A novel role of sphingosine 1-phosphate receptor S1pr1 in mouse thrombopoiesis

Lin Zhang,1,3 Martin Orban,1,3 Michael Lorenz,1,3 Verena Barocke,1,3 Daniel Braun,1,3 Nicole Urz,1,3 Christian Schulz,1,3 Marie-Luise von Brühl,1,3 Anca Tirniceriu,1,3 Florian Gaertner,1,3 Richard L. Proia,4 Thomas Graf,5,6 Steffen-Sebastian Bolz,7 Eloi Montanez,8 Marco Prinz,9,10 Alexandra Müller,9 Louisa von Baumgarten,2 Andreas Billich,11 Michael Sixt,8 Reinhard Fässler,8 Ulrich H. von Andrian,12 Tobias Junct,11 and Steffen Massberg1,3,13

1Medizinische Klinik und Poliklinik I and Neurologische Klinik, Klinikum der Universität, Ludwig-Maximilian-Universität München, 81337 Munich, Germany
2Deutsches Herzzentrum München, Technische Universität München, 80636 Munich, Germany
3Genetics of Development and Disease Branch, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, MD 20892
4Centre for Genomic Regulation, 08010 Barcelona, Spain
5Department of Physiology, University of Toronto, Toronto, Ontario, Canada M5S 1A8
6Max Planck Institute of Biochemistry, 82152 Martinsried, Germany
7Pathologisches Institut, Abteilung Neuropathologie, Universitätsklinikum Freiburg, 79106 Freiburg, Germany
8BIOSS Centre for Biological Signalling, University of Freiburg, 79108 Freiburg, Germany
9Novartis Institutes for BioMedical Research, 4002 Basel, Switzerland
10Department of Pathology, Harvard Medical School, Boston, MA 02115
11Max Planck Institute of Biochemistry, 82152 Martinsried, Germany
12Department of Pathology, Harvard Medical School, Boston, MA 02115
13Munich Heart Alliance, German Cardiovascular Research Centre, 80802 Munich, Germany

Millions of platelets are produced each hour by bone marrow (BM) megakaryocytes (MKs). MKs extend transendothelial proplatelet (PP) extensions into BM sinusoids and shed new platelets into the blood. The mechanisms that control platelet generation remain incompletely understood. Using conditional mutants and intravital multiphoton microscopy, we show here that the lipid mediator sphingosine 1-phosphate (S1P) serves as a critical directional cue guiding the elongation of megakaryocytic PP extensions from the interstitium into BM sinusoids and triggering the subsequent shedding of PPs into the blood. Correspondingly, mice lacking the S1P receptor S1pr1 develop severe thrombocytopenia caused by both formation of aberrant extravascular PPs and defective intravascular PP shedding. In contrast, activation of S1pr1 signaling leads to the prompt release of new platelets into the circulating blood. Collectively, our findings uncover a novel function of the S1P–S1pr1 axis as master regulator of efficient thrombopoiesis and might raise new therapeutic options for patients with thrombocytopenia.
the perivascular niche, where they interact with sinusoidal BM endothelial cells (Avecilla et al., 2004; Patel et al., 2005a). Once they have settled in the perivascular microenvironment, mature MKs form dynamic transendothelial pseudopods, which extend into the lumen of BM sinusoids. These intravascular pseudopodial extensions, termed proplatelets (PPs), continue to elongate and become tapered into multiple platelet-size beads connected to each other and with their maternal MKs by thin cytoplasmic bridges (Italiano et al., 1999; Patel et al., 2005a). The release of platelets, the final step of platelet formation, then occurs within the blood, where new platelets are shed as fragments from the tips of intravascular PPs (Stenberg and Levin, 1989; Choi et al., 1995; Italiano et al., 1999; Junut et al., 2007).

MKs are a rare cell population, constituting <0.01% of all BM cells. This contrasts with the high demand of platelet production, implying that the differentiation of MKs (termed megakaryocytopoiesis) and the subsequent assembly and release of platelets by MKs (termed thrombopoiesis) are highly efficient and tightly controlled processes. Among the factors that modulate megakaryocytopoiesis, thrombopoietin (TPO) is the major regulator of MK expansion from hematopoietic stem and progenitor cells, whereas chemokines, including stromal-derived factor-1 (SDF–1), primarily initiate the relocation of maturing MKs to the perivascular microenvironment (Avecilla et al., 2004). In contrast, the molecular pathways that control the final steps of thrombopoiesis, particularly the guidance signals that direct megakaryocytic pseudopodial extensions into the vascular lumen and trigger the intravascular release of new platelets, are entirely unknown.

The bioactive sphingolipid sphingosine 1-phosphate (S1P) and the receptors responsive to this mediator regulate important biological functions of various hematopoietic cell types (Spiegel and Milstien, 2003, 2011; Schwab et al., 2005; Massberg et al., 2007), including cell migration in the BM compartment (Ishi et al., 2009; Allende et al., 2010). Here we report that S1P and the MK S1P receptor S1pr1 receptor are indispensable for normal BM thrombopoiesis. Using mouse mutants and by multiphoton intravitral microscopy (MP–IVM), we demonstrate that a transendothelial S1P gradient navigates megakaryocytic PP extensions into the lumen of BM sinusoids. In the blood, PP extensions are exposed to high S1P concentrations, which initiate the subsequent shedding of platelets into the circulation. Both processes involve the S1P receptor S1pr1, triggering activation of the Gi/Rac GTPase signaling. Correspondingly, lack of S1pr1 on MKs, but not of other S1P receptors, results in severe thrombocytopenia. Thus, we have identified the S1P–S1pr1 pathway as a key nodal point integrating guidance cues that navigate directional PP elongation and enabling the final step of thrombopoiesis, the shedding of new platelets into the blood stream.

