Serum Lipid Profiles and Cancer Risk in the Context of Obesity: Four Meta-Analyses

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Review Article

Serum Lipid Profiles and Cancer Risk in the Context of Obesity: Four Meta-Analyses

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The objective here was to summarize the evidence for, and quantify the link between, serum markers of lipid metabolism and risk of obesity-related cancers. PubMed and Embase were searched using predefined inclusion criteria to conduct meta-analyses on the association between serum levels of TG, TC, HDL, ApoA-I, and risk of 11 obesity-related cancers. Pooled relative risks (RRs) and 95% confidence intervals were estimated using random-effects analyses. 28 studies were included. Associations between abnormal lipid components and risk of obesity-related cancers when using clinical cutpoints (TC ≥ 6.50; TG ≥ 1.71; HDL ≤ 1.03; ApoA-I ≥ 1.05 mmol/L) were apparent in all models. RRs were 1.18 (95% CI: 1.08–1.29) for TC, 1.20 (1.07–1.35) for TG, 1.15 (1.01–1.32) for HDL, and 1.42 (1.17–1.74) for ApoA-I. High levels of TC and TG, as well as low levels of HDL and ApoA-I, were consistently associated with increased risk of obesity-related cancers. The modest RRs suggest serum lipids to be associated with the risk of cancer, but indicate it is likely that other markers of the metabolism and/or lifestyle factors may also be involved. Future intervention studies involving lifestyle modification would provide insight into the potential biological role of lipid metabolism in tumorigenesis.

1. Introduction

Obesity is a major worldwide problem, over 30% of adults in Western populations are obese, and there is growing evidence of the associated health risks associated [1–5]. The link between obesity and cancer risk has been studied extensively, but the results of individual studies do not suggest a consistent association [1, 6, 7]. Common cancers studied in the context of obesity include colorectal, breast, prostate, endometrial, pancreatic, liver, ovarian, kidney, gallbladder, leukaemia, and oesophageal cancers [4, 7–17].

The underlying mechanisms of action are not clear [1, 18–20]. A solid understanding could translate directly to patient benefit through implementation of therapeutic strategies to reduce cancer risk and mortality [19]. Assuming that the lipid metabolism plays a role in the biological processes driving the development of cancer, this could be easily modified by existing methods such as exercise, medication, or diet.
Increased physical activity levels improve cardiovascular and overall mortality in healthy populations [21–24]. Also, physical activity after cancer diagnosis is associated with improvements in cancer outcomes [25, 26] and metabolic markers such as cholesterol [27, 28]. As such, improvements in lipid levels through uptake of physical activity may also translate into improvements in cancer-specific survival. Experimental evidence largely suggests that statins, a commonly used drug to lower cholesterol levels, reduce cancer risk, though further trials are needed [29]. However, a large meta-analysis by the Cholesterol Treatment Trials’ Collaboration showed no statistically significant associations between statin use and cancer risk [30]. Nevertheless, due to heterogeneity of plasma lipid profiles in overweight and obese people, there may be some inconsistency in the associations for serum lipids and cancer risk [31]. Moreover, study populations were often small, insufficient information was collected (e.g., lack of BMI measurement, few lipid components measured), and timing of blood sampling in relation to diagnosis varied widely [32, 33]. In addition, it is thought that tissue types are influenced differently by lipid components and to varying degrees [2, 32].

With these meta-analyses, we aimed to summarize and quantify the evidence for the link between markers of lipid metabolism and risk of obesity-related cancers. We examined the associations between four components of the lipid profile measured in serum (total cholesterol (TC), triglycerides (TGs), high-density lipoprotein (HDL), and apolipoprotein A-I (ApoA-I)) and risk of cancers previously shown to be linked with obesity.

2. Methods

2.1. Literature Search Strategy. We used computerised literature search databases (PubMed search followed by an Embase search) to identify full text and abstracts published to date. Searches were conducted both with and without MeSH terms (“neoplasms/epidemiology,” “cancer,” “hyperlipidemias,” “lipoproteins, HDL,” “hypertriglyceride-mia,” “lipoproteins, apo A”) except for English language, human subjects, adults, and publications within the last 10 years no additional restrictions were added to the search. We also included “grey literature,” such as letters and abstracts presented in relevant conference meetings. All references of the selected articles were checked, including hand searches.

