The Calcium-Sensing Receptor Mediates Bone Turnover Induced by Dietary Calcium and Parathyroid Hormone in Neonates

Citation

Published Version
doi:10.1002/jbmr.300

Permanent link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:10357609

Terms of Use
This article was downloaded from Harvard University’s DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

Share Your Story
The Harvard community has made this article openly available. Please share how this access benefits you. Submit a story .

Accessibility
The Calcium-Sensing Receptor Mediates Bone Turnover Induced by Dietary Calcium and Parathyroid Hormone in Neonates

Lei Shu,1 Ji Ji,1 Qi Zhu,1 Guofan Cao,1 Andrew Karaplis,2 Martin R Pollak,3 Edward Brown,3 David Goltzman,2 and Dengshun Miao1,4

1Laboratory of Reproductive Medicine, Research Center for Bone and Stem Cells, Nanjing Medical University, Nanjing, Jiangsu, People’s Republic of China
2Department of Medicine, McGill University, Montreal, Quebec, Canada
3Division of Endocrinology, Diabetes and Hypertension, Department of Medicine, Brigham and Women’s Hospital, Harvard Medical School, Boston, MA USA
4Department of Geriatrics, First Affiliated Hospital of Nanjing Medical University, Nanjing, Jiangsu, People’s Republic of China

ABSTRACT
We have investigated, in neonates, whether the calcium-sensing receptor (CaR) mediates the effects of dietary calcium on bone turnover and/or modulates parathyroid hormone (PTH)–induced bone turnover. Wild-type (WT) pups and pups with targeted deletion of the Pth (Pth–/–) gene or of both Pth and CaR (Pth–/–CaR–/–) genes were nursed by dams on a normal or high-calcium diet. Pups nursed by dams on a normal diet received daily injections of vehicle or of PTH(1–34) (80 μg/kg) for 2 weeks starting from 1 week of age. In pups receiving vehicle and fed by dams on a normal diet, trabecular bone volume, osteoblast number, type 1 collagen–positive area, and mineral apposition rate, as well as the expression of bone-formation-related genes, all were reduced significantly in Pth–/– pups compared with WT pups and were decreased even more dramatically in Pth–/–CaR–/– pups. These parameters were increased in WT and Pth–/– pups but not in Pth–/–CaR–/– pups fed by dams on a high-calcium diet compared with pups fed by dams on a normal diet. These parameters also were increased in WT, Pth–/–, and Pth–/–CaR–/– pups following exogenous PTH treatment; however, the percentage increase was less in Pth–/–CaR–/– pups than in WT and Pth–/– pups. In vehicle-treated pups fed by dams on either the normal or high-calcium diet and in PTH-treated pups fed by dams on a normal diet, the number and surfaces of osteoclasts and the ratio of RANKL/OPG were reduced significantly in Pth–/– pups and less significantly in Pth–/–CaR–/– pups compared with WT pups. These parameters were further reduced significantly in WT and Pth–/– pups from dams fed a high-calcium diet but did not decrease significantly in similarly treated Pth–/–CaR–/– pups, and they increased significantly in PTH-treated pups compared with vehicle-treated, genotype-matched pups fed by dams on the normal diet. These results indicate that in neonates, the CaR mediates alterations in bone turnover in response to changes in dietary calcium and modulates PTH-stimulated bone turnover.

KEY WORDS: CALCIUM-SENSING RECEPTOR; PARATHYROID HORMONE; DIETARY CALCIUM; BONE TURNOVER

Introduction

The calcium-sensing receptor (CaR) plays a central role in controlling systemic calcium homeostasis, predominately through its effects on regulation of parathyroid hormone (PTH) secretion by the parathyroid glands and urinary calcium excretion by the kidney.(1-3) Recent evidence clearly has shown that the CaR is expressed and modulates cell function in a variety of bone cells, including osteoblasts,(4-8) stromal cells,(9) osteoclasts,(6,10,11) and chondrocytes.(4) Previous studies on cultured osteoblasts have shown that CaR activation not only stimulates the proliferation of these cells but also promotes their differentiation and mineralization. Cellular proliferation and expression of osteoblast differentiation markers, including Cbfa-1, alkaline phosphatase (Alp), type 1 collagen, and osteocalcin, and mineralized nodule formation were increased significantly in rat calvarial osteoblasts treated with high calcium. Inhibition of the CaR by NPS 89636 (a CaR antagonist) blocked responses in vitro to CaR agonists, suggesting that high-calcium-induced osteoblastic differentiation was mediated by the CaR.(5) Abolition of
CaR function by stably transfecting MC3T3-E1 cells with either a CaR antisense vector or a vector containing the dominant-negative R185Q variant of the CaR resulted in diminishing ALP activity, osteocalcin expression, and mineralization in mouse osteoblastic cells. More recently, mice with conditional knockout of the CaR in early committed osteoblasts were generated by breeding a novel mouse line carrying a CaR gene allele in which exon 7 had been floxed with transgenic mice expressing Cre recombinase under the control of a 2.3-kb type 1 collagen promoter. Smaller, undermineralized skeletons with significant reductions in bone volume and bone mineral density (BMD) were observed in the femur and vertebrae from the conditional knockout mice. This study demonstrated a critical role for the CaR at early stages of osteoblastic differentiation and bone development potentially by promoting osteoblast maturation.

Previous studies failed to find a direct role for CaR in mice homozygous for targeted disruption of exon 5 of the CaR gene, which encodes a segment of the extracellular domain of this receptor. These observations may have been confounded by the severe hyperparathyroidism and accompanying hypercalcemia and hypophosphatemia in this animal model. To better understand the direct effects of the CaR on bone and cartilage function, correction of hyperparathyroidism is required in this CaR-deficient mouse model. Therefore, a double-knockout model was established by ablating the Pth gene in the CaR-deficient mice to correct the severe hyperparathyroidism, hypercalcemia, and hypophosphatemia observed in the CaR-deficient mice. The results of this study showed that elimination of hyperparathyroidism rescued the mice from the increased neonatal mortality as well as the rickets-like skeletal abnormality in the CaR−/− mice. However, any essential, nonredundant role for CaR in regulating chondrogenesis or osteogenesis was not identified based on analysis of the skeleton of this adult double-knockout model. It is unclear why there are discrepancies between the findings from the mice with the osteoblastic-specific knockout of exon 7 of the CaR gene and those from mice with conventional global knockout of exon 5 of both the CaR and Pth genes regarding the physiologic significance of the CaR in osteoblasts. It is possible, however, that the alternatively spliced CaR lacking exon 5 that is expressed in the global exon 5 knockout mice may compensate to some extent in bone for loss of the full-length CaR.

