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MetR-Regulated *Vibrio cholerae* Metabolism Is Required for Virulence

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**ABSTRACT** LysR-type transcriptional regulators (LTTRs) are the largest, most diverse family of prokaryotic transcription factors, with regulatory roles spanning metabolism, cell growth and division, and pathogenesis. Using a sequence-defined transposon mutant library, we screened a panel of *V. cholerae* El Tor mutants to identify LTTRs required for host intestinal colonization. Surprisingly, out of 38 LTTRs, only one severely affected intestinal colonization in the suckling mouse model of cholera: the methionine metabolism regulator, MetR. Genetic analysis of genes influenced by MetR revealed that *glyA1* and *metJ* were also required for intestinal colonization. Chromatin immunoprecipitation of MetR and quantitative reverse transcription-PCR (qRT-PCR) confirmed interaction with and regulation of *glyA1*, indicating that misregulation of *glyA1* is likely responsible for the colonization defect observed in the *metR* mutant. The *glyA1* mutant was auxotrophic for glycine but exhibited wild-type trimethoprim sensitivity, making folate deficiency an unlikely cause of its colonization defect. MetJ regulatory mutants are not auxotrophic but are likely altered in the regulation of amino acid-biosynthetic pathways, including those for methionine, glycine, and serine, and this misregulation likely explains its colonization defect. However, mutants defective in methionine, serine, and cysteine biosynthesis exhibited wild-type virulence, suggesting that these amino acids can be scavenged in vivo. Taken together, our results suggest that glycine biosynthesis may be required to alleviate an in vivo nutritional restriction in the mouse intestine; however, additional roles for glycine may exist. Irrespective of the precise nature of this requirement, this study illustrates the importance of pathogen metabolism, and the regulation thereof, as a virulence factor.

**IMPORTANCE** *Vibrio cholerae* continues to be a severe cause of morbidity and mortality in developing countries. Identification of *V. cholerae* factors critical to disease progression offers the potential to develop or improve upon therapeutics and prevention strategies. To increase the efficiency of virulence factor discovery, we employed a regulator-centric approach to multiplex our in vivo screening capabilities and allow whole regulons in *V. cholerae* to be interrogated for pathogenic potential. We identified MetR as a new virulence regulator and serine hydroxymethyltransferase GlyA1 as a new MetR-regulated virulence factor, both required by *V. cholerae* to colonize the infant mouse intestine. Bacterial metabolism is a prerequisite to virulence, and current knowledge of in vivo metabolism of pathogens is limited. Here, we expand the known role of amino acid metabolism and regulation in virulence and offer new insights into the in vivo metabolic requirements of *V. cholerae* within the mouse intestine.
how members of its regulon are critical for 
*V. cholerae* host colonization. These findings underscore the importance of pathogen metabolism, and the regulation thereof, as a virulence factor.

**RESULTS**

**MetR is essential for colonization of the infant mouse intestine.**

We chose to screen the LysR-type transcriptional regulator (LTTR) family because it is the largest and most diverse family of prokaryotic transcriptional regulators with regulatory roles spanning metabolism, cell growth and division, and pathogenesis (16–19). Several LTTRs are conserved among bacteria; thus, virulence regulators identified in *V. cholerae* may also contribute to the virulence of other pathogens.

A panel of 38 *Vibrio cholerae* El Tor mutants representing all annotated LTTRs available from our sequence-defined transposon insertion library was selected (20). The ability of each LTTR mutant to colonize the intestine was examined using the suckling mouse model (21, 22), which compares the *in vivo* fitness of a mutant strain to the wild-type parental strain, yielding a ratio termed the competitive index (CI). Only two LTTR mutants were significantly impaired in colonization (Fig. 1): the strain with the mutation for the methionine regulator MetR showed a highly significant, severe defect (CI = 0.028 ± 0.021; *P* < 0.001), while the strain with the mutation for a regulator of unknown function encoded by the locus VC1947 exhibited a less significant defect (CI = 0.719 ± 0.632; *P* < 0.05). The CI values for well-studied regulators such as GcvA (glycine cleavage regulon), CysB (cysteine regulon), IlvY (branched amino acid regulon), LeuO (leucine regulon), and OxyR (oxidative stress regulon) were not significantly different from that of the wild type (Fig. 1).

