A Homogeneous, High-Throughput Assay for Phosphatidylinositol 5-Phosphate 4-Kinase with a Novel, Rapid Substrate Preparation

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Abstract
Phosphoinositide kinases regulate diverse cellular functions and are important targets for therapeutic development for diseases, such as diabetes and cancer. Preparation of the lipid substrate is crucial for the development of a robust and miniaturizable lipid kinase assay. Enzymatic assays for phosphoinositide kinases often use lipid substrates prepared from lyophilized lipid preparations by sonication, which result in variability in the liposome size from preparation to preparation. Herein, we report a homogeneous 1536-well luciferase-coupled bioluminescence assay for PISP4Kz. The substrate preparation is novel and allows the rapid production of a DMSO-containing substrate solution without the need for lengthy liposome preparation protocols, thus enabling the scale-up of this traditionally difficult type of assay. The Z'-factor value was greater than 0.7 for the PISP4Kz assay, indicating its suitability for high-throughput screening applications. Tyrphostin AG-82 had been identified as an inhibitor of PISP4Kz by assessing the degree of phospho transfer of γ-32P-ATP to PISP; its inhibitory activity against PISP4Kz was confirmed in the present miniaturized assay. From a pilot screen of a library of bioactive compounds, another tyrphostin, I-Ome tyrphostin AG-538 (I-Ome-AG-538), was identified as an ATP-competitive inhibitor of PISP4Kz with an IC50 of 1 μM, affirming the suitability of the assay for inhibitor discovery campaigns. This homogeneous assay may apply to other lipid kinases and should help in the identification of leads for this class of enzymes by enabling high-throughput screening efforts.

Introduction
Phosphatidylinositol (PI) signaling has been shown to impact a variety of fundamental cellular processes, including intracellular membrane trafficking, cytoskeletal rearrangement, cell proliferation, survival and growth. Dysregulation of these pathways can lead to cancer and other diseases [1,2,3,4,5,6]. Phosphoinositides contain two fatty-acid chains linked through a diacylglycerol moiety and phosphodiester bond to an inositol headgroup. PIs are an important class of lipids that are regulated by reversible phosphorylation of the inositol headgroup. Phosphatidylinositols have three main phosphorylation sites on the inositol (positions 3, 4 and 5) that are regulated by different classes of phosphoinositide kinases and phosphatases. The three phosphatidylinositol mono-phosphates (PIPs) are PI3P, PI4P and PI5P. Importantly, these regioisomers have distinct roles in vivo, and there are three types of kinases (PIPKs) that distinguish and phosphorylate specific PIPs [7,8,9]. Type I PIP kinases (PI4P 3-kinases/PI4P5Ks) preferentially phosphorylate PI4P on the 5 position, Type III PIP kinases (PI5P 3-kinases/PI3P5Ks/Fab1/PIK5ye) preferentially phosphorylate PI3P on the 5 position, and Type II kinases (PI5P 4-kinases/PI5P4K), which are the focus of the present report, preferentially phosphorylate PI5P on the 4 position as shown in Figure 1 [10].

There has been a massive expansion in the investigation of the function and role of PI5K after the discovery of the phosphoinositide-3-kinase (PI3K) inhibitors wortmannin and LY294002 [11,12]. While PI5K is known to play a role in insulin signaling, megakaryocyte development, and Vitamin-D signaling [13,14,15,16], to date there are no commercially-available...
PI5P4K inhibitors, which hinders the advancement of this field. Recently, Demian et al. have demonstrated that selective inhibitors against PI5P4K can be obtained from screening [17]. However, potent PI5P4K inhibitors are still not available and such compounds would serve as valuable research tools to investigate the physiological role of PI5P4K activity.