PTH can stimulate bone formation in adult and aged animals of either sex and in animals with osteopenia induced by disuse, denervation, and immobilization. The anabolic action of PTH on bone has been validated in humans and its antifracture efficacy established in postmenopausal osteoporotic women. At present, however, the physiologic basis of PTH anabolism remains unclear. A large body of evidence demonstrates that sufficient calcium intake augments bone gain during growth, retards age-related bone loss, and reduces osteoporotic fracture risk, but the mechanisms underlying the positive effect of increased calcium intake on bone mass are unknown. Our recent study indicates that the skeletal anabolic action of PTH in neonates results not only from its direct action via the PTH receptor to increase the osteoblast pool but also by an indirect action mediated through PTH-induced increases in the extracellular calcium concentration. However, it is unclear whether the increase in extracellular calcium caused by PTH administration acts cooperatively with a skeletal anabolic action mediated via the CaR.

In this study, we attempted to determine whether deletion of CaR in PTH-deficient mice (1) leads to a skeletal phenotype distinct from that caused by deletion of Pth alone before weaning, (2) abolishes skeletal responses to dietary calcium supplementation, and (3) reduces skeletal responses to exogenous PTH administration. To explore these issues, we examined pups from wild-type (WT), Pth+/−, and Pth−/− CaR−/− mice nursed by dams on a normal calcium diet or on a high-calcium diet. In addition, we treated some pups nursed by dams on a normal calcium diet with daily injections of vehicle or of PTH(1–34) for 2 weeks starting from 1 week of age and assessed skeletal homeostasis.

Materials and Methods

Derivation of Pth−/− and Pth−/−CaR−/− mice

The derivation of the two parental strains of Pth−/− mice and CaR−/− mice by homologous recombination in embryonic stem cells was described previously by Miao and colleagues and Ho and colleagues. Pth−/− mice were bred with CaR−/− mice to generate Pth−/−CaR−/− mice. Pth−/−CaR−/− mice were bred among themselves to generate Pth−/−CaR−/− pups. These mice were maintained on a mixed genetic background with contributions from C57Bl/6, 129/SvJ, and 129/SvEv strains. The genotypes of the animals were determined by polymerase chain reaction (PCR).

In vivo experiments

All animal experiments were approval by the Institutional Animal Care and Use Committee. Mice were maintained in a virus- and parasite-free barrier facility and exposed to a 12-hour light/dark cycle. To determine whether PTH and calcium interact in neonatal calcium and skeletal homeostasis, WT, Pth−/−, or Pth−/−CaR−/− dams were fed with rodent diets from Harlan Teklad (Madison, WI, USA) containing either normal levels of calcium and phosphate (1% Ca, 0.6% phosphate) or high-calcium and moderate-phosphate levels (2% Ca and 0.4% phosphate). Serum biochemistry was not significantly different in the Pth−/−CaR−/− mice than in the WT mice. Dietary phosphate content in the high-calcium diet was reduced moderately to decrease phosphate absorption and resulting hyperphosphatemia and to facilitate increasing the blood calcium concentration in the hypoparathyroid Pth−/− and Pth−/−CaR−/− mice. The high-calcium, moderate-phosphate diet was found to normalize hypocalcemia and hyperphosphatemia in Pth−/− and Pth−/−CaR−/− mice and is referred to as the high-calcium diet. Litter size was equalized to 5 to 6 pups per dam to equalize suckling intensity. On day 7 postpartum, pups nursed by dams on the normal calcium diet received daily injections of vehicle or of PTH(1–34) (80 μg/kg) for 2 weeks. At the end of experiments, milk of dams and blood of pups were collected and used for biochemical analysis, and long bones of pups then were removed for the analyses described below.
Biochemistry and hormone analyses

Milk calcium (milk was diluted 1:100 in distilled water) was measured with an atomic absorptiometer as described previously.\(^{30}\) Serum calcium and phosphorus levels were determined by an autoanalyzer (Beckman Synchron 67; Beckman Coulter, Inc., Fullerton, CA, USA). Milk calcium levels were determined using the 0-cresolphthalein-complexone method (Sigma-Aldrich, St Louis, MO, USA); urine creatinine was measured by the colorimetric alkaline picate method (Sigma Kit 555, Sigma-Aldrich), and urine calcium/creatinine ratio was calculated. Milk and serum 1,25-dihydroxyvitamin D\(_3\) [1,25(OH)\(_2\)D\(_3\)] was measured by RIA (Immuno-Diagnostic Systems, Bolden, UK), and PTH-related protein (PTHRP) was measured by a two-site immunoradiometric assay (Immutopics, San Clemente, CA, USA). Serum PTH was measured using an ELISA (Immutopics).

Radiography and measurement of BMD

For radiography, tibias were removed and dissected free of soft tissue, and X-ray images were taken with a Faxitron machine (Model 805; Faxitron X-Ray Corp., Wheeling, IL, USA) under constant conditions (22 kV, 4-minute exposure) using Kodak X-OMat TL film (Eastman Kodak Co., Rochester, NY, USA). For measurement of tibial BMD, a PIXImus densitometer (Lunar PIXImus Corp., Madison, WI, USA) was used (5-minute image acquisition with the precision of 1% coefficient of variation for skeletal BMD). The PIXImus software automatically calculated the BMD and recorded the data in Microsoft Excel files (Microsoft Corp., Redmond, WA, USA).