**glyA1 and metJ are essential for colonization of the infant mouse intestine.** Since MetR is a transcriptional activator required for the biosynthesis of methionine, we focused our secondary search for the MetR-regulated virulence factor(s) on a panel of transposon mutants with disruption of genes involved in methionine metabolism. The metabolic pathways in *V. cholerae* remain understudied; however, the well-conserved nature of metabolic genes combined with the well-defined pathways in *Escherichia coli* allowed bioinformatic extrapolation and annotation of the *V. cholerae* genome (23, 24). The KEGG and VchoCyc databases proved useful in determining which pathways may exist in *V. cholerae* and subsequently which mutants to select for the panel (25–27). Colonization assays on our selected mutant panel (Fig. 2) revealed that mutants defective in *glyA1*, which encodes serine hydroxymethyltransferase, and *metJ*, which encodes the methionine repressor, displayed significant colonization defects with CI values of 0.087 ± 0.075 (*P* < 0.001) and 0.252 ± 0.016 (*P* < 0.001), respectively. Disruption of *metA*, *metB*, and *metC* of the homocysteine biosynthetic branch, *metF* of the folate branch, and the methionine synthase genes *metE* and *metH* did not result in a detectable colonization defect. Additionally, disruption of genes involved in the uptake of methionine, *metL* and *metT*, or cobalamin, *mtsB* and *btuB*, had no impact on *in vivo* fitness. Interestingly, *V. cholerae* possesses a heterologous copy of serine hydroxymethyltransferase encoded by *glyA2* in the small chromosome; however, disruption of this gene did not result in a colonization defect. We hypothesized that the colonization defect displayed by the *glyA1* mutant was the result of glycine limitation; however, the possibility that *glyA1* disruption impedes biosynthesis of a vital glycine-dependent metabolite required further investigation. We decided to examine genes of metabolic pathways involved in glycine processing: the glycine cleavage system genes *gcvP*, *gcvH*, and *gcvT*, the serine biosynthesis gene *serB*, and the cysteine biosynthesis gene *cysE*. All of these mutants displayed normal colonization properties in the infant mouse intestine (Fig. 2).

**Disruption of *glyA1* results in glycine auxotrophy but not trimethoprim sensitivity.** To phenotypically validate transposon disruption of essential metabolic genes, we examined the methionine mutant panel for growth in M9 minimal medium with gluo-
cose as the sole carbon source (see Fig. S1 in the supplemental material). In agreement with their annotated functions, we found that disruption of genes in the homocysteine biosynthesis and folate branches, as well as the cobalamin-independent methionine synthase gene metE, resulted in the inability to grow in minimal medium. Furthermore, the cobalamin-dependent methionine synthase gene metH and the methionine uptake transporter genes were dispensable, while disruption of the cobalamin uptake transporter gene inhibited growth. The glycine cleavage system genes gcvH, gcvP, and gcvT were not required for growth in minimal medium, while the serine, threonine, cysteine, and homoserine biosynthesis genes serB, thrB, cysE, and thrA, respectively, were all required. In agreement with the regulatory roles of MetR and MetJ, disruption of metR resulted in lack of growth while the metJ mutant exhibited wild-type growth. The glyA2 mutant grew in minimal medium, consistent with the dispensability of glyA2 for in vivo growth.

