/978-1-4419-6366-6_18))
285
- 286 14. Brown IE, Loeb GE. 1999. Measured and modeled properties of mammalian skeletal
287 muscle: I. The effects of post-activation potentiation on the time course and velocity
288 dependencies of force production. *J. Muscle Res. Cell Motil.* **20**, 443-456. (doi:
289 [10.1023/a:1005590901220](https://doi.org/10.1023/a:1005590901220))
290
- 291 15. Josephson RK. 1985. Mechanical power output from striated muscle during cyclic
292 contraction. *J. Exp. Biol.* **114**, 493-512.
293
- 294 16. Altringham JD, Johnston IA. (1990). Modeling muscle power output in a swimming fish.
295 *J. Exp. Biol.* **148**, 395-402.
296
- 297 17. Askew GN, Young IA, Altringham JD. (1997). Fatigue of mouse soleus muscle using the
298 work loop technique. *J. Exp. Biol.* **200**, 2907-2912.
299
- 300 18. Biewener AA, Corning WR. 2001. Dynamics of mallard (*Anas platyrhynchos*)
301 gastrocnemius function during swimming versus terrestrial locomotion. *J. Exp. Biol.* **204**,
302 1745-1756.
303
- 304 19. Prange HD, Schmidt-Nielsen K. 1970. The metabolic cost of swimming in ducks. *J. Exp.*
305 *Biol.* **53**, 763-777.
306
- 307 20. Abourachid A. 2000. Bipedal locomotion in birds: the importance of functional
308 parameters in terrestrial adaptation in Anatidae. *Can. J. Zool.* **78**, 1994-1998.
309
- 310 21. Colomo F, Rocchi P. 1965. Eserine effects on single twitches and staircase phenomenon
311 in frog nerve-single muscle fibre preparations. *Arch. Fisiol.* **65**, 24-51.
312
- 313 22. Torrella JR, Fouces V, Palomeque J, Viscor G. 1996. Capillarity and fibre types in
314 locomotory muscle of wild mallard ducks (*Anas platyrhynchos*). *J. Comp. Physiol. B.* **166**,
315 164-177.
316
- 317 23. Behm DG. 2004. Force maintenance with submaximal fatiguing contractions. *Can. J. Appl.*