Micro–computed tomography (\(\mu\)CT)

Tibias obtained from 3-week-old mice were dissected free of soft tissue, fixed overnight in 70% ethanol, and analyzed by \(\mu\)CT with a SkyScan 1072 scanner and associated analysis software (SkyScan, Antwerp, Belgium) as described previously.\(^{31}\) Briefly, image acquisition was performed at 100 kV and 98 \(\mu\)A with a 0.9-degree rotation between frames. During scanning, the samples were enclosed in tightly fitting plastic wrap to prevent movement and dehydration. Thresholding was applied to the images to segment the bone from the background. 2D images were used to generate 3D renderings using the 3D Creator software supplied with the instrument (SkyScalculator). The resolution of the \(\mu\)CT images is 18.2 \(\mu\)m.

Quantitative real-time RT-PCR

Reverse-transcription (RT) reactions were performed using the SuperScript First-Strand Synthesis System (Invitrogen, Carlsbad, CA, USA) as described previously.\(^{32}\) Real-time polymerase chain reaction (PCR) was performed using a LightCycler system (Roche Molecular Biochemicals, Indianapolis, IN, USA). The conditions were 2 \(\mu\)L of LightCycler DNA Master SYBR Green I (Roche), 0.25 \(\mu\)M of each 5’ and 3’ primer (Table 1), and 2 \(\mu\)L of sample and/or \(\mathrm{H}_2\mathrm{O}\) to a final volume of 20 \(\mu\)L. Samples were amplified for 35 cycles with a temperature transition rate of 20 °C/s for all three steps, which were denaturation at 94 °C for 10 seconds, annealing for 5 seconds, and extension at 72 °C for 20 seconds. SYBR green fluorescence was measured to determine the amount of double-stranded DNA. To discriminate specific from nonspecific cDNA products, a melting curve was obtained at the end of each run. Products were denatured at 94 °C for 30 seconds; the temperature was then decreased to 55 °C for 15 seconds and raised slowly from 55 to 94 °C using a temperature transition rate of 0.18 °C/s. To determine the number of copies of target DNA in the samples, purified PCR fragments of known concentration were serially diluted and served as external standards for each experiment. Data were normalized to GAPDH levels.

Histology

Tibias were removed and fixed in 2% paraformaldehyde containing 0.075 M lysine and 0.01 M sodium periodate overnight at 4 °C and processed as described previously.\(^{33}\) Proximal ends of tibias were decalcified in EDTA-glycerol solution for 5 to 7 days at 4 °C. Decalcified right tibias were dehydrated and embedded in paraffin, after which 5-\(\mu\)m sections were cut on a rotary microtome. The sections were stained with hematoxylin and eosin (H&E) or histochemically for tartrate-resistant acid phosphatase (TRACP) activity or immunohistochemically, as described below. For calcein labeling, mice were injected intraperitoneally with 10 \(\mu\)g calcein per gram of body weight (C-0875; Sigma) at 8 and 3 days prior to killing, as described previously.\(^{34}\) Left tibiae were embedded in LR white acrylic resin (London Resin Co., Ltd., London, UK), and 1-\(\mu\)m sections were viewed and imaged using fluorescence microscopy. The double-calcein-labeled widths of trabeculae were measured in the secondary spongiosa of the proximal ends of tibias using Northern Eclipse image-analysis software Version 6.0 (Empix Imaging, Inc., Mississauga, Ontario, Canada), and the mineral apposition rate (MAR) was calculated as the interlabel width/labeling period. TRACP staining was performed using a modification of a previously described protocol.\(^{32}\) In brief,

Table 1. Real-Time RT-PCR Primers Used with Their Name, Orientation (S = Sense; AS = Antisense), Sequence, Annealing Temperature (Tm), and Length of Amplicon (bp)

<table>
<thead>
<tr>
<th>Name</th>
<th>S/AS</th>
<th>Sequence</th>
<th>Tm(°C)</th>
<th>bp</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cbfa-1</td>
<td>S</td>
<td>GTCAGACCGTGTCAGCAAG</td>
<td>55</td>
<td>356</td>
</tr>
<tr>
<td></td>
<td>AS</td>
<td>GGAACAGGGAGTTGGGAC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rankl</td>
<td>S</td>
<td>GTGGGCGGAATGGCATG</td>
<td>57</td>
<td>813</td>
</tr>
<tr>
<td></td>
<td>AS</td>
<td>GGAATACAAAAATGACGAG</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Opg</td>
<td>S</td>
<td>TGGAGATGCAATCTGCTTG</td>
<td>57</td>
<td>719</td>
</tr>
<tr>
<td></td>
<td>AS</td>
<td>TCAAGTGCTTAGGGGCTAC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alp</td>
<td>S</td>
<td>CTTGCTGGTAGAGGAGGAGG</td>
<td>55</td>
<td>393</td>
</tr>
<tr>
<td></td>
<td>AS</td>
<td>GGAACAGGAATTGGGGAC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>type 1 collagen</td>
<td>S</td>
<td>TCTCCACCTTCCTAGTCTCT</td>
<td>55</td>
<td>269</td>
</tr>
<tr>
<td></td>
<td>AS</td>
<td>TTGGTATCTTCCACAGGTAC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>osteocalcin</td>
<td>S</td>
<td>CAAGTCCCACACAGCACTT</td>
<td>55</td>
<td>370</td>
</tr>
<tr>
<td></td>
<td>AS</td>
<td>AAAGCCGAGCTGCCAGATTT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GAPDH</td>
<td>S</td>
<td>GGTCCGTTGGTAAGCGATTG</td>
<td>55</td>
<td>508</td>
</tr>
<tr>
<td></td>
<td>AS</td>
<td>ATAGCCCTTCCACACTG</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
dewaxed sections were preincubated for 20 minutes in buffer containing 50 mM sodium acetate and 40 mM sodium tartrate at pH 5.0. Sections then were incubated for 15 minutes at room temperature in the same buffer containing 2.5 mg/mL of naphthol AS-MX phosphate (Sigma) in dimethylformamide as substrate and 0.5 mg/mL of fast garnet GBC (Sigma) as a color indicator for the reaction product. After washing with distilled water, the sections were counterstained with methyl green and mounted in Kaiser’s glycerol jelly.