The growth kinetics of glyA1, metR, and metJ mutants in M9 minimal medium with and without supplementation with methionine, serine, threonine, aspartate, or glycine (Fig. 3) were analyzed to determine nutritional auxotrophy status of each metabolic mutant. As confirmed previously, MetR and GlyA1 were required for growth in minimal medium, while MetJ was dispensable. The metJ mutant exhibited growth inhibition relative to the wild type when glycine was added. The metR mutant exhibited methionine auxotrophy that could be reversed with the addition of methionine. In fact, addition of methionine caused increased growth of the metJ mutant compared to the wild type. The addition of glycine was unable to rescue the metR mutant; however, this was likely the result of methionine limitation. The glyA1 mutant exhibited glycine auxotrophy that could be reversed by the addition of serine, threonine, or glycine but not methionine. Neither the glyA1 mutant nor the metR mutant could be rescued by the addition of aspartate.

GlyA1 not only catalyzes the reversible reaction of serine to glycine but simultaneously regenerates tetrahydrofolate to N5,N10-methylene tetrahydrofolate by transfer of a methyl group. Disruption of tetrahydrofolate regeneration creates a “methyl trap,” a type of folate deficiency, and consequently growth inhibition. In order to determine if disruption of glyA1 would result in folate deficiency, we compared trimethoprim sensitivities of the wild type and the glyA1 mutant. Trimethoprim inhibits dihydrofolate reductase, resulting in blockage of tetrahydrofolate acid synthesis. We hypothesized that if the in vivo limitation of tetrahydrofolate was responsible for the colonization defect of the glyA1 mutant, then this mutant should also exhibit increased sensitivity to, or a reduced MIC of, trimethoprim. The glyA1 mutant and the wild type exhibited equal sensitivities to trimethoprim, as determined by MIC determination (data not shown) and growth kinetics (see Fig. S2 in the supplemental material).

Chromatin immunoprecipitation of MetR confirms its association with glyA1. The regulon of MetR in V. cholerae is unknown; thus, a direct regulatory relationship between MetR and glyA1 is lacking. To determine whether MetR is capable of binding to the promoter of glyA1, we performed chromatin immunoprecipitation (ChiP) using epitope-tagged MetR overexpressed in V. cholerae from an arabinose-inducible promoter. This method was used in previous studies to map the genomic binding locations of the V. cholerae transcriptional regulators Fur and ToxT (11, 28). After conducting ChiP on C6706(pBAD) or C6706(pMetR-3xV5) grown in LB, we performed quantitative PCR (qPCR) validation to identify enrichment of promoter targets bound by MetR using primers specific to the promoter regions of metA, metE, metH, metR, glyA1, glyA2, hmpA, and tcpA (Fig. 4). The promoter of toxin-coregulated pilus encoded by tcpA served as a negative control. Enrichment of metE, metR, and glyA1 promoter regions in C6706(pMetR) relative to C6706(pBAD) were 45-fold, 102-fold, and 45-fold, respectively. Conversely, the promoter regions for tcpA, metA, metH, and hmpA were not enriched.

The ChiP results confirmed direct association of MetR with the promoters of glyA1 and metE. To assay MetR transcriptional control at these sites, RNA samples were obtained from C6706(pMetR) incubated in M9 minimal medium or LB medium, with or without arabinose induction of metR. The relative abundance of mRNA transcripts from metA, metC, metE, metH, metJ, metR, glyA1, tcpA, and 16S rRNA transcripts were determined using quantitative reverse transcription-PCR (qRT-PCR). The 16S rRNA served as an internal control to normalize samples to total RNA, and tcpA served as a negative control. Our expression analysis compared all samples to the noninduced LB medium sample to yield the relative change in expression (see Table S2 in the supplemental material). We observed 343-fold and 505-fold increases in abundance of metR transcript levels under arabinose induction conditions compared to noninduction upon growth in M9 and LB medium, respectively. Furthermore, metR induction with arabinose resulted in a 2.0-fold repression of glyA1 and metE in M9 medium and a 3.5-fold upregulation of metE in LB medium. All other genes examined showed a <2-fold difference in expression when arabinose induction and noninduction were compared. Analysis of growth in LB relative to M9 medium showed that all genes experienced strong nutrient-dependent repression that ranged from approximately 10-fold to 700-fold regardless of metR induction.

