



Blood Lead Levels and Serum Insulin-Like Growth Factor 1 Concentrations in Peripubertal Boys

Citation

Fleisch, Abby F., Jane S. Burns, Paige L. Williams, Mary M. Lee, Oleg Sergeyeve, Susan A. Korrick, and Russ Hauser. 2013. "Blood Lead Levels and Serum Insulin-Like Growth Factor 1 Concentrations in Peripubertal Boys." *Environmental Health Perspectives* 121 (7): 854-858. doi:10.1289/ehp.1206105. <http://dx.doi.org/10.1289/ehp.1206105>.

Published Version

doi:10.1289/ehp.1206105

Permanent link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:11717530>

Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA>

Share Your Story

The Harvard community has made this article openly available.
Please share how this access benefits you. [Submit a story](#).

[Accessibility](#)

Blood Lead Levels and Serum Insulin-Like Growth Factor 1 Concentrations in Peripubertal Boys

Abby F. Fleisch,¹ Jane S. Burns,² Paige L. Williams,³ Mary M. Lee,^{4,5} Oleg Sergeev,^{6,7} Susan A. Korrick,^{2,8} and Russ Hauser²

¹Department of Endocrinology, Children's Hospital Boston, Boston, Massachusetts, USA; ²Department of Environmental Health, and ³Department of Biostatistics, Harvard School of Public Health, Boston, Massachusetts, USA; ⁴Department of Pediatrics, and ⁵Department of Cell Biology, Pediatric Endocrine Division, University of Massachusetts Medical School, Worcester, Massachusetts, USA; ⁶Department of Physical Education and Health, Samara State Medical University, Samara, Russian Federation; ⁷Chapaevsk Medical Association, Chapaevsk, Russian Federation; ⁸Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, USA

BACKGROUND: Childhood lead exposure has been associated with growth delay. However, the association between blood lead levels (BLLs) and insulin-like growth factor 1 (IGF-1) has not been characterized in a large cohort with low-level lead exposure.

METHODS: We recruited 394 boys 8–9 years of age from an industrial Russian town in 2003–2005 and followed them annually thereafter. We used linear regression models to estimate the association of baseline BLLs with serum IGF-1 concentration at two follow-up visits (ages 10–11 and 12–13 years), adjusting for demographic and socioeconomic covariates.

RESULTS: At study entry, median BLL was 3 µg/dL (range, < 0.5–31 µg/dL), most boys (86%) were prepubertal, and mean ± SD height and BMI z-scores were 0.14 ± 1.0 and –0.2 ± 1.3, respectively. After adjustment for covariates, the mean follow-up IGF-1 concentration was 29.2 ng/mL lower (95% CI: –43.8, –14.5) for boys with high versus low BLL (≥ 5 µg/dL or < 5 µg/dL); this difference persisted after further adjustment for pubertal status. The association of BLL with IGF-1 was stronger for mid-pubertal than prepubertal boys (*p* = 0.04). Relative to boys with BLLs < 2 µg/dL, adjusted mean IGF-1 concentrations decreased by 12.8 ng/mL (95% CI: –29.9, 4.4) for boys with BLLs of 3–4 µg/dL; 34.5 ng/mL (95% CI: –53.1, –16.0) for BLLs 5–9 µg/dL; and 60.4 ng/mL (95% CI: –90.9, –29.9) for BLLs ≥ 10 µg/dL.

CONCLUSIONS: In peripubertal boys with low-level lead exposure, higher BLLs were associated with lower serum IGF-1. Inhibition of the hypothalamic–pituitary–growth axis may be one possible pathway by which lead exposure leads to growth delay.

KEY WORDS: cohort studies, growth, insulin-like growth factor 1, lead; puberty.

Environ Health Perspect 121:854–858 (2013). <http://dx.doi.org/10.1289/ehp.1206105> [Online 26 April 2013]

The Centers for Disease Control and Prevention (CDC) recently revised the reference value for childhood lead exposure downward from 10 to 5 µg/dL (Betts 2012). This has occurred in response to evidence showing consistent associations between blood lead levels (BLLs) < 10 µg/dL and neurocognitive (Bellinger et al. 1992; Lanphear et al. 2005), cardiovascular (Gump et al. 2005; Menke et al. 2006), and immunologic (Karmaus et al. 2005) outcomes, as well as pubertal (Kafourou et al. 1997; Naicker et al. 2010; Selevan et al. 2003; Shukla et al. 1991; Williams et al. 2010) and growth delay (Ballew et al. 1999; Little et al. 2009; Schwartz et al. 1986).