The PI5P4Ks share very little sequence homology with protein or other lipid kinases, which may facilitate the design of selective inhibitors. PI5P4Kβ has been crystallographically characterized and shows a flattened lysine-containing basic patch that is expected to bind to the phospholipid headgroup on the surface of the lipid membrane [18,19]. PI5P4Kα and PI5P4Kγ also have crystal structures available (PDB ID:2YBX and 2GK9), and they show a similar overall structure. A binding assay and an enzymatic assay that utilized an elaborate liposome-based substrate preparation have been previously reported [17,10]. Additionally, low-throughput radiolabeled enzymatic thin layer chromatography (TLC) assays were developed where the substrate was prepared in an isotonic KCl solution or as a liposome mixture [20,21]. The radiometric assay uses γ-32P-ATP and PI5P and measures radiolabeled enzymatic product, PI(4,5)P2 after the separation by TLC [10,22,23]. There is, however, a need for additional assay types with simplified reproducible substrate preparations that are amenable to high-throughput screening.

We sought to establish a 1536-well compatible high-throughput enzymatic assay for PI5P4Kα to enable large multi-day library screens. A novel substrate preparation was developed that yielded a stable solution amenable to large-scale screening and that could be prepared reproducibly in house with common laboratory tools. Furthermore, the new assay was employed in a small library screen, resulting in the identification of an ATP-competitive tyrphostin (TYRosine PHOSphorylation INhibitors) inhibitor of PI5P4Kα (IC50 = 1 μM). This new high-throughput screening methodology should enable large library screening to help identify selective inhibitors of PI5P4Kα and related enzymes.

Materials and Methods

Reagents and Consumables

Hepes, pH 7.4 (4-[2-Hydroxyethyl]-1-piperazine ethanesulfonic acid) and MgCl2 were purchased from GIBCO (Carlsbad, CA, USA) and Quality Biological Inc. (Gaithersburg, MD, USA), respectively. EGTA, tyrphostin I-OMe-AG-538 and CHAPS (3-[3-Cholamidopropyl]dimethylammonio)propane-sulfonic acid) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Dimethyl sulfoxide (DMSO, certified ACS grade) was obtained from Thermo-Fisher Scientific (Pittsburg, PA, USA). Tyrphostin 25 (AG-82) was purchased from Cayman Chemical Company (Ann Arbor, MI, USA). Medium binding white solid-bottom 1536-well plates (assay plates), 1536-well polystyrene plates (compound plates) and 384-well white solid-bottom plates were purchased from Greiner Bio One (Monroe, NC, USA). The 384-well polypropylene V-bottom plates (compound storage) were from Matrix/Thermo Scientific (Hudson, NH, USA). Bioluminescent assay detection used the Promega ADP-Glo kit (Madison, WI, USA), which came with 10 μM Ultrapure ATP that was used for the assay. Per the manufacturer’s protocol, this kit can be used for reactions containing between 1 μM to 1 mM ATP, PI5P4Kα with a GST tag was expressed and purified as described previously [10]. A 0.4 mg/mL stock was stored at −80°C and used for this work.

Compound Library

The library of pharmacologically active compounds (LO-PAC1280, Sigma-Aldrich) contains 1280 known bioactives that were received as 10 mM DMSO solutions. The compound library was plated in 1536-well format with an Evolution P3 plate (Perkin-Elmer, Shelton, CT, USA). The library was formatted into columns 3–48 of 1536-well compound plates at 4 stock concentrations (1: 5 dilution, spanning the 10000 to 80 μM range) and 5 μL per well. Preparation of the compound library for quantitative high-throughput screening (qHTS) has been described previously [24]. A control plate was made in a compound plate in columns 1–4 using a Cybi-Well (CyBio, Jena, Germany) to transfer solutions from a 384 compound storage plate to the 1536 plate compound plate. Columns 1, 3, and 4 contained DMSO and column 2 contained a 1:2 serial dilution of the control compound tyrphostin AG-82 (16 points with N = 2 and starting concentration of 100 μM).