**DISCUSSION**

Transcriptional regulators of the LysR-type family are found in all prokaryotes and regulate virulence in a wide range of bacterial species (29–33). Interestingly, many of the virulence-associated LTRTs found in these bacteria regulate metabolism and virulence factors concomitantly. Of the 38 LTRTs mutated, only two exhibited an intestinal colonization defect: that with a mutation in the methionine biosynthesis regulator MetR and that with a mutation in a regulator encoded by the locus CV1947, which exhibited a mild in vivo defect that was considered significant, although to a much lesser extent than the MetR defect. Upon binding of homocysteine at the N terminus, MetR becomes activated, resulting in transcriptional activation of methionine metabolism genes and negative feedback autorepression of itself (34–36). In pathogenic *E. coli* and *Salmonella enterica* serovar Typhimurium, MetR and the S-adenosylmethionine (SAM)-dependent repressor MetJ are key regulators of a complex metabolite feedback network that maintains proper intracellular methionine levels (36–41); however, regulation of methionine metabolism in *V. cholerae* remains less well understood.

Methionine metabolism is one facet of a highly interconnected and interdependent network of amino acid metabolism (Fig. 5). The biosynthesis of methionine involves two essential pathways: the homocysteine biosynthesis pathway, containing *metA*, *metB*, and *metC*, and the folate pathway, containing *glyA1* and *metF*.
The pathways converge at the gene for methionine synthase, **metH** (cobalamin dependent) or **metE** (cobalamin independent), whose product catalyzes the transfer of a methyl group from 5-methyltetrahydrofolate to homocysteine, yielding L-methionine and tetrahydrofolate (40, 42). The work presented here suggests that the SAM-dependent repressor MetJ and serine hydroxymethyltransferase GlyA1 are required for intestinal colonization, while the folate and homocysteine branches of methionine biosynthesis are dispensable in vivo. In agreement with this, a previous study using signature-tagged mutagenesis in *V. cholerae* also identified **glyA1** as being required for colonization of the infant mouse (43). In *E. coli*, MetJ functions as a repressor of methionine biosynthesis genes, including **metR** (41), while MetR is known to activate **glyA1** in a homocysteine-dependent manner (44, 45). Misregulation of **metR** or **glyA1** in the **metJ** repressor mutant may explain its in vivo colonization defect. Likewise, misregulation of genes required for virulence, such as **glyA1**, likely explains the colonization defects of the **metR** mutant. As part of the folate branch, GlyA1 catalyzes the

**FIG 3** Growth kinetics of wild-type *Vibrio cholerae* (red) and **metR** (green), **metJ** (yellow), and **glyA1** (blue) mutants in M9 minimal medium or M9 supplemented with 0.3 mM concentrations of the indicated amino acids.
reversible interconversion of serine to glycine while simultaneously regenerating $N^5,N^{10}$-methylene tetrahydrofolate (46, 47). The regeneration of methylated tetrahydrofolate is critical not only for the synthesis of methionine but also for a wide array of intracellular methylation events (42, 46, 48). However, the wild-type trimethoprim sensitivity of the $\text{glyA1}$ mutant indicates that GlyA1 does not play a critical role in the generation of one-carbon (C1) units, unlike in $\text{E. coli}$ (49). Taken together, these results strongly suggest that the production of glycine, rather than methionine or methylated tetrahydrofolate, is required for colonization of the infant mouse intestine by $\text{V. cholerae}$.