RESULTS
S1pr1 expression in MKs intrinsically regulates platelet homeostasis

We observed here that cultured mouse and human MKs, as well as the human megakaryocytic cell lines Meg01 and CMK, each express the S1P receptor subtypes 1, 2, and 4 (encoded by S1pr1 and S1PR1, S1pr2 and S1PR2, and S1pr4 and S1PR4 in mice and humans, respectively; Fig. 1, A–E; and Table S1). To directly test whether S1P receptors play a role for megakaryo- or thrombopoiesis, we determined platelet counts in peripheral blood of WT mice and mice lacking the S1P receptors expressed by MKs. Loss of S1pr2 or S1pr4 on hematopoietic cells had no significant effects on peripheral platelet counts or platelet size (Fig. 1 F and Table S2), and not depicted). In contrast, ablation of the S1pr1 gene was associated with dramatically reduced platelet numbers. Loss of one S1pr1 allele (S1pr1+/− mutants) already resulted in a significant reduction in the number of circulating platelets (Fig. 1 G and Table S3). Loss of both alleles (S1pr1−/− mice) was embryonically lethal (Liu et al., 2000); thus, to circumvent embryonic lethality, we generated chimaeras by transferring fetal liver (FL) cells from S1pr1−/−, S1pr1+/−, or S1pr1+/+ donors into irradiated WT mice. 6–8 wk after reconstitution, BM cells from S1pr1−/−, S1pr1+/−, or S1pr1+/+ FL chimaeras were isolated and further transplanted into irradiated secondary recipient mice. Platelet counts in S1pr1−/− and S1pr1+/− chimaeras were reduced by >50% and 70% compared with S1pr1+/+ chimaeras, respectively (Fig. 1 G and Table S2). Collectively, these results indicate that S1pr1 on hematopoietic cells controls blood platelet homeostasis, whereas S1pr2 and S1pr4 are dispensable for this process.

Next we evaluated whether S1pr1 expressed by MKs or by other hematopoietic lineages regulates the number of blood platelets. To this end, we reconstituted irradiated mice with BM cells carrying two floxed S1pr1 alleles (S1pr1fl/fl) and transduced with a lentivirus expressing Cre recombinase under the MK-specific GpIIb promoter (GpIIb-Cre S1pr1fl/fl) to delete S1pr1 in the MK/platelet progeny (Fig. 1 H; Allende et al., 2003). Importantly, platelet counts became significantly reduced in GpIIb-Cre S1pr1fl/fl BM recipients as compared with S1pr1fl/fl control chimaeras (Fig. 1 H). Moreover, lentiviral reexpression of S1pr1 under the MK-specific GpIIbα promoter rescued S1pr1−/− FL cells to reconstitute the blood platelet compartment in lethally irradiated mice (Fig. 1 I). These findings demonstrate that S1pr1 expressed by the MK lineage intrinsically controls platelet homeostasis.

Normal MK development, platelet life span, and serum TPO levels in S1pr1-deficient mice

What could be the reason for the severe thrombocytopenia in the absence of S1pr1? First we showed that the life spans of platelets from S1pr1+/+, S1pr1−/−, or S1pr1−/− chimaeras and between WT and S1pr1+/− mutant mice was similar, excluding a reduced life span as cause for the reduced platelet counts (Fig. 2 A). We also excluded a defect in the release of TPO, the principle regulator of thrombopoiesis (Kaushansky, 2005a), as cause for the thrombocytopenia in S1pr1-null mutants (Fig. 2 B). Finally, we also could not find evidence for a gross defect in MK development, as we found similar numbers of megakaryocytic progenitor cells in WT and S1pr1−/− FL cells populations.
Figure 1. MKs express S1pr1, and S1pr1-deficient mice display severe thrombocytopenia. (A) Relative expression of S1P receptor mRNA by FL-derived mature and immature MKs. (B) Relative expression of S1P receptor mRNA in human megakaryocytic cell lines. (A and B) Data are representative of three independent experiments with triplication. (C) Representative immunostaining of S1pr1 in immature and mature FL-derived MKs. WT MKs stained
Loss of S1pr1 increases MK size but has no effect on positioning and motility of MKs in vivo

Next we examined whether S1pr1 controls platelet biogenesis for example by modulating MK motility or their positioning within the BM compartment. To address this question, we performed MP-IVM of calvarial BM (Junt et al., 2007) of two different sets of S1pr1+/+ or S1pr1−/− chimaeras, in which MKs and their progeny were genetically marked: (a) S1pr1+/+ or S1pr1−/− CD41-YFP<sup>fl/fl</sup> FL chimaeras, in which MKs and platelets express the YFP driven from the endogenous CD41 gene locus (Zhang et al., 2007) and (b) S1pr1<sup>+/+</sup> or S1pr1<sup>−/−</sup> lenti-GpIbα–enhanced GFP (EGFP) BM chimaeras, in which MKs and platelets express EGFP under the transcriptional control of the murine GpIbα promoter (Lavenu-Bomble et al., 2007). The experiments revealed neither differences in MK size nor in their positioning or motility when we compared S1pr1<sup>+/+</sup> CD41-YFP<sup>fl/fl</sup> or S1pr1<sup>−/−</sup> lenti-GpIbα–EGFP chimaeras and naive (nontransplanted) S1pr1<sup>+/+</sup> CD41-YFP<sup>fl/fl</sup> or platelet factor 4 (Pf4)<sup>+</sup>-EGFP transgenic mice, in which EYFP is driven by the MK-specific Pf4 promoter, which allowed excluding a major influence of irradiation and BM transplantation (Fig. 3 A–D). As reported previously (Junt et al., 2007), S1pr1<sup>+/+</sup> MKs were large, mostly sessile cells always located in close proximity to BM sinusoids (Fig. 3 A and D–G). In S1pr1<sup>−/−</sup> chimaeras and S1pr1<sup>+/+</sup> mice, MKs were significantly larger compared with S1pr1<sup>+/+</sup> chimaeras, whereas the position and motility of MKs was similar among all genotypes (Fig. 3, E–G). The aforementioned results suggest that in contrast to other cells in the BM (Ishii et al., 2009), neither positioning nor migration of MKs or their committed progenitors in marrow spaces is controlled by S1pr1.