**Immunohistochemical staining**

Immunohistochemical staining for type 1 collagen was performed using affinity-purified goat anti-mouse type 1 collagen antibody (Southern Biotechnology Associates, Birmingham, AL, USA) as described previously. Briefly, dewaxed and rehydrated paraffin-embedded sections were incubated with methanol–hydrogen peroxide (1:10) to block endogenous peroxidase activity and then washed in Tris-buffered saline (pH 7.6). The slides then were incubated with the primary antibody overnight at room temperature. After rinsing with Tris-buffered saline for 15 minutes, tissues were incubated with secondary antibody (biotinylated goat anti-rabbit IgG or biotinylated goat anti-mouse IgG; Sigma). Sections then were washed and incubated with the Vectastain Elite ABC reagent (Vector Laboratories, Burlington, Ontario, Canada) for 45 minutes. Staining was developed using 3,3-diaminobenzidine (2.5 mg/mL) followed by counterstaining with Mayer’s hematoxylin.

**Bone marrow cell cultures**

Tibias and femurs of 3-week-old WT, Pth+/–, and Pth+/– CaR–/– pubes fed by dams on the normal diet or the high-calcium diet that were administrated the exogenous PTH(1–34) were removed under aseptic conditions, and bone marrow cells were flushed out with DMEM containing 10% fetal calf serum (FCS), 50 μg/mL of ascorbic acid, 10 mM β-glycerophosphate, and 10–8 M dexamethasone. Cells were dispersed by repeated pipetting, and a single-cell suspension was achieved by forceful expulsion of the cells through a 22-gauge syringe needle. Total bone marrow cells (10⁶) were cultured in 36-cm² Petri dishes in 5 mL of calcium-deficient medium (Sigma) overnight. These cells then were treated with various reagents as described in the figure legends. After treatment, cells were washed in PBS (20 mM NaH₂PO₄, 0.9% NaCl, pH 7.4) and lysed for 20 minutes with lysis buffer (20 mM Tris-Cl, pH 7.4, 150 mM NaCl, 0.1% Nonidet P-40, 1% glycerol, 0.2 mM sodium vanadate, and a protease mixture tablet/10 mL of buffer). The samples were collected and microcentrifuged at 14,000 rpm for 5 minutes, and the supernatants were collected, assayed for protein, and prepared for Western blot analysis with antibody against the active phosphorylated form of mitogen-activated protein kinase (MAPK). Membranes were stripped and reprobed with polyclonal antibodies against MAPK (ERK-1 and ERK-2).

**Western blot analysis**

Proteins were extracted from the duodenum of 2-week-old WT or CaR–/– mice and quantitated by a kit (Bio-Rad, Mississauga, Ontario, Canada). In total, 30 μg of protein samples was fractionated by SDS-PAGE and transferred to nitrocellulose membranes. Immunoblotting was carried out as described previously using TRPV5 (ECaC1), calbindin-D₂₈K, calbindin-D₉₉K, and the Na⁺/Ca²⁺ exchanger (NCX1) as markers (Swant, Bellinzona, Switzerland) and β-tubulin (Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA). Bands were visualized using ECL chemiluminescence (Amersham, Piscataway, NJ, USA) and quantitated by Scion Image Beta 4.02 (Scion Corporation, Frederick, MD, USA).

**Computer-assisted image analysis**

After H&E staining or histochemical or immunohistochemical staining of sections from six mice of each genotype, images of fields were photographed with a Sony (Tokyo, Japan) digital camera. Images of micrographs from single sections were digitally recorded using a rectangular template, and recordings were processed and analyzed using Northern Eclipse image-analysis software (Empix Imaging Inc.), as described previously.

**Statistical analysis**

Data from image analysis are presented as means ± SEM. Statistical comparisons were made using a two-way ANOVA, with p < .05 being considered significant.

**Results**

**Effect of dietary calcium on milk calcium content and calcium-regulating hormones in milk**

To determine whether dietary calcium affects milk calcium content and calcium-regulating hormones, milk calcium, 1,25(OH)₂D₃, and PTHrP levels were examined in lactating WT and CaR–/– dams fed either the normal diet or the high-calcium diet. The high-calcium diet increased milk calcium and decreased PTHrP and 1,25(OH)₂D₃ levels (Fig. 1A–C) but did not affect milk protein concentrations (Fig. 1D).
Effects of dietary calcium and exogenous PTH on serum calcium, phosphorus, PTH, and 1,25(OH)₂D₃ levels in Pth⁻/⁻ and Pth⁻/⁻CaR⁻/⁻ pups

To determine whether maternal dietary calcium and exogenous PTH affect mineral homeostasis in the suckling pups, serum calcium, phosphorus, PTH, and 1,25(OH)₂D₃ levels were measured in 3-week-old pups fed by Pth⁺/⁻CaR⁺/⁻ dams on either the normal diet or the high-calcium diet or administrated exogenous PTH. Serum calcium levels were decreased in Pth⁻/⁻ and Pth⁻/⁻CaR⁻/⁻ pups fed by Pth⁺/⁻CaR⁺/⁻ dams on either the normal diet or the high-calcium diet or administrated exogenous PTH and were decreased more dramatically in Pth⁻/⁻CaR⁻/⁻ pups fed by Pth⁺/⁻CaR⁺/⁻ dams on the high-calcium diet or...
administered exogenous PTH (Fig. 1E). Serum phosphorus levels were still elevated relative to WT mice in both mutants when fed by mothers on either the normal diet or the high-calcium diet and were reduced further in the genotype-matched pups fed by dams on the high-calcium diet compared with pups fed by dams on the normal diet (Fig. 1F). Serum phosphorus levels were normalized by exogenous PTH treatment of both mutant animals (Fig. 1F). Serum PTH levels in WT pups were decreased when the dams were on the high-calcium diet. Administration of PTH to pups increased PTH levels in WT pups fed by dams on the normal diet. Basal serum PTH levels in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups were undetectable (Fig. 1G). Although exogenous PTH administration increased serum PTH levels in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups, the levels were significantly lower than in their WT littermates (Fig. 1G). Serum 1.25(OH)<sub>2</sub>D<sub>3</sub> levels were reduced in Pth<sup>−/−</sup> pups and were even more reduced in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups fed by dams on the normal diet. Feeding dams a high-calcium diet decreased serum 1.25(OH)<sub>2</sub>D<sub>3</sub> levels significantly in WT and Pth<sup>−/−</sup> pups but insignificantly in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups compared with genotype-matched pups from dams fed the normal diet. PTH increased BMD was determined using a PIXImus densitometer (Fig. 2A). Tibias then were examined by either the normal or the high-calcium diet, or pups were administrated exogenous PTH. Two weeks of PTH treatment increased tibial length and BMD in WT and Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups, but the percentage increase was less in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups than in the other two genotypes (Fig. 1H).