2.2. Inclusion Criteria. The final collection of selected studies was chosen based on the following criteria: the publication pertained to an epidemiological study (cohort or case-control studies), which measured the serum concentration of at least one of the selected lipid components (TC, TGs, HDL, and ApoA-I) prior to cancer diagnosis; the analytical methods were well described, with sufficient data available; the cancers included must have previously been linked to an increased risk associated with obesity. Those included were colorectal, breast, prostate, endometrial, pancreatic, liver, ovarian, kidney, gallbladder, leukaemia, and oesophageal cancer [7]. To include studies with large enough power, only those with at least 20 cancer cases were included. Initially, titles and abstracts of articles were reviewed in order to ascertain whether they potentially fit the inclusion criteria. If there was doubt over whether an article met the relevant criteria, it was subjected to a thorough assessment. After this first selection, all articles underwent detailed evaluations of the methods and results. Figure 1 illustrates the study exclusion process.

2.3. Data Extraction. The following details were recorded for each study: author, year of publication, country where the study was undertaken, serum lipid component levels (mmol/l), study type (case-control or cohort), cancer type, number of cases and total subjects for each level of the lipid component(s) measured. To allow for ease of comparison, all values in conventional units (mg/dL) were converted into SI units (mmol/l) using conversion factors [34].

2.4. Statistical Methods. The effect of each lipid component on cancer risk was evaluated by calculating the random effects summary relative risk to allow for possible heterogeneity between study results. The analyses were conducted for dichotomised values of TC, TGs, HDL, and apoA-I. The following clinical cutpoints were used: TC ≥ 6.50; TG ≥ 1.71; HDL ≤ 1.03; ApoA – 1 ≤ 1.05 mmol/L, all of which mirrored the NCEP and WHO guidelines as closely as practicable [35–39]. Some studies presented with dichotomised serum lipid levels, but for those that did not, participants from each study were divided into two groups based on their serum lipid level (“high” and “low”), and this mirrored the NCEP and WHO guidelines as closely as practicable. A first meta-analysis used all cancers from all studies. Potential heterogeneity of the study results was assessed with weighted forest plots, which display the relative risk estimates of cancer risk for each lipid component. Heterogeneity of the study results was also statistically evaluated using the Q-statistic as well as the I² statistic [40]. Sufficient data allowed for an individual analysis (using the methods described above) of prostate, colorectal, and breast cancers. Finally, we performed a metaregression to evaluate the effect of study design (i.e. prospective cohort versus case-control studies). No other potential confounders were included in the metaregression due to the nature in which the data was presented through the papers. For example, some papers acknowledge having data for fasting status of the individuals at time of sampling, but did not provide the actual data by individuals. Potential publication bias and effect modification (by country and year) were assessed using Begg’s Test and Egger’s funnel plot. All analyses were performed using STATA (version 11.2).

3. Results

The initial searches produced a total of 701 articles. 33 studies were selected for further evaluation based on information from abstracts. Of these, 17 were excluded and a further 12 were added from hand searches and references of included studies, giving a total of 28 studies used in the primary data analysis. Eight studies were conducted in Asia, 12 in Europe, and eight in the USA (Table 1). All studies reported on at least one of the four lipid components under investigation. The nine included cancers were studied in association with cancer
703 search results
47 potentially relevant studies

14 excluded based on abstract:
- Cancer risk not main outcome: 4
- Exposure not lipid components of interest: 7
- Review article: 2
- Study design: 1

33 potentially appropriate for inclusion

17 excluded based on full article:
- Cancer risk not main outcome: 3
- Exposure not lipid components of interest: 3
- Insufficient data included: 10
- Diseased study population: 1

3 studies included from references of included studies

8 studies included from hand searches

27 included in primary analysis
(20 for TC, 16 for TGs, 16 for HDL, and 7 for ApoA)*

*NB: some studies were used in more than one analysis.

For each lipid component studied we looked at the risk of developing cancer in those with abnormal versus normal levels. The random-effects analysis, comparing overall cancer risk and total cholesterol level, showed a pooled effects relative risk of 1.18 (95% CI 1.08–1.29) (Figure 2). The Q-statistic and $I^2$-statistic suggested heterogeneity ($Q = 250.02; \ df = 18; P = < 0.000; I^2 = 92.8\%$), which warranted the use of a random-effects model. The association between TGs and overall cancer risk resulted in a pooled relative risk of 1.20 (95% CI 1.07–1.35). The pooled relative risk was 1.15 (95% CI 1.01–1.32) when studying the association between HDL...
respectively.