Lipid Preparation

DPPS (1,2-dipalmitoyl-sn-glycero-3-phosphoserine) and PI5P (D-myo-phosphatidylinositol 5-phosphate diC16) were purchased from Echelon Biosciences (Salt Lake City, UT, USA). DPPS was dissolved in DMSO and sonicated for one minute (using a sonicating water bath) and mixed by vortexing for 30 seconds, thereby forming a clear solution (333 μL DMSO per 1 mg DPPS). PI5P was suspended in DMSO and mixed by sonication and vortexing for several minutes (333 μL DMSO per 1 mg PI5P). A 2:1 ratio of DPPS to PI5P was then made (500 μL of PI5P, 1000 μL of DPPS plus an additional 1500 μL of DMSO), and the resulting lipid mixture was sonicated and mixed by vortexing for

![Figure 1. Schematic representation of the PI5P4K reaction using PI5P as the substrate.](image-url)
several minutes. The resulting solution can be stored at -20°C and was found to be stable for at least six cycles of freeze/thaw.

On the day of the experiment, the lipid mix was thawed and mixed by sonication and vortexing for one minute. For each step up until the enzyme addition, each reagent addition was followed by a brief sonication and vortex mixing step. First, 1259 µL of the lipid mix was added to 315 µL of DMSO. Then, 5827 µL of buffer 1 (30 mM Hepes pH 7.4, 1 mM EGTA, 0.1% CHAPS) was added and lastly, 13525 µL of buffer 2 (46 mM Hepes pH 7.4, 0.1% CHAPS) was added. This lipid reagent was used for the assay as described below. All lipid mixtures were prepared in glass vessels to minimize surface absorption.

Miniaturized Enzyme Activity Assay

P54K (32.4 µL) was added to the lipid reagent (3 mL, described above) to form the enzyme-lipid mixture. First, 2 µL of the enzyme/lipid substrate reagent (2 µg/mL P54K final concentration) were dispensed into a white solid-bottom 1536-well plate in columns 1–2, 4–48 using a Flying Reagent Dispenser (FRD, Beckman Coulter, Fullerton, CA, USA). Into columns 3 and 4, 2 µL of a no-lipid and no-enzyme control were dispensed, respectively, where the lipid was replaced by DMSO or the enzyme was replaced by buffer (20 mM Hepes 7.4, 0.1% CHAPS) in the enzyme-lipid mixture. Then, 25 µL of the enzyme/lipid mixture and control compounds were transferred by pintool into a 1536 pintool (Kalypsys Systems, San Diego, CA, USA) into wells 5–48 and 1–4, respectively. To initiate the reaction, 1 µL of the ATP solution (20 mM Hepes pH 7.4, 60 mM MgCl2, 0.015 mM ATP and 0.1% CHAPS) was added. The final concentration of DMSO in the reaction was 5%. The resulting mixture was incubated at room temperature in the dark for one hour, at which time 2 µL of ADP-Glo reagent 1 were added to stop the reaction and remove any remaining ATP. After a 45-minute incubation, 4 µL of the ADP-Glo reagent 2 were added and allowed to incubate for 30 minutes. The luminescence was then read with a ViewLux high-throughput CCD imager (Perkin Elmer, Waltham, MA, USA). A total of six plates were assayed: one DMSO plate for monitoring throughput CCD imager (Perkin Elmer, Waltham, MA, USA). A total of six plates were assayed: one DMSO plate for monitoring throughput CCD imager (Perkin Elmer, Waltham, MA, USA).

Data Analysis

Screening data were corrected and normalized, and concentration-response curves were derived using in-house algorithms [25]. Overall assay performance, including trends in Z’ factor, % CV, and S/B, were recorded. Percent activity was computed after normalization using the median values of the uninhibited enzyme control (32 wells in column 1) and the no-enzyme, or 100% inhibited, control (32 wells, column 4). Additionally, the compound structures were evaluated, and the long chain compounds such as the C18 containing compounds from LOPAC®1280 that are likely to act by disrupting the lipid vesicles were eliminated. Selected actives were procured for re-testing in the miniaturized ADP-Glo enzyme assay.