S1pr1 is essential for intravascular PP formation (PFF)

During thrombopoiesis, mature MKs extend transendothelial protrusions, termed PPs, into BM microvessels (Junt et al., 2007). To test whether S1p/S1pr1 receptor signaling plays a role during PPF, we cultured MKs in vitro (Lecine et al., 1998) and found that on average, 9 out of 100 WT MKs spontaneously formed PPs as assessed by phase-contrast microscopy. MKs isolated from S1pr2<sup>−/−</sup> and S1pr4<sup>−/−</sup> mice generated similar number of PPs (unpublished data). In sharp contrast, in vitro PPF was reduced by >70% in S1pr1<sup>−/−</sup> MKs, as <2 out of 100 S1pr1<sup>−/−</sup> MKs formed PPs (Fig. 4 A). Importantly, lentiviral reexpression of GpIbα promoter–driven S1pr1 in S1pr1<sup>−/−</sup> MKs corrected PPF in vitro (Fig. 4 B). These results clearly indicate that S1pr1 plays a critical and intrinsic role for PPF by MKs.

When we examined how S1pr1 might control PPF, we could exclude a primary lack of the invaginated demarcation membrane system (DMS), the predominant reservoir for PP membranes (Radley and Haller, 1982; Schulze et al., 2006), in S1pr1<sup>−/−</sup> MKs, as electron microscopy of S1pr1<sup>−/−</sup> BM MKs did not reveal abnormalities of the DMS when compared with S1pr1<sup>+/+</sup> BM MKs (Fig. 4 C). Next we tested whether S1P serves as a chemoattractant for polarizing MKs and for inducing the formation of PP protrusions. Within the normal BM compartment, S1P is rapidly degraded by lyases and phosphatases expressed by most hematopoietic cells. Thus, the local S1P concentrations in the BM (with its densely packed hematopoietic cells) are exceedingly low (unpublished data), reflecting similar concentrations reported for other tissues such as lymph nodes (Schwab et al., 2005; Pappu et al., 2007). In contrast, high S1P concentrations exist in the blood stream (Caligan et al., 2000; Berdyshev et al., 2005; Pappu et al., 2007). Because of their positioning at the vascular interface, MKs are therefore exposed to a steep transendothelial S1P gradient. To mimic the situation in the BM, we exposed cultured MKs to a gradient of S1P in vitro. Notably, PP extensions developed preferentially toward increasing concentrations of S1P but not of vehicle (Fig. 4 D). A similar result was also obtained with S1pr2<sup>−/−</sup> and S1pr4<sup>−/−</sup> MKs (Fig. 4 D). VPC23019, a previously described S1pr1 and S1pr3 antagonist (Davis et al., 2005), was used in our study to selectively block megakaryocytic S1pr1 signaling because MKs do not express S1pr3 (Fig. 1 A). Inhibition of the megakaryocytic S1pr1 using VPC23019 abolished this directionality of PPF; MKs projected PP extensions into random directions (Fig. 4 D). These findings suggest that S1P–S1pr1 signaling is essential for PPF by providing a chemoattractant stimulus that controls the polarization of PP processes generated by MKs in culture.

Next we defined the signaling downstream of S1pr1 involved in the regulation of PPF and polarization. Because Rac GTPase activity controls actin dynamics leading to membrane protrusion/extensions (Aspenström et al., 2004), we tested whether activation of Rac GTPases via
indicating that activation of Rac GTPases downstream of S1pr1 is required for PPF.

To evaluate the in vivo relevance of S1P and its receptors for PPF, we examined CD41-YFP<sup>ki/+</sup> mice by MP-IVM. In CD41-YFP<sup>ki/+</sup> mice, 59% of all MKs extended plump or long PP protrusions into BM sinusoids (Fig. S1A and Video 1), indicating active participation in platelet biogenesis. PP protrusions S1pr1 in MKs controls PPF. As reported previously for endothelial cells (Paik et al., 2004), we found that S1P, and also the S1pr1-specific agonist SEW2871, enhances Rac GTPase activity via S1pr1 in megakaryocytic cell lines (Fig. 4, E and F). Conversely, we observed that pharmacological inhibition of Rac GTPases by NSC23766 completely abolished PPF in response to S1P in vitro (Fig. 4 G), indicating that activation of Rac GTPases downstream of S1pr1 is required for PPF.