ChIP analysis of MetR in $\text{V. cholerae}$ validated its binding capacity to the promoters of genes involved in methionine biosynthesis. As in other bacteria, the loci for metR and metE are oriented divergently (23), with an ~400-bp intergenic region. Bioinformatic analysis of this intergenic region with the Multiple Expect Maximization for Motif Elicitation (MEME) tool identified a common, but not identical, binding motif that occurs twice (50). The motifs are oriented correctly for divergent transcription of metR and metE and are separated by ~100 bp. The qPCR primers used in the MetR ChIP analysis were designed to amplify the promoter region (~150 to 200 bp) just upstream of the coding region for each locus; therefore, the fact that metE possesses half the enrichment of metR indicates differential binding to each promoter. No enrichment for the metH promoter was observed; however, the relationship between MetR and its metabolite activator homocysteine is complex and varies greatly with nutrient conditions. In addition to metE and metR, we confirmed that MetR bound the promoter region of glyA1 equally well as that of metE. Subsequent qRT-PCR analysis supported the ChIP results and validated the regulation of metE and glyA1 by MetR. The regulatory role of MetR was found to be a repressor of glyA1 and metE under M9 medium growth and an activator of metE during growth in LB medium. These results are in contrast to the previously reported regulatory control by MetR for $\text{E. coli}$; however, regulation in

![Metabolic pathways of V. cholerae showing methionine, aspartate, threonine, glycine, serine, and cysteine biosynthesis. The metabolic transcriptional activator MetR and repressor MetJ are highlighted in green and red, respectively.](https://example.com/fig5)

**FIG 4** Relative enrichment of promoter targets in ChIP DNA from C6706(pMetR-3xV5) relative to C6706(pBAD) grown in LB medium with arabinose induction.

**FIG 5** Metabolic pathways of $\text{V. cholerae}$ showing methionine, aspartate, threonine, glycine, serine, and cysteine biosynthesis. The metabolic transcriptional activator MetR and repressor MetJ are highlighted in green and red, respectively.
**V. cholerae** may differ. Regulation of amino acid metabolism is carried out with a dynamic network or regulatory circuits with metabolic intermediates serving as coinducers for several transcriptional regulators. In order to maintain amino acid homeostasis, both MetR and MetJ are constantly in flux between induction states but typically reach a state of equilibrium. Another possible explanation for the difference in regulatory control of MetR between our results and those of previous literature is that overexpression of metR may result in abnormally high intracellular concentrations that sequester available homocysteine via binding to its coinducer domain. The result would be an artificial reduction in SAM and homocysteine accessibility and subsequently an excess of uninduced, unbound MetJ and/or MetR with unexpected regulatory consequences.

Nutritional opportunities within the host environment require specialized metabolism by the pathogen in order to access and utilize them. Furthermore, these nutritional opportunities may even drive evolutionary pressure to acquire and maintain genetic elements encoding metabolic factors deemed beneficial to survival, replication, or transmission (51). *Vibrio* pathogenicity island 2 (VPI2), encoding a neuraminidase and containing a cluster of genes involved in the scavenging, transport, and utilization of sialic acids, is required for metabolic activity that has been found to influence intestinal colonization (52, 53). While this specialized cluster of metabolic genes was horizontally acquired, other, less specialized metabolic genes from the ancestral genome may be considered to encode virulence factors as well. In this study, we found that glycine biosynthesis by GlyA1 is required by *V. cholerae* for successful colonization of the infant mouse intestine. Intriguingly, the essentiality of GlyA1 and no other related, interdependent, or interconnected metabolic enzymes suggests that glycine is limited in the host intestinal environment, at least relative to its demand by *V. cholerae* for *in vivo* biosynthetic activity or other needs.

