A 600 kb Deletion Syndrome at 16p11.2 Leads to Energy Imbalance and Neuropsychiatric Disorders

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Accessibility
A 600 kb deletion syndrome at 16p11.2 leads to energy imbalance and neuropsychiatric disorders

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INTRODUCTION

The 16p11.2 locus (figure 1) encompasses several distinct genomic structural variants, including a recurrent interstitial deletion, of a ∼600 kb region containing 29 genes. 1 This deletion defined by breakpoints 4 and 5 (BP4-BP5) has a population prevalence of approximately 1/2000, 2 and reaches 0.5% in autism spectrum disorders (ASD). 2–7 It is one of the most frequent known single locus aetiologies of neurodevelopmental disorders and ASD. 8

We and others have demonstrated that this deletion predisposes to a highly penetrant form of obesity with a 43-fold increased risk of developing morbid obesity. 1 2 9 Increased head circumference (HC) has also been associated with the deletion. 2 10 A mirror phenotype is observed in carriers of the reciprocal duplication who present a high risk of being underweight and microcephalic. 1 2 10

The diversity of published clinical features, 10–14 together with the report of asymptomatic (but not fully evaluated) transmitting parents, 10 15 16 demonstrated the need to assess systematically the impact of the deletion on neurocognitive development and behaviour. 15 To identify the essential clinical traits accurately and assist with genetic counselling, through a collaborative effort we have collected the largest dataset of 16p11.2 BP4-BP5 ∼600 kb deletion carriers. It combines deletion carriers from both the 16p11.2 European and Simons Variation in Individuals Project (Simons VIP) consortia (n=116 and n=52, respectively). We present in this report the natural history, frequency, and range of phenotypes of this rearrangement. We demonstrate that the 16p11.2 deletion consistently impacts cognitive functioning, behaviour, growth, and body mass index (BMI).

PATIENTS AND METHODS

Patients

This study was reviewed and approved by the institutional review board of each site conducting the study. Signed consents were obtained from participants who underwent full assessments. For the data collected through questionnaires, information was gathered retrospectively and anonymously by
physicians who had ordered comparative genomic hybridisation (CGH) analyses performed for patient care purposes only. Consequently, research based informed consent was not required by the institutional review board of the University of Lausanne which granted an exemption for this part of the data collection. Carriers were ascertained through several cohorts (table 1). Details on ascertainment and data collection for the participants have been previously published. All participants in the Simons VIP cohort were mapped using whole genome oligonucleotide arrays, and were found to have the common, Table 1: Ascertainment of deletion carriers

<table>
<thead>
<tr>
<th>Cohorts‡</th>
<th>Mean age (y)</th>
<th>Carriers</th>
<th>M</th>
<th>F</th>
<th>Sex ratio p Value</th>
<th>Patients screened</th>
</tr>
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<tbody>
<tr>
<td>Europe†</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Probands</td>
<td>10.7</td>
<td>85**</td>
<td>56</td>
<td>28</td>
<td>1.49×10⁻³</td>
<td>30635**</td>
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<tr>
<td>Carriers siblings</td>
<td>13.9</td>
<td>9</td>
<td>6</td>
<td>3</td>
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<tr>
<td>Transmitting parents</td>
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<td>22</td>
<td>8</td>
<td>13</td>
<td>0.19</td>
<td>NA</td>
</tr>
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<td>Simons VIP</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Probands</td>
<td>8.2</td>
<td>45***</td>
<td>26</td>
<td>19</td>
<td>0.19</td>
<td>15749***</td>
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<tr>
<td>Carriers siblings</td>
<td>8.7</td>
<td>4</td>
<td>0</td>
<td>4</td>
<td>0.62</td>
<td>NA</td>
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<tr>
<td>Transmitting parents</td>
<td>42.7</td>
<td>3</td>
<td>2</td>
<td>1</td>
<td>–</td>
<td>NA</td>
</tr>
<tr>
<td>Literature</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>DD/ID§</td>
<td>9.8</td>
<td>84</td>
<td>56</td>
<td>28</td>
<td>1.49×10⁻³</td>
<td>5635§</td>
</tr>
<tr>
<td>General population†</td>
<td>45.1</td>
<td>18*</td>
<td>8</td>
<td>10</td>
<td>0.41</td>
<td>NA</td>
</tr>
<tr>
<td>Obesity‡</td>
<td>26.5</td>
<td>15</td>
<td>4</td>
<td>11</td>
<td>0.59</td>
<td>2579</td>
</tr>
<tr>
<td>Carriers TOTAL</td>
<td>–</td>
<td>285</td>
<td>166</td>
<td>117</td>
<td>2.12×10⁻³</td>
<td>107598</td>
</tr>
</tbody>
</table>

