Augmented Pulmonary Responses to Acute Ozone Exposure in Obese Mice: Roles of TNFR2 and IL-13

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BACKGROUND: Acute ozone (O\textsubscript{3}) exposure results in greater inflammation and airway hyper-responsiveness (AHR) in obese versus lean mice.

OBJECTIVES: We examined the hypothesis that these augmented responses to O\textsubscript{3} are the result of greater signaling through tumor necrosis factor receptor 2 (TNFR2) and/or interleukin (IL)-13.

METHODS: We exposed lean wild-type (WT) and TNFR2-deficient (TNFR2\textsuperscript{-/-}) mice, and obese Cpe\textsuperscript{-/-} and TNFR2-deficient Cpe\textsuperscript{-/-} mice (Cpe\textsuperscript{-/-}/TNFR2\textsuperscript{-/-}), to O\textsubscript{3} (2 ppm for 3 hr) either with or without treatment with anti–IL-13 or left them unexposed.

RESULTS: O\textsubscript{3}-induced increases in baseline pulmonary mechanics, airway responsiveness, and cellular inflammation were greater in Cpe\textsuperscript{-/-} than in WT mice. In lean mice, TNFR2 deficiency ablated O\textsubscript{3}-induced AHR without affecting pulmonary inflammation; whereas in obese mice, TNFR2 deficiency augmented O\textsubscript{3}-induced AHR but reduced inflammatory cell recruitment. O\textsubscript{3} increased pulmonary expression of IL-13 in Cpe\textsuperscript{-/-} but not WT mice. Flow cytometry analysis of lung cells indicated greater IL-13–expressing CD4\textsuperscript{+} cells in Cpe\textsuperscript{-/-} versus WT mice after O\textsubscript{3} exposure. In Cpe\textsuperscript{-/-} mice, anti–IL-13 treatment attenuated O\textsubscript{3}-induced increases in pulmonary mechanics and inflammatory cell recruitment, but did not affect AHR. These effects of anti–IL-13 treatment were not observed in Cpe\textsuperscript{-/-}/TNFR2\textsuperscript{-/-} mice. There was no effect of anti–IL-13 treatment in WT mice.

CONCLUSIONS: Pulmonary responses to O\textsubscript{3} are not just greater, but qualitatively different, in obese versus lean mice. In particular, in obese mice, IL-13 induces IL-13–dependent AHR and regulates inflammatory cell recruitment but not AHR.

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Ozone (O\textsubscript{3}), an air pollutant, causes respiratory symptoms and reductions in lung function (Alexis et al. 2000). O\textsubscript{3} is also a trigger for asthma. Asthma-related emergency room visits increase on days of high ambient O\textsubscript{3} (Fauroux et al. 2000; Gent et al. 2003). O\textsubscript{3} activates the innate immune system causing pulmonary inflammation and neutrophil and airway hyper-responsiveness (AHR), a characteristic feature of asthma (Garantziotis et al. 2009).

Two-thirds of the U.S. population is obese or overweight (National Center for Health Statistics 2012), and obesity is a risk factor for asthma (Shore and Johnston 2006). Nevertheless, our understanding of how obesity impacts pulmonary responses to O\textsubscript{3} is still rudimentary. O\textsubscript{3}-induced decrements in lung function are greater in obese and overweight than lean human subjects (Alexeoff et al. 2007; Bennett et al. 2007). Obese mice also exhibit greater pulmonary inflammation and greater AHR than lean mice after acute O\textsubscript{3} exposure (Johnston et al. 2006; Shore et al. 2003). The mechanistic basis for the effect of obesity on responses to O\textsubscript{3} is unknown.

Tumor necrosis factor-\alpha (TNF\textalpha) promoter polymorphisms that augment TNF\textalpha expression are associated with an increased obesity-related risk of asthma, especially non-atopic asthma (Castro-Giner et al. 2009), suggesting that TNF\textalpha may also be relevant for obesity-related asthma. The role of TNF\textalpha in the augmented responses to acute O\textsubscript{3} observed in obese mice has not been established.

TNF\textalpha binds to two receptors, TNFR1 and TNFR2, which differ in their ability to induce inflammation and apoptosis and in their affinity for cleaved versus membrane-associated TNF\textalpha (Naude et al. 2011). In lean mice, O\textsubscript{3}-induced AHR requires TNFR2 (Shore et al. 2001), though TNFR1 may also play a role (Cho et al. 2001). TNFR2 is also required for the innate AHR that is observed in obese mice (Williams et al. 2012), but the role of TNFR2 in the augmented responses to O\textsubscript{3} observed in obese mice is not established.

The first purpose of this study was to examine the hypothesis that TNFR2 is required for the augmented response to acute O\textsubscript{3} exposure in obese mice. To test this hypothesis, we bred Cpe\textsuperscript{-/-} mice that were genetically deficient in the TNFR2 receptor (Cpe\textsuperscript{-/-}/TNFR2\textsuperscript{-/-} mice). Cpe\textsuperscript{-/-} mice lack carboxypeptidase E (Cpe), an enzyme involved in appetite regulation and energy expenditure (Leibelt et al. 1997). Lack of Cpe leads to obesity (Johnston et al. 2006, 2010). We assessed airway responsiveness and pulmonary injury and inflammation in Cpe\textsuperscript{-/-}/TNFR2\textsuperscript{-/-} mice along with wild-type (WT), Cpe\textsuperscript{-/-}, and TNFR2\textsuperscript{-/-} mice.

Interleukin (IL)-13 also plays an important role in AHR. In mice, exogenous administration of IL-13 to the lungs results in AHR, and IL-13–blocking reagents inhibit allergen-induced AHR (Grunig et al. 1998; Wills-Karp et al. 1998). IL-13 may also play a role in responses to acute O\textsubscript{3} exposure. In lean BALB/c mice, IL-13 deficiency reduces O\textsubscript{3}-induced AHR and inflammation (Pichavant et al. 2008; Williams et al. 2008), but the role of IL-13 in responses to O\textsubscript{3} in obese mice is not established. Consequently, to examine the hypothesis that IL-13 contributes to obesity-related differences in the response to O\textsubscript{3}, we measured IL-13 expression and examined the effects of anti–IL-13 antibodies in obese and lean mice. Because TNF and IL-13 can synergize to promote the expression of chemokines that may contribute to the effects of O\textsubscript{3} (Reibman et al. 2003), we examined the effect of anti–IL-13 treatment in both TNFR2-sufficient and -deficient mice.

Methods

Animals. This study was approved by the Harvard Medical Area Standing Committee on Animals. Animals were treated humanely and with regard for alleviation of suffering. We bred Cpe\textsuperscript{-/-}/TNFR2\textsuperscript{-/-}, TNFR2\textsuperscript{-/-}, Cpe\textsuperscript{-/-}, and WT mice as previously described (Williams et al. 2012). Female mice were on a C57BL/6 background, were fed standard mouse chow diets, and were 10–12 weeks old.

Protocol. We exposed mice to 2 ppm O\textsubscript{3} for 3 hr. Twenty-four hours after exposure, we measured pulmonary mechanics and airway responsiveness, performed bronchoalveolar lavage (BAL), harvested blood by cardiac puncture, and collected lungs for preparation.
of RNA. Controls for these mice were not exposed to O₃ but were otherwise treated identically and studied simultaneously. Pulmonary mechanics and airway responsiveness for these unexposed mice have been previously reported (Williams et al. 2012). We treated other mice with anti–IL-13 antibody (2 µg/g body weight intraperitoneally) 24 hr before O₃ exposure. In a final cohort of O₃-exposed mice, we harvested lungs at 24 hr, enzymatically digested the lungs, and isolated lung cells for flow cytometry to quantitate IL-13–expressing CD4⁺ cells.

**Measurement of pulmonary mechanics and airway responsiveness.** We generated quasi-static pressure volume (PV) curves, assessed baseline pulmonary mechanics using the forced oscillation technique, and measured airway responsiveness to aerosolized methacholine as previously described (Williams et al. 2012). We assessed Newtonian resistance (Rn), which largely reflects the resistance of the conducting airways, and the coefficients of lung tissue damping (G) and lung tissue elastance (H), which reflect changes in the small airways and pulmonary parenchyma.

**Bronchoalveolar lavage.** We lavaged lungs and counted BAL cells as previously described (Williams et al. 2012). We measured BAL protein and counted BAL cells as previously described. We measured BAL cytokines, chemokines, and hyaluronan by ELISA (R&D Systems Inc., Minneapolis, MN; bionscience, San Diego, CA; and Echelon Biosciences, Salt Lake City, UT). We measured BAL protein by Bradford assay (BioRad, Hercules, CA).

**RNA extraction and real-time polymerase chain reaction (PCR).** We used real-time PCR to quantitate IL-13 and IL-17A mRNA expression as described (Shore et al. 2011). We subtracted Ct values for a housekeeping gene, 36B4 (rplp0) (which codes for a ribosomal protein), from Ct values for IL-13 or IL-17A to obtain ΔCt values. We expressed changes in mRNA relative to values from the WT unexposed mice, using the ΔΔCt method.

**Flow cytometry.** We flushed the lungs to remove blood cells, and then excised, minced, and digested lung tissue as previously described (Kasahara et al. 2012). We cultured lung cells either with or without PMA (phorbol myristate acetate) and ionomycin, in the presence of Golgi Stop (BD Bioscience, Franklin Lakes, NJ), for 5 hr before staining for flow cytometry. Cells were fixed with 0.1% Triton X-100, incubated with anti-Fcγ blocking mAb (clone 93; Biolegend, San Diego, CA), and washed. We stained the cells with Alexa Fluor 488-conjugated CD4 mAb (clone GK1.5; Biolegend) and Alexa Fluor 647-conjugated anti-mouse IL-13 (clone eBio13A; bionscience). We passed the cells through a BD Canto flow cytometer (BD Bioscience), and analyzed the data with FlowJo software (Tree Star Inc., Ashland, OR).

**Statistics.** We used factorial analysis of variance to assess the significance of differences in outcome indicators, as previously described (Williams et al. 2012). We performed analyses using Statistica version 6 (StatSoft, Tulsa, OK).

**Results**

**Body mass.** There was a significant effect of Cpe (p < 0.001) but not TNFR2 genotype on body mass. Whether or not they were deficient in TNFR2, Cpe⁺/⁻ mice weighed about twice as much as lean controls, consistent with previous observations (Williams et al. 2012).

**Pulmonary mechanics and airway responsiveness.** In WT mice, no significant differences in the PV curve of the lungs were observed in O₃-exposed versus unexposed mice (Figure 1A). However, in Cpe⁺/⁻ mice, there was a rightward shift and widening of the PV loop indicative of increased hysteresis in O₃-exposed versus unexposed mice (Figure 1B). To quantitate these changes, we measured the area of the PV loop, and normalized it by A, the difference in volume

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Figure 1. O₃-induced changes in pulmonary mechanics of mice 24 hr after O₃ exposure. Mice were unexposed, exposed to O₃, or treated with anti–IL-13 24 hr before O₃ exposure. Pressure volume (PV) curves of WT (A) and Cpe⁺/⁻ (B) mice. The areas of the PV curves were normalized for volume A (the differences between total lung capacity and end expiratory volume, i.e., the height of the PV curves (O), baseline values for airway resistance (Rn) (D), the coefficient of lung tissue damping, G (E), and the coefficient of lung tissue elastance, H (F). Values shown are mean ± SE of data from 6–9 mice in each group.

* p < 0.05, compared with unexposed genotype-matched mice. ** p < 0.05, compared with TNFR2 genotype-matched lean mice with the same exposure. *** p < 0.05, compared with O₃-exposed genotype-matched mice not treated with anti–IL-13.
between total lung capacity and end expiratory lung volume (height of the PV loops in Figure 1). Area/A is a measure of the thickness of the PV loop. A was lower in obese versus lean mice, as previously reported (Williams et al. 2012), but there was no difference in A in O3-exposed versus unexposed mice (data not shown). Consistent with the increased hysteresis induced by O3 in Cpefat/– but not WT mice (Figure 1A,B), Area/A was greater in O3-exposed versus unexposed Cpefat/– mice, whereas O3 had no effect on Area/A in WT mice (Figure 1C). Area/A was also greater in O3-exposed versus unexposed Cpefat/–/TNFR2–/– mice (Figure 1C).

Even in unexposed mice, baseline Rn, G, and H were elevated in the lungs of obese mice (Figure 1D,E,F). O3 exposure had no effect on Rn in mice of any genotype, but O3 exposure increased baseline G and H in Cpefat/– mice (Figure 1D,E,F). In contrast, no significant changes in baseline G and H were observed in O3-exposed versus unexposed Cpefat/–/TNFR2–/– mice.

Airway responsiveness was greater in unexposed Cpefat/– versus WT mice, and this difference was abolished by TNFR2 deficiency (see Williams et al. 2012). In lean WT mice, O3 exposure caused AHR (Figure 2A). This effect of O3 was observed when G, a measure of the lung tissue response, was used as the outcome indicator (Figure 2A). A similar trend was observed for H, but did not reach statistical significance [see Supplemental Material, Figure S1D (http://dx.doi.org/10.1289/ehp.1205880)]. There was no effect of O3 on methacholine-induced changes in Rn (see Supplemental Material, Figure S1A).

O3-induced AHR was absent in TNFR2–/– mice (Figure 2B). O3 did cause AHR in Cpefat/– mice (Figure 2C), and the magnitude of the O3-induced AHR was significantly greater than in WT mice. O3 also induced AHR in Cpefat/–/TNFR2–/– mice (Figure 2D). Indeed, O3-induced AHR was actually greater in Cpefat/–/TNFR2–/– than in Cpefat/– mice, and O3-induced AHR was observed even when changes in Rn were used as the outcome indicator, whereas this was not the case in mice of any other genotype (see Supplemental Material, Figure S1D). O3-induced AHR was also observed in Cpefat/–/TNFR2–/– and Cpefat/– mice when H was used as the index of response, and AHR based on changes in H was also greater in Cpefat/–/TNFR2–/– than in Cpefat/– mice (see Supplemental Material, Figure S1G,H).

Because IL-13 has been shown to contribute to O3-induced AHR in lean BALB/c mice (Pichavant et al. 2008; Williams et al. 2008), we determined whether there were obesity- and/ or TNFR2-dependent differences in IL-13 expression. O3 caused an increase in IL-13 mRNA in obese but not lean mice (Figure 3A). There was a nonsignificant trend toward increased IL-13 mRNA expression in Cpefat/–/TNFR2–/– versus Cpefat/– mice. O3 exposure also increased BAL IL-13 in Cpefat/– but not WT mice (Figure 3B). Furthermore, both the percentage and the total number of IL-13–expressing CD4+ cells were higher in Cpefat/– than WT mice exposed to O3 (Figure 3D,E). Consequently, we also measured BAL concentrations of another Th2 cytokine, IL-5. O3-induced increases in IL-5 were significantly greater in obese versus lean mice, and TNFR2 deficiency significantly increased BAL IL-5 levels in obese but not lean mice (Figure 3F), consistent with the trend observed for IL-13 (Figure 3B). IL-17A has also been linked to O3-induced injury (Bhalla 1999)]. O3 exposure significantly increased BAL IL-13 mRNA expression in Cpefat/–/TNFR2–/– versus Cpefat/– mice (Figure 4). There was a nonsignificant trend toward greater IL-17A in Cpefat/–/TNFR2–/– than Cpefat/– mice. To examine the functional impact of this IL-13 expression, we treated mice with antibodies to IL-13. In WT mice, there was no effect of anti–IL-13 treatment, likely because pulmonary IL-13 was negligible in these mice (Figure 3A,B). In Cpefat/– mice, anti–IL-13 treatment reversed the effects of O3 exposure on the PV curve (Figure 1B,C), and it prevented the O3-induced increase in baseline G and H (Figure 1E,F) but did not affect O3-induced AHR (Figure 2C). In Cpefat/–/TNFR2–/– mice, anti–IL-13 treatment also prevented O3-induced changes in the PV curve (Figure 1C) but had no effect on baseline pulmonary mechanics or O3-induced AHR (Figure 2D).

**Pulmonary inflammation and injury.** O3 exposure significantly increased BAL macrophages, neutrophils, and protein [a marker of O3-induced injury (Bhalla 1999)] (Figure 4). Lymphocytes and eosinophils were not observed in BAL fluid of most mice regardless of exposure, genotype, or obesity status. BAL neutrophils and protein were increased in Cpefat/– versus WT mice exposed to O3 (Figure 4). In lean mice, TNFR2 deficiency had no effect on these responses to O3. However, in obese mice, BAL neutrophils and macrophages, but not BAL
protein, were significantly reduced in TNFR2-deficient versus -sufficient mice (Figure 4). The TNFR2-dependent changes in neutrophil recruitment were not the result of differences in the expression of IL-6, KC (keratinocyte chemotactrant), or G-CSF (granulocyte colony stimulating factor) [Figure 5A; see also Supplemental Material, Figure S2 (http://dx.doi.org/10.1289/ehp.1205880)], cytokines and chemokines reported to be important for O3-induced neutrophil recruitment to the lungs (Johnston et al. 2005a, 2005b; Kasahara et al. 2012). These chemokines were greater in O3-exposed obese versus lean mice, but they were either not affected (i.e., IL-6 and KC) or actually augmented (i.e., G-CSF) by TNFR2 deficiency in obese mice. Similarly, TNFR2-dependent changes in macrophage recruitment are not the result of changes in MCP-1, a chemokine required for O3-induced macrophage recruitment (Zhao et al. 1998). BAL MCP-1 was greater in obese versus lean mice, but TNFR2 deficiency had no effect on BAL MCP-1 (Figure 5B). Surprisingly, O3-induced increases in BAL TNFα were significantly reduced in obese versus lean mice (Figure 5C).

O3 induces fragmentation of the matrix glycoprotein, hyaluronan, leading to AHR (Garantziotis et al. 2009). Such hyaluronan fragments are thought to be induced via oxidative stress caused by O3. O3 exposure caused an increase in BAL hyaluronan (Figure 5D) as well as protein carbonyls (Figure 5E), a marker of oxidative stress, although protein carbonyls were only increased in obese and not lean mice. Furthermore, O3-induced increases in BAL hyaluronan and protein carbonyls were significantly greater in obese than lean mice (Figure 5D,E), suggesting...

Figure 3. IL-13 mRNA expression in lungs (A) and IL-13 concentration in BAL (B) from mice that were either unexposed or exposed to O3. Example of flow cytometry data from one Cpefat mouse (C) showing histograms of CD4+ lung cells stained with isotype control antibody or stained with anti–IL-13 and either unstimulated or stimulated with PMA (10 ng/mL) and ionomycin (500 ng/mL) for 5 hr to induce cytokine expression. Shown also are the percentages (D) and total number (E) of PMA and ionomycin stimulated CD4+ cells isolated from lungs of O3-exposed mice that expressed IL-13. BAL IL-5 concentrations (F) and IL-17A mRNA expression (G) in unexposed and O3-exposed mice. Values shown are mean ± SE of data from 4–8 mice per group. Results for IL-13 and IL-17A mRNA are normalized to 36B4 expression.

Figure 4. Macrophages (A), neutrophils (B), and protein (C) in BAL from mice that were unexposed, exposed to O3, or treated with anti–IL-13 24 hr before O3 exposure. Values shown are mean ± SE of data from 4–9 mice per group.
that elevations in BAL hyaluronan may contribute to greater O₃-induced AHR in obese versus lean mice. However, BAL hyaluronan does not appear to account for the greater O₃-induced AHR in Cpefat/TNFR2⁻/⁻ versus Cpewt because BAL hyaluronan was not different in these two strains (Figure 5D).

In Cpefat mice, anti–IL-13 treatment reduced O₃-induced increases in BAL neutrophils and macrophages (Figure 4A,B), but not BAL protein (Figure 4C), whereas there was no effect of anti–IL-13 treatment in Cpewt/TNFR2⁻/⁻ mice (Figure 4A,B). O₃-induced increases in MIP-3α (CCL20; macrophage inflammatory protein 3 α) and LIX (CXCL5; lipopolysaccharide-induced CXC chemokine) mRNA are reduced in IL-13–deficient versus WT BALB/c mice, suggesting that changes in these chemokines may contribute to the effects of IL-13 deficiency on inflammatory cell recruitment (Williams et al. 2008).

O₃-induced increases in BAL LIX, a neutrophil chemotactic factor, were not affected by obesity, TNFR2 deficiency, or anti–IL-13 treatment (Figure 6A) indicating that this chemokine does not account for the observed effects on BAL neutrophils (Figure 4B). However, TNFR2 deficiency did attenuate O₃-induced increases in MIP-3α regardless of obesity status (p < 0.001) (Figure 6B). Anti–IL-13 also caused a significant reduction in MIP-3α in TNFR2-sufficient (p < 0.01) but not -deficient mice (Figure 6B), suggesting that IL-13 and TNFα synergize in the induction of MIP-3α after O₃ exposure.

Discussion

We observed greater effects of acute O₃ exposure in obese versus lean mice consistent with previous observations (Johnston et al. 2006; Shore et al. 2003). Importantly, our results also indicated obesity-related differences in the mechanisms governing the pulmonary effects of O₃ (see Table 1 for summary). In lean mice, TNFR2 deficiency reduced O₃-induced AHR but had no effect on O₃-induced inflammation; whereas in obese mice, TNFR2 deficiency enhanced O₃-induced AHR while attenuating O₃-induced inflammation (Figures 2, 4). In

![Figure 5. BAL G-CSF (A), MCP-1 (B), TNFα (C) hyaluronan (D), protein carbonyls (E) from mice that were either unexposed or exposed to O₃. Values shown are mean ± SE of data from 4–10 mice per group.](image-url)

* p < 0.05, compared with unexposed genotype-matched mice. ** p < 0.05, compared with obesity-matched TNFR2-deficient mice with the same O₃ exposure. *** p < 0.05, compared with obesity-matched TNFR2-sufficient mice with the same O₃ exposure.

![Figure 6. BAL LIX (A) and MIP-3α (B) from mice that were unexposed, exposed to O₃, or treated with anti–IL-13 24 hr before O₃ exposure. Values shown are mean ± SE of data from 5–8 mice per group.](image-url)

* p < 0.05, compared with unexposed genotype-matched mice. ** p < 0.05, compared with obesity-matched TNFR2-deficient mice with the same O₃ exposure. # p < 0.05, compared with O₃-exposed genotype-matched mice not treated with anti–IL-13.

Table 1. Effect of IL-13 blockade or TNFR2 deficiency on O₃-induced increases in airway responsiveness, BAL neutrophil numbers, and pulmonary mechanics in lean and obese mice.

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<thead>
<tr>
<th>Intervention</th>
<th>Lean mice</th>
<th>Obese mice</th>
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<tr>
<td></td>
<td>Airway responsiveness</td>
<td>BAL neutrophils</td>
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<tr>
<td>IL-13 blockade</td>
<td>No change</td>
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<td>TNFR2 deficiency</td>
<td>Decrease</td>
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Outcomes (no change, increase, decrease) indicate the qualitative change in the response to O₃ relative to the absence of IL-13 blockade or TNFR2 deficiency.
lean mice, IL-13 was not induced by O3, and there was no impact of anti–IL-13 treatment on responses to O3. However, in obese mice, O3 did induce IL-13 expression (Figure 3). Importantly, obesity-related differences in IL-13 expression accounted for the ability of O3 to induce changes in the PV curve of lung and to increase baseline G and H in obese Cpe−/− but not lean WT mice (Figure 1). IL-13 also contributed to the greater recruitment of inflammatory cells to the lungs of Cpe−/− versus WT mice (Figure 4). Moreover, in obese mice, IL-13 and TNFR2 appeared to synergize to exacerbate O3-induced inflammation and changes in pulmonary mechanics because anti–IL-13 treatment reduced IL-13 mRNA expression and BAL IL-13 in TNFR2-deficient mice but not WT mice (Figure 3A,B). CD4+ T cells appeared to be the source of this IL-13 (Figure 3C-E), and another CD4+ T-cell–derived cytokine, IL-5, was also induced by O3 exposure in obese mice (Figure 3F).

To determine whether IL-13 contributed to obesity- and/or TNFR2-dependent changes in the response to O3, we treated mice with anti–IL-13 before O3 exposure. Anti–IL-13 attenuated the O3-induced increase in the hysteresis of the PV curve that was induced by O3 exposure in obese mice (Figure 1B,C). Lung surfactant is important in limiting lung hysteresis and IL-13 reduces the pulmonary expression of surfactant protein C (Ito and Mason 2010), which is important for the surface activity of surfactant. Coupled with the observations that O3-induced changes in lung hysteresis were not observed in WT mice (Figure 1A,C), which lacked IL-13 (Figure 3), the results suggest that O3 caused changes in pulmonary surfactant activity in Cpe−/− mice and that these changes were mediated by IL-13. Changes in lung hysteresis in obese mice may be aggravated by the ability of O3 exposure to cause oxidation of phospholipids important for surfactant activity (Pulfer and Murphy 2004). O3-induced changes in baseline G and H, measures of the lung tissue, were also ablated by anti–IL-13 treatment (Figure 1). Thus obesity-related differences in the ability of O3 to induce IL-13 expression (Figure 3) appear to account for the ability of O3 to induce changes in baseline pulmonary mechanics in Cpe−/− but not WT mice. TNFR2 deficiency also reduced O3-induced changes in pulmonary mechanics in Cpe−/− mice, and anti–IL-13 treatment had no effect on mechanics in Cpe−/−TNFR2−/− mice (Figure 1), suggesting that synergy between IL-13 and TNFR2 contributes to these changes.

In contrast to the effects on baseline pulmonary mechanics (Figure 1), we observed no effect of anti–IL-13 treatment on O3-induced AHR in Cpe−/− mice (Figure 2C), nor was there any effect on AHR in Cpe−/−TNFR2−/− mice (Figure 2D), indicating that the enhanced AHR in Cpe−/−TNFR2−/− mice was not the result of increased IL-13 signaling. Thus, other factors must account for obesity-related increases in O3-induced AHR and for the augmented AHR observed in Cpe−/−TNFR2−/− mice. IL-5 can induce AHR even in the absence of eosinophils (Borchers et al. 2001) (eosinophils were not observed in these animals). IL-5 was augmented in O3-exposed Cpe−/− versus WT mice and further augmented in Cpe−/−TNFR2−/− mice (Figure 3F), consistent with the changes in AHR (Figure 2). IL-17A can also induce AHR (Kudo et al. 2012). IL-17A was expressed after O3 exposure (Figure 3G), especially in the obese mice, and could also contribute to the augmented O3-induced AHR observed in these mice (Figure 2).

In lean mice, TNFR2 deficiency had no effect on O3-induced inflammatory cell recruitment (Figure 4), consistent with previous observations (Cho et al. 2001; Shore et al. 2001). In contrast, in obese mice, TNFR2 deficiency reduced O3-induced neutrophil and macrophage recruitment and ablated obesity-related differences in these outcomes (Figure 4). Anti–IL-13 treatment also had no effect on O3-induced inflammation in lean WT mice, which lacked IL-13 (Figure 3A,B), but anti–IL-13 treatment significantly reduced O3-induced inflammatory cell recruitment in obese Cpe−/− mice (Figure 4A,B). A similar reduction in O3-induced inflammation occurs in lean IL-13-deficient BALB/c mice (Williams et al. 2008), which are more thrombolytic than the C57Bl/6 mice used in the present study. Importantly, in obese mice, TNFR2 and IL-13 appeared to interact to promote inflammatory cell recruitment because anti–IL-13 treatment had no effect on BAL neutrophils or macrophages in Cpe−/−TNFR2−/− mice even though these mice had at least as much IL-13 expression as Cpe−/− mice (Figure 4A,B). This interaction may occur at the level of MIP-3α expression. In obese Cpe−/− mice, both TNFR2 deficiency and anti–IL-13 treatment resulted in a significant decrease in BAL MIP-3α (Figure 6B), consistent with observations of others that TNFα and IL-13 can both induce the expression of MIP-3α in airway epithelial cells (Reibman et al. 2003). Importantly, anti–IL-13 treatment inhibited MIP-3α expression in TNFR2−/− sufficient but not -deficient Cpe−/− mice (Figure 6B). Dendritic cells and T cells typically express CCR6, the receptor for MIP-3α, but neutrophils can be induced to express CCR6 in the presence of TNFα (Yamashiro et al. 2000). In addition, MIP-3α can induce migration of IL-17A expressing cells that may contribute to neutrophil recruitment (Li et al. 2011). IL-17A was induced by O3 (Figure 3G).

O3-induced changes in lung function (i.e., G and H) and cellular inflammation occurred in concert (Table 1). O3-induced increases in BAL neutrophils and in both G and H were greater in Cpe−/− versus WT mice. Furthermore, both TNFR2 deficiency and anti–IL-13 treatment reduced O3-induced changes in G and H as well as BAL neutrophil numbers in obese mice (Table 1). The results suggest that effects of O3 on lung function and inflammation may be mechanistically related. In contrast, O3-induced AHR
and inflammations were dissociated. For example, TNFR2 deficiency and anti-IL-13 treatment either augmented or had no effect on O3-induced mice, but they attenuated O3-induced inflammation in obese mice.

Two technical issues require consideration. First, obese mice have a slightly higher minute ventilation (Shore and Johnston 2006) and consequently a slightly higher inhaled dose of O3. Their lungs are also smaller (Williams et al. 2012), so the dose per gram of lung tissue may be higher. We do not think this issue contributed substantially to the outcome because neither TNFR2 deficiency nor anti–IL-13 treatment had any effect on lung volume (data not shown), but both reduced the augmented effects of O3 on BAL cells and pulmonary mechanics in obese mice. Second, we used the same volume of fluid for lavage in obese and lean mice. Because the lungs of the obese mice were smaller and presumably had correspondingly less lung lining fluid, there should have been greater dilution of substances in that lining fluid and thus lower concentrations of BAL moieties in the obese than the lean mice. In fact, the opposite was true for most BAL cytokines/chemokines examined [Figures 3, 5, 6; see also Supplemental Material, Figure S2 (http://dx.doi.org/10.1289/ehp.1205880)]. Thus, issues related to normalization of the BAL sampling are unlikely to explain the observed results.

Conclusions

Our results indicate differences in the mechanisms regulating pulmonary responses to O3 in lean and obese mice. In particular, TNFR2 deficiency had opposing effects on O3-induced AHR in lean and obese mice. In addition, O3 increased the pulmonary expression of IL-13 in obese but not lean mice, and this IL-13 appeared to account for the augmented ability of O3 to induce changes in pulmonary mechanics and inflammation in obese mice via synergetic effects with TNFR2. The majority of the population of the United States is either obese or overweight. Our results emphasize the need for improved understanding of the effects of O3 in this population.

References