In cross-sectional studies conducted in prepubertal children, mean decreases of 1–1.5 cm in height have been estimated for each 10-µg/dL increase in BLL (Ballew et al. 1999; Little et al. 2009; Schwartz et al. 1986). BLL has also been negatively associated with height in two pubertal cohorts (Burns et al. 2012; Selevan et al. 2003). In our prospective cohort of Russian boys followed annually from 8 to 13 years of age, boys with BLL ≥ 5 µg/dL had significantly lower mean height z-score (–0.44) than boys with BLL < 5 µg/dL (Burns et al. 2012).

A mechanistic explanation for the association between lead exposure and growth delay in childhood has not been firmly established. Studies to this end have been conducted almost exclusively in animal models with high lead exposures, and results have suggested multiple possible mechanisms, perhaps operating simultaneously, including lead-induced reduction in food consumption (Hammond et al. 1989, 1990) and direct growth plate effects (Hass et al. 1967; Hicks et al. 1996). Several rodent studies have also implicated lead-mediated suppression of growth hormone (GH) release from the pituitary (Camoratto et al. 1993; Lau et al. 1991; Ronis et al. 1998a), and consistent with this finding, high lead exposure has been associated with decreased serum insulin-like growth factor 1 (IGF-1) concentration in rodents (Ronis et al. 1998a) and prepubertal children (Huseman et al. 1992).

The physiologic mechanisms that mediate health outcomes such as growth delay following low lead exposure may be different from those operating at higher exposures. Mechanistic differences in low- versus high-dose effects have been demonstrated for several environmental chemicals, including

bisphenol A, atrazine, dioxin, and perchlorate (Vandenberg et al. 2012). Because exposures to lead and other heavy metals are increasingly regulated, a better understanding of the physiologic effects of low-level exposures is needed. With regard to growth delay, the relationship between low-level lead exposure and serum IGF-1 concentration has not been examined.

The present analysis was conducted to evaluate the association between childhood BLL and longitudinally measured serum IGF-1 concentrations and to assess whether there is a dose–response relationship between BLL and IGF-1 in a large cohort of peripubertal boys with low-level lead exposure.

Methods

Study population. From 2003–2005, we recruited 8- to 9-year-old boys in Chapaevsk, Russia, to participate in the Russian Children's Study, as previously described (Burns et al. 2012). Enrollment exclusion criteria included being institutionalized or having severe cerebral palsy. A total of 499 boys were thus identified, and they have been followed annually since recruitment. For the present analysis, 10 boys with severe chronic illnesses that could affect growth were excluded. Of the remaining 489, two pubertal boys were also excluded

Address correspondence to R. Hauser, Department of Environmental Health, Harvard School of Public Health (Bldg 1, 14th floor), 665 Huntington Ave., Boston, MA 02115 USA. Telephone: (617) 432-3326. E-mail: rhauser@hsph.harvard.edu

We thank the former chief of Chapaevsk Central Hospital, V. Zeilert, and the staff of the Chapaevsk Medical Association. We also thank our colleagues B. Revich from the Institute for Forecasting, RAS, Moscow; A. Safronova and M. Starovoytov from the Russian Institute of Nutrition, Moscow; and staff of EFiS, Moscow laboratory. We thank L.E. Cohen for helpful discussions.

We have received support from the following grants: R82943701 from the U.S. Environmental Protection Agency; ES014370, ES000002, and ES017117 from the National Institute of Environmental Health Sciences; and T32 HS000063 from the Agency of Healthcare Research and Quality. M.M.L. is a member of the National Institute of Diabetes and Digestive and Kidney Disease–funded UMass Diabetes Research Center (DRC P3DK032520).

The authors declare they have no actual or potential competing financial interests.

Received 5 October 2012; accepted 24 April 2013.

from the analysis because of implausibly low IGF-1 concentrations (< 50 ng/mL) despite normal height and body mass index (BMI) z -scores. Of the remaining 487 boys, 394 met inclusion criteria for this analysis, specifically, by availability of a baseline BLL (at 8–9 years) and follow-up serum IGF-1 concentration at both the 2-year (at 10–11 years) and the 4-year (at 12–13 years) follow-up visits. None of the participants had IGF-1 measured at baseline or BLL measured at follow-up. The Russian Children's Study was approved by the human studies institutional review boards (IRBs) of the Chapaevsk Medical Association, Harvard School of Public Health, Brigham and Women's Hospital, and University of Massachusetts Medical School. The parent or guardian of each Russian Children's Study participant signed an informed consent form, and each boy signed an assent form. The present analysis was a secondary data analysis that was exempt from requirement for IRB review of already collected, deidentified data under federal and Children's Hospital Boston policies.