To evaluate the in vivo relevance of S1P and its receptors for PPF, we examined CD41-YFP<sup>ki/+</sup> mice by MP-IVM. In CD41-YFP<sup>ki/+</sup> mice, 59% of all MKs extended plump or long PP protrusions into BM sinusoids (Fig. S1A and Video 1), indicating active participation in platelet biogenesis. PP protrusions.
Figure 3. Loss of S1pr1 increases the size but has no effect on the positioning and motility of MKs in vivo. (A) Representative MP-IVM images of YFP⁺ or EGFP⁺ MKs (green) in BM. BM microvasculature was visualized by intravenous injection of TRITC-dextran (red). Left, naive (nontransplanted) S1pr1⁺⁻;CD41-YFPki/⁺; middle left, S1pr1⁺⁻;Pf4-EYFP; middle right, S1pr1⁺⁺;lenti-GPib-EGFP BM chimaeras; right, S1pr1⁺⁺;CD41-YFPki/⁺ FL chimaeras. Bars, 20 µm. (B–D) Volumes (B), distances from sinusoids (C), and the instantaneous lateral (x-y) velocity (D) of MKs in the indicated groups. Red lines indicate medians; error bars represent SEM. n = 13–46 MKs per genotype. Data are pooled from three mice each group. P-values among the different groups in B–D are >0.05. (E) Surface area of MKs in BM. (F) Distance of MKs from BM sinusoids. (G) Instantaneous lateral (x-y) velocity of MKs. Red lines indicate medians. (E–G) Data are pooled from three mice each group.
Figure 4. S1P regulates PPF. (A) The percentage of MKs displaying PPF. PPF is expressed as the percentage of MKs carrying PPs (8,000–10,000 MKs per experiment, five independent experiments with triplications). (B) The percentage of MKs displaying PPF in S1pr1+/+ or S1pr1−/− MKs transduced with lenti-GpIbα-S1pr1 or empty control vectors (3,000–8,000 MKs per experiment, two independent experiments with triplications). (C) Representative electron micrographs of WT and S1pr1−/− MKs in BM. Arrowheads indicate the DMS. Red color highlights the DMS. N, nucleus. (D) The percentage of MKs with polarized PPF in the presence or absence of S1P and the S1pr1-specific inhibitor VPC23019 (VPC). n = 127–265 MKs per group. Data are pooled from three to five independent experiments. (E) Y10/L8057 cells were incubated with 10 µM S1P or vehicle for 2 min. The activities of Rac-GTP were quantified by pull-down assay (n = 5 independent experiments). (F) Y10/L8057 cells were incubated with 1 µM S1pr1 agonist, SEW2871, or vehicle for 5 min. The activities of Rac-GTP were quantified by pull-down assay (n = 3 independent experiments). (G) The percentage of MKs displaying PPF in the presence or
extends almost exclusively into marrow sinusoids of CD41-YFP
ki/+ mice, whereas we rarely detected extravascular PP
processes (Fig. 4, H and I; and Video 2). To determine whether
the S1P receptors expressed by MKs provide the guidance
information necessary to direct PP processes into BM
sinusoids, we examined S1pr1+/- CD41-YFPki/+; S1pr2-/-
CD41-YFPki/+; and S1pr4-/- CD41-YFPki/+ mice as well as
lethi-Gfpbo-EGFP BM chimaeras. Consistent with our in
vitro findings (Fig. 4 D), loss of S1pr2 or S1pr4 did not affect
the formation or polarization of PPs in vivo (Fig. 4, H and I).
In contrast, loss of S1pr1 disrupted PPF and polarization;
correspondingly, S1pr1+/- or S1pr1-/- MKs projected PP
extensions in random directions (Fig. 4, H and I; and Video 2).
As a consequence, we found aberrant PP processes in the
marrow interstitial space, whereas intrasinusoidal PPs were rarely
detected in S1pr1-deficient chimaeras (Fig. 4, H and I; and
Video 2). Likewise, when we treated S1pr1+/- CD41-YFPki/+ mice with the S1pr1-specific antagonist W146 for 24 h, the
physiological directionality of PPF was entirely disrupted.
We frequently observed long cytoplasmic extensions outside si-
nusoids in mice treated with W146 but not in vehicle-treated
animals (Fig. 4, H and I; and Video 3). In addition, inhibition
of S1pr1 also retarded PP growth in vivo, resembling the re-
duced PPF of cultured S1pr1+/- MKs in vitro (Fig. 4, A and J).
These results indicate that S1pr1 signaling supports PPF and
elongation along the physiological S1P gradient between BM
interstitium and BM sinusoids and controls the entry of PPs
into the marrow blood stream in vivo.

S1P enhances PP fragmentation via S1pr1 in vitro
Once MK PP processes have entered the blood, they are ex-
posed to significantly higher S1P concentrations compared
with the BM interstitium (unpublished data). To our surprise,
when we mimicked the situation in the blood by incubating
cultured MKs to allow PPF and then adding a high concen-
tration of S1P (instead of exposing MKs to an S1P gradient),
we found a significant reduction in the number of MKs dis-
playing PPF extensions (Fig. 5 A). Using differential interfer-
ence contrast (DIC) microscopy of cultured MKs, we observed
that exposure of MKs to a high, homogenous concentration of
S1P results in almost immediate shedding of platelet-like
particles from PPs (Fig. S1 B and Video 4). Within 1 h, platelet-
like particles were shed from 26% of PPs in response to
S1P but only from 3% of PPs treated with vehicle (Fig. 5 B).
To further quantify the effect of S1P on PP shedding in vitro,
we determined the number of fragmentation events by flow
cytometry (Fig. 5 C). S1P, but not vehicle, increased PP
fragmentation at high S1P concentrations, mimicking S1P
plasma levels but not at low concentrations prevailing in the
BM interstitium (Fig. 5, C and D).
In vivo, blood flow–induced shear stress might facilitate
the separation of intravascular cell fragments from MKs (Junt
et al., 2007). We therefore evaluated whether S1P also plays
a role for PP fragmentation under flow conditions. Cultured
MKs exposed to the physiological shear stress of BM sinu-
soids (4 dynes/cm²; Junt et al., 2007) in the absence of S1P
(serum-free buffer) rarely shed PPs from their MK stems.
In contrast, in the presence of 5 μM S1P, PPs were rapidly re-
leased (Fig. 5, E and F; and Video 5), indicating that S1P is
required for PP shedding under static as well as flow condi-
tions. Loss of S1pr2 or S1pr4 did not affect S1P-induced PP
shedding (Fig. 5 B, Fig. S1 B, and Video 6). However, lack of
the megakaryocytic S1pr1 receptor completely abolished
S1P-induced release of PPs (Fig. 5 B, Fig. S1 B, and Video 6).
This indicates that S1pr1, but not S1pr2 or S1pr4, plays the
predominant role for S1P-driven PP shedding. To further
clarify the involved signaling pathway, we used pertussis toxin
and NSC23766 to inhibit Gi and Rac GTPase activity, re-
spectively. Both inhibitors blocked S1P-induced fragmenta-
tion of PPs (Fig. 5 B and Fig. S1 B). The observation that
S1P activates Rac GTPase in MKs via S1pr1 (Fig. 4, E and F)
together with the aforementioned findings suggests that
S1P-induced PP fragmentation depends on S1pr1/Gi/Rac
GTPase signaling.