We also conducted a stratified analysis by study type and found that the pooled RRs for case-control studies were slightly different (i.e., RR (95% CI) = 1.20 (0.90, 1.59) and found that the pooled RRs for case-control studies were 1.04 (95% CI: 0.98–1.10) and 1.08 (95% CI: 0.98–1.20) for case-control studies, respectively.

Finally, we also performed a meta-analysis specifically for TC and risk of prostate, breast, and colorectal cancer. The pooled relative risk for prostate cancer was 1.04 (95% CI: 0.98–1.10), whereas it was 1.08 (95% CI: 0.98–1.20) and 1.20 (95% CI: 0.44–3.26) for breast and colorectal cancers, respectively.

### 4. Discussion

These meta-analyses summarize the current evidence for a link between serum markers of lipid metabolism and risk of obesity-related cancers. All pooled models showed evidence for an association between abnormal lipid components and risk of obesity-related cancers when using clinical cutpoints.

The precise etiology of the link between obesity and risk of cancer has yet to be determined, but there has been growing evidence for a role of lipid metabolism in tumour development [18]. Apart from the studies listed on the link between serum lipids and cancer risk (Table 1), there is also preclinical evidence. For instance, it is thought that androgens stimulate prostate tumor growth via activation of androgen receptor-dependent pathways that regulate lipogenic gene expression, resulting in lipid accumulation [41]. Hyperlipidemia has also been shown to be involved in colorectal tumour development and initiation and progression of breast and prostate cancers [42–44]. Moreover, there is experimental evidence that fatty acid synthase (FAS), the enzyme that synthesizes fatty acids de novo,
is involved in tumorigenesis [45–47]. For example, prostate cancers overexpressing FAS display aggressive behavior, with the highest expression in patients with bony metastatic disease [47, 48]. In addition, nutritional studies showed that diets high in fat are linked to accelerated tumour growth and metastasis [42, 49]. Furthermore, cholesterol-lowering drugs such as statins have been shown to reduce the formation and spread of metastatic cancer cells [50, 51]. Finally, the immune system is thought to play a role in the link between HDL, ApoA-I, and tumorigenesis [52]. These lipid components decrease free proinflammatory cytokines such as tumour necrosis factor-α (TNF-α) which consequently reduces tissue damage, infiltration of macrophages and neutrophils, and attenuates tumour formation [53]. Therefore, low levels of HDL and Apo A-I may contribute to an inflammatory process linked to tumour biology.

Recent years have seen a multitude of reviews and meta-analyses comparing obesity and specific cancer risks; results varied widely depending on the type of cancer investigated, with relative risks ranging from 1.02 to 4.10 (breast cancer [11, 54–56], endometrial cancer [57], pancreatic [58–60], liver [17], prostate cancer [61], and colon and rectal cancer [15, 16]). As a result, our findings for an association between serum lipid components and risk of cancer also varied by type of cancer, as can be seen from our results for HDL (Figure 2), show that there may be stronger correlation between serum HDL levels and cancer risk, dependent on cancer type. Those results focusing on a single cancer showed more consistent results, suggesting that even among obesity-related cancers there may be a different association with serum lipid levels.

4.1. Strengths and Limitations of This Study. The greatest strength of this study is that we examined four different components of the lipid profile in relation to risk of developing cancer in the context of obesity. We also made all possible efforts to include all relevant available publications, including searching the two main online databases (PubMed and Embase). Additionally, our clearly defined objective criteria for exposure, outcome, and other study characteristics were specified a priori. There was no evidence of publication bias in these analyses.

A number of the studies subdivided levels of lipid components, but this was not performed consistently across the studies. Studies which had not dichotomised serum lipid levels from the outset were divided into two groups based on their serum lipid level (“high” and “low”) to mirror the NCEP and WHO guidelines as closely as practicable [38, 62]. This crude categorization may have compromised the accuracy and resulted in miscategorising of individuals, but given the rather small differences in cutoffs we do not believe that this has had a major impact on our analyses.

Heterogeneity among studies may also arise from different method of assessment of serum lipids. By performing random-effects analyses, we have taken into account between-study variation. Within-person variation is a likely interference with results as the one measurement taken may not be representative for a person’s average, or previous lipid levels. However, this variation will be present in all studies using a single measurement. In addition, adjustments made for confounding factors (e.g., gender or age) were not consistent across included studies and some sample sizes were relatively small or excluded one gender. Again, random effects analyses take into account this heterogeneity and in addition we included a metaregression analysis for study type.