1Distribution of clinical indications for referral is detailed in the previous publication. Artikel of anthropometric data were available and previously published. Artikel of cases (including 7 relatives) reviewed from the literature. Artikel of the copy number variant detection methods used in the literature are: 19K bacterial artificial chromosome (BAC) microarray, Affymetrix 500K, Affymetrix 6.0, Illumina HumanHap300 BeadChip, 38K BAC microarray (Swegene), 44K, 105K and 244K Agilent, Affymetrix 500K, Affymetrix 6.0, Illumina 1M, BAC microarray, Affymetrix 6.0, Agilent 105K, NimbleGen HD2, Affymetrix 6.0, Affymetrix 500K or ROMA 85K, 244K Agilent or Affymetrix 6.0. Data were available on 18/25*, 85/113** and 45/67*** deletion carriers.

Due to missing data, there may be differences between the total number of cases and the sum of cases in the various columns. Statistically significant values are in bold.

Figure 1: The 16p11.2 locus. Highly homologous blocks of low copy repeats (LCRs) may act as substrates for non-allelic homologous recombination, predisposing to genomic disorders. Five LCRs have been defined as mediators of recurrent and clinically relevant imbalances within the 16p11.2 chromosomal band. To clarify the terminology, we propose to number these 'recombination hotspots’ from telomere to centromere as breakpoints BP1 to BP5. The current study describes only features associated with the proximal 600 kb recurrent deletion, delineated by BP4 and BP5 at genome sequence coordinates 29.5 and 30.1 Mb, respectively. Distal BP2-BP3 and BP1-BP3 mediated rearrangements, of respectively 220 and 550 kb, containing the SH2B1 gene, have also been reported in individuals with early onset obesity and variable degrees of developmental delay. Several recurrent rearrangements overlap the proximal BP4-BP5 region studied here including the 1.7 Mb deletions and duplications from BP1 to BP5 which should be considered as distinct entities. (A) Rearrangements are schematically pinpointed with reddish bars while grey bars and striated blocks indicate intervals of recurrent polymorphisms reported in the Database of Genomic Variants (http://projects.tcag.ca/variation) and common sequence stretches, respectively. (B) Genes encompassed by the genomic region between BP4 and BP5 are shown. All genomic positions are given according to the human genome build hg18/NCBI 36.

DD/ID, developmental disorders/intellectual disability; NA, non-available or not applicable; Simons VIP, Simons Variation in Individuals Project.
Copy-number variation

recurrent 16p11.2 BP4-BF5 deletion. Data for 96 carriers (76 probands and 20 relatives) from the European consortium were obtained by completion of a questionnaire by referring clinicians. Fifty-four probands and 18 relatives (n=72) were extensively evaluated on site by investigators of one of the consortia. Data on 117 deletion carriers ascertained for developmental disorders/intellectual disability (DD/ID) (n=84), obesity (n=15) and from the general population cohorts (n=18), were collected from our previously published studies, as well as from the literature.1 2 4 5 7 10 11 13–16 19–23

In the Simons VIP cohort, four patients were excluded based on the presence of additional pathological copy number variants (CNV), other genetic diagnoses, birth asphyxia, fetal alcohol syndrome, and/or prematurity <30 weeks. In the European series, known additional variants (three carriers with a CNV >500 kb and a pair of twins with an FGFR3 mutation) did not represent an exclusion criterion. The inclusion or exclusion of any of these five carriers did not have an impact on any results (supplementary table S1). They were included in the final analysis because of the arbitrariness of this filtering that only takes into account visible rearrangements and/or known point mutations. We have to assume that many additional mutational events not detectable by CGH array may be present in this dataset.