S1P controls PP shedding into blood via S1pr1 in vivo
To address whether S1P–S1pr1 signaling is also essential for
PP fragmentation in vivo, we examined PP shedding in live
mice by MP-IVM. PP shedding from MKs was a frequent
event in naive (nontransplanted) S1pr1+/- CD41-YFPki/+ transgenic mice (Junt et al., 2007) but also in S1pr1+/- CD41-
YFPki/+ BM chimaeras (Video 7). Most MKs shed PP frag-
ments that consist of beaded platelet-like structures (Fig. 6,
A and B; and Video 7), which generate mature platelets by
undergoing consecutive fragmentation steps (Behnke and
Forer, 1998; Junt et al., 2007). More than 60% of the S1pr1+/-
MKs carrying intravascular PP processes showed fragmentation
within 1 h (Fig. 6, A and B; and Video 7). We did not find
any defect in PP fragmentation in S1pr2-/- or S1pr4-/-
mice (Fig. 6 A), whereas this process was severely impaired in
S1pr1 mutants. In S1pr1-/- chimaeras, we barely observed
intravascular PP processes because of the aberrant interstitial
PPF reported above (Fig. 4, H and I). However, 70–100% of
the PP processes that had eventually made their way into BM
absence of 50 μM NSC23766, a Rac GTPase inhibitor (4,000–7,000 MKs per experiment; three independent experiments with triplications). (H) Representative
MP-IVM images of MKs with YFP° or EGFP° PPs. Green indicates MKs and PPs; red indicates sinusoids. Arrowheads indicate extravascular YFP°
or EGFP° PPs; arrows indicate interstitial YFP° or EGFP° PPs. The inset shows magnification for the dotted box. The arrow in the inset indicates the connect-
tion between extravascular YFP° PPs. Bars: (C) 2 μm; (H) 20 μm. (I) MKs displaying intrasinusoidal PPF in vivo presented as a percentage of all MKs carry-
ning PPs (20–30 MKs per group; five independent experiments for WT; three independent experiments for other genotypes). (J) The lateral (x-y) speed of PP
elongation was measured in naive (nontransplanted) S1pr1+/-CD41-YFPki/+ mice, S1pr1+/-CD41-YFPki/+ mice, and S1pr1+/-CD41-YFPki/+ mice treated with
W146. Red lines indicate means. Data are pooled from three mice each group. All error bars indicate SEM.
Figure 5. The effect of S1P on PP fragmentation in vitro. (A) The number of MKs displaying PPF in the absence or presence of 10 µM S1P (230–590 MKs per experiment; three independent experiments with triplications). (B) The number of PPs with or without fragmentation observed by DIC microscopy in vitro over 1 h in the indicated groups. Data are pooled from 4–10 independent experiments for each group (n = 30–60 per group). (C) Representative dot plots show flow cytometric analyses of PP fragmentation. The first two panels show the gates for PPs. The CD41⁺CD61⁺ population was analyzed for the distribution of PPs according to FSC and SSC. MKs are G3; PPs with higher and lower FSC values are G2 and G1, respectively. The three representative microphotographs in the right show a representative brightfield image, as well as tubulin and CD41 stainings of fragments sorted using the gating strategy illustrated in the two plots. (D) Flow cytometric analyses of the PP fragmentation index in the presence or absence of various concentrations of S1P. The PP fragmentation index was calculated as described in Materials and methods. Data are representative of six independent experiments with triplication. (E) PP fragmentation by MKs exposed to shear stress. The efficiency of dynamic PP fragmentation was calculated as described in Materials and methods. Data are pooled from five independent experiments for each group. (F) Representative time-lapse video microscopy of PPs in the presence or absence of 5 µM S1P under shear stress (4 dynes/cm²). Arrows indicate direction of flow; arrowheads indicate PP shedding events. All error bars indicate SEM. Bars: (C) 10 µm; (F) 20 µm.
Figure 6. The effect of S1P on PP fragmentation in vivo. (A) Percentage of PP fragmentation events observed by MP-IVM over 1 h in the indicated groups. n = 13–33 per group. Data are pooled from three to seven independent experiments. (B) Role of S1pr1 for PP shedding in vivo visualized by MP-IVM. Representative MP-IVM sequences show that WT MKs frequently shed PPs as shown in the first and the third rows. The inset shows a magnification of a shed PP particle. Asterisks show embedded platelet-like particles. Inhibition or loss of S1pr1 abolishes PP shedding (second and fourth rows). Arrowheads indicate intrasinusoidal PPs, and arrows show extrasinusoidal PPs in S1pr1−/− chimaeras. The dashed lines highlight the sinusoids. Green or yellow
sinusoids remained firmly attached to their MK stems; only in rare instances did MKs release PP fragments (Fig. 6, A and B; and Video 7). Together, these data indicate that S1pr1 is critical for both directional PPF and for proper intravascular PP fragmentation. Defective PP shedding is likely to explain the increase in size of S1pr1−/− MKs (Fig. 3 E). Interestingly, the frequency of intravascular PP shedding was only moderately reduced in CD41-YFP+/− S1pr1+/− mice (Fig. 6 A), suggesting that a single S1pr1 allele is sufficient to maintain intravascular PP shedding and that the mild thrombocytopenia observed in S1pr1+/− mice is mostly caused by a defect in navigating PP processes into BM sinusoids (Fig. 4, H and I).