Substrate Competition Assay

Seven concentrations of ATP (concentration in final reaction was 0.25 Km, 0.5 Km, 1 Km, 1.5 Km, 3 Km, 5 Km, and 10 Km) were tested in the qHTS ADP-Glo P54K assay, the compounds were transferred for their effect on the detection reagent. A single kinase assay was available and has been previously established a 384-well enzymatic assay for P54K [10,22,23]. Overall assay performance, including trends in Z’ factor, % CV, and S/B, were recorded. Percent activity was computed after normalization using the median values of the uninhibited enzyme control (32 wells in column 1) and the no-enzyme, or 100% inhibited, control (32 wells, column 4). Additionally, the compound structures were evaluated, and the long chain compounds such as the C18 containing compounds from LOPAC®1280 that are likely to act by disrupting the lipid vesicles were eliminated. Selected actives were procured for re-testing in the miniaturized ADP-Glo enzyme assay.

Substrate Preparations

Preparation of the substrate is crucial for the development of a robust and miniaturizable lipid kinase assay. Phosphatidylinosine (DPPS = 1,2-dipalmitoyl-sn-glycero-3-phosphoinositol) is often used as a carrier for lipid substrates and was used in combination with P54K here. Phosphatidylinositol (P5P = D-myo-phosphatidylinositol 5-phosphate diC16) [17,22,27]. We aimed to make a DPPS/P5P lipid mix in which the reagents would be stable for at least 8 hours and would not be subject to settling or clumping during the repeat dispensing steps. The main challenge originated from the diverging solubility and substrate competency trends by the lipids under consideration: P5P and DPPS can have different alkyl chain lengths and the shorter ones (C4 and C8) are soluble in buffer but are not substrates for P54K [data not shown and [17]), while the longer C16 P54K substrates are not soluble in buffer but are soluble in DMSO.

Previously, a binding assay was developed with sucrose-loaded unilamellar vesicles that were then prepped by multiple freeze-thaw cycles followed by extrusion [18]. The study investigated a variety of lipid substrates and showed the importance of the negative charge for binding of the lipid to P54K. Demian et al. previously established a 384-well enzymatic assay for P54K and P54KB using a translucent liposome-based presentation of the substrate (2:1 ratio of phosphatidylinositol to P5P) [17]. They had used a liposome suspension in buffer for their C16 lipid substrate that was prepared by a time-consuming and technically challenging method that involved mixing the two lipids in acidified liposomal bilayers.

Countscreen Assays

After hit confirmation of the selected and reacquired compounds in an 11-point retest in the qHTS ADP-Glo P54K assay, the compounds were tested for their effect on the detection reagent. A single kinase assay was available and has been previously established a 384-well enzymatic assay for P54K [10,22,23]. Overall assay performance, including trends in Z’ factor, % CV, and S/B, were recorded. Percent activity was computed after normalization using the median values of the uninhibited enzyme control (32 wells in column 1) and the no-enzyme, or 100% inhibited, control (32 wells, column 4). Additionally, the compound structures were evaluated, and the long chain compounds such as the C18 containing compounds from LOPAC®1280 that are likely to act by disrupting the lipid vesicles were eliminated. Selected actives were procured for re-testing in the miniaturized ADP-Glo enzyme assay.
Design and Miniaturization of the High-throughput PI5P4Kα Enzyme Assay

To enable large-scale library screening, a 1536-well luciferase-coupled biofluorescence PI5P4Kα assay was pursued. Our goal was to design an enzymatic reaction whereby the enzyme, lipid substrate, and ATP could be dispensed in two, rather than three, distinct steps to minimize the overall variability of dispense, which in turn required the combination of two of the reagents into one vessel. Many kinases, including PI5P4Kα have low levels of substrate-independent consumption of ATP, which precluded the enzyme and ATP from being stored together. There was also a spontaneous phospho transfer from ATP (by a very small amount) to the lipid when these two reagents are stored together (data not shown). The stability of the enzyme/lipid solution was tested, and it was found that there was a less than 5% change in activity for a premixed solution maintained at 4°C for 16 hours versus a freshly prepared stock (see Figure 3A). Stability over 8 hours is ideal for the large robotic screens to allow for facile reagent exchanges to be scheduled in a continuous robotic run. This reagent configuration (enzyme/lipid followed by ATP) was used in the assay.