In the VIP cohort, ethnicity was 75% Caucasian, 5.8% African American, 1.9% Native American, 7.7% other (mixed ancestry), 9.6% unknown (adopted). For the 72 patients evaluated on site in the European cohort, all carriers were of European descent.

All available data on patients ascertained for DD/ID from the 16p11.2 European cohort, Simons VIP and the literature (n=285) were pooled for statistical analysis. Due to missing data for some phenotypes, the denominator changes, which is specified throughout the text. Data collected through questionnaires or by direct assessment are presented separately in supplementary tables S2 and S3. The 35 deletion carriers from the general population and obesity cohorts, for which only anthropometric data were available, are used in the analysis of growth parameters alone. Full-scale intelligence quotient (FSIQ) was assessed in 68 intrafamilial controls. The Simons Simplex Collection (SSC)24 was used as an ASD reference population to investigate the effect of obesity on the frequency of macrocephaly in patients with a neurodevelopmental disorder.

Cognitive functioning, psychiatric and behavioural assessment

Overall cognitive functioning was assessed using either the Mullen Scales of Early Learning, the Differential Ability Scales—Second Edition (Early Years and School Age, DAS-II), the Wechsler Intelligence Scales (WPPSI-III; WISC-IV; WAIS-III) or the Wechsler Abbreviated Scales of Intelligence, depending on the age and ability level of the individual.25 Vineland Adaptive Behaviour Scales (VABS)26 were used to measure adaptive skills in daily life. Intellectual disability (ID) was diagnosed as having an FSIQ of 70 or below on a standardised IQ test as well as concurrent deficits in adaptive functioning as defined by the Diagnostic and Statistical Manual of Mental Disorders, 4th edition, text revision (DSM-IV-TR) criteria.27

Different standardised neuropsychological measures were used to assess global cognitive functioning of carriers in the literature.5 11 15 16 25 For the purpose of this study, throughout the text we use the term FSIQ to refer to normalised cognitive levels (general population mean=100; standard deviation (SD)=15). ASD was diagnosed by experienced, research-reliable clinicians using the Autism Diagnostic Interview—Revised (ADI-R)28 and the Autism Diagnostic Observation Schedule (ADOS).29 The Diagnostic Interview for Genetic Studies (DIGS)30 was performed for adults, while children’s behavioural and emotional problems were assessed with parent report questionnaires.31–33 Additional DSM-IV-TR diagnoses were made by licensed psychologists and psychiatrists using history, parent report and in-person interview. For all aforementioned evaluations, the clinicians were not blinded to the genetic status of the participants.

Statistical analyses

To prevent bias due to intrafamilial correlations, deletion-carrying relatives of probands were excluded as required to avoid including more than one member of the same family in a single analysis. Obesity and morbid obesity in adults are defined throughout the study as BMI ≥30 and 40 kg/m², respectively. Z scores were computed for all data using gender, age, and geographically matched reference populations as previously described.1 Obesity in children and macrocephaly were defined as BMI and HC Z score ≥2, respectively.

One-tailed Fisher’s exact test was used to compare frequencies of the deletion in patients and controls. Two-tailed Student’s t test was performed to assess whether BMI, height, weight, and HC Z scores of deletion carriers were different from zero (general population mean). Correlation between these features and FSIQ were examined using Spearman correlation. The binomial test was performed to test for gender bias.

A linear model was applied to correct HC Z scores for BMI, age, and gender effects, using Matlab function regress. A two-tailed Student t test was performed to test the residual effects being different from zero. Linear mixed effect model was used to analyse the longitudinal and cross-sectional data and properly handle autocorrelations. Model fitting was performed to obtain the mean Z score and the corresponding p value for a given time window. The calculations were done using the lme function from the R package nlme.

For each of the W age windows the mean Z scores were computed for each of the P patients resulting in a W×P matrix. Since W tests were carried out, we applied a multiple testing correction that takes into account the correlation structure of the test statistics. As a first step, the effective number of tests (W_eff) was derived based on the Pearson’s correlation matrix of the W×P dataset.34 We then applied Bonferroni correction of the linear mixed effect model p values, but using W_eff instead of W tests to correct for multiple testing.