Host-microbe metabolism within the mammalian intestine creates a gradient of nutritional niches that as a whole contain a wide diversity of nutrients (54). Interestingly, while most amino acids are rapidly catabolized by the small intestine for fuel, glycine is readily synthesized (55). However, the luminal availability of glycine remains to be determined. A recent *in vivo* transcriptome expression analysis of *V. cholerae* isolated from the rabbit or mouse intestine found no difference in expression of glyA1 compared to that in the transcriptional profile of LB-grown *V. cholerae* (56). Additionally, in comparisons of expression analyses between conditions (*in vivo* sample versus LB culture; *in vivo* sample versus M9 minimal medium culture), the expression levels of glyA1 in LB-grown and *in vivo*-isolated *V. cholerae* were similar, while that of M9-cultured *V. cholerae* was approximately 10-fold greater. While glyA1 was highly expressed under all conditions, the reduced expression of glyA1 in *in vivo* relative to that in minimal medium suggests adequate glycine availability in the host intestinal environment. This raises the possibility that glycine biosynthesis may play additional roles in virulence besides fulfillment of nutritional requirements. Bacteria are known to export amino acids to the extracellular environment via specific transporters (57, 58), and excretion of glycine by *E. coli* has been shown to elicit a quorum-based social behavior (59). The chemoreceptors Tar and Tsr sense serine and glycine or aspartate, respectively, and the bacteria respond collectively by congestion or dispersion depending upon temperature (60). Interestingly, several amino acids, including glycine, have been found to bind to chemoreceptors in *V. cholerae* and serve as strong chemotactic attractants (61). One of these chemoreceptors, Mlp24 (McpX), is required for expression of ctxAB in the infant mouse model; however, the underlying mechanism of genetic control is unknown (62). It is interesting to speculate that *V. cholerae* may possess a glycine excretion and response system similar to *E. coli* that coordinates chemotactic responses required for virulence. The dependency of glycine biosynthesis for host colonization by *V. cholerae* El Tor highlights the importance of understanding the *in vivo* metabolism of pathogens.

**MATERIALS AND METHODS**

**Ethics statement.** The animal experiments were performed according to protocols approved by Harvard Medical School Office for Research Protection Standing Committee on Animals. The Harvard Medical School animal management program is accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care, International (AAALAC) and meets National Institutes of Health standards as set forth in the Guide for the Care and Use of Laboratory Animals (63). The institution also accepts as mandatory the PHS Policy on Humane Care and Use of Laboratory Animals by Awardee Institutions and NIH Principles for the Utilization and Care of Vertebrate Animals Used in Testing, Research, and Training. There is on file with the Office of Laboratory Animal Welfare (OLAW) an approved Assurance of Compliance (A3431-01).

**Bacterial strains.** Strains and plasmids are listed in Table S3A in the supplemental material. *V. cholerae* El Tor biotype strain C6706 and a spontaneous lacZ derivative of C6706 were used as parental (wild-type [WT]) strains. *E. coli* DH5α *Apr* was used for cloning. Antibiotics used were streptomycin (Sm; 50 μg/ml), kanamycin (Kan; 50 μg/ml), trimethoprim (Tmp; ~39 ng/ml to 5 μg/ml), and chloramphenicol (Cm; 2.5 μg/ml for C6706 and 10 μg/ml for *E. coli* DH5α *Apr*). LB was used for normal growth conditions [10 g/liter of tryptone (Bacto), 5 g/liter of NaCl, and 2.5 g/liter of yeast extract (Bacto), and 5 g/liter of NaCl and was supplemented with 16 g/liter of agar (Bacto) for growth on plates. Filter-sterilized M9 minimal medium was used for auxotrophy determination assays [1.28 g/liter M9 minimal salts (prepared as instructed by the manufacturer; Sigma), 20 mM glucose, 2 mM MgSO₄, and 0.1 mM CaCl₂]. Arabidopsis was used at a final concentration of 0.02% for ChIP and metR expression experiments. X-Gal (5-bromo-4-chloro-3-indolyl-β-d-galactopyranoside) was used at 40 mg/ml.

**DNA manipulations and cloning.** To construct pMetR or pMetR-3xV5, the LysR-type transcriptional regulator metR was PCR amplified (NEB Phusion HF Master Mix) from *V. cholerae* genomic DNA and cloned into plasmid pBAD18 or pBAD18 carrying a C-terminal 3x V5 epitope tag, respectively, after digestion with KpnI and Sall (NEB). All cloned products were sequence verified (Dana Farber). Primers are listed in Table S3B in the supplemental material. The vectors pMetR and pMetR-3xV5 were then electroporated using standard techniques into *V. cholerae* C6706 *metR:*Tn to generate C6706(pMetR) and C6706(pMetR-3xV5), respectively.