318 *Physiol.* **29**, 274-290. (doi: [10.1139/h04-019](https://doi.org/10.1139/h04-019))





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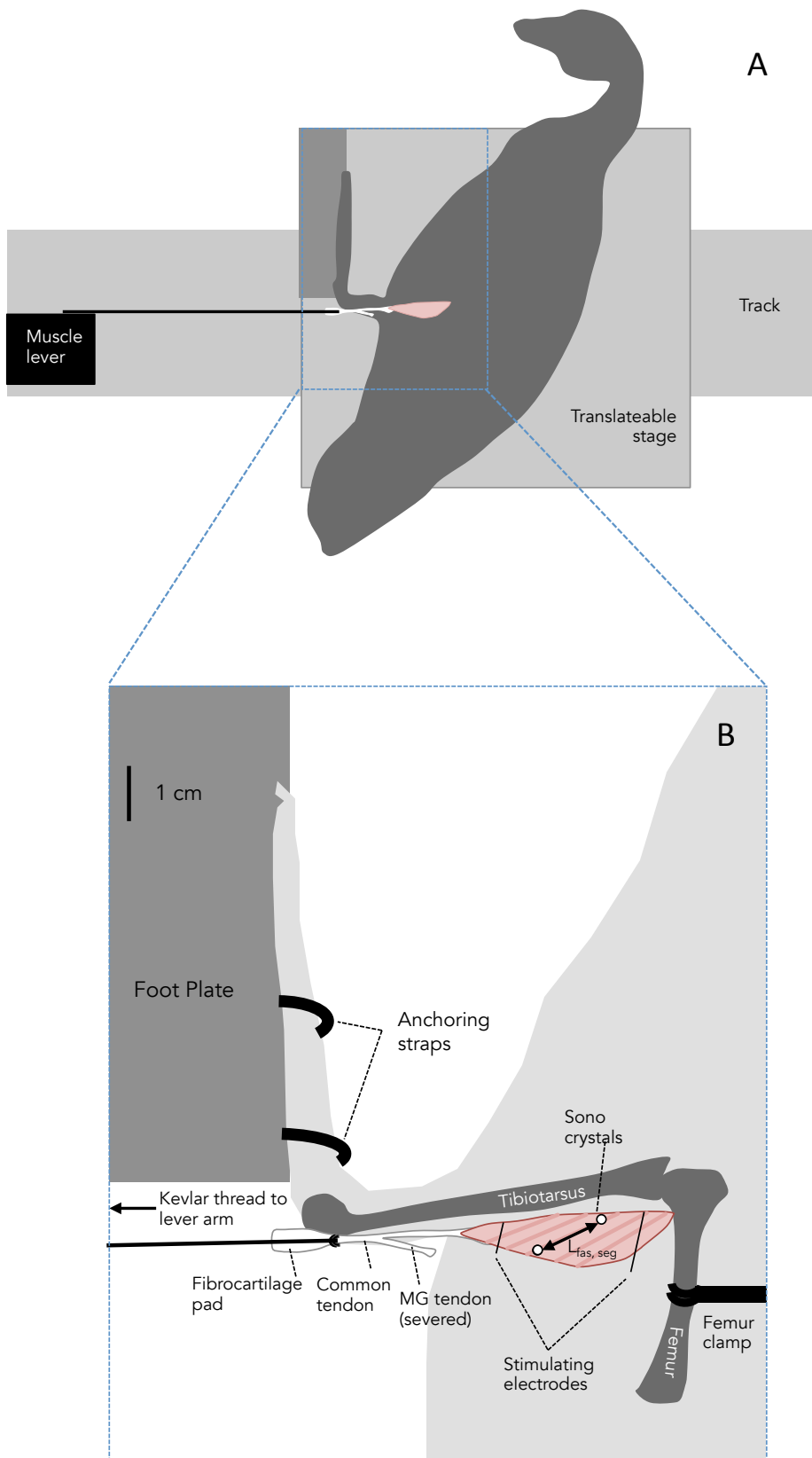


Figure S1. In situ experimental setup to measure force and length in the mallard lateral gastrocnemius (LG). (A) Schematic showing the *in situ* rig. The animal was placed on a stage that could be translated along a track to change the distance between the animal and muscle lever. (B) Larger view of a portion of the *in situ* rig (blue box). The femur was clamped mid-shaft and the foot was anchored to a plate to hold the leg and muscle origin stationary. The LG tendon was severed below the fibrocartilage pad, and a Kevlar thread was secured above the pad and connected to a muscle ergometer lever arm. The medial gastrocnemius (MG) tendon was severed and the muscle was freed from connective tissue attachments to surrounding tissues except for the point of origin on the femur. A pair of sonomicrometry crystals was inserted along the fascicle to measure a segment of fascicle length ($L_{fas, seg}$). An effort was made to measure as much of the fascicle as possible. We assumed the shortening in the segment was representative of shortening along the full length of the fascicle. Post-mortem 'resting' measurements of the segment ($L_{fas, seg, rest}$) and the full fascicle length ($L_{fas, rest}$) permitted us to correct sonomicrometry measurements of segment length ($L_{fas, seg}$) to fascicle lengths ($L_{fas, calculated} = L_{fas, seg} * L_{fas, resting} / L_{fas, seg, rest}$). Two pairs of stainless steel stimulating electrodes were inserted through the muscle belly with one polarity as far proximal and the other as far distal as possible. Although only one pair is shown, two pairs were inserted in medial and lateral sagittal planes.