The $K_m$ of ATP (5–6 μM) had been determined previously [17,30], and the ATP concentration was set at the $K_m$ to ensure that the assay would be sensitive to ATP-competitive inhibitors. The substrate concentration was determined such that a reasonable % ATP conversion (<20%) and robust assay statistics could be obtained with a one-hour incubation time by 10 nM of PI5P4Kα (see Figure 3B and Figure 4). A PI5P concentration of 75 μM was determined to be optimal because 50 μM resulted in lower enzyme activity and 100 μM gave nearly the same results as 75 μM. The $K_m$ of this substrate is unknown and could not be measured here because the use of lipid concentrations much higher than 100 μM, required for the accurate derivation of $K_m$, would lead to a DMSO concentration that is not well tolerated by the enzyme (see following section). By having excess lipid relative to the $S_{0.5}$ (~50 μM), a slight loss of lipid sticking to the storage bottle, dispenser tubing or assay plate during the HTS can be tolerated without an appreciable change in assay performance. The two-step ADP-Glo kit was used to detect the ADP product, which allows for the development of a sensitive assay without the need for high levels of conversion [31]. The detailed assay protocol can be found in the Materials and Methods and is summarized in

![Figure 2. Schematic representation of the 2:1 DPPS:PI5P lipid preparation protocol.](doi:10.1371/journal.pone.0054127.g002)
Table 1. The signal to background of the PI5P4Kα assay was quite comparable when the substrate was prepared by the method of Demian et al. and the method described here (S/B = 12 -13, Figure S1) [17], indicating that the new lipid preparation could function as a substrate for PI5P4Kα. Additionally, the lipid mixture was found to be stable to at least six rounds of freeze/thaw (data not shown), which allows the substrate to be prepared and validated in large batches and used for multiple experiments.

DMSO Tolerance

The effect of DMSO on the enzyme reaction was tested, and it was found that up to 7.25% DMSO there was little effect on the enzyme activity and assay performance. At high DMSO levels, the concentration of DMSO used to make the enzyme-lipid reagent was found to impact the ratio of lipid-coupled kinase ATPase activity to lipid-independent kinase ATPase activity: 5% DMSO was found to have very little lipid-independent kinase ATPase activity (~4%) while allowing the lipid substrate to be readily accessible to the enzyme active site; however, at DMSO concentrations of greater than 15%, the amount of lipid-independent phosphorylation increased dramatically (See Figure S2). Therefore, DMSO at 5% was able to help solubilize the substrate while retaining the desired substrate-coupled enzyme activity, and this concentration of DMSO was used for the miniaturized luciferase-coupled assay.

Pilot Screen

Validation of the PI5P4Kα assay was performed by a qHTS (quantitative HTS, [25]) against the library of pharmacologically active compounds (LOPAC1280, Sigma-Aldrich) arrayed as a five-point titration series (152, 76, 15, 3, and 0.61 μM final compound concentration). Additionally, a vehicle only DMSO plate was run and used to monitor the background. The validation experiment showed excellent performance as measured by a stable signal:background ratio (12.6), stable Z' factor (0.77) and a low CV (9.3%) (Figure 4A–C). Tyrphostin AG-82 was identified as a weak inhibitor of PI5P4Kα (decreases the enzyme activity by 75%) by a radiometric assay that uses γ-32P-ATP and PIP and measures the radiolabeled enzymatic product, PI(4,5)P2, after the separation by thin layer chromatography. Five additional compounds were tested and found not to significantly inhibit PI5P4Kα (AG17 = tyrphostin AG-17, AG18 = tyrphostin AG-18, MP = mycophenolate, PB = purvalanol B and SU6668). All compounds were tested at 100 μM, except for PBV, which was tested at 10 μM due to solubility limitations at higher concentrations. The raw image and the extracted data are shown in (C) and (D), respectively. The commercial PI5P substrate predominantly contains two palmitate groups with a very small amount of deacylated lipid lyso-PI5P that contains only one palmitate group. The intense top spots in (C) represent the PI(4,5)P2 product and the faint spots below represent the product with just one palmitate group.