Effects of dietary calcium and exogenous PTH on bone growth, BMD, and bone volume in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups

To determine whether CaR deficiency influences the effects of dietary calcium and exogenous PTH on bone growth and BMD, 3-week-old pups were fed by Pth<sup>−/−</sup>CaR<sup>−/−</sup> dams that received either the normal or the high-calcium diet, or pups were administrated exogenous PTH. Tibias then were examined by radiography (Fig. 2A), tibia lengths were measured (Fig. 2B), and BMD was determined using a PIXImus densitometer (Fig. 2C) in the 3-week-old pups. Pth<sup>−/−</sup> pups fed by dams receiving either a normal or a high-calcium diet had a significant reduction in bone length and BMD compared with WT pups. The reduction was more severe in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups. Administering the high-calcium diet to dams increased serum 1.25(OH)<sub>2</sub>D<sub>3</sub> levels significantly in WT and Pth<sup>−/−</sup> pups but insignificantly in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups compared with genotype-matched pups. Basal serum PTH levels in WT and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups were still elevated relative to WT mice in both mutants when fed by dams on the high-calcium diet. Administration of PTH to pups increased serum PTH levels in WT pups fed by dams on the normal diet. Basal serum PTH levels in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups were undetectable (Fig. 1G). Although exogenous PTH administration increased serum PTH levels in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups, the levels were significantly lower than in their WT littermates (Fig. 1G). Serum 1.25(OH)<sub>2</sub>D<sub>3</sub> levels were reduced in Pth<sup>−/−</sup> pups and were even more reduced in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups fed by dams on the normal diet. Feeding dams a high-calcium diet decreased serum 1.25(OH)<sub>2</sub>D<sub>3</sub> levels significantly in WT and Pth<sup>−/−</sup> pups but insignificantly in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups compared with genotype-matched pups from dams fed the normal diet. PTH increased serum 1.25(OH)<sub>2</sub>D<sub>3</sub> levels in all WT, Pth<sup>−/−</sup>, and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups, but the percentage increase was less in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups than in the other two genotypes (Fig. 1H).

Effects of dietary calcium and exogenous PTH on osteoblastic bone formation in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups

To determine whether or not alterations of trabecular bone volume in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups are associated with changes in osteoblast function, bone-formation parameters were assessed by staining with H&E (Fig. 3A), immunostaining for type 1 collagen (Fig. 3B), double calcein labeling (Fig. 3C) and histomorphometric analysis for osteoblast number (Fig. 3D), type 1 positive area (Fig. 3E), and mineral apposition rate (MAR; Fig. 3F). Pth<sup>−/−</sup> pups fed by dams on the normal diet displayed a significantly reduced osteoblast number, type 1 collagen immunopositive area, and MAR compared with WT pups, and these parameters were decreased further in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups. These bone-formation parameters were increased in WT and Pth<sup>−/−</sup> pups but not in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups fed by dams on the high-calcium diet compared with the pups fed by dams on the normal calcium diet. PTH treatment for 2 weeks increased bone formation in WT and Pth<sup>−/−</sup> pups and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups to a lesser extent. Exogenous PTH had a greater stimulatory effect on bone formation than in the pups fed by dams on the high-calcium diet in both WT and Pth<sup>−/−</sup> pups (Fig. 3A–F). We also assessed whether these alterations of osteoblastic bone formation were associated with similar changes in the expression of several osteoblastic genes. The expression levels of Cbfa-1 (Fig. 4A), Alp (Fig. 4B), type 1 collagen (Fig. 4C), and osteocalcin (Fig. 4D) were examined by real-time RT-PCR. The alterations in the expression of transcripts for Cbfa-1, Alp, type 1 collagen, and osteocalcin, as assessed by real-time RT-PCR, were consistent with the alterations observed by histomorphometry.

Effects of dietary calcium and exogenous PTH on osteoprogenitor commitment in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups

To investigate whether the reduction in osteoblast numbers in Pth<sup>−/−</sup> and Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups is due to impaired commitment of bone marrow mesenchymal stem cells (MSCs) into osteogenic cells, colony-forming assays were performed using bone marrow stromal cells from these animals. Consistent with the bone-formation data, the total number of colony-forming units fibroblastic (CFU-F) and ALP<sup>−</sup> CFU-F (CFU-F<sub>AP</sub>) in bones from Pth<sup>−/−</sup>pups fed by dams on either the normal or the high-calcium diet were reduced significantly compared with WT pups, with a further decrease in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups (Fig. 5). The high-calcium diet increased the numbers of CFU-F and CFU-F<sub>AP</sub> in WT and Pth<sup>−/−</sup> pups but not in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups. Similar to the bone-formation parameters, PTH treatment increased CFU-F numbers in WT and Pth<sup>−/−</sup> pups, with a lesser increase in Pth<sup>−/−</sup>CaR<sup>−/−</sup> pups. The percentage increase in these osteogenic parameters was
Fig. 2. Effects of dietary calcium and exogenous PTH on bone growth, BMD, and bone volume in Pth\(^{−/−}\) and Pth\(^{−/−}\)CaR\(^{−/−}\) pups. (A) X-rays of tibias, (B) the lengths of tibias, (C) BMD values, (D) representative longitudinal sections of the proximal ends of tibias by \(\mu\)CT and 3D reconstruction, and (E) trabecular bone volume (BV/TV) that were determined in the metaphyseal regions, including both primary and secondary spongiosa, of tibias from 3-week-old WT, Pth\(^{−/−}\), and Pth\(^{−/−}\)CaR\(^{−/−}\) pups fed by dams on normal Ca or high Ca or were administered exogenous PTH(1–34). Each value is the mean ± SEM of determinations in 5 mice of each genotype. *\(p < .05\); **\(p < .01\); ***\(p < .001\) compared with WT littermates of the same group; *\(p < .05\); **\(p < .01\); ***\(p < .001\) compared with Pth\(^{−/−}\) pups of the same group; ~\(p < .05\); ~~\(p < .01\); ~~~\(p < .001\) compared with the genotype-matched pups fed by dams on the normal diet.
higher in PTH-treated pups than in pups fed by dams on the high-calcium diet in both WT and Pth–/– pups (Fig. 5A–D).