In addition, the studies did not provide age-specific data, so it was not possible to conduct age-specific meta-analyses which presents us with a limitation. Persons younger than middle aged more rarely have abnormal lipid profiles and are also considerably less likely to be diagnosed with the cancers of interest than in those people aged over 50. Thus, this leaves our study population with a relatively low probability of having both sufficient exposure and number of cases in the lower age range. We do not believe that this will have had a major effect on our results, although it is worth considering that this may have diluted the strength of our findings somewhat.

Due to the information provided in the included studies, we had no means to adjust our analyses for cancer screening practices. Undoubtedly, these practices vary around the world and thus the differences could lead to the introduction of detection bias.

Finally, the analyses of the three individual cancers (prostate, breasts and colorectal cancers) did not produce statistically significant relative risks, which most likely follows from a lack of power due to the limited number of studies available for inclusion. Future research, with larger sample sizes, repeated measurements, and consistent adjustments for confounding could provide information to inform a more reliable estimate of links between serum lipid components and cancer risk.

4.2. Conclusions. Abnormal levels of all lipid components studied were statistically significantly associated with an increased risk of obesity-related cancers, with the strongest association for serum ApoA-I. Despite a suggestion for a link between the lipid metabolism and risk of cancer, the magnitude of the pooled relative risk was relatively small. This may be because the studied lipid components are markers of obesity or because they are markers of other lifestyle factors potentially associated with tumorigenesis.
### Table 1: Details of included studies.

<table>
<thead>
<tr>
<th>Author (y)</th>
<th>Country</th>
<th>Cases/controls</th>
<th>Study Type</th>
<th>Cancer(s) included</th>
<th>Mean age (SE)/age range</th>
<th>Timing of measurement</th>
<th>Measurement method</th>
<th>Cholesterol measure</th>
<th>Adjusted for</th>
</tr>
</thead>
<tbody>
<tr>
<td>Inoue et al. (2009) [63]</td>
<td>Japan</td>
<td>1263/26461</td>
<td>Cohort</td>
<td>Colon/rectum and breast</td>
<td>56.0 (8.2)</td>
<td>Fasting and nonfasting</td>
<td>NS</td>
<td>TGs, HDL</td>
<td>Adjusted for age, study area, smoking status, weekly ethanol intake, and total serum cholesterol</td>
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<tr>
<td>Iso et al. (2009) [64]</td>
<td>Japan</td>
<td>1093/32275</td>
<td>Cohort</td>
<td>Colorectal, prostate, leukemia, breast and cervical</td>
<td>54.0</td>
<td>Fasting and nonfasting</td>
<td>Enzymatic</td>
<td>TC</td>
<td>Age, BMI, pack year of smoking, ethanol intake, hypertension, diabetes, hyperlipidemia medication use, total vegetable intake, coffee intake, and public health centre</td>
</tr>
<tr>
<td>Kitahara et al. (2011) [65]</td>
<td>Korea</td>
<td>44935/1144784</td>
<td>Cohort</td>
<td>Oesophagus, colon/rectum, pancreas, prostate, kidney, and gallbladder</td>
<td>30–95</td>
<td>Fasting</td>
<td>NS</td>
<td>TC</td>
<td>Cigarette smoking, alcohol consumption, BMI, physical activity, hypertension, and fasting serum glucose</td>
</tr>
<tr>
<td>Mainous et al. (2005) [66]</td>
<td>USA</td>
<td>203/3075</td>
<td>Cohort</td>
<td>Cancer</td>
<td>30+</td>
<td>12 hr fasting</td>
<td>NS</td>
<td>HDL</td>
<td>Age, gender, smoking status, and BMI</td>
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<td>Melvin et al. (2012) [67]</td>
<td>Sweden</td>
<td>6871/227603</td>
<td>Cohort</td>
<td>Breast and ovarian</td>
<td>25+</td>
<td>Fasting and nonfasting</td>
<td>Enzymatic and immunoturbidimetric</td>
<td>TC, TGs, HDL, ApoA</td>
<td>Glucose, TGs, TC, age, parity fasting status, and SES</td>
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<td>Chang et al. (2007) [68]</td>
<td>Turkey</td>
<td>150/71</td>
<td>Case</td>
<td>Breast</td>
<td>49.2 (11.8)</td>
<td>12 hr fasting</td>
<td>Enzymatic</td>
<td>TC, TGs, HDL, ApoA</td>
<td>HDL, apoA-I, apoB, ApoA-I/ApoB ratio, and VLDL</td>
</tr>
<tr>
<td>Furberg et al. (2004) [69]</td>
<td>Norway</td>
<td>1287/27912</td>
<td>Cohort</td>
<td>Breast</td>
<td>43.6 (0.1)</td>
<td>Nonfasting</td>
<td>Enzymatic</td>
<td>HDL</td>
<td>Age, country of residence, parity, height, TC, recreational and occupation activity. Some models also included blood pressure, BMI, TGs, age at first birth, time since last meal, dietary energy, and fat intake</td>
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<tr>
<td>Ha et al. (2009) [70]</td>
<td>Korea</td>
<td>714/169660</td>
<td>Cohort</td>
<td>Breast</td>
<td>55.9 (5.0)</td>
<td>Fasting</td>
<td>NS</td>
<td>TC</td>
<td>Age and BMI</td>
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<td>Hayashi et al. (2012) [71]</td>
<td>Japan</td>
<td>377/528</td>
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<td>&gt;60</td>
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<td>NS</td>
<td>TGs</td>
<td>Age, PSA level, prostatic volume, BMI, and TGs level</td>
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<td>Author(s)</td>
<td>Country</td>
<td>Cases/controls</td>
<td>Study Type</td>
<td>Cancer(s) included</td>
<td>Mean age (SE)/age range</td>
<td>Timing of measurement</td>
<td>Measurement method</td>
<td>Cholesterol measure</td>
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<td>Kaye et al. (2002)</td>
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<td>158/725</td>
<td>Case</td>
<td>Breast</td>
<td>50–79</td>
<td>NS</td>
<td>NS</td>
<td>TC</td>
<td>HDL, age, family history of breast cancer, age at menarche, age at the first full-term pregnancy, and total cholesterol.</td>
</tr>
<tr>
<td>Kim et al. (2009)</td>
<td>Korea</td>
<td>690/1380</td>
<td>Case</td>
<td>Breast</td>
<td>48.5 (7.0)</td>
<td>8 hr fasting</td>
<td>Enzymatic</td>
<td>TC, TGs, HDL, TC, TGs, HDL, Total cholesterol, Age, other lipids, BMI, and smoking.</td>
<td></td>
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<tr>
<td>Lindemann et al. (2009)</td>
<td>Norway</td>
<td>100/31273</td>
<td>Cohort</td>
<td>Endometrial</td>
<td>56.1</td>
<td>NS</td>
<td>Nonfasting</td>
<td>Enzymatic</td>
<td>HDL, age, family history of breast cancer, age at menarche, and total cholesterol.</td>
</tr>
<tr>
<td>Cast et al. (2007)</td>
<td>France</td>
<td>286/546</td>
<td>Case</td>
<td>Endometrial</td>
<td>56.9</td>
<td>8 hr fasting</td>
<td>Enzymatic</td>
<td>TC, TGs, HDL, TC, TGs, HDL, Total cholesterol, Age, other lipids, BMI, and smoking.</td>
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<tr>
<td>Seth et al. (2012)</td>
<td>Sweden</td>
<td>242/288</td>
<td>Cohort</td>
<td>Endometrial</td>
<td>25.0</td>
<td>Nonfasting</td>
<td>Enzymatic and immunoturbidimetric</td>
<td>TC, TGs, HDL, Total cholesterol, Age, other lipids, BMI, and smoking.</td>
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<td>Maguire et al. (2008)</td>
<td>USA</td>
<td>312/319</td>
<td>Case</td>
<td>Prostate</td>
<td>50–74</td>
<td>NS</td>
<td>NS</td>
<td>Enzymatic</td>
<td>Age, family history of prostate cancer, BMI, total cholesterol, smoking status, intake of meat, dairy, tomato products and alcohol, and use of diabetes medications.</td>
</tr>
<tr>
<td>Platz et al. (2009)</td>
<td>USA</td>
<td>1251/4335</td>
<td>Cohort</td>
<td>Prostate</td>
<td>63.1</td>
<td>NS</td>
<td>Nonfasting</td>
<td>Enzymatic</td>
<td>Age, race, first-degree family history of prostate cancer, BMI, history of total cholesterol, smoking status, intake of meat, dairy, tomato products and alcohol, and use of diabetes medications.</td>
</tr>
<tr>
<td>Author (y)</td>
<td>Country</td>
<td>Cases/controls</td>
<td>Study Type</td>
<td>Cancer(s) included</td>
<td>Mean age (SE)/age range</td>
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<td>Van Hemelrijcket et al. (2011) [81]</td>
<td>Sweden</td>
<td>2008/65719</td>
<td>Cohort</td>
<td>Prostate</td>
<td>35+</td>
<td>Fasting and nonfasting</td>
<td>Enzymatic and immunoturbidimetric</td>
<td>HDL, ApoA</td>
<td>Age, glucose, TGs, and TC, fasting status, and SES.</td>
</tr>
<tr>
<td>Van Hemelrijcket et al. (2011) [82]</td>
<td>Sweden</td>
<td>5112/195548</td>
<td>Cohort</td>
<td>Prostate</td>
<td>45–75</td>
<td>Fasting and nonfasting</td>
<td>Enzymatic and immunoturbidimetric</td>
<td>TC, TGs</td>
<td>Glucose and/or TGs and/or TC, SES, fasting status, and time between measurements and entry</td>
</tr>
<tr>
<td>Bravi et al. (2005) [83]</td>
<td>Italy</td>
<td>1294/1451</td>
<td>Case</td>
<td>Prostate</td>
<td>46–74</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>Age, centre, education, BMI, physical activity, tobacco smoking, alcohol consumption, and family history of prostate cancer</td>
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<td>Tsushima et al. (2005) [84]</td>
<td>USA</td>
<td>1004/21255</td>
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<td>Colorectal</td>
<td>NS</td>
<td>Nonfasting</td>
<td>NS</td>
<td>NS</td>
<td>Glucose and/or TGs and/or TC, SES, fasting status, and time between measurements and entry</td>
</tr>
<tr>
<td>Van Duijnhoven et al. (2011) [85]</td>
<td>W. Europe</td>
<td>939/939</td>
<td>Cohort</td>
<td>Colorectal</td>
<td>35–70</td>
<td>Fasting and nonfasting</td>
<td>Enzymatic</td>
<td>TC, TGs, HDL, ApoA</td>
<td>Age, centre, education, BMI, height, weight, smoking habits, physical activity, education, consumption of fruit, vegetables, meat, fish, and alcohol, intake of fibre, energy from fat, and energy from nonfat</td>
</tr>
<tr>
<td>Chung et al. (2006) [86]</td>
<td>Korea</td>
<td>105/105</td>
<td>Case</td>
<td>Colorectal</td>
<td>58.6 (8.3)</td>
<td>12 hr fasting</td>
<td>Enzymatic</td>
<td>TC, TGs</td>
<td>Age, gender, BMI, glucose, triglycerides, and total cholesterol</td>
</tr>
<tr>
<td>Van Hemelrijcket et al. (2012) [87]</td>
<td>Sweden</td>
<td>156/82391</td>
<td>Cohort</td>
<td>Kidney</td>
<td>20+</td>
<td>Fasting and nonfasting</td>
<td>Enzymatic and immunoturbidimetric</td>
<td>TC, TGs, HDL, ApoA</td>
<td>Age, gender, glucose, TGs and TC, creatinine levels, fasting status, and SES</td>
</tr>
<tr>
<td>Andreotti et al. (2008) [88]</td>
<td>USA</td>
<td>264/1839</td>
<td>Case</td>
<td>Biliary</td>
<td>34–75</td>
<td>Overnight fasting</td>
<td>NS</td>
<td>NS</td>
<td>Age, gender, BMI, waist-to-hip ratio, cigarette smoking, alcohol drinking, hypertension, diabetes, and gallstone status</td>
</tr>
<tr>
<td>Asano et al. (2008) [89]</td>
<td>Japan</td>
<td>97/2507</td>
<td>Cohort</td>
<td>Gastric</td>
<td>59.2 (0.3)</td>
<td>Fasting and nonfasting</td>
<td>Enzymatic</td>
<td>TC</td>
<td>Age and gender</td>
</tr>
</tbody>
</table>

NS: not specified; BMI: body mass index; TC: total cholesterol; TGs: triglycerides; HDL: high-density lipoprotein; ApoA: apolipoprotein A-I; ApoB: apolipoprotein B; VLDL: very low-density lipoprotein.
Since lipid components are easily modified through lifestyle interventions such as diet or exercise, research into serum lipid components and cancer risk presents a prime opportunity for intervention studies to help provide the desired insight into their biological role.

Abbreviations

(TC): Total cholesterol  
(TGs): Triglycerides  
(HDL): High-density lipoprotein  
(ApoA-I): Apolipoprotein A-I  
(BMI): Body mass index.

Conflicts of Interests

The authors declare that they have no potential conflict of interests to disclose.

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References