To examine whether S1pr1 regulates the dynamic process of PP shedding independently from its effects on PP invasion into BM sinusoids, we next tested the consequences of short-term pharmacological inhibition of S1pr1. We treated naive (nontransplanted) S1pr1+/+ CD41-YFP+/+ mice with a single dose of the selective S1pr1 antagonist W146 and visualized PP shedding immediately thereafter. In contrast to protracted inhibition or genetic ablation of S1pr1 (Fig. 4, H and I), this did not affect the overall number of MKs with intravascular PP protrusions. However, <20% of MKs with established intravascular protrusions managed to release PP fragments into the blood stream within 6 h after administration of W146; the vast majority of the intrasinusoidal processes remained attached to their MK stems (Fig. 6, A and B; and Video 8). Repetitive treatment of mice with W146 for 24 h resulted in a significant reduction in circulating young reticulated platelets with an elevated RNA content (Fig. 6 C), consistent with a central role of the S1P–S1pr1 pathway for PP fragmentation and release of platelets. Short-term treatment with W146 also reduced platelet counts in CD1 mice (Fig. 6 D), suggesting that S1pr1 controls thrombopoiesis across different strains of mice. W146 maintains an adequate in vivo receptor blockade for only 5–6 h (Sanna et al., 2006), and shedding reoccurred 6 h after a single dose of W146, suggesting that S1pr1 inhibition does not affect the viability of MKs (Fig. 6 A and Video 8). In rare instances, where PP shedding occurred in the presence of the S1pr1 inhibitor W146, the time required until an intravascular fragment dissociated from its MKs stem was significantly prolonged (Fig. 6 E). The failure to properly shed PPs resulted in the formation of abnormal, thick intravascular PP processes (Video 8). In line with this observation, the few PP fragments that were released despite the presence of W146 were significantly bigger compared with those in vehicle-treated control mice (Fig. 6 F), reminiscent of the large platelets observed in S1pr1-null chimaeras (Fig. 6 G and Table S2).

S1pr1 agonists enhance platelet production

Modulation of S1P receptors by FTY720 (fingolimod) has become a promising strategy for the treatment of patients with multiple sclerosis (Kappos et al., 2006). Here, we show that treatment of mice with a single dose of FTY720 leads to a prompt and transient increase in circulating platelets (Fig. 7 A). When we used MP-IVM to examine MKs before and after treatment with a single dose of FTY720, we found that FTY720 accelerates the shedding of intravascular PP extensions into the blood stream. As a consequence, the number of MKs carrying intravascular PPs significantly decreased immediately after a single dose of FTY720 compared with vehicle (Fig. 7, B and C). This suggests that FTY720 represents an agonist for megakaryocytic S1pr1 receptors and has the potential to rapidly mobilize PPs into the blood, most likely by supporting fragmentation of intravascular PPs (Fig. 7, B and C). Treatment with the S1pr1-specific agonist SEW2871 also caused an increase in circulating blood platelets (Fig. 7 D), further supporting that activation of S1P–S1pr1 receptor signaling enhances thrombopoiesis.

DISCUSSION

Our results assign a new role for S1P and its receptor S1pr1 as master regulators of thrombopoiesis. In a dose-dependent and sequential manner, S1P controls two key steps in the cascade of thrombopoiesis by BM MKs: (1) the polarized development of PP extensions into the blood stream and (2) the subsequent shedding of PPs from their transendothelial stems. As a consequence, loss of S1pr1 is not compatible with normal thrombopoiesis. Collectively, our findings uncover the molecular pathway that enables the final steps of thrombopoiesis.

S1P navigates PP extensions into BM sinusoids and initiates platelet release

Mature MKs form intravascular PP extensions that grow from the MK cell body at a mean speed of 10 μm/min under shear conditions in vivo (Fig. 4 J), with the DMS functioning as the membrane reservoir for PP elongation (Schulze et al., 2006). During elongation, PPs are equipped with specific proteins associated with platelets, including von Willebrand factor (vWF) and fibrinogen receptors. Microtubules, assembled from α/β-tubulin dimers, are the primary structural component of the engine that drives the elongation of PPs. Correspondingly, PPs fail to form when cultured MKs are exposed to agents that inhibit microtubule assembly (Italiano et al., 1999) or sliding (Patel et al., 2005b), and mice lacking β1-tubulin,
fragments break down further in the circulation giving rise to mature platelets of 2–3-µm diameter within the circulation (Stenberg and Levin, 1989). Blood shear stress contributes to the shedding of PPs (Junt et al., 2007; Dunois-Lardé et al., 2009; Thon et al., 2010); however, whether additional signals are required for efficient PP shedding was completely unknown. In this study, we show that hydrodynamic forces alone are not sufficient to allow the release of new platelets from MK PP extensions. Instead, we found that high concentrations of the bioactive lipid S1P prevailing in the sinusoidal blood, but not in the BM interstitium, are mandatory for the release of new platelets from MKs.

From a teleological point of view, the S1P-dependent sequential guidance of thrombopoiesis comprising (a) directional PPF along a transendothelial S1P gradient and (b) subsequent S1P-dependent intravascular PP shedding leads to the introduction of naive platelets into the circulating blood and prevents aberrant platelet production within the BM interstitium. S1P guidance of intravascular PPF, elongation, and shedding therefore provides grounds for efficient thrombopoiesis, which seems instrumental given the relative paucity of MKs.