Breakpoint mapping with short arm of chromosome 16 custom array CGH

To confirm 16p11.2 deletions and ensure that the breakpoints are within the BP4 and BF5 low copy repeats (figure 1), we hybridised Cy5-labelled DNA of the European patients to custom made Nimblegen arrays. These arrays contained 71 000 probes spread across the short arm of chromosome 16 from 22.0 to 32.7 Mb (at a median space of 45 bp between 27.5 and 31.0 Mb) and 1000 control probes situated in invariable region of the X chromosome.3 Cy5-labelled DNA from the GM12042 CEPH (Centre d’Etude du Polymorphisme Humain) cell line was invariably used as reference. DNA labelling, hybridisation and washing were performed according to Nimblegen protocols. Scanning was performed using an Agilent G2565BA Microarray Scanner. Image processing, quality control, and data extraction were performed using the NimbleScan software V2.5.

RESULTS
The extent of the 16p11.2 rearrangements was assessed by aCGH through standard medical diagnostic procedures and confirmed by a high density custom made array as published (see Methods). Only carriers of the ∼600 kb 16p11.2 BP4-BP5 deletion were considered for further phenotyping; we gathered clinical information on 285 such carriers (figure 1). Cognitive functioning was evaluated in 71 carriers (mean FSIQ=76.1; SD=16.4; figure 2A) and was on average 32 points lower in de novo carriers when compared to their non-carrier family members (~−2 SD, p=3.96×10^−27; table 2A). There is a trend towards higher FSIQ in de novo (n=32) versus inherited (n=14) carriers (FSIQ 83 vs 74; p=0.13; table 2A and supplementary table S1). Of note, parents of de novo deletion carriers who directly enrol their child through a web based questionnaire (VIP cohort) have a higher IQ than the mean of the general population. Among carriers, 20% met DSM-IV-TR criteria for ID (65% mild FSIQ 55–70 and 35% moderate FSIQ 40–55). There are no differences in FSIQ between probands referred for neurodevelopmental disorders and carriers who were not medically ascertained (relatives who carry the deletion) (table 2, supplementary table S1). Carriers’ verbal IQ (mean=74; SD=17.5; range 25–107; n=42) is significantly lower than non-verbal IQ (mean=83.3; SD=18.0; range 47–160; n=45) (p=0.02). Of the 24 carriers evaluated in Europe, 20 (83%) had speech and language therapy during childhood.

The 16p11.2 BP4-BF5 deletion has been repeatedly associated with ASD, and is one of its most frequent known aetiologies. Of the fully assessed carriers, ∼15% (8/55) of the children and no adults met criteria for ASD by ADOS and ADI-R. More than 70% (51/70) of non-ASD carriers were found to have other DSM-IV-TR diagnoses including attention deficit and disruptive behaviour disorders, anxiety disorders, mood disorders, and substance related disorders (table 2B). There is a significant excess of males among carriers ascertained for neurodevelopmental disorders (138 M/75 F; p=2.4×10^−4) in contrast to other criteria (table 1). Gender, however, is not a significant covariate of FSIQ, adaptive level (Vineland), behavioural scores (Social Responsiveness Scale, SRS), neurological symptoms or any other trait (supplementary table S1).

### Table 2
<table>
<thead>
<tr>
<th>Carriers</th>
<th>Probandss</th>
<th>Relatives</th>
<th>Non-carriers</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Inherited</td>
<td>De novo</td>
<td>Unknown</td>
</tr>
<tr>
<td>Mean age (years)</td>
<td>14.7</td>
<td>10.9</td>
<td>15.7</td>
</tr>
<tr>
<td>Mean FSIQ</td>
<td>74</td>
<td>82.7</td>
<td>64.9</td>
</tr>
<tr>
<td>Number of cases</td>
<td>14</td>
<td>32*</td>
<td>14</td>
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### 16p11 Europe n (%)†

<table>
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<tr>
<th>DSM-IV-TR diagnosis</th>
<th>Children</th>
<th>Adults</th>
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<tr>
<td>ASD†</td>
<td>1 (12.5)</td>
<td>0</td>
</tr>
<tr>
<td>Any DSM-IV-TR diagnosis other than ASD</td>
<td>4 (50)§</td>
<td>6 (50)§</td>
</tr>
<tr>
<td>No diagnosis</td>
<td>3 (37.5)</td>
<td>6 (50)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Simons VIP n (%)‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>Children</td>
</tr>
<tr>
<td>7 (15)</td>
</tr>
<tr>
<td>39 (83)</td>
</tr>
<tr>
<td>1 (2)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Total cohorts n (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>8</td>
</tr>
<tr>
<td>47</td>
</tr>
<tr>
<td>70</td>
</tr>
</tbody>
</table>