Effects of dietary calcium and exogenous PTH on osteoclastic bone resorption in Pth–/– and Pth–/–CaR–/– pups

To determine whether alterations in osteoclast function also contribute to decreased bone volume in Pth–/– and Pth–/–CaR–/– pups, osteoclast number and surface were determined by histomorphometric analysis on TRACP-stained sections (Fig. 6).

We also examined the expression levels of Rankl and Opg mRNA in bony tissues by real-time RT-PCR and calculated the ratio of Rankl/Opg. The number and surfaces of TRACP+ osteoclasts and the ratio of Rankl/Opg were reduced significantly in Pth–/– pups and less significantly in Pth–/–CaR–/– pups compared with WT pups fed by dams on either the normal or the high-calcium diet.

Fig. 3. Effects of dietary calcium and exogenous PTH on osteoblastic bone formation in Pth–/– and Pth–/–CaR–/– pups. Representative micrographs of sections that were (A) stained with H&E, (B) immunostained for type 1 collagen (Col I), or (C) double calcein labeled from trabecular bone from 3-week-old WT, Pth–/–, and Pth–/–CaR–/– pups fed by dams fed with normal Ca or high Ca or pups that were administered exogenous PTH(1–34). Scale bars in A, B, and C represent 50, 50, and 25 μm, respectively. (D) Numbers of osteoblasts per square millimeter (N.Ob/T.Ar, n/mm²) that were counted in the metaphyseal regions of H&E-stained tibial sections. (E) The type 1 collagen–positive area as a percentage of the tissue area was measured in the metaphyseal regions of tibial sections, and (F) MAR of trabecular bone. Each value is the mean ± SEM of determinations in 5 mice of each genotype. *p < .05; **p < .01; ***p < .001 compared with WT littermates of the same group; ▲p < .05; ▲▲p < .01; ▲▲▲p < .001 compared with Pth–/– pups of the same group; ≈p < .05; ≈≈p < .01; ≈≈≈p < .001 compared with the genotype-matched pups fed by dams on the normal diet.
These parameters were reduced significantly in WT and Pth\(^{-/-}\) pups but did not decrease significantly in Pth\(^{-/-}\)CaR\(^{-/-}\) pups fed by dams on the high-calcium diet compared with genotype-matched pups fed by dams on the normal diet. These parameters increased significantly in PTH-treated WT and Pth\(^{-/-}\) pups and less significantly in PTH-treated Pth\(^{-/-}\)CaR\(^{-/-}\) pups compared with vehicle-treated genotype-matched pups fed by dams on the normal diet (Fig. 6A–D).

**Effect of CaR deficiency on osteoprogenitor intracellular signaling, intestinal calcium transporter expression, and urinary calcium excretion**

To determine whether bone marrow MSCs from CaR-deficient mice respond to signaling changes in extracellular calcium (Ca\(^{2+}\)) concentrations, cells were treated with either a high concentration of Ca\(^{2+}\) or with the nonpermeant CaR agonist gadolinium (Gd\(^{3+}\)), and ERK1/2 phosphorylation was determined by Western blots with an antibody against the active, phosphorylated form of ERK1/2. Basal levels of ERK1/2 in the presence of 0.5 mM Ca\(^{2+}\) were significantly lower in cells from CaR\(^{-/-}\) mice than in cells from WT mice. Elevated Ca\(^{2+}\) or Gd\(^{3+}\) treatment significantly induced ERK1/2 activation in the bone marrow MSCs from both WT and CaR\(^{-/-}\) mice at 5 minutes. However, the high-Ca\(^{2+}\)-induced ERK1/2 activation was markedly more pronounced in bone marrow MSCs from WT mice than from CaR\(^{-/-}\) mice. The Gd\(^{3+}\)-induced ERK1/2 activation in bone marrow MSCs from WT mice was even more pronounced than in cells from CaR\(^{-/-}\) mice, where Gd\(^{3+}\)-stimulated ERK1/2 activation barely reached the basal levels of the WT mice (Fig. 7A, B). High-Ca\(^{2+}\)-induced ERK1/2 activation was time-dependent in the cells from both WT and CaR\(^{-/-}\) mice. However, ERK1/2 activation from 2 to 10 minutes was more markedly stimulated by 5 mM Ca\(^{2+}\) in cells from WT mice than in cells from CaR\(^{-/-}\) mice (Fig. 7C, D).

To determine whether the ability of Ca\(^{2+}\) transport in the duodenum is altered in the CaR-deficient mice, the expression of intestinal calcium transporters was examined in duodenums from WT and CaR\(^{-/-}\) mice by Western blots. The results revealed that protein expression levels of TRPV5, calbindin-D28K, calbindin-D9K, and NCX1 in the duodenum were reduced in CaR\(^{-/-}\) mice compared with their WT littermates (Fig. 7E, F). When urine calcium and creatinine levels were measured and the urine calcium/creatinine ratio was calculated, we found that the urine calcium/creatinine ratio was lower in CaR\(^{-/-}\) mice than in their WT littermates (Fig. 7G).

**Discussion**

We employed a genetic approach to determine whether deficiency of PTH alone or in combination with loss of CaR
leads to alterations in calcium and skeletal homeostasis in neonatal animals in vivo, and we investigated the involvement of dietary calcium supplementation or exogenous PTH administration in these processes using Pth<sup>-/-</sup> and Pth<sup>-/-</sup>CaR<sup>+-/-</sup> pups that were fed by dams on normal Ca or high Ca or were administered exogenous PTH(1–34) were cultured in osteogenic differentiation medium for 18 days and stained with methylene blue for total number of colonies (CFU-F) or cytochemically for ALP. Each value is the mean ± SEM of determinations in 5 mice of each genotype. *p < 0.05; **p < 0.01; ***p < 0.001 compared with WT littermates of the same group; ▲p < 0.05; ▲▲p < 0.01; ▲▲▲p < 0.001 compared with Pth<sup>-/-</sup> pups of the same group; ▼p < 0.05; ▼▼p < 0.01; ▼▼▼p < 0.001 compared with the genotype-matched pups fed by dams on the normal diet.