Figure 3. Lipid dependence, overnight stability and control compound. (A) The overnight (16 hour) stability of the assay reagents at 4°C when the enzyme and lipid were premixed, stored separately or made up fresh as compared to a no enzyme and 5 μM ADP (representing 0% and 100% conversion, respectively). The error bars represent the standard deviation (N = 2). (B) The PI5P lipid dependence of the PI5P4Kα enzyme reaction. The error bars represent the standard deviation (N = 2) and are not discernable on the plot. (C) and (D) Tyrphostin AG-82 (AG82) was identified as a weak inhibitor of PI5P4Kα (decreases the enzyme activity by 75%) by a radiometric assay that uses γ-32P-ATP and PIP and measures the radiolabeled enzymatic product, PI(4,5)P2, after the separation by thin layer chromatography. Five additional compounds were tested and found not to significantly inhibit PI5P4Kα (AG17 = tyrphostin AG-17, AG18 = tyrphostin AG-18, MP = mycophenolate, PB = purvalanol B and SU6668). All compounds were tested at 100 μM, except for PBV, which was tested at 10 μM due to solubility limitations at higher concentrations. The raw image and the extracted data are shown in (C) and (D), respectively. The commercial PI5P substrate predominantly contains two palmitate groups with a very small amount of deacylated lipid lyso-PI5P that contains only one palmitate group. The intense top spots in (C) represent the PI(4,5)P2 product and the faint spots below represent the product with just one palmitate group.

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thin layer chromatography. AG-82 displayed ~75% inhibition of activity at 100 μM in the radiometric assay (See Figure 3C) and was used as a control compound here. It exhibited an excellent MSR (Minimum Significant Ratio, \cite{32}) of 1.29 (Figure 4D) in the LOPAC\textsuperscript{1280} screen and yielded an IC\textsubscript{50} of 93 μM, which is consistent with the observed 75% inhibition at 100 μM in the radioassay. Importantly, the enzyme-substrate solution was not prone to settling and did not require mixing during dispense of multiple plates, as evidenced by the successful scale-up experiment performed on a fully-automated robotic platform (See Figure S3).

**Table 1.** Steps of the High-Throughput PI5P4K\textalpha{} Assay.

<table>
<thead>
<tr>
<th>Step</th>
<th>Parameter</th>
<th>Value</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Enzyme/Lipid</td>
<td>2 μL</td>
<td>Enzyme/lipid, no enzyme and no lipid solutions; reagent bottles are kept on ice</td>
</tr>
<tr>
<td>2</td>
<td>Centrifugation</td>
<td>10 sec</td>
<td>Spin 300 xg</td>
</tr>
<tr>
<td>3</td>
<td>Library and Control Compounds</td>
<td>23 nL</td>
<td>152, 76, 15, 3 and 0.61 μM final concentration titration series of library and 769 μM AG-82 1:2 16-point control titration.</td>
</tr>
<tr>
<td>4</td>
<td>Substrate</td>
<td>1 μL</td>
<td>ATP, room temperature</td>
</tr>
<tr>
<td>5</td>
<td>Incubation time</td>
<td>60 min</td>
<td>Room temperature</td>
</tr>
<tr>
<td>6</td>
<td>ADP-Glo Reagent 1</td>
<td>2 μL</td>
<td>ADP-Glo detection reagent 1 at room temperature</td>
</tr>
<tr>
<td>7</td>
<td>Incubation time</td>
<td>40 min</td>
<td>Room temperature</td>
</tr>
<tr>
<td>8</td>
<td>ADP-Glo Reagent 2</td>
<td>4 μL</td>
<td>ADP-Glo detection reagent 2 at room temperature; reagent bottle is protected from light</td>
</tr>
<tr>
<td>9</td>
<td>Incubation time</td>
<td>30 min</td>
<td>Room temperature</td>
</tr>
<tr>
<td>10</td>
<td>Assay Readout</td>
<td>Luminescence</td>
<td>ViewLux in end-point mode: 20 second exposure</td>
</tr>
</tbody>
</table>