Study assessment protocol. At study entry, boys underwent a physical examination and blood collection. We used a validated Russian Institute of Nutrition semiquantitative food-frequency questionnaire to estimate dietary intakes during the previous year (Martinchik et al. 1998; Rockett et al. 1997). Mothers or guardians also completed nurse-administered health and lifestyle questionnaires that included information on birth, family and child medical histories, occupational and residential history, and measures of socioeconomic status (SES) such as household income and parental education. We obtained birth weight and gestational age from medical records.

Physical examination. A single study nurse who did not have knowledge of the boys' BLLs performed standardized anthropometric examinations, and a single investigator (O.S.) performed pubertal assessments at study entry and at annual follow-up visits. We measured height to the nearest 0.1 cm with a stadiometer. We measured weight to the nearest 100 g with a metric scale. We calculated age-adjusted percentiles for BMI (kilograms per meter squared) using the World Health Organization (WHO) standards (WHO 2011). For this analysis, pubertal status was based on testicular volume measured by Prader beads (orchidometer).

Blood lead levels. Venous blood samples (3.0 mL) were collected in trace metal-free Vacutainer tubes (Becton-Dickinson, Franklin Lakes, NJ, USA), after cleansing the venipuncture site with alcohol. Whole-blood samples were diluted with a matrix modifier solution and analyzed using Zeeman background corrected, flameless graphite furnace, atomic absorption spectrometry (ESA Laboratories,

Chelmsford, MA, USA). BLLs below the limit of detection (1 μ g/dL) were imputed as 0.5 μ g/dL for 9 (2.2%) of 394 boys.

Serum IGF-1 concentrations. Serum IGF-1 concentrations were measured by a chemiluminescent immunometric assay using Siemens Immulite 2000 (Siemens AG, Munich, Germany). The assay is highly specific for IGF-1 with undetectable cross-reactivity with insulin, pro-insulin, luteinizing hormone (LH), thyroid-stimulating hormone, or insulin-like growth factor-2. The detection limit was 20 ng/mL; no IGF-1 values were below the limit of detection. The intra-assay coefficient of variation (CV) was $< 3.9\%$, and the inter-assay CV was $< 8.1\%$ for the Immulite 2000 kit.

Statistical analysis. We used repeated measures analysis to estimate the association between BLL measured at 8–9 years of age and serum IGF-1 concentration at both the 2-year (at 10–11 years) and 4-year (at 12–13 years) follow-up visits. The distribution of BLLs was right-skewed with outliers. We considered several different ways of evaluating BLLs, including dichotomized as high (≥ 5 μ g/dL) versus low (< 5 μ g/dL) based on the new CDC threshold, as a continuous measure (log-transformed), and categorized as 0–2, 3–4, 5–9, or ≥ 10 μ g/dL.

We fit linear regression models using a generalized estimating equation (GEE) approach to account for the repeated measures and slight skewness of IGF-1 concentrations. We first fit GEE linear regression models to evaluate unadjusted associations of high versus low BLL (≥ 5 or < 5 μ g/dL) with serum IGF-1 concentrations. Next, we created a full multivariable model that included dichotomized BLL, birth weight (continuous), gestational age at birth (continuous), breastfeeding duration (< 12 , 12–24, or > 24 weeks), maximum parental education (secondary education or less, junior college/technical training, or university graduate), monthly household income ($< US\$175$, $\$175$ – 250 , or $> \$250$), nutritional intake (total caloric intake and percent calories from protein, fat, and carbohydrate as continuous variables), baseline and follow-up age (continuous), and baseline and follow-up BMI [underweight (BMI < 10 th percentile), overweight (BMI > 85 th percentile), or normal weight]. We decided *a priori* not to consider height as a covariate due to its strong correlation with IGF-1 during puberty (Silbergeld et al. 1986). We then reduced this model by excluding covariates that did not predict the outcome with $p \leq 0.10$, or did not change the estimated association between BLL and IGF-1 by $> 10\%$ when removed from the model, resulting in a final model that included baseline parental education, birth weight, nutritional intake, and baseline and follow-up age and BMI.

Because pubertal status is influenced by lead exposure (Selevan et al. 2003; Williams et al. 2010) and may be considered in the causal pathway between BLL and serum IGF-1 concentration, we fit the final reduced model both with and without adjustment for pubertal status [categorized based on testicular volume (TV) as prepubertal, ≤ 3 mL TV; early pubertal, > 3 – 6 mL TV; or mid-pubertal, > 6 – 15 mL TV]. No boys in this analysis had a TV > 15 mL. We also evaluated whether the association between BLL and IGF-1 concentration differed by pubertal stage by including an interaction term between pubertal status (prepubertal, early pubertal, or mid-pubertal) and BLL (≥ 5 or < 5 μ g/dL).

Sensitivity analyses included the following modifications to the final reduced model: *a*) inclusion of maternal and paternal heights as potential confounding variables for the subset of boys who had these values available ($n = 337$), *b*) inclusion of 44 additional boys with only one follow-up IGF-1 concentration, *c*) BLL categorized as ≤ 2 , 3–4, 5–9, or ≥ 10 μ g/dL, and *d*) BLL modeled as a natural log-transformed continuous variable. We conducted all analyses using SAS version 9.3 (SAS Institute Inc., Cary, NC, USA), and we considered two-sided p -values ≤ 0.05 statistically significant.

Results

Baseline growth and demographic characteristics of the 394 boys included are shown in Table 1. The 95 eligible boys excluded from the present analysis were not significantly different from those included with regard to the characteristics shown in Table 1, except that they tended to have a higher percent protein intake at baseline and were more likely to have parents that had junior college or technical training (data not shown).

The median BLL at 8–9 years of age among boys included in the analysis was 3 μ g/dL (25th, 75th percentiles: 2 μ g/dL, 5 μ g/dL; range, 0.5–31 μ g/dL). Most boys were prepubertal at baseline. The mean baseline height z -score was slightly above the WHO average, and the mean baseline BMI z -score was slightly below the WHO average (Table 1).

In unadjusted GEE models that accounted only for correlation among study visits, mean serum IGF-1 concentration during follow-up was 24.3 ng/mL lower (95% CI: -39.3 , -9.3) among boys with BLL ≥ 5 μ g/dL compared with those with < 5 μ g/dL. A significantly lower mean IGF-1 concentration for boys with high versus low BLL was also estimated based on the full multivariable model (-28.0 ng/mL; 95% CI: -43.1 , -12.9) and the final reduced model (-29.2 ng/mL; 95% CI: -43.8 , -14.5)

Table 1. Baseline and follow-up characteristics of 394 boys from Chapaevsk, Russia, with baseline blood lead levels and two longitudinal measures of serum IGF-1.

Variable	8–9 years old (baseline)	10–11 years old	12–13 years old
Age (years) [median (range)]	8.1 (7.8–9.4)	10.1 (9.9–11.5)	12.1 (11.9–13.5)
BMI (WHO z-score)			
Mean ± SD	−0.2 ± 1.3	−0.2 ± 1.3	−0.2 ± 1.4
≤ 10th percentile [n (%)]	67 (17)	83 (21)	75 (19)
> 85th percentile [n (%)]	58 (15)	83 (21)	70 (18)
Height (WHO z-score) (mean ± SD)	0.14 ± 1.0	0.14 ± 1.0	0.03 ± 1.1
Testicular volume (mL) [n (%)]			
≤ 3 (prepubertal)	336 (86) ^a	213 (54)	52 (13) ^a
> 3–6	55 (14)	153 (39)	104 (27)
> 6	0 (0)	28 (7)	235 (60)
IGF-1 (ng/mL) (mean ± SD)		146.9 ± 52.1	253.5 ± 115.9
Birth weight (kg) (mean ± SD) ^b	3.34 ± 0.52		
Gestational age (weeks) (mean ± SD) ^a	39.01 ± 1.74		
Breastfeeding duration (weeks) [median (IQR)] ^c	13.0 (30.3)		
Baseline nutritional intake (mean ± SD) ^d			
Total kcal/day	2,837 ± 972		
Percent fat	34.1 ± 5.8		
Percent protein	11.5 ± 1.6		
Percent carbohydrate	54.4 ± 6.5		
Monthly household income (US\$) [n (%)] ^d			
< 175	136 (35)		
175–250	107 (27)		
> 250	150 (38)		
Maximal parental education [n (%)] ^a			
Secondary education or less	25 (6)		
Junior college/technical training	244 (62)		
University graduate	122 (31)		
Blood lead level (μg/dL)			
Median (IQR)	3.0 (3.0)		
< 5 [n (%)]	285 (72)		
≥ 5 [n (%)]	109 (28)		

IQR, interquartile range.

^aFive subjects missing. ^bTwo subjects missing. ^cFive subjects missