Absorption of Manganese and Iron in a Mouse Model of Hemochromatosis

Citation

Published Version
doi:10.1371/journal.pone.0064944

Permanent link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:11377981

Terms of Use
This article was downloaded from Harvard University’s DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

Share Your Story
The Harvard community has made this article openly available. Please share how this access benefits you. Submit a story.

Accessibility
Absorption of Manganese and Iron in a Mouse Model of Hemochromatosis

Jonghan Kim1,2*, Peter D. Buckett2, Marianne Wessling-Resnick2*
1 Department of Pharmaceutical Sciences, Northeastern University, Boston, Massachusetts, United States of America, 2 Department of Genetics and Complex Diseases, Harvard School of Public Health, Boston, Massachusetts, United States of America

Abstract

Hereditary hemochromatosis, an iron overload disease associated with excessive intestinal iron absorption, is commonly caused by loss of HFE gene function. Both iron and manganese absorption are regulated by iron status, but the relationships between the transport pathways of these metals and how they are affected by HFE-associated hemochromatosis remain poorly understood. Loss of HFE function is known to alter the intestinal expression of DMT1 (divalent metal transporter-1) and Fpn (ferroportin), transporters that have been implicated in absorption of both iron and manganese. Although the influence of HFE deficiency on dietary iron absorption has been characterized, potential effects on manganese metabolism have yet to be explored. To investigate the role of HFE in manganese absorption, we characterized the uptake and distribution of the metal in Hfe−/− knockout mice after intravenous, intragastric, and intranasal administration of 54Mn. These values were compared to intravenous and intragastric administration of 59Fe. Intestinal absorption of 59Fe was increased and clearance of injected 59Fe was also increased in Hfe−/− mice compared to controls. Hfe−/− mice displayed greater intestinal absorption of 54Mn compared to wild-type Hfe+/+ control mice. After intravenous injection, the distribution of 59Fe to heart and liver was greater in Hfe−/− mice but no remarkable differences were observed for 54Mn. Although olfactory absorption of 54Mn into blood was unchanged in Hfe−/− mice, higher levels of intranasally-instilled 54Mn were associated with Hfe−/− brain compared to controls. These results show that manganese transport and metabolism can be modified by HFE deficiency.

Introduction

High levels of manganese impair neurobehavior in both humans and animal models [1–5]. Fortunately, manganese loading due to ingestion is relatively rare since hepatic first-pass elimination of the metal provides a protective mechanism against toxicity [6]. However, intake of airborne manganese bypasses the biliary excretion route and inhaled manganese is efficiently transported into the body including the brain through the nasal epithelium [7–9]. High levels of airborne manganese are common in occupational settings of mining, manganese ore processing, dry battery manufacture and organochemical fungicide use [10,11], raising concerns about public and occupational health problems. Recent work by Haynes et al. [12] determined hair and blood manganese levels in residents living near Marietta OH and a ferromanganese refinery that is a major US airborne emission source. The relationship between these biomarkers and ambient air levels of manganese became significant when iron metabolism genes, including HFE (hyperferremia) alleles, were incorporated in their models [12]. Our group recently uncovered a relationship between HFE status and manganese metabolism by demonstrating that Hfe−/− knockout mice have reduced levels of blood manganese [13]. This observation validated an epidemiological study of demonstrating human carriers of disease-associated HFE(C282Y) or HFE(H63D) alleles also have lower blood manganese [13]. The HFE(C282Y) and HFE(H63D) variants in the iron regulatory HFE gene are the leading cause of adult onset hereditary hemochromatosis (HH), one of the most common genetic diseases in the North American Caucasian population. C282Y and H63D have prevalence in North American populations of 7–17% and 10–32%, respectively [14]. HFE-associated HH is the most common Mendelian inherited trait of northern Europeans, with a prevalence of 1:200 to 1:500 [15,16]. Defects in the HFE gene promote increased intestinal iron absorption and progressive tissue deposition of the metal resulting in liver damage and disease, congestive heart failure, and premature death. Mice with either the orthologous mutations or null allele display the same iron-loading HH phenotype observed in humans [17].

The effects of iron loading on manganese in vivo have been well established [13,18,19]. Recent molecular studies have documented a role for divalent metal transporter-1 (DMT1) in manganese uptake [20–23]. DMT1 functions in dietary iron absorption across the apical surface of the intestinal mucosa [24,25] and transports iron from endocytosed transferrin to enable heme synthesis by erythroid cells [26]. Because impaired DMT1 function also results in reduced manganese transport [20], the transporter appears to play an important physiological role in the metabolism of this
metal as well. Emerging new evidence indicates that the iron exporter ferroportin (Fpn) [27,29] also transports manganese [29,30]. There is strong evidence in the literature that HFE deficiency alters levels of both transporters [31–33]. However, while the influence of HFE deficiency on dietary iron absorption has been characterized, its potential effects on manganese metabolism have not been explored. Therefore, we undertook this investigation to characterize the uptake and distribution of the metal in Hfe<sup>−/−</sup> knockout mice after intravenous, intragastric, and intranasal administration of 54Mn.

**Results and Discussion**

**Iron loading characteristics of HFE deficiency**

Several studies have characterized the iron-loading phenotype of Hfe knockout (Hfe<sup>−/−</sup>) mice [17,34–36]. In our hands, Hfe<sup>−/−</sup> mice also displayed an age-dependent increase in liver non-heme iron levels, which were elevated as early as 4 weeks of age (Figure 1A; P<0.001; n = 9–14 per group). These data are consistent with findings by other investigators [37,38]. To characterize the effect of dietary iron on the iron-loading phenotype, weaning mice were fed iron deficient (5 mg/kg), control (50 mg/kg) or high iron (20,000 mg/kg) diet. Non-heme iron levels in both liver and serum increased in a manner corresponding to dietary iron content regardless of genotype (Figure 1B and C). Liver non-heme iron levels were greater in Hfe<sup>−/−</sup> mice than in wild-type control (Hfe<sup>+/+</sup>) mice fed control diet (Figure 1B; P<0.001; n = 8–12 per group), while the other two diets did not show significant differences between the two strains. Similarly, Hfe<sup>−/−</sup> mice fed the control diet displayed significantly higher serum iron concentrations compared with Hfe<sup>+/+</sup> mice (Figure 1C; P<0.001; n = 8–12 per group). It is notable that this pattern was also observed in mice fed iron-deficient diet and high iron diet (Figure 1C; P<0.001 and P = 0.042, respectively; n = 6–8 per group). In contrast to results reported by others [39,40], Hfe<sup>−/−</sup> and Hfe<sup>+/+</sup> mice fed a high iron diet had similar liver non-heme iron levels, but this difference could be due to strain variation (129/SvJ vs C57BL/6) or the different duration of dietary iron (2–6 weeks) [39–41]. To control for the influence of body iron status on the metal uptake studies described below, both Hfe<sup>−/−</sup> and Hfe<sup>+/+</sup> mice were fed the control diet containing 50 mg/kg iron for 5 weeks after the time of weaning.

**HFE deficiency enhances intestinal uptake of 59Fe and clearance of 59Fe from circulation**

To examine the influence of HFE deficiency on intestinal iron uptake, 59Fe was administered to Hfe<sup>−/−</sup> and Hfe<sup>+/+</sup> mice by intragastric gavage. Iron was reduced to the ferrous form using freshly dissolved ascorbate immediately prior to instillation. Blood levels of 59Fe were determined 4 h after administration (Figure 2A). Hfe<sup>−/−</sup> mice accumulated a greater amount of iron in blood over the 4-hour period compared to Hfe<sup>+/+</sup> wild-type controls (P = 0.034; n = 4–5 per group). This observation is consistent with the hyperabsorption of iron from the gut of Hfe<sup>−/−</sup> mice [36,37,42]. However, we considered the additional possibility that increased appearance of 59Fe in the blood after gavage could be due to decreased clearance. Thus, we also characterized blood clearance following intravenous injection of 59Fe (Figure 2B). Four hours after intravenous injection, the amount of 59Fe in the blood was less in Hfe<sup>−/−</sup> compared to Hfe<sup>+/+</sup> mice (P = 0.008; n = 5 per group), indicating that loss of HFE function contributes to enhanced uptake of iron by peripheral tissues and/or excretion from the body. The accelerated blood clearance promoted by HFE deficiency could be explained by an increased fraction of non-transferrin-bound iron, which displays faster clearance kinetics than transferrin-bound iron [43]. Trinder et al [39] have shown that 59FeTt uptake is similar in Hfe knockout and wild-type mice. Despite more rapid clearance of injected 59Fe, blood 59Fe levels after gavage were still greater in Hfe<sup>−/−</sup> mice (Figure 2A). These combined data suggest an even larger extent of “intrinsic” intestinal uptake (bioavailability) of iron in the absence of HFE. The calculated bioavailability after correcting for blood clearance of iron during absorption over the 4 hour time period was 3-fold greater in Hfe<sup>−/−</sup> mice compared with Hfe<sup>+/+</sup> mice (Table 1, Figure 1. **HFE deficiency and iron-loading.** Weaning mice were fed facility chow (220 mg/kg iron), euthanized with isoflurane overdose at the indicated ages. Liver non-heme iron levels (A) were determined for Hfe<sup>+/+</sup> (empty bar) and Hfe<sup>−/−</sup> (closed bar) mice. Data were presented as means ± SEM (n = 6–12 per group). * P<0.05 between Hfe<sup>+/+</sup> and Hfe<sup>−/−</sup> mice (Table 1, Figure 2A). Four hours after intravenous injection, the amount of 59Fe in the blood was less in Hfe<sup>−/−</sup> compared to Hfe<sup>+/+</sup> mice (P = 0.008; n = 5 per group), indicating that loss of HFE function contributes to enhanced uptake of iron by peripheral tissues and/or excretion from the body. The accelerated blood clearance promoted by HFE deficiency could be explained by an increased fraction of non-transferrin-bound iron, which displays faster clearance kinetics than transferrin-bound iron [43]. Trinder et al [39] have shown that 59FeTt uptake is similar in Hfe knockout and wild-type mice. Despite more rapid clearance of injected 59Fe, blood 59Fe levels after gavage were still greater in Hfe<sup>−/−</sup> mice (Figure 2A). These combined data suggest an even larger extent of “intrinsic” intestinal uptake (bioavailability) of iron in the absence of HFE. The calculated bioavailability after correcting for blood clearance of iron during absorption over the 4 hour time period was 3-fold greater in Hfe<sup>−/−</sup> mice compared with Hfe<sup>+/+</sup> mice (Table 1,
HFE deficiency promotes delivery of $^{59}$Fe to liver and heart

To further examine the influence of HFE on clearance of iron, mice were euthanized 4 h after the injected dose and radioactivity in tissue samples was measured to determine the degree of $^{59}$Fe uptake (Figure 3A). $^{59}$Fe was observed as a 1.4-fold increase in $^{59}$Fe uptake in the liver compared with Hfe+/+ mice ($P<0.003; n = 5 per group$). Moreover, $^{59}$Fe in the heart after intravenous injection was 0.5-fold greater in Hfe+/+ mice than in Hfe−/− mice ($P=0.033; n = 5 per group$). Thus, loss of HFE function results in increased tissue uptake to the liver and heart. These observations are of interest since the major causes of death related to iron overload hemochromatosis are liver toxicity and cardiomyopathy [44]. Previous studies by Ajioka et al. [37] reported increased liver distribution of $^{59}$Fe after gavage. The idea that loss of HFE function leads to hepatic iron loading is consistent with other studies [17,34–36]. Turosz et al. [45] investigated iron loading of cardiac tissue in Hfe−/− mice and showed increased iron deposition and reactive oxygen species (ROS). These data support the notion that preferential uptake of iron into these two major organs is associated with metal-related oxidative stress and tissue damage. While the presence of increased non-transferrin bound iron could promote these effects [43], it is possible that differential expression of certain iron transporters contributes to the specific uptake of $^{59}$Fe; for example, Zip14 in liver [46]. In our experiments, the total radioactivity remaining in the carcass was similar in the two groups (data not shown), suggesting that excretion of iron cannot account for increased blood clearance in HFE deficiency. We conclude that the deposition of iron into key target tissues, particularly the liver, is promoted by HFE deficiency.

Figure 3. Uptake of injected $^{59}$Fe and $^{54}$Mn to brain, heart and liver. Levels of $^{59}$Fe in brain, heart and liver (as nCi/g tissue) were characterized 4 h post-dose of $^{59}$FeCl3 (200 μCi/kg body weight) to mice by intravenous injection (A). Levels of $^{54}$Mn in brain, heart and liver (as nCi/g tissue) were characterized 1 h post-dose of $^{54}$MnCl2 (200 μCi/kg body weight) to mice by intravenous injection (B). Empty and closed bars represent Hfe+/+ and Hfe−/− mice, respectively. Data were presented as mean ± SEM (n = 4–5 per group). * P<0.05 between Hfe+/+ and Hfe−/− mice determined by two-sample t-test.

doi:10.1371/journal.pone.0064944.g003

Table 1. Intestinal bioavailability of iron and manganese.

<table>
<thead>
<tr>
<th></th>
<th>Wild-type (%)</th>
<th>HFE-deficient (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>$^{59}$Fe</td>
<td>19.0 ± 4.2</td>
<td>81.1* ± 7.3</td>
</tr>
<tr>
<td>$^{54}$Mn</td>
<td>2.3 ± 0.2</td>
<td>3.6* ± 0.4</td>
</tr>
</tbody>
</table>

Mice were euthanized after intravenous injection or intragastric gavage of the indicated isotopes (200 μCi/kg body weight). Blood and tissues were collected 4-hour after $^{59}$Fe dose or 1-hour after $^{54}$Mn dose. Blood concentration after intragastric gavage was divided by the concentration determined after intravenous injection to estimate time-variant intestinal bioavailability of each metal. Data are presented as the mean ± SEM.

$^*$ P<0.05 between Hfe+/+ and Hfe−/− mice determined by two-sample t-test.

doi:10.1371/journal.pone.0064944.t001
Loss of HFE function increases intestinal manganese uptake

To determine the role of HFE in manganese absorption, we also measured the amount of $^{54}$Mn in blood after intragastric gavage or intravenous injection of the radioisotope (Figure 2). $\text{Hfe}^{-/-}$ mice displayed higher blood $^{54}$Mn levels than $\text{Hfe}^{+/+}$ mice 1 h after gavage (Figure 2A; $P = 0.036; n = 4$ per group), indicating that like iron, intestinal absorption of manganese is enhanced by HFE deficiency (Table 1). Unlike iron, there was no difference in blood clearance of $^{54}$Mn administered by injection (Figure 2B; $P = 0.488; n = 7–8$ per group). Tissue distribution of $^{54}$Mn was similar between $\text{Hfe}^{-/-}$ and $\text{Hfe}^{+/+}$ mice with no difference in uptake by heart or liver (Figure 3B). The observations that manganese clearance from circulation is not altered despite high serum iron and accelerated iron clearance in HFE deficiency (Figure 2B) is interesting. It is possible there are potential differences in the kinetics of manganese clearance from circulation by both $\text{Hfe}^{+/+}$ and $\text{Hfe}^{-/-}$ mice earlier than the 1 h time point of our tracer study. We chose 1 h for $^{54}$Mn sampling since manganese absorption reached a plateau level in the brain 1 h following intranasal instillation of $^{54}$Mn and we therefore monitored blood clearance over the same time frame (see below). Further study is necessary to more fully characterize manganese from circulation in $\text{Hfe}^{-/-}$ mice, including the effects of continuous administration and chronic inhalation of manganese [9,47,48].

**Figure 4. Effect of HFE deficiency on manganese uptake after intranasal instillation.** One hour after intranasal instillation of $^{54}$MnCl$_2$ (200 μCi/kg), blood (A) and brain (B) samples were weighed and radioactivity was measured to calculate isotope level as nCi/g blood or brain. Empty and closed bars represent $\text{Hfe}^{+/+}$ and $\text{Hfe}^{-/-}$ mice, respectively. Data were presented as means ± SEM (n=7–8 per group). * $P<0.05$ between $\text{Hfe}^{+/+}$ and $\text{Hfe}^{-/-}$ mice determined by two-sample t-test.
doi:10.1371/journal.pone.0064944.g004

Olfactory manganese uptake into the brain is enhanced by HFE deficiency

Absorption of airborne manganese is an increasing concern due to neurotoxicity of the metal [49]. To study the influence of HFE deficiency on olfactory absorption, uptake of $^{54}$MnCl$_2$ was examined in $\text{Hfe}^{-/-}$ and $\text{Hfe}^{+/+}$ mice after intranasal instillation. Preliminary studies demonstrated that $^{54}$Mn absorption to the brain was maximal 1 h post-instillation. Levels of DMT1 (A) and Fpn (B) were determined by western blot. Data were presented as means ± SEM (n=6 per group). Olfactory bulbs from 3 male and 5 female $\text{Hfe}^{+/+}$ mice and 4 male and 4 female $\text{Hfe}^{-/-}$ mice were pooled and determined for iron and manganese levels by ICP-MS (C). Data were presented as means.
doi:10.1371/journal.pone.0064944.g005

**Figure 5. Levels of transporters and metals in olfactory bulbs of wild-type and HFE knockout mice.** Weanling mice were fed control diet and euthanized to collect olfactory bulbs. Levels of DMT1 (A) and Fpn (B) were determined by western blot. Data were presented as means ± SEM (n=6 per group). DMT1 levels of $\text{Hfe}^{-/-}$ mice determined by 2-sample t-test.
doi:10.1371/journal.pone.0064944.g005

**Manganese Transport in HFE Deficiency**
confirming that inhalation is a critical route of manganese intoxication, especially in occupational settings [5,6]. Moreover, our data show that uptake to the brain after intranasal instillation was increased in Hfe<sup>-/-</sup> mice compared with Hfe<sup>+/+</sup> mice (Figure 4B; P = 0.028; n = 7–8 per group). This effect appears to be specific for brain uptake by the olfactory pathway since no differences were detected between the groups after intragastric gavage or intravenous injection (Figure 2 and 3B). Notably, the uptake of manganese (Figure 3B) and iron (Figure 3A) across the blood-brain barrier in HFE deficiency is unaffected. Combined, these observations suggest different mechanism(s) of metal uptake between olfactory and vascular transport to the brain. The latter may be primarily mediated by transferrin. Nonetheless, the observed increase in uptake to the brain of Hfe<sup>-/-</sup> mice suggests that individuals with HFE-associated hemochromatosis may be more vulnerable to neurotoxicity of inhaled manganese.

**DMT1 and Fpn expression in olfactory bulb**

It is known that increased absorption of iron across the intestinal mucosa is promoted by up-regulation of DMT1 and Fpn [24,28,50]. Both transporters are post-transcriptionally regulated by iron-responsive elements [51,52]. In the case of HFE deficiency, post-translational modification also is imparted by the iron regulatory hormone hepcidin [53,54], which is inappropriately down-regulated to enhance Fpn levels [55,56]. We have previously found that systemic iron deficiency induces DMT1 up-regulation in the olfactory epithelium [20] and olfactory bulbs [57], an effect that is associated with increased olfactory transport of manganese to the brain [20,21]. Therefore, we compared DMT1 and ferroportin levels in the olfactory bulbs from Hfe<sup>-/-</sup> with Hfe<sup>+/+</sup> mice (Figure 5A and B). Despite the iron overload status of Hfe<sup>-/-</sup> mice (Figure 1), levels of both transporters were similar in the two groups. However, ICP-MS analysis of the metal content of the olfactory bulbs from wild-type and knockout mice showed no differences that would otherwise promote changes in the expression pattern of the transporters at the tissue level (Figure 5C). Levels of iron were 50-fold greater than manganese. These results indicate that there could be other factors independent of DMT1 or Fpn involved in manganese transport across nasal cavity into brain that are affected by the loss of HFE function. For example, local regulation of hepcidin may play an important role in metal uptake in the brain [58]. Other transport pathways could also influence brain uptake of airborne manganese, including newly emerging roles for SLC30A10 transporters [59,60] and manganese binding proteins like calprotectin [61]. Further work is necessary to define the manganese pathways affected in Hfe<sup>-/-</sup> mice and the functional consequences due to the increased uptake of manganese from the nasal cavity to the brain.

**Conclusions**

- DMT1 and Fpn are expressed in olfactory bulb, however, levels of both transporters were similar in HFE-deficient and wild-type mice.
- HFE deficiency was not associated with altered levels of iron or manganese in olfactory bulbs; levels of iron were 50-fold greater than manganese.
- Loss of HFE increases uptake of manganese to the brain by the olfactory pathway, suggesting individuals with HFE-associated hemochromatosis may be at greater risk for inhalation exposures.

**Materials and Methods**

**Ethics statement**

This study was performed in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. The protocol was approved by the Harvard Medical Area Animal Care and Use Committee (Animal Experimentation Protocol AEP #04545 and #04692).

**Animal care and procedures**

HFE-deficient (Hfe<sup>-/-</sup>) mice and wild-type control (Hfe<sup>+/+</sup>) mice were kindly provided by Dr. Nancy Andrews (Duke University, NC). All mice used for these studies were on the 129S6/SvEvTac background. Weanling mice were fed a diet containing 50 mg iron/kg (as ferric citrate, TD07800, Harlan Teklad) for 5 weeks. For experiments testing the effect of dietary iron, groups of knockout and wild-type mice were also placed on iron-deficient diet (5 mg iron/kg diet; TD99397), or iron-overload diet (20,000 mg/kg; as carbonyl iron, TD08714). Mice were euthanized under isoflurane inhalation (5%) to obtain serum and liver, and non-heme iron concentrations in these tissues were determined by spectrophotometric methods [62,63]. All experiments were carried out between 12-3 p.m. to avoid circadian effects on iron metabolism. No obvious gender differences were noted in the transport studies described below.

To characterize gastrointestinal absorption of iron, 8-week-old mice were fasted for 4 hours, anesthetized with up to 2% vaporized isoflurane, and administered 59FeCl<sub>3</sub> (200 μCi/kg body weight) or 54MnCl<sub>2</sub> (20 μCi/kg body weight) by intragastric gavage using a 20 gauge 1.5-inch gavage needle. 59Fe was diluted in Tris-buffered saline containing ascorbic acid (10 mM) at 1.5 mL/kg immediately prior to administration; 54Mn was diluted in phosphate-buffered saline (PBS). Mice were euthanized by isoflurane overdose 4 h post-gavage to collect blood via the inferior vena cava and tissues were dissected. Radioactivity was determined using a WIZARD 1410 gamma counter (Perkin Elmer). Data were analyzed and expressed as nCi/g tissue. Since the blood levels of 59Fe represent the amount absorbed from the gut as well as the amount cleared from the circulation, similar cohorts of mice were intravenously injected with the same dose of 59Fe or 54Mn via the tail vein to account for the contribution of clearance to the blood level. To study intranasal absorption, 54Mn was instilled into the right nostril at 0.02 mL/kg using a thin polyurethane catheter via the inferior nasal cavity. As noted in the transport studies described below.

**Western blots**

Olfactory bulbs from male Hfe<sup>+/+</sup> and Hfe<sup>-/-</sup> mice were homogenized in 10 mM Tris, pH 7.4, 150 mM NaCl, 1.0 mM...
EDTA, 0.1% SDS, 1.0% Triton X-100, 1.0% sodium deoxycholate containing protease inhibitors (Complete Mini, Roche). Aliquots 30 µg (DMT1) or 40 µg (ferroportin) were prepared in sample buffer without boiling and electrophoresed on 10% gels. After transfer to nitrocellulose membranes, blots were blocked with 5% non-fat milk and immunoblotted using rabbit anti-HDII (all four isoforms) antibody (1:1,000; a kind gift from Dr. Jerry Kaplan, The University of Utah, Salt Lake City, UT) or rabbit anti-ferroportin antibody (1:500; Alpha Diagnostics). Blots were probed with mouse anti-actin (MP Biomedicals) as a loading control. Secondary antibodies were goat anti-rabbit IRDye800 (Actin; LI-COR, Immunoreactivity was detected using an Odyssey Infrared Imaging System (DMT1 and actin; LI-COR) or ECL (ferroportin; West Pico, Thermo Scientific). Relative intensities of protein bands were determined using Odyssey (version 2.1; LI-COR) or Image J software (version 1.43; NIH).

ICP-MS

Olfactory bulbs from 3 male and 5 female Hfe+/+ mice and 4 male and 4 female Hfe−/− mice were pooled and analyzed for iron and manganese levels by ICP-MS (Trace Element Analysis Laboratory, Dept. of Earth Sciences, Dartmouth College, Hanover, NH).

Statistical analyses

Values reported were expressed as means ± SEM. For multi-group comparison (gene and diet effect), two-way ANOVA test followed by Tukey’s post-hoc comparison was used. Two-sample t-test was employed to compare the parameters between Hfe−/− and Hfe+/+ mice using Systat 13 (Systat). Differences were considered significant at P<0.05.

Acknowledgments

The authors are grateful to Ms. Dorothy Vargas, Matthew Cozzolino, and Ashley Henry for technical assistance on pharmacokinetic experiments.

Author Contributions

Conceived and designed the experiments: JK MWR. Performed the experiments: JK PDB. Analyzed the data: JK. Wrote the paper: JK MWR.

References


PLOS ONE | www.plosone.org 6 May 2013 | Volume 8 | Issue 5 | e64944