S1P controls thrombopoiesis via megakaryocytic S1pr1 receptors

Our study shows that MKs robustly express three different S1P receptors, S1pr1, S1pr2, and S1pr4. Loss of S1pr1 on hematopoietic cells and also conditional deficiency of S1pr1 in MKs were associated with severe thrombocytopenia. Moreover, gain of S1pr1 function in S1pr1−/− MKs rescued their...
We measured blood cell \( \text{fl/fl} \) (fingolimod) has emerged as a promising immunosuppressive strategy and is currently being used in patients with relapsing multiple sclerosis (Kappos et al., 2006). After administration, FTY720 is metabolized to phosphorylated FTY720 (FTY720P), an agonist for four of the five S1P receptors including S1pr1. FTY720 limits effector lymphocyte egress from lymph nodes (Matloubian et al., 2004), contributing to its immunosuppressive actions. However, FTY720 has not been examined for its potential effects on megakaryo- and thrombopoiesis. In this study, we show that treatment of mice with a single dose of FTY720 leads to shedding of intravascular PP extensions into the blood stream, paralleled by a prompt, but transient increase in circulating platelets. This suggests that FTY720 acts as an agonist on megakaryocytic S1pr1 receptors and has the potential to rapidly mobilize PPs into the blood, most likely by supporting fragmentation of intravascular PPs. Whereas lymphocyte S1pr1 engagement by phosphorylated FTY720 within secondary lymphoid organs triggers down-modulation of the receptor, resulting in functional antagonism of the S1pr1 pathway, an agonistic effect of FTY720 similar to the one observed here for MK has recently been reported to promote the recirculation of BM osteoclast precursor monocytes from the bone surface (Ishii et al., 2009). This indicates that FTY720 predominantly exerts agonist effects in cells of the myeloid lineage. Because activation of S1P–S1pr1 receptor signaling enhances thrombopoiesis in mice, future studies will have to evaluate potential clinical implications of S1pr1 agonists, in particular in the treatment of thrombocytopenia.

Collectively, the present study reveals that S1P, a signaling lipid circulating in the blood, regulates dynamic intravascular PP elaboration and PP shedding without affecting MK maturation and positioning. Tonic S1P–S1pr1 signaling is critical for normal thrombopoiesis in mice. Although the exact role of S1P–S1pr1 signaling for human thrombopoiesis still needs to be defined, our findings could have clinical implications and provide new approaches to treat thrombocytopenia.

S1P receptor agonist increases blood platelet counts

Recently, modulation of S1P receptor signaling by FTY720 (fingolimod) has emerged as a promising immunosuppressive strategy and is currently being used in patients with relapsing multiple sclerosis (Kappos et al., 2006). After administration, FTY720 is metabolized to phosphorylated FTY720 (FTY720P), an agonist for four of the five S1P receptors including S1pr1. FTY720 limits effector lymphocyte egress from lymph nodes (Matloubian et al., 2004), contributing to its immunosuppressive actions. However, FTY720 has not been examined for its potential effects on megakaryo- and thrombopoiesis. In this study, we show that treatment of mice with a single dose of FTY720 leads to shedding of intravascular PP extensions into the blood stream, paralleled by a prompt, but transient increase in circulating platelets. This suggests that FTY720 acts as an agonist on megakaryocytic S1pr1 receptors and has the potential to rapidly mobilize PPs into the blood, most likely by supporting fragmentation of intravascular PPs. Whereas lymphocyte S1pr1 engagement by phosphorylated FTY720 within secondary lymphoid organs triggers down-modulation of the receptor, resulting in functional antagonism of the S1pr1 pathway, an agonistic effect of FTY720 similar to the one observed here for MK has recently been reported to promote the recirculation of BM osteoclast precursor monocytes from the bone surface (Ishii et al., 2009). This indicates that FTY720 predominantly exerts agonist effects in cells of the myeloid lineage. Because activation of S1P–S1pr1 receptor signaling enhances thrombopoiesis in mice, future studies will have to evaluate potential clinical implications of S1pr1 agonists, in particular in the treatment of thrombocytopenia.

Collectively, the present study reveals that S1P, a signaling lipid circulating in the blood, regulates dynamic intravascular PP elaboration and PP shedding without affecting MK maturation and positioning. Tonic S1P–S1pr1 signaling is critical for normal thrombopoiesis in mice. Although the exact role of S1P–S1pr1 signaling for human thrombopoiesis still needs to be defined, our findings could have clinical implications and provide new approaches to treat thrombocytopenia.

MATERIALS AND METHODS

**Mice.** C57BL/6J (CD45.2), B6.SJL-Ptprc-Pep3/J/BoyCrl (CD45.1), and CD1 mice were purchased from Charles River. B-Actin–EGFP mice were provided by A. Wagers (Harvard Medical School, Boston, MA). S1pr1−/− and S1pr2−/− mice were generated as described previously (Lau et al., 2009; Kono et al., 2004). S1pr4−/− mice were provided by D. Guerini (Novartis Institutes for BioMedical Research, Basel, Switzerland). CD41–YFPϕ/ϕ mice were generated as described previously (Zhang et al., 2007). P4–cre and ROSA26-flox-stop-flox-EYFP mice were obtained from the Jackson Laboratory and crossed to get P4-EYFP transgenic mice, in which EYFP is driven by the MK-specific P4 promoter. Fl. chimaeras and BM chimaeras were generated as described previously (Massberg et al., 2007). Cytometric analysis showed that >95% of the blood cells were derived from donors in all the BM chimaeras. S1pr1fl/fl mice were obtained from R.L. Proia. Age- and gender-matched mice in a C57BL/6 background were used in all experiments. All experimental procedures performed on animals met the requirements of the German legislation on the protection of animals.

**Blood cells and serum TPO measurements.** We measured blood cell counts in the mice before and 12 h after a single injection of FTY720 (3 mg/kg i.p.; Cayman) or DMSO (Sigma-Aldrich) as vehicle. Platelet counts were assessed in the mice before and 12 h after a single injection of SEW2871 (20 mg/kg i.p.; Cayman) or dimethyl formamide (Sigma-Aldrich). Serum TPO was measured using the Quantikine murine TPO Immunoassay kit.
BM samples were fixed in 2.5% formaldehyde and permeabilized with 0.1% Triton X-100/L8057 mouse megakaryocytic cells were cultured in IMDM supplemented with 10% FCS and 25 ng/ml TPO for 1 d and then incubated with FITC-CD41 (BD) and PE-CD34 (BioLegend) antibodies. Immature MKs were identified as a CD41/CD34 double-positive population and enriched with biotinylated anti-CD41 (BD), and anti-CD42c (Emfret Analytics) antibodies were used for immunostaining. The samples were examined using Leica microscopy equipped with 4x0 objective lens (NA = 0.7) or 20x0 objective lens (NA = 0.5) and commercial charge-coupled device camera (AxioCam; Carl Zeiss). Images were acquired by Axiovision software (Carl Zeiss). For quantification of MK number and size in BM, we counted the total number of MKs in five randomly selected 2x0 microscopic fields, and the area of MK cell body was analyzed by Imagej software (National Institutes of Health).