Fig. 5. Effects of dietary calcium and exogenous PTH on osteoprogenitor commitment in Pth<sup>-/-</sup> and Pth<sup>-/-</sup>CaR<sup>+-/-</sup> pups. Bone marrow cells from WT, Pth<sup>-/-</sup>, and Pth<sup>-/-</sup>CaR<sup>+-/-</sup> pups that were fed by dams on normal Ca or high Ca or were administered exogenous PTH(1–34) were cultured in osteogenic differentiation medium for 18 days and (A) stained with methylene blue for total number of colonies (CFU-F) or (B) cytochemically for ALP. (C) Total CFU-F number per dish. (D) ALP<sup>+</sup> CFU-F (CFU-F<sub>AP</sub>) number per dish. Each value is the mean ± SEM of determinations in 5 mice of each genotype. *p < 0.05; **p < 0.01; ***p < 0.001 compared with WT littermates of the same group; ▲p < 0.05; ▲▲p < 0.01; ▲▲▲p < 0.001 compared with Pth<sup>-/-</sup> pups of the same group; ▼p < 0.05; ▼▼p < 0.01; ▼▼▼p < 0.001 compared with the genotype-matched pups fed by dams on the normal diet.
kidney and possibly in the gut also may contribute significantly to the development of the bone phenotype in vivo.

We further investigated whether CaR deficiency affects skeletal responses to dietary calcium supplementation. Because the calcium source for suckling neonates is maternal milk, we altered the dietary content of the dams to improve calcium intake in the neonates. Our results show that a high-calcium diet increases the calcium levels in milk of the lactating mothers. In contrast, milk PTHrP and 1,25(OH)2D3 levels were reduced in dams fed the high-calcium diet compared with dams fed the normal calcium diet. Previous study has shown that the CaR stimulates calcium transport into milk and inhibits PTHrP secretion by the breast. In view of the fact that the protein levels in milk were not substantially different, dietary calcium appeared to influence milk calcium independent of changes in protein. We also demonstrate that serum calcium, phosphorus, PTH, and 1,25(OH)2D3 levels are altered in suckling pups by altering the calcium in the diet of the lactating mothers. These results suggest that the high-calcium diet can elevate milk calcium significantly in lactating mothers, which subsequently results in changes of serum ion homeostasis and calcitrophic hormones in suckling neonates. These findings are consistent with our previous report related to the effect of dietary calcium supplementation on milk calcium content and calcium-regulating hormones in dams and on serum calcium, phosphorus, and 1,25(OH)2D3 in sucking pups.

We found that the loss of CaR in Pth−/−CaR−/− pups abolished dietary calcium supplement–mediated augmentation of osteoblastic bone formation. Thus, compared with genotype-matched pups fed by dams on the normal diet, skeletal growth and bone–formation parameters were increased in the WT and Pth−/− pups but not in Pth−/−CaR−/− pups fed by dams on the high-calcium diet. These results, demonstrating that CaR deficiency in vivo abolishes skeletal responses to dietary calcium supplementation in suckling neonates, are consistent with previously published in vitro findings in which exposure of primary osteoblasts or a variety of osteoblast-like cells to high calcium or the polycationic CaR agonists neomycin and gadolinium stimulates their

Fig. 6. Effects of dietary calcium and exogenous PTH(1–34) on osteoclastic bone resorption in Pth−/− and Pth−/−CaR−/− pups. Representative micrographs of sections of the tibias from WT, Pth−/−, and Pth−/−CaR−/− pups that were fed by dams on normal Ca or high Ca or were administered exogenous PTH(1–34) (A) stained histochemically for TRACP. Scale bar = 50 μm. (B) Number of TRACP+ osteoclasts related to tissue area (N.Oc/T.Ar, /mm2) and (C) osteoclast surface relative to bone surface (Oc.S/BS, %) were counted in the metaphyseal regions of TRACP-stained tibial sections. (D) Real-time RT-PCR was performed on bone extracts for Rankl and Opg mRNA as described in “Materials and Methods.” Messenger RNA expression assessed by real-time RT-PCR analysis is calculated as a ratio to the GAPDH mRNA level and expressed relative to levels of WT mice. Ratio of Rankl/Opg relative mRNA levels was calculated. Each value is the mean ± SEM of determinations in 5 mice of each group. *p < .05; **p < .01; ***p < .001 compared with WT littermates of the same group; Δp < .05; ΔΔp < .01; ΔΔΔp < .001 compared with Pth−/− pups of the same group; ^p < .05; ^^p < .01; ^^^p < .001 compared with the genotype-matched pups fed by dams on the normal diet.
proliferation, differentiation, and mineralization by activating p42/44 MAPK, p38 MAPK, and/or JNK pathways. (5,7,41,42) Although an alternate skeletal calcium receptor, GPRC6A, has been described(43,44) through which calcium might act, its role in skeletal physiology is unclear because conflicting reports on the skeletal phenotype of Gprc6a null mice have been reported,(44,45) and no calcium channel or transporter in bone has been reported consistently to alter osteoblast and osteoclast activity in response to changes in serum calcium. Consequently, the calcium effect appears to be via CaR. Although serum calcium