**Step Notes.**
1. Dispensed into white solid-bottom 1536-well MB Greiner plates with a Flying Reagent Dispenser (FRD). Reagent is kept on ice. Final concentration is 10 nM PI5P4K\textalpha{}, 75 μM PI5P and 150 μM DPPS.
2. Plates centrifuged at 300 xg for 10 seconds.
3. DMSO compound solutions transferred with a Kalypsys pintool.
4. Dispensed with a FRD. Final concentration is 5 μM ATP.
5. Plates incubated at room temperature.
6. Dispensed with a FRD.
7. Plates incubated at room temperature.
8. Dispensed with a FRD.
9. Plates incubated at room temperature.
10. Luminescence was measured with a ViewLux CCD Imager with a 20 second exposure time.

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Curve fitting of the concentration responses using the Hill equation was performed using in-house methods and is detailed in the Materials and Methods section. Of the 1,280 LOPAC compounds screened, a total of 32 compounds were identified with upper and lower asymptotes in their concentration responses and an efficacy of over 80%.

The LOPAC library contains many tyrphostins and indeed several analogues of the AG-92 (Tyrphostin 25) control with a range of potencies IC50 of 2.5 µM to over 100 µM, Figure 5A) were identified. The most potent analogue, I-OMe-tyrphostin AG-538 (I-OMe-AG-538), had an IC50 of 2.5 µM. Additionally, long-chain compounds, such as 1-O-Octadecyl-2-O-methyl-sn-glycero-3-phosphorylcholine, were identified as inhibitors. These compounds may be acting through prevention of lipid substrate binding to PISP4K.

Seven of the initial hits were reacquired and tested as 11-point titrations in the PISP4K enzyme assay. The activity of all of these inhibitors was reconfirmed indicating the reproducibility of the assay. The IC50 was determined to be 1 µM for a freshly plated stock of I-OMe-AG-538.

**Counterscreen Assay on Selected Hits**

When developing a screen, it is important to assess the type of compounds that could interfere with the detection. For the assay developed here, the formation of the product ADP is detected by a coupled assay system that contains several enzymes [31,33,34]. Compounds that impact these detection components could be erroneously identified as actives, so a facile assay was developed to test for compounds that interfere with this detection system. Previously, compounds have been identified that interact with the firefly luciferase system, one of the components in the ADP-Glo kit [35]. It is possible and indeed recommended to screen apparent hits in a luciferase-coupled assay against the luciferase enzyme itself [36]. Furthermore, ADP-Glo contains additional enzymes that may also be subject to compound interference, making it a counterscreen for any apparent hits using the full ADP-Glo kit very important. The selected compounds from the screen were tested in an assay where a fixed concentration of ADP/ATP, representing 20% conversion, is present in the absence of the enzyme and lipid, and the detection system was used to quantitate the ADP. If the compound does not act on the detection system, the amount of luminescence, which correlates with the amount of ADP, should not differ between wells with compound and wells with DMSO. If, however, the compound interferes with the detection system, apparent IC50 values would be obtained for the compound titration. The compounds were found to be free of effects against the detection in the ADP-Glo detection system (Figure 5B), indicating that the compound IC50 values that were obtained in the PISP4K assay were due to the effect of the compound on the enzyme reaction itself and not on the detection reagents.