SIP level measurement. Levels of SIP were determined by high-performance liquid chromatography with mass detection as described previously (Berdyshhev et al., 2005).

Transmission electron microscopy. BM samples were fixed in 2.5% glutaraldehyde, embedded in epon, and analyzed using an electron microscope (EM 902; Carl Zeiss).

Quantitative RT-PCR. Quantitative real-time PCR was performed using the MaximaTM SYBR Green PCR Master mix (Thermo Fisher Scientific) and an ABI PRISM 7700 Sequence Detection System (Applied Biosystems) according to the manufacturers’ instructions. The data were normalized to the mRNA level of glyceraldehyde 3-phosphate dehydrogenase (Gapdh or Gapdh). Primers are listed in Table S1.

Western blot analyses. Y10/L8057 mouse megakaryocytic cells were cultured in IMDM supplemented with 10% FCS and 25 ng/ml TPO for 1 d and then starved overnight on 100-mm dishes coated with 0.5% fatty acid–free BSA (Sigma-Aldrich). The starved Y10/L8057 cells were simulated with 10 µM SIP or vehicle for 2 min or 1 µM SEW2871 or vehicle for 5 min. Rac-GTP activities were measured using Rac assay kit (Cell Biolabs). Platelet lysates were subjected to SDS-PAGE and then immunoblotted with antibodies recognizing murine S1pr1 (Imgenex) or β-actin (Abcam) as loading controls.

Multiphoton intravital imaging of the BM. We prepared the mouse calvaria BM as described previously (Junt et al., 2007). We used a BioTech TriMscape system (LaVision BioTec) and Ti:Sa laser (MaiTai) to capture images through a 20x water immersion objective lens (NA = 0.95; Olympus). Images were acquired with ImSpectorPro (LaVision BioTec). For three-dimensional (3D) acquisition, the stacks were acquired at a 920-nm wavelength at vertical spacing of 2–3 μm to cover an axial depth of 30–100 μm (for YFP or EGFP). Subsequently, the same stacks were acquired at a wavelength of 800 nm (for TRITC–dextran). The distances between MKs and the vasculature were measured in the reconstructed 3D structure using Volocity software (PerkinElmer). If MKs were outside the vessels, the closest distance from MKs to vessels was measured and represented as negative values. If MKs were in direct physical contact with the vessels, the distance was regarded as positive.
zero. For analysis of PP shedding, four-dimensional acquisitions were performed at 920 nm by capturing 3D image stacks at an interval of 60 s for 60 min. Videos were generated as maximum intensity projections representing a “top” (x-y) view of the volume using Velocity. The centroid positions (x-y) of MKs or PP tips from a series of top-view (x-y) images were measured using ImageJ, and instantaneous lateral (x-y) velocity, a measure of cell motility was determined by dividing the change in cell displacement between each frame by the time interval between frames and was quantified by the Chemotaxis and Migration Tool plugin (ibidi). All mice were treated with 8 µg/kg/d mPTP (ImmunoTools) for 3 d before imaging as described previously (Junt et al., 2007). W146 (Avanti Polar Lipids, Inc.) or vehicle was injected (i.p.) 3 mg/kg body weight every 8 h for 24 h before imaging. To evaluate PP shedding, the mice were injected (i.p.) with W146 (3 mg/kg body weight) or vehicle and immediately visualized using MP-IVM. For FTY720 experiments, the same MKs were visualized in mice before and after a single injection of FTY720 (3 mg/kg i.p.) or DMSO using MP-IVM.

**PP shedding under shear stress.** MKs from β-actin–EGFP mice were seeded in μ-slides VI coated with 100 µg/ml of human fibrinogen (Sigma-Aldrich). The slides were then connected to a pump system (ibidi). A laminar shear stress of 4 dynes/cm² was applied to the cells in the presence of 5 µM S1P or vehicle. Image stacks were acquired at 2 µm in z to cover a 20-µm vertical distance at 60-s intervals for 20 min. The efficiency of PP fragmentation was determined by (L_{0 min} − L_{20 min})/L_{0 min} × 100%. L_{0 min} and L_{20 min} represent the length of PPs at 0 min and 20 min, respectively.

**Live cell imaging.** Mature MKs were starved in serum-free medium in 20 min MKs from V. Barocke, D. Braun, N. Urtz, C. Schulz, M.-L. von Brühl, A. Tirniceriu, F. Gaertner, L. Zhang, M. Orban, M. Lorenz, S. Massberg came up with the conception and study design and wrote the manuscript; L. Zhang, M. Orban, M. Lorenz, V. Barocke, D. Braun, N. Urtz, C. Schulz, M.-L. von Brühl, A. Tirniceriu, F. Gaertner, S.-S. Bolz, and A. Billich performed MP-IVM, generated chimaeras, generated lentiviral constructs, performed in vitro MK assays, generated F-deriv. MKs, and performed S1P measurements; R.L. Proia and T. Graf generated and provided mutant mice and helped with data interpretation; M. Prinz and A. Müller performed electron microscopy; E. Montagne and M. Sist performed in vitro shedding assay and examined S1p1 downstream signaling; L. von Baumbarten, R. Fässler, M. Sist, U.H. von Andrian, and T. Junt helped with MP-IVM, data interpretation, and discussion.

Submitted: 22 May 2012
Accepted: 5 October 2012

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