R5-SHIV Induces Multiple Defects in T Cell Function during Early Infection of Rhesus Macaques Including Accumulation of T Reg Cells in Lymph Nodes

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R5-SHIV Induces Multiple Defects in T Cell Function during Early Infection of Rhesus Macaques Including Accumulation of T Reg Cells in Lymph Nodes

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Abstract

**Background:** HIV-1 is a pathogen that T cell responses fail to control. HIV-1gp120 is the surface viral envelope glycoprotein that interacts with CD4 T cells and mediates entry. HIV-1gp120 has been implicated in immune dysregulatory functions that may limit anti-HIV antigen-specific T cell responses. We hypothesized that in the context of early SHIV infection, immune dysregulation of antigen-specific T-effector cell and regulatory functions would be detectable and that these would be associated or correlated with measurable concentrations of HIV-1gp120 in lymphoid tissues.

**Methods:** Rhesus macaques were intravaginally inoculated with a Clade C CCR5-tropic simian-human immunodeficiency virus, SHIV-1157ipd3N4. HIV-1gp120 levels, antigen-specificity, levels of apoptosis/anergy and frequency and function of Tregs were examined in lymph node and blood derived T cells at 5 and 12 weeks post inoculation.

**Results/Conclusions:** We observed reduced responses to Gag in CD4 and gp120 in CD8 lymph node-derived T cells compared to the peripheral blood at 5 weeks post-inoculation. Reduced antigen-specific responses were associated with higher levels of PD-1 on lymph node-derived CD4 T cells as compared to peripheral blood and uninfected lymph node-derived CD4 T cells. Lymph nodes contained increased numbers of Tregs as compared to peripheral blood, which positively correlated with gp120 levels; T regulatory cell depletion restored CD8 T cell responses to Gag but not to gp120. HIV gp120 was also able to induce T regulatory cell chemotaxis in a dose-dependent, CCR5-mediated manner. These studies contribute to our broader understanding of the ways in which HIV-1 dysregulates T cell function and localization during early infection.

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Introduction

It is increasingly clear that a single HIV virion or a small founder virus population infects a target cell in the mucosa and initiates a localized infection that is followed by systemic spread of the virus [1,2,3,4]. Early HIV infection is characterized by a reduction of CD4 T cells and may limit anti-HIV antigen-specific T cell responses. We hypothesized that in the context of early SHIV infection, immune dysregulation of antigen-specific T-effector cell and regulatory functions would be detectable and that these would be associated or correlated with measurable concentrations of HIV-1gp120 in lymphoid tissues. HIV proteins, including the surface envelope glycoprotein gp120, perform critical functions during the viral life cycle as well as playing a direct role in the immune pathogenesis of HIV/AIDS. For example, it has been shown that HIV-1 gp120 can mediate both viral entry and dysregulate immune cell function through its well-described interaction with cellular receptors, including CD4 and the chemokine co-receptors, CXCR4 and CCR5 (for X4 and R5 tropic viruses, respectively). HIV gp120 has been extensively studied *in vitro* with respect to its effects on a variety of both stromal and immune cell types [10,11,12]. HIV gp120 can impair dendritic cell maturation or induce dendritic cells to become immunosuppressive [12,13,14]. HIV gp120 has been shown *in vivo* to dysregulate T cell functions, including TCR desensitization, interference with co-stimulation, induction of apoptosis, and T cell migration [15,16,17,18,19,20,21]. The impairment of these
functions may have a dramatic effect on the formation and stability of effective anti-HIV immunity beyond the characteristic depletion of CD4 T cells seen in HIV/AIDS.

In vitro, gp120-induced immune dysfunction is more difficult to quantify, although human studies have consistently shown that gp120 is poorly targeted by both humoral and cellular immune responses [22,23,24,25]. In addition, there is a growing body of evidence to support the view that levels of gp120 required for immune dysregulation in vivo are present in lymphoid tissues in vivo [26]. Accumulation of high levels of gp120 in lymphoid organs of early SHIV-infected rhesus monkeys (RM) have been demonstrated [27], and high levels of the envelope protein have been found in the lymphoid tissues of HIV-1-infected humans with very low or undetectable viral loads [28,29]. These data support the view that the earliest events after primary infection, including exposure of immune cells to high local or systemic levels of gp120, play a major role in the long-term outcome of infection [1,3,30]. Taken together, these studies support the view that HIV-1 gp120 may play a role in immune dysfunction in lymphoid tissues as infection progresses.