**Suckling mouse colonization competition assays.** A modified version of the protocol described by Sigel et al. (22) was performed for infection and recovery of C6706-derived strains. C6706, C6706 lac2, and transposon mutant strains were grown on LB agar plates with Sm overnight at 37°C. Wild-type and mutant strains (10 μl each) were mixed together in 1 ml LB. Fifty microliters of this competition mixture (~50,000 bacteria) was inoculated into a 5-day-old CD1 mouse pup (Charles River Company). Serial dilutions of the competition mixture were plated on LB containing Sm (LB + Sm) and enumerated to determine the input ratio of wild-type and mutant strains. After incubation at 30°C for 12 h, the mouse pups were sacrificed, and small intestines were removed and homogenized in 10 ml of LB. Serial dilutions were plated on LB + Sm and enumerated to determine the output ratio of wild-type and mutant strains. The competitive index for each mutant is defined as the input ratio of...
of mutant to wild-type strains divided by the output ratio of mutant to wild-type strains. On average, four mice were assayed for each mutant strain. The results of in vivo experiments for the transposon strains were the cumulative results of experiments performed on different days. The average competitive index was reported for ease of communication.

**Growth in minimal medium with or without amino acid supplementation.** Overnight cultures were diluted (1:1000) in LB and grown to log phase at 30°C with aeration. One milliliter of culture was pelleted by centrifugation, washed 3× in M9 minimal medium, and then diluted to an optical density at 600 nm (OD$_{600}$) of 0.1 in M9 minimal medium with or without supplementation with 0.3 mM methionine, aspartate, threonine, glycine, or serine. Aurotrophix was determined by the absence or presence of growth. Growth kinetics of strains in LB or M9 minimal medium with or without amino acids was determined by OD$_{600}$ measurement using a 96-well-plate-format spectrophotometer (Spectramax Plus 384; Molecular Devices). Environmental parameters were set to 30°C with aeration, and readings were taken every 15 min for 10 h. Studies were conducted in triplicate.

**Trimethoprim sensitivity assay.** Overnight cultures were diluted (1:1000) in LB and grown to log phase at 30°C with aeration. Cultures were diluted to an OD$_{600}$ of 0.1 in fresh LB containing twofold serially diluted trimethoprim (1 μg/μl). Sensitivity to trimethoprim, as determined by growth kinetics of mutant strains relative to WT, was determined by OD$_{600}$ measurement using a 96-well-plate-format spectrophotometer (Spectramax Plus 384; Molecular Devices). Environmental parameters were set to 30°C with aeration, and readings were taken every 15 min for 10 h. Studies were conducted in triplicate. Trimethoprim MIC determination was conducted by washing cells and diluting them (1:100) in 5 ml LB with twofold serial dilutions of trimethoprim. Cultures were grown at 30°C with aeration and monitored for the presence or absence of growth, as indicated by turbidity, after 36 h.