Fig. 7. Effect of CaR deficiency on osteoprogenitor intracellular signaling, intestinal calcium transporter expression, and renal calcium excretion. (A) Second-passaged bone marrow MSCs from WT and CaR−/− (KO) mice were treated with 0.5 and 5 mM CaCl2 or 50 μM Gd3+ for 5 minutes, and cells were analyzed by Western blot for ERK phosphorylation activation (p-ERK) and total ERK (ERK). (B) p-ERK level relative to total ERK level was assessed by densitometric analysis and expressed relative to levels of cells from WT mice treated with 0.5 mM CaCl2. (C) The cells were treated with 5 mM CaCl2 for various times and were analyzed by Western blot for ERK phosphorylation activation (p-ERK) and total ERK (ERK). (D) p-ERK level relative to total ERK level was assessed by densitometric analysis and expressed relative to levels of cells from untreated cultures. (E) Western blots of extracts of duodenums from WT and CaR−/− (KO) mice for expression of TRPV5, calbindin-D28K (CB28K), calbindin-D9K (CB9K), and NCX1. β-Tubulin was used as a loading control for Western blots. (F) TRPV5, CB28K, CB9K and NCX protein levels relative to β-tubulin protein level were assessed by densitometric analysis and expressed relative to levels of WT mice. (G) Urine calcium and urine creatinine were measured as described in “Materials and Methods,” and the urine calcium/creatinine ratio was calculated. *p < .05; **p < .01; ***p < .001 compared with WT mice.
increases were more modest in the Pth<sup>+/−</sup>CaR<sup>+/−</sup> mice than in the WT and Pth<sup>−/−</sup> mice, our conclusion would be that even if serum calcium were elevated to the same degree in all phenotypes we employed in this study, the absence of the CaR would be the factor reducing skeletal responsiveness. Our results therefore strongly indicate that CaR plays an important role in regulating dietary calcium-induced changes in both bone formation and resorption in neonates in vivo.

We demonstrated previously that the skeletal anabolic action of PTH in neonates results not only from its direct action via the PTH receptor to increase the osteoblast pool but also by an indirect action mediated through increasing extracellular calcium concentration. In this study, we further demonstrated that increasing the extracellular calcium concentration acts cooperatively with PTH to promote osteoblastic bone formation in the neonate and that the effect of the former is mediated via the CaR. When 1-week-old WT, Pth<sup>−/−</sup>, and Pth<sup>+/−</sup>CaR<sup>+/−</sup> pups were injected with PTH(1–34) daily for 2 weeks, PTH(1–34) administration raised serum calcium and 1,25(OH)<sub>2</sub>D<sub>3</sub> levels and reduced serum phosphorus levels significantly in WT and Pth<sup>−/−</sup> pups and less significantly in Pth<sup>+/−</sup>CaR<sup>+/−</sup> pups. The length of long bones; BMD values; trabecular volume; osteoblast number; type 1 collagen deposition in bone matrix; MAR; and the gene expression of Cbfa-1, Alp, type 1 collagen, and osteocalcin, as well as CFU-F forming potential from ex vivo bone marrow cultures, all were increased significantly in WT and Pth<sup>−/−</sup> pups and less significantly in Pth<sup>+/−</sup>CaR<sup>+/−</sup> pups. Thus it is likely that the CaR is required for an optimal skeletal anabolic action of PTH in neonates.

Previous studies have revealed the importance of regulation of osteoclast function by extracellular calcium. Exposing osteoclasts to very high calcium, as seen only in the bone microenvironment, results in dramatic cell retraction followed by a profound inhibition of bone resorption. It has been shown that such very high calcium levels inhibit bone-resorbing activity of osteoclasts by directly acting on the CaR, which is expressed by osteoclast precursors and mature osteoclasts. Furthermore, mature osteoclasts undergo apoptosis in the presence of very high calcium, and a dominant-negative CaR construct abrogates this effect, indicating the role of CaR in osteoclast apoptosis. The signaling pathways that are associated with induction of osteoclast apoptosis by the CaR likely involve PLC and NF-κB. Evidence also has been provided recently to support the role of CaR in osteoclast differentiation, and using CaR<sup>+/−</sup> mice, Mentaverri and colleagues have shown that osteoclast differentiation from bone marrow precursor cells in vitro is 70% less in CaR<sup>+/−</sup> mice compared with WT mice. Taken together, these studies suggest that CaR activation is linked to physiologic responses of bone-resorbing activity, although it is not clear if the calcium concentrations required for these processes are equivalent. In this study, we found that TRACP<sup>+/−</sup> osteoclast numbers and surfaces, as well as the ratio of Rankl/OPG, were reduced more dramatically in Pth<sup>−/−</sup> pups than in Pth<sup>+/−</sup>CaR<sup>+/−</sup> pups fed by dams on the normal diet and, unlike pups from dams fed a high-calcium diet, were not further reduced significantly in Pth<sup>−/−</sup>CaR<sup>+/−</sup> pups fed by dams on the same diet. Taken together, our observations suggest that in the model used, CaR is needed to support the calcemic action of PTH and to mediate calcium-induced inhibition of osteoclastic bone resorption, as suggested by earlier studies.

Our findings are relevant to neonates but may differ in adult CaR-deficient mice. Thus our previous studies had shown that the bone phenotypes of Pth<sup>−/−</sup> mice are different before and after weaning. Trabecular bone volume was reduced significantly in newborn and 2- and 3-week-old Pth<sup>−/−</sup> pups compared with their WT littermates. In contrast, trabecular bone volume was increased significantly in 4-month-old Pth<sup>−/−</sup> mice compared with their WT littermates. Although it is unclear why PTH deficiency results in such different bone phenotypes before weaning and in adulthood, such changes may be related to the different forms of food intake in these two settings. Mice are breast-fed before weaning but eat mouse chow thereafter. Breast milk not only contains a higher calcium concentration than the normal diet fed after weaning but also contains low levels of 1,25(OH)<sub>2</sub>D<sub>3</sub> both of which will have an impact on intestinal calcium absorption. Physiologic adaptation to these environmental changes may involve alterations in the function and/or efficacy of calcium-regulating hormones, including PTH, vitamin D, and calcium.

In summary, this study describes the role of the CaR in skeletal turnover in neonates and for the first time demonstrates that in the neonate, the CaR is essential to mediate changes in bone turnover resulting from alterations in dietary calcium as well as to facilitate the increased bone turnover elicited by PTH.

**Disclosures**

All the authors state that they have no conflicts of interest.

**Acknowledgments**

This work was supported by the Key Project grants from the National Natural Science Foundation of China (No. 30830103) and by Program for Changjiang Scholars and Innovative Research Team in University (to DM), a grant from the Canadian Institutes for Health Research (to DG), and a grant from the National Institutes of Health (DK078331 to EB and DK070756 to MP).

**References**