*Two mosaic cases.
†10 probands and 10 relatives in the 16p11.2 European cohort and 44 probands and 6 relatives in Simons VIP.
‡ADOS and Autism Diagnostic Interview criteria
§Include attention deficit and disruptive behaviour disorders (n=4), anxiety disorders (n=5), mood disorders (n=3) and substance related disorders (n=2). Patients can have more than one diagnosis.

ADOS, Autism Diagnostic Observation Schedule; ASD, autism spectrum disorder; DSM-IV-TR, Diagnostic and Statistical Manual of Mental Disorders, 4th edition, text revision; FSIQ, full scale intelligence quotient.
We observed characteristic anthropometric patterns in deletion carriers. Birth weight is below average (n=124, Z score=-0.61; supplementary table S4), whereas Z scores for BMI are significantly higher by the age of 3.5 (Z score=1.01, p=0.05) (figure 3A). Longitudinal data show that this increase in BMI can be sudden and dramatic (figure 3D, supplementary figure S1). By the age of 7 years, obesity is a major comorbidity present in more than 50% of the carriers (supplementary figure S2). Excluding carriers ascertained for obesity, the frequency reaches 75% in adults (36/48), and among all adult obese patients 45% are morbidly obese (figure 2B). The variance of BMI in deletion carriers is higher than in the general population (p=2.06×10^-5). Hyperphagia was recorded (through parental questionnaires or reports) in all carriers (n=14) with obesity examined at the European site. The most severe cases require strict control of food access through locked cupboards and refrigerator.

BMI correlates neither with FSIQ (supplementary table S5) nor behavioural or adaptive scores (SRS, VABS). Furthermore, it is independent of ascertainment methods, inheritance, gender, or the presence of neurological features (supplementary table S1). Childhood obesity is, however, often associated with accelerated prepubertal linear growth, although the mechanism underlying this phenomenon remains unclear.\(^{25}\) In obese deletion carriers, a similar association is observed (figure 3A) in children whose height is increased (n=57, Z score=0.54, p=6.8×10^-5) and peaks at 9 years of age (Z score=1.08, p=1.5×10^-6). This is to be contrasted with short stature, which is apparent in probands (n=182, Z score=1.08, p=3.2×10^-8), newborns (n=88, Z score=-0.49, p=1.3×10^-6), non-obese children (n=91, Z score=-0.59, p=4.4×10^-6), and adults (n=32, Z score=1.1, p=8.0×10^-5) (figure 3A, supplementary tables S1, S4, supplementary figure S3).

An overall increase in HC is observed in probands (n=146, Z score=0.56, p=4.0×10^-4) with macrocephaly (HC Z score≥2) present in 29/170 (17%) of the carriers. HC correlates positively with BMI (r=0.45, p=2.8×10^-5) (supplementary table S5) and remains elevated after correcting for BMI, age, and gender (linear model; mean corrected Z score=0.60, p=1.1×10^-5). Obese deletion carriers show an increased frequency of macrocephaly when compared to non-obese carriers (32% and 10%, respectively; p=1.8×10^-5). Longitudinal data (figure 3B,C, supplementary figure S4) show that HC, which is lower at birth by 0.57 Z scores (n=58), increases during infancy (+1.74 Z scores, p=4.8×10^-4), several years before the elevation of BMI Z scores. In the Simon Simplex Collection of ASD children, 41.3% of patients with obesity have macrocephaly when compared to 12.5% in non-obese patients (p=5.4×10^-25) (supplementary figure S5).

Neurological features observed in patients referred for DD/ID include a wide range of findings (supplementary table S2). Gross motor delay was reported in 57.6% of the patients (32/58 ascertained for DD/ID in the European cohorts), which is consistent with previous series,\(^{10, 23, 37}\) and mean age of walking is significantly delayed (mean=20.5 months, SD=8.6, n=50, p=8.65×10^-5). Epilepsy is a frequent feature reported in 47/195 (24%) of the probands. While this frequency is not significantly different in non-medically ascertained carriers (5/38, 13%, p=0.2), it is higher in the fully assessed probands (22/54, 41%, p=1.6×10^-4). A small fraction of carriers (12/233) received a diagnosis of paroxysmal dyskinesia syndrome (OMIM 123200). MRI performed in a subset of carriers (n=65) showed mostly mild and non-recurrent features. Posterior fossa

Figure 3: Height, body mass index (BMI), and head circumference (HC) in 16p11.2 BP4-BP5 deletion carriers through development. Height (panel A), BMI (panel B) and HC (panel C) mean Z scores (and corresponding p values in red) for each age window were computed using a mixed effect model to analyse longitudinal and cross-sectional data together. p Values are derived from a two-sided t test of the fixed effects estimates probing whether they are significantly different from 0. Full red dots are p values surviving multiple testing correction (significance’s threshold at 6.3×10^-3 for height in both obese and non-obese, at 5.6×10^-3 for BMI, and at 7.1×10^-3 for HC) as opposed to empty red dots. Number of cases N is indicated for each age category. Panel A: Deletion carriers were classified in two groups; either the ‘obese group’ (squares) if they presented obesity at least once during their development, or the non-obese group (triangles). Height is significantly increased in prepubertal obese carriers while non-obese children remain slightly shorter than the general population. Panel B: BMI is significantly elevated by 3.5 years of age. Panel C: HC follows a rapid increase (+1.74 Z score, p=4.8×10^-4) during infancy, and remains high throughout life. Panel D: Longitudinal measures of BMI in a subset of 12 carriers illustrating different age onsets of BMI acceleration. The grey area specifies the interval between the 3rd and 97th centile as defined by the WHO data (http://www.who.int/childgrowth/en) between 0–2 years and the Centre for Disease Control and Prevention data above 2 years of age. The white line marks the 50th centile. All available longitudinal data are included in supplementary figure S2.
and/or craniocervical junction related abnormalities (eg, Chiari I malformation, cerebellar tonsillar ectopia, platybasia) were present in 19 of 41 fully reviewed MRIs.

Malformations or major medical problems are present in 76/130 (58%) of the probands from both cohorts, of which 55 have only a single pathological feature (supplementary tables S3 and S6). In non-medicallyascertained carriers identified in the families through cascade testing, this frequency is much lower (10/38, 26.3%, p=7.6×10^{-4}). No specific recurrent malformation sequence or multisystemic involvement is observed (supplementary table S6). There are, however, features likely to be associated with the deletion which, in most cases, do not require treatment such as vertebral abnormalities (hemivertebrae or kyphoscoliosis affect ~20% of carriers). Facial dysmorphism was visible in 69/150 (46%) carriers, yet no recurrent facial gestalt was observed in either cohort.

The frequency of this BP4-BPS deletion (table 1) among patients referred for clinical chromosome hybridisation microarray is similar for both consortia (0.28%). Though the two cohorts were assembled in different contexts and healthcare systems, as well as using different ascertainment criteria, they share striking similarities with the exception of the rate of inherited cases. Among Simons VIP patients, the deletion arose almost exclusively de novo (table 1). Globally, for participants with available parental data (51%), the deletion occurred de novo in 92/145 cases (including two mosaic cases) (64%) and was inherited in the remaining cases (36%). Assuming a stable prevalence of the deletion, this high rate of de novo events implies decreased fitness of this deletion (0.52 and 0.20 in the European and Simons VIP cohorts, respectively).

DISCUSSION

Our study demonstrates that the recurrent 600 kb BP4-BPS deletion, which has a prevalence of ~0.05% in the general population, has a consistent impact on traits affecting adaptive skills. FSIQ is decreased by ~2 SD in carriers of both genders, about 20% of them meeting criteria for ID. Since the FSIQ variance remains identical to that of the general population (p=0.25), approximately 30% of deletion carriers fall within the normal range (FSIQ ≥85). Bias in these estimates is unlikely since there were no differences in FSIQ scores of probands and non-medically ascertained carriers.