**Chromatin immunoprecipitation (ChIP).** ChIP was carried out as previously described (11, 28); however, changes were made to optimize the protocol. Fifty milliliters of exponentially growing culture [C6706(pMetR-3xV5)] in LB was incubated with 0.02% arabinose for 30 min at 37°C with aeration. Formaldehyde (37%, Sigma) was added to a final concentration of 1% and incubated at RT for 20 min with occasional swirling. Cross-linking was quenched by adding glycine to 0.5 M. Cell pellets were washed in 1× Tris-buffered saline (TBS), resuspended in lysis buffer [10 mM Tris (pH 8.0), 100 mM NaCl, 1 mM EDTA, 0.5 mM EGTA, 0.1% deoxycholate (DOC), 0.5% N-lauroylsarcosine] plus bacterial protease inhibitor cocktail (Sigma P8845) and 50,000 U Ready-Lyse lysozyme (Epicentre), and then incubated at 30°C for 30 min. The cells were subjected to nucleotide sonication for 60 s and unlysed debris was removed by centrifugation. To shear DNA, the lysate was sonicated for 10 min with a 10-s on/10-s off cycle (MISONIX). A sample was taken as a sequencing input control. Following clarification by centrifugation, 1/10 volume of 10% Triton X-100 in lysis buffer was added to each sample followed by 50 μl of Dylight-Protein G beads coated with anti-V5 monoclonal antibody (Sigma), and samples were incubated overnight at 4°C with rotation. The beads were washed five times with radioimmunoprecipitation assay (RIPA) buffer [50 mM HEPES (pH 7.5), 500 mM LiCl, 1 mM EDTA, 1% Nonidet P-40, 0.7% deoxycholate (DOC)] and once with Tris-EDTA buffer [50 mM Tris-Cl (pH 8.0) plus 50 mM NaCl and then resuspended in 100 μl elution buffer [50 mM Tris-HCl (pH 7.5), 10 mM EDTA, 1% sodium dodecyl sulfate (SDS)]]. Samples were incubated at 65°C for 30 min, and then the beads were removed by magnet. Supernatants were incubated at 65°C overnight to reverse cross-links. Each sample’s volume was increased by the addition of 100 μl Tris-EDTA (pH 8.0), and then samples were incubated with 8 μl of 10 mg/ml RNase A for 2 h at 37°C. Samples were then treated with 4 μl of 20 mg/ml proteinase K at 55°C for 2 h, purified with a MinElute reaction cleanup kit (Qiagen), eluted into 50 μl, and quantified with a Pico green kit (Invitrogen). Experiments were performed in triplicate.

**Quantitative PCR and RT-PCR.** For ChIP-Seq peak validation, relative-abundance quantitative PCR (qPCR) was performed with Kapa Biosystems Fast SYBR green Master Mix using 16S rRNA genes and 3cpA targets as controls. Relative target levels were calculated using the $\Delta \Delta C_T$ method, with normalization of ChIP targets to the 16S RNA gene signal (64). For gene expression analysis, total RNA was extracted using a RiboPure kit (Invitrogen), contaminating was DNA removed using a Turbo DNA-free kit (Ambion), quantification was conducted on a NanoDrop spectrophotometer (Thermo Scientific), and abundance was normalized among samples. Relative-expression qRT-PCR was performed with a Power SYBR green RNA-to-C$_T$ one-step kit (Applied Biosystems). Relative expression levels were calculated using the $\Delta \Delta C_T$ method with normalization of gene targets to the 16S RNA signal (64). For qRT-PCR experiments, cells of C6706(pMetR) were grown to log phase in 5 ml of LB medium at 37°C, washed three times in M9 minimal medium, resuspended in 5 ml of M9 medium, incubated at 37°C for 1 h with aeration, and then induced with arabinose for 30 min at 37°C, and RNA was extracted. Primers used are listed in Table S3B in the supplemental material.

**SUPPLEMENTAL MATERIAL**

Supplemental material for this article may be found at http://mbio.asm.orglookup/suppl/doi:10.1128/mbio.00236-12/-/DCSupplemental.

Figure S1, TIF file, 10.7 MB.
Figure S2, TIF file, 2.6 MB.
Table S1, XLSX file, 0.1 MB.
Table S2, XLSX file, 0.1 MB.
Table S3, XLSX file, 0.1 MB.

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**REFERENCES**

14. Champion GA, Neely MN, Brennan MA, DiRita VJ. 2003. A branch in...


34. LaMonte BL, Hughes JA. 2006. *In vivo* hydrolysis of S-adenosylmethionine induces the met regulon of *Escherichia coli*. Microbiology 152:1451–1459.