**ATP Competition**

To help identify the mode of inhibition of I-OMe-AG-538, the IC50 was measured using the miniaturized ADP-Glo assay in the presence of seven concentrations of ATP spanning 0.25xKm to 10xKm of ATP. By the slope of the resulting data plotted as IC50 vs. [ATP]/Km, the inhibitor can be determined to be a non- or competitive inhibitor with ATP [37]. Figure 6 shows a positive relationship between the [ATP]/Km and the IC50 indicating competitive inhibition, i.e. that the inhibitor and ATP cannot bind to the enzyme in an orthosteric mode. Additionally, the Ks was determined to be 0.5 µM. Tyrphostin AG-538 (AG-530) inhibits insulin-like growth factor 1 receptor (IGF1R) with an IC50 of 60 nM and is competitive with respect to substrate for IGF1R [38]. Both AG-530 and I-OMe-AG-538 inhibited IGF1R in a similar dose-dependent manner in a cell assay [39]. Modeling showed that the tyrosinates in the substrate overlaid with the catechols in AG-530 and that AG-530 mimics the substrate [38]. For PISP4K, however, I-OMe-AG-538 does not resemble the substrate PISP. Indeed, tyrphostins have been shown previously to be ATP competitive, substrate competitive and competitive with both substrate and ATP [39,40]. Although we did identify a few compounds such as oleamide and OMDM-2 ((R)-N-oleoyl Tyrosinol), which were found to be weak inhibitors (IC50 ~30 µM), whose potency did not vary with ATP, and could therefore interact with the lipid pocket, we were not able to vary lipid in the present assay configuration, due to the limited solubility of the lipid substrate. As well, if the lipid concentration was increased to look for competition, the DMSO concentration would increase, and the enzyme would uncouple from its substrate, making it impossible to conduct this experiment. To assess lipid competition, either a higher concentration of substrate in DMSO would need to be tried or the commercial liposome

**Figure 5. Confirmation of inhibitors.** (A) i, The IC50 inhibition curves of the tyrphostin analogues identified from the Lopac library are shown. Numbers refer to the tyrphostin analog and IC50 was determined to be as follows: tyrphostin I-OMe-AG-538 (2 µM), tyrphostin 51 (5 µM), tyrphostin AG 112 (13 µM), tyrphostin AG 538 (14 µM), tyrphostin AG 808 (18 µM), tyrphostin 47 (20 µM), tyrphostin AG 537 (32 µM), tyrphostin 23 (45 µM), tyrphostin AG 555 (50 µM), tyrphostin AG 698 (60 µM), tyrphostin AG 490 (89 µM), tyrphostin AG 494 (89 µM), tyrphostin AG 1478 (100 µM), ii, Structures of the four most potent tyrphostin analogs. (B) The IC50 curves with standard deviation error bars (N = 2) of tyrphostin I-OMe-AG-538 in the PISP4K assay (squares) and the counterscreen (open circles). doi:10.1371/journal.pone.0054127.g005
preparation process could be utilized. Alternatively, methods, such as surface plasmon resonance using lipid-coupled sensor chips, could be employed to determine the mechanism of action in regards to substrate competition.

Conclusions

In conclusion, this new and validated luciferase-coupled bioluminescence 1536-well PI5P4Kα assay entails a lipid preparation that is more facile and less costly than earlier assay designs for the important drug targets PI5P4Kα/PI5P4Kβ. The lipid could be solubilized in DMSO with just sonication and vortex mixing on the benchtop. The enzyme/lipid mixture is stable overnight and did not require additional stirring during the course of our screens. The IC_{50} of the control compound measured in this assay was corroborated by the orthogonal thin layer chromatography of the lyophilized lipid cakes sonicated in buffer method (black) and the DMSO method (checkered) are shown. Standard deviation error bars are shown (N = 2).

(TIF)

Figure S2 DMSO effect on the performance of the luciferase-coupled assay system. DMSO concentrations were tested from 5–30%. Testing below 5% DMSO was not feasible due to the requirement of DMSO for solubilization of the substrate. Standard deviation error bars are shown (N = 2).

(TIF)

Figure S3 Assay performance on robotic system. Stable performance of the PI5P4Kα assay was obtained in the 1536-well assay scale-up experiment performed on a fully-automated robotic platform [41] (147 plates tested). The Z' factor is shown as a function of assay plate.

(TIF)

Author Contributions

Conceived and designed the experiments: MID ATS BME NT SM DSA LCS AS KS KT. Performed the experiments: MID ATS BME NT SM RP MB DSA ZL LCC AS KS KT. Analyzed the data: MID ATS MS BME NT SM RP MB DSA ZL LCC AS KS KT. Contributed reagents/materials/analysis tools: MID ATS MS BME NT SM KS KT. Performed the experiments: MID ATS BME NT SM RP MB DSA ZL LCC AS KS KT. Wrote the paper: MID ATS MS BME NT SM RP MB DSA ZL LCC AS.

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