FSIQ does not capture the full range of disabilities experienced by deletion carriers. A history of speech therapy is frequent (83%) and psychiatric comorbidities affect >80% of carriers. The penetrance of ASD is 15% in our cohorts (table 2B), supporting the association of this deletion with ASD. Increased growth velocity of HC during infancy recapitulates the well documented pattern in idiopathic ASD. This shared head growth pattern, which has attracted considerable attention as a marker of abnormal brain development, has been linked to an increase in white matter volume, brain weight and numbers of neurons in the prefrontal cortex of ASD patients.

Obesity is a major comorbidity of 16p11.2 deletion carriers, with a penetrance among adults of >70%. Durable weight loss in adolescents or adults has not yet been documented: (1) two adults treated by bariatric surgery relapsed several years later; and (2) an adolescent dropped from 39.5 to 27.7 kg/m² by dieting and engaging in intense physical activity, but returned to his initial weight 2 years after discontinuing this regimen. Weight control is therefore recommended although no data are available on the efficacy of early intervention in deletion carriers. The reported association of ID with obesity has led to the proposal that impaired cognition may result in abnormal eating behaviour and obesity. In 16p11.2 deletion carriers, however, obesity occurs independently of cognitive function, adaptive or social behaviour scores.

We hypothesise that the deletion directly affects the neural circuitry involved in all these phenotypes, including energy balance. The early increase in HC precedes the onset of obesity (supplementary figure S4). This increase in HC is also observed later in childhood and adolescence (figure 3C) when correlation with brain volume is lower and contribution of skull thickness higher. Furthermore we demonstrate that childhood obesity in 16p11.2 deletion carriers, as well as in patients with autism (SSC cohort), is a confounding factor contributing to increased HC (supplementary figure S5). This deletion lowers final adult height by 1 SD (supplementary figure S3), though the well documented association between idiopathic childhood obesity and prepubertal height acceleration is maintained in obese children carrying the deletion (figure S3). In conclusion, our study demonstrates in two independent datasets that the 16p11.2 deletion should not be regarded primarily as a malformation syndrome. Indeed, the majority of these abnormalities are infrequent, suggesting either fortuitous associations or low penetrance (eg, coloboma/microphthalmia) possibly through unmasking of recessive mutations. Vertebral and spinal related anomalies (~20%, supplementary table S3) seem, however, to be strongly associated with the deletion, suggesting that spine x-ray and orthopaedic evaluations should be routinely performed in deletion carriers. 

TBX6, which maps within the interval, is a candidate gene for vertebral malformations since mice homozygous for a Tbx6 mutation showed rib and vertebral body abnormalities. Additionally, Tbx6 polymorphisms were associated with congenital scoliosis in the Han population.

In conclusion, our study demonstrates in two independent datasets that the 16p11.2 BP4-BPS 600 kb deletion consistently and quantitatively impacts cognitive functioning, HC, BMI, and growth. A range of behavioural disorders affects the vast majority of carriers. Probands referred for neurodevelopmental disorders or morbid obesity and carriers who were not medically ascertained (relatives of a proband) show similar values for FSIQ and BMI, suggesting that ‘asymptomatic’ carriers are uncommon. Five carriers within our cohorts presented an FSIQ >100 and may therefore represent the higher end tail of
the FSIQ distribution of deletion carriers. All five presented language disorders, autism, disruptive behaviour or obesity. The deleterious impact of the deletion is further highlighted by its low fitness reflected by the rarity of multigenerational carrier families. In contrast to BMI, the variance of global cognitive functioning (FSIQ) is the same among carriers and control population, suggesting that the factors determining its variability are identical to those at play in the general population and unrelated to the 16p11.2 locus.

This comprehensive study of the 16p11.2 BP4-BFP phenotype helps to guide clinical monitoring and counselling of patients and families and to potentially overcome the genetic counselling challenge posed by its variability. It illustrates that the study of rare variants causing common diseases lacking pathogenic mechanisms requires the assembly and detailed clinical characterisation of large cohorts, recruited using multiple ascertainment criteria.

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REFERENCES


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Corrections


Fassone E, Rahman S. Complex I deficiency: clinical features, biochemistry and molecular genetics. J Med Genet. 2012;49:578–90. In the above article updated figures were not included in the proof. These will be uploaded with the article as a data supplement.

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