We examined the hypothesis that a biologically relevant R5 HIV-1 gp120, in the context of early mucosal SHIV challenge, results in multimodal dysregulation of T cell-mediated immune function that in some cases, are associated with persistently high levels of gp120 in lymphoid tissues. Mucosal R5-SHIV inoculation of RM was chosen as a model system because 90 percent of all HIV infections among humans involve mucosal exposure to an R5-tropic virus. In this non-human primate study, we demonstrate early immune dysregulation of CD8 gp120-specific and CD4 Gag-specific T cells in the lymph nodes (LN), as compared to the peripheral blood. We found no enhancement of antigen-induced T cell proliferation in lymphoid tissues in vivo [16,17]. Accumulation of early HIV-1 gp120 and SIV p27 in lymphoid tissues and peripheral blood (PB) was associated with HIV-1 infection, including axillary LN were sampled at 5 weeks and 12 weeks post-inoculation. Some LN were frozen and stored at -80°C for later quantification of gp120. Mononuclear cells (MNC) were isolated from PB via Ficoll gradient enrichment as previously described [27]. MNC were obtained from the LN via physical disruption with cell lifters in a Petri dish and enriched by Ficoll gradient centrifugation. Samples were frozen at -80°C prior to use in vitro assays.

**Quantitation of HIV gp120 and SIV p27 in tissue**

LN biopsy specimens from infected and non-infected animals were thawed, weighed and suspended in 3.5 ml of lysis buffer (RIPA containing protease inhibitors), per gram. After being subjected to two freeze/thaw cycles, samples were spun at 16,000×g for 10 min and supernatants were frozen at −80°C prior to analysis as previously described [29]. Thawed samples were diluted in RPMI1640 containing 10% fetal bovine serum (FBS) and aliquots were assayed using the HIV-1 gp120 Antigen Capture Assay (Advanced Bioscience Laboratories, Inc., Kendall, MD) and a signal amplification kit ELAST ELISA Amplification (Perkin Elmer, San Jose, CA). Briefly, the modifications to the assay consisted of diluting the HIV-1 gp120 Antigen Capture Assay kit conjugate 1:1 with 1% BSA PBS-Tween 20 (0.05%) followed by 60 min incubation at 37°C. Next, 100 μl of Biotinyl Tyramide Solution (ELAST ELISA Amplification) was added to each well, followed by a 20 min incubation at room temperature. Detection consisted of adding 100 μl of diluted Streptavidin HRP Concentrate from the ELAST ELISA Amplification kit; 1000 fold with 1% BSA PBS-Tween 20 (0.05%) to each well and incubating the plate at room temperature for 30 min. The peroxidase substrate and the stop solution from the HIV-1 gp120 Antigen Capture Assay kit were used as described by the manufacturer. After stopping the reaction, absorption was measured at 450 nm as previously described [29]. SIV p27 was measured via a Retrotek SIV p27 antigen ELISA Amplification kit. The studies were conducted in accordance with National Institute of Health guidelines on the care and use of laboratory animals at the Yerkes National Primate Research Center (YNPRC), Emory University, Atlanta, GA, which is fully accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International. The Animal Care and Use Committee of YNPRC and DFCI approved all animal experiments.

**Materials and Methods**

**Ethics Statement**

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**Animals**

Six rhesus macaques (RM) of Indian origin were exposed intravaginally to various dilutions of a SHIV-1157ipd3N4 (R5-SHIV) stock grown in RM peripheral blood mononuclear cells (PBMC) (227 ng/ml of p24; 4×10^6 /ml) 50% tissue culture infectious doses (TCID50) as titrated in TZM-bl cells. This virus is exclusively R5-tropic [31,32]. Two animals that received the lowest mucosal dose (1:50) of SHIV did not seroconvert and were challenged subsequently with 1 ml of viral stock via the intravenous route (Table 1). Peripheral blood (PB) and peripheral nodes, including axillary LN were sampled at 5 weeks and 12 weeks post-inoculation. Some LN were frozen and stored at -80°C for later quantification of gp120. Mononuclear cells (MNC) were isolated from PB via Ficoll gradient enrichment as previously described [27]. MNC were obtained from the LN via physical disruption with cell lifters in a Petri dish and enriched by Ficoll gradient centrifugation. Samples were frozen at -80°C prior to use in vitro assays.

**Table 1. Levels of gp120 and p27 in the Plasma, Lymph Node and Spleen of RM 12 weeks post-inoculation with R5-SHIV.**

<table>
<thead>
<tr>
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<th>Tissue</th>
<th>gp120 (pg/ml)</th>
<th>p27 (pg/ml)</th>
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<tr>
<td>Rbo-6</td>
<td>Plasma</td>
<td>0.00</td>
<td>21.53</td>
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<td>LN</td>
<td>71.81</td>
<td>1832.04</td>
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<tr>
<td>RBw-8</td>
<td>Spleen</td>
<td>297.63</td>
<td>545.57</td>
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<tr>
<td></td>
<td>Plasma</td>
<td>0.00</td>
<td>62.82</td>
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<tr>
<td></td>
<td>LN</td>
<td>170.37</td>
<td>447.72</td>
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<tr>
<td></td>
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<td>278.16</td>
<td>603.89</td>
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<td>Rby-6</td>
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<td>0.00</td>
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<td>LN</td>
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<tr>
<td></td>
<td>Spleen</td>
<td>451.38</td>
<td>1064.68</td>
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LN: Lymph Node.

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Flow cytometry and intracellular cytokine staining (ICS)

PB and LN derived MNC were stimulated for 5 hours in the presence of an inhibitor of Golgi function (Golgi plug, BD
Biosciences, San Jose, CA) and anti-CD28 and anti-CD49d (clone CD28.2 and 9F10, BD Biosciences). Antigen-specific stimulation utilized 2 μg/ml of HIV clade C (kindly provided by Dr. Christian Brandt), or from SIVmac239 Gag (AIDS Research and Reference Reagent Program, Division of AIDS, NIAID, NIH, 15mers). Multi-parameter surface staining for CD3, CD4, CD8, CD25, CD127, (CD3 clone Sp34-2, CD4 clone L200, CD8 clone SK1, CD25 clone M-A251, CD127 clone hIL-7R-M21 BD Biosciences) and PD1 (clone EH12.2H7, Biologend, San Diego, CA) were carried out, followed by permeabilization and intracellular staining for IFN-γ or FoxP3 as per manufacturer’s instructions (IFN-γ clone B27 BD Biosciences, FoxP3 clone PHC101 eBiosciences). Analysis was completed using FlowJo Software (TreeStar, Ashland, OR) and gating was completed using a ‘fluorescence minus one’ (FMO) approach.

**Activation-Induced Cell Death (AICD)**

A 96 well round-bottom plate was coated with 1 μg of anti-CD3 monoclonal antibody (clone Sp34-2, BD Biosciences) overnight at 4°C. Previously frozen samples from PB and LN were cultured in the presence or absence of plate-bound anti-CD3 with the addition of anti-CD28, anti-CD49d for 5 hours at 37°C and 5% CO2.

**CD25 Depletion and CD4+CD25+ Isolation**

PB and LN derived MNC were depleted via a MACS CD25 depletion kit (Miltenyi Biotec, Auburn, CA) as per the manufacturer’s instructions. CD25 depletion was confirmed via flow cytometry and was 78±2.9%. Depleted or non-depleted cells were then stimulated and stained as described above to examine the role of CD25 cells on T-effector cell functions. For migration studies, human PBMC were obtained via a Ficoll spin and CD4+CD25+ T regulatory cells were isolated via a Human CD4+CD25+ MACS isolation kit (Miltenyi Biotec, Auburn, CA), as previously described [27]. Cell subpopulation purity was confirmed via flow cytometry to be greater than 80%.

**Transmigration Assay**

T cell migration was measured using Transwells (96-well format, 3-mm pore; ChemoTx System, Neuro Probe Inc, Gaithersburg, MA) as previously described [33]. Briefly, 7,000 CD4+CD25+ Tregs were loaded into the upper chamber, and 30 μl of medium alone or media supplemented with CCR5 tropic YU2 gp120 (ImmunoDiagnostics, Woburn, MA) was added to the lower or media supplemented with CCR5 tropic YU2 gp120 and when cells were exposed to media alone in upper and lower chamber at the concentrations of 500 pg/ml, 5 ng/ml, and 342.4±54.7 pg/ml, vs 12 week LN; p<0.05 non-paired T-test), in spite of the fact that half of the animals (3/6) had non-detectable viral loads in their PB (<1,300 copies/ml) [33]. Furthermore, we observed considerably lower levels of p27 than the expected Gag:p27 ratio would lead one to predict. Previous studies of the protein content of HIV viruses have demonstrated a ratio of p24 to gp120 to vary between 6:1 and 60:1 (table 1, 12 weeks post-inoculation). We have previously demonstrated that gp120 is found in the secondary lymphoid organs of RM at 12 weeks post mucosal R5-SHIV inoculation

We have previously demonstrated that gp120 is found in the secondary lymphoid organs of individuals with chronic HIV infection at levels that were disproportionate in comparison to both local p24 concentrations or plasma viral loads[29]. We sought to examine if during early mucosal R5-SHIV challenge, R5 gp120 would also be found at high levels in LN and spleen. At 12 weeks post-inoculation, we observed measurable amounts of gp120 in LN as compared to PB (Table 1, 139.7±34 pg/ml). Interestingly, at 12 weeks, we also observed significantly more gp120 in the spleen of infected RM than in the LN (342.4±54.7 pg/ml vs 12 week LN; p<0.05 non-paired T-test)

**Reduced Gag-specific CD4 and gp120 specific CD8 T cell responses in the LN of RM during early infection**

We examined responses of T cells derived from PB and LN upon stimulation with overlapping peptide pools of gp120 and Gag antigens at both 5 and 12 weeks post-inoculation. All animals were productively infected and had measurable viral loads at 5 weeks post inoculation. Half (3/6) of the RM subsequently had viral loads that were below the level of detection at 12 weeks post inoculation (Figure 1A). HIV-1gp120 specific IFN-γ+ CD4 T cells were lower in LN compared to PB, although this was not significant (Figure 1B &1C). However, the frequency of gp120-specific IFN-γ+ CD8 T cells was lower in LN than in PB at both 5 and 12 weeks post-inoculation (CD8 p<0.05, Figure 1D & 1E). Furthermore, in general gp120-specific CD4 and CD8 IFN-γ+ T cell responses in LN did not increase over time post R5-SHIV inoculation, whereas anti-gp120 CD4 and CD8 IFN-γ+ T cell responses in the PB increased over time between 5 and 12 weeks post-inoculation(Figure 1C & E). We also observed a different temporal pattern of Gag-specific responses. We observed a significant reduction with respect to Gag-specific CD4 T cells in LN as compared to PB at 5 weeks but not at 12 weeks post R5-SHIV inoculation (5 weeks: p<0.01, Figure 1F). Gag-specific CD4 and CD8 T cells increased over time regardless of anatomic location (Figure 1F & G). These data suggest that fewer gp120-specific T cells reside or are maintained in LN than in PB and that this effect is stable over time.

**Increased levels of Activation-Induced Cell Death (AICD) occur during early infection but are not associated with HIV-1 gp120 in lymph nodes**

Previous *in vitro* studies have demonstrated that gp120 can enhance T cell sensitivity to AICD and therefore we hypothesized...
that the relative depletion of anti-gp120 responses might be due to enhanced activation of LN-derived T cells [16,20,21,37]. We examined the ability of T cells to undergo AICD after polyclonal stimulation and compared the results between T cells derived from either PB or LN from naive RM or RM during early infection at 5 and 12 weeks post-inoculation (B-G). PB and LN lymphocytes were stimulated with overlapping clade C gp120 peptide pools at 5 and 12 weeks post inoculation and IFN-γ gp120-specific CD4 (B,C) and CD8 (D,E) T cell were assayed. F & G) PB and LN derived cells were stimulated with overlapping SIVmac239 Gag peptide pools at 5 and 12 weeks post inoculation and IFN-γ Gag-specific CD4 (F) and CD8 (G) T cell responses were assayed. Data representative of frequency of total (C,E-G) or frequency of parent (B,D) are shown. Error bars ± SEM, *p<0.05, ** p<0.01.

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immune dysregulation observed in LN of RM during early infection.

Enhanced expression of PD-1 in LN derived T cells from RM during early infection

Although, we observed no differences in apoptosis that were associated with reduced antigen-specific LN responses we proposed that T cell anergy may in part account for the observed immune dysregulation. Previous studies have demonstrated that during chronic HIV infection, levels of PD-1 are upregulated and correlated with anergic T cell responses to HIV antigens [38,39,40]. Furthermore, the blockade of PD-1 restores T cell function in animal models [41,42]. To further elucidate immune dysregulation in LN during early infection, we examined the level of PD-1 on T cells derived from PB and LN of naive and infected RM at 5 and 12 weeks post-R5-SHIV inoculation. We observed no major differences with respect to the proportion of PD-1 in any of the groups (data not shown). However, we also observed more PD-1 on CD4 T cells derived from LN at both 5 and 12 weeks post-inoculation as compared to PB-derived CD4 T cells (5 weeks p<0.005, 12 weeks p<0.05).

Figure 2. Enhanced basal T cell apoptosis from PB, but not LN of R5-SHIV-infected RM. Naive and R5-SHIV challenged RM were sampled at 5 and 12 weeks post-inoculation. PB and LN samples from either naive or 5 and 12 weeks post-inoculation samples were either non-stimulated (NS) or stimulated with plate-bound anti-CD3 (CD3) for 24 hours. A) Percent of apoptotic CD4 T cells from naive RM or infected RM at 5 or 12 weeks post inoculation comparing non-stimulated to CD3 stimulated is shown (naive PBMC non-stimulated vs naive PBMC stimulated p<0.005; naive PBMC non-stimulated vs 5 weeks infected PBMC non-stimulated p<0.005; naive PBMC non-stimulated vs 12 weeks infected non-stimulated p<0.05). B) Percent of apoptotic T cells from naive RM or infected RM at 5 or 12 weeks post inoculation comparing non-stimulated to CD3 stimulated, (naive PBMC non-stimulated vs naı¨ve PBMC stimulated p<0.005; naive PBMC, non-stimulated vs 5 weeks infected PBMC, non-stimulated p<0.05).

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phenomenon with respect to LN-derived CD8 T cells, where PD-1 receptor expression was upregulated at 5 weeks post-inoculation compared to cells from naïve RM (p < 0.005, Figure 3B). These data suggest that upregulation of PD-1 can be observed in LN-derived CD4 T cells of R5-SHIV challenged RM during early infection in LN and not in PB.

Increased frequency of CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>low</sup> T cells in LN of RM during early infection correlate with tissue levels of gp120

We hypothesized that lower frequencies of gp120-specific CD4 and CD8 T cells and to some extent Gag-specific CD4 and CD8 T cells seen in LN compared to PB, resulted from antigen-specific Treg suppression. To address this, we examined the proportion of CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>low</sup> Treg in PB and LN of SHIV challenged RM during early infection. We observed CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>low</sup> T cells in both PB and LN of RM during early infection (Figure 4A & B). We found consistently higher frequencies of CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>low</sup> T cells at 5 weeks post-inoculation in LN compared to PB (>85% of all CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>low</sup> T cells were FoxP3<sup>+</sup>) (5 weeks: p < 0.01, Figure 4C, E). In order to determine whether the observation of increased frequencies of CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>low</sup> T cells was associated with decreased frequencies of IFN-γ<sup>+</sup> T cells in LN, we examined the effect of Tregs on antigen-specific T cell production of IFN-γ. To accomplish this, we depleted 5 week post-inoculation PB and LN mononuclear cell samples of CD4<sup>+</sup>CD25<sup>+</sup> cells, where we observed significant differences in the frequencies of Treg and the greatest difference between IFN-γ producing, antigen-specific T cells (gp120 and Gag) from PB and LN. CD4<sup>+</sup>CD25<sup>+</sup> depletion reduced the level of this subpopulation by 78±8%. We observed a trend toward more Gag-stimulated CD8 T cells producing IFN-γ in the CD4<sup>+</sup>CD25<sup>+</sup>-depleted samples than in the non-depleted samples, as shown by the difference between depleted compared to non-depleted cells (p = 0.06, Figure 4F). Interestingly, gp120-stimulated CD8 T cells were not impacted by the depletion of CD4<sup>+</sup>CD25<sup>+</sup> T cells (data not shown), which is similar to observations in mouse models where the effects of Tregs are most potent on immunodominant epitopes [43,44,45]. Although Treg depletion had no impact on the restoration of CD4 T cell responses, these results may be difficult to interpret due to the fact that the Treg depletion step may have reduced the proportion of
RS-SHIV Causes Multiple Defects in T Cell Function

A

PBMC

LN

0.42%

1.95%

CD25

CD127

B

CD127 FMO

CD25 FMO

C

Frequency of CD4+CD25+CD127Low

D

Frequency of CD4+CD25+CD127Low

E

Frequency of CD4+CD25+CD127Low

F

Relative change of CD8+ T cells

G

Treg [% freq of total]

LN HIV-1gp120 [pg/ml]

PB

LN

0.2

0.4

0.6

0.8

1

5 Weeks 12 Weeks

5 Weeks 12 Weeks

gp120 stimulated Gag stimulated

p=0.06

0

0.1

0.2

0.3

0.4

0.5
effector T cells in our samples. Finally, we observed that increasing levels of CD4^+CD25^+CD127^low T cells in LN of RM at 5 weeks and 12 weeks post-inoculation positively correlated with the measured concentration of gp120 in lymphoid tissues (Figure 4G, \(r^2 = 0.7\), two-tailed \(p = 0.035\)).

Tregs migrate towards R5 gp120 in a CCR5 and G-protein coupled receptor manner

In view of the finding that LN levels of gp120 correlated with the number of Tregs in this tissue, we hypothesized that gp120 may play a direct role in Treg recruitment. To examine this, we investigated the migration of Tregs from human HIV naive donors in response to recombinant R5 HIV-1 gp120. We observed that Tregs migrated with increasing frequency to an R5 (YU2) gp120 in a dose-dependent manner (Figure 5A). Furthermore, the specific CCR5 antagonist, TAK-779, inhibited Treg migration demonstrating the dependence of this directional migration on CCR5. Additionally, pertussis toxin, an inhibitor of G-protein coupled receptor signaling, potently inhibited Treg migration toward R5 gp120 (Figure 5A). Finally, we observed that Tregs do not migrate away from R5 gp120 at any concentration of envelope protein that we examined (Figure 5B). These results suggest that recruitment of Tregs to lymphoid tissues during HIV infection may be in part due to the chemoattractant activity of R5 gp120 for this T cell subpopulation.

Discussion

In this study, we demonstrate a significant impact of early R5-SHIV infection on critical aspects of T cell function at and beyond the 5-week time point of maximal CD4 T cell depletion and viral load, including upregulation of PD-1 expression, differential suppression of gp120-specific T cell responses and preferential accumulation of Treg cells in lymphoid tissues. These immune dysregulatory effects are associated with high levels of the envelope protein (gp120) of HIV in lymphoid tissues when viral load is low or undetectable in PB (12 weeks). These data are consistent with a growing literature describing the effects of gp120 on T and B-cell function \[10,11,12,13,14,15,16,17,19,23\] and gp120-mediated dysregulation of immune cell function and localization \[22,27,29,46,47\].

Our results reveal consistent differences between the measurements of immune activation and regulation of PB versus LN-derived CD4 and CD8 T cells, regardless of route of infection (intravenous or mucosal). These differences are also evident despite the small sample size and the inclusion of two animals in this study that were challenged via a non-mucosal route and are consistent with a similar study performed in the dual-tropic SHIV-KB9 model using intravenous transmission [27]. In addition, these data imply that the anti-HIV immune response during early infection could easily be overestimated if the responses generated by circulating T cells was the only measurement made in this model.
context. The examination of PB T cell function as a surrogate marker of immune activation in lymphoid tissues in HIV has been the gold standard in both basic science and clinical trials. Interestingly, there is an increasingly consistent lack of correlation between PB T cell responses and immune protection in RM models [24,27,48,49]. Although immune cells residing in LN are more difficult to sample, they may provide a deeper understanding of the mechanisms used by the virus to subvert and evade host immune responses.

The presence of gp120 in LN of RM during early infection was shown to be associated with dysregulated IFN-γ responses of CD4 and CD8 T cells [27]. Previously, our laboratory demonstrated that the addition of exogenous gp120 to PB CD4 and CD8 T cells reduced HIV-specific IFN-γ responses to those levels observed in LN [27]. In the current study, increased levels of gp120 and the impaired IFN-γ response observed in LN were associated with increased levels of the T cell exhaustion marker, PD-1. Moreover, we observed enhanced basal apoptosis in PB of infected RM, suggesting that apoptosis may play a role in immune dysregulation, but this did not correlate with LN gp120 levels, PD-1 levels or impaired IFN-γ responses. The lack of correlation to apoptosis may be due to the fact that tissues have differential apoptotic rates during early infection [50]. Although HIV gp120 has been demonstrated to upregulate T cell death in vivo, we were unable to find a direct correlation to this parameter in vivo in this model [16,19,21].

Regulatory T cells have been demonstrated to have either a deleterious effect or no effect on HIV infection [51,52,53,54,55]. Here, we show that increased numbers of CD4⁺CD25⁺CD127low Treg correlate to the levels of gp120 in the LN at 12 weeks post infection and that the LN resident Tregs are in part responsible for the dampening of ex vivo CD8 IFN-γ responses against Gag antigen. Treg depletion resulted in a trend toward higher Gag-specific T cell responses. This finding carries the caveat that it is difficult to interpret the effect of Treg depletion on Gag and gp120-specific CD4 T cells as we deplete a population of CD4⁺CD25⁺ that may include T effector/memory cells [36]. These results support previous findings in mouse models, where immunodominant epitopes are preferentially targeted by Tregs [43,44,45]. Our data also suggest that other mechanisms, such as direct suppression via gp120, may play a role in immune dysregulation. In vitro studies suggest that gp120 itself can suppress the immune response independent of Treg [57]. Interestingly, a recent study by Becker et al. reported that a single dose of HIV-1 gp120 was able to antagonize granzyme B host disease in a mouse via the specific activation of Tregs [38]. Our in vitro data demonstrate that Tregs migrate toward R5 gp120 in a CCR5/Gαi-protein coupled receptor-dependent manner. We propose that the accumulation of Tregs in lymphoid tissue during acute R5-SHIV infection may be completely or partially driven by HIV-1 gp120 induced Treg cell chemotraction.

This study is the first demonstration of multimodal dysregulation of T cell function that occurs in vivo during early mucosal challenge with R5-SHIV. Furthermore, these data support the view that the persistence of HIV-1 gp120 in lymphoid tissues during early infection is associated with dysregulation of T cell function beyond CD4 T cell depletion that is emblematic of HIV/AIDS. Further examination of the effects of the virus and its envelope protein on HIV-1 antigen specific responses in lymphoid tissues in vivo at early time points following virus inoculation will assist in the broader understanding of the pathogenesis of HIV infection and those aspects of the disease which will need to be prevented or reversed by a vaccine or viral eradication approach.

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Author Contributions

Conceived and designed the experiments: MS MCP. Performed the experiments: MS ER PRI EDH HM BK NBS ALC LS. Analyzed the data: MS NBS. Contributed reagents/materials/analysis tools: RMR MG SLH. Wrote the paper: MS MCP. Discussions and editing: AHH NBS RMR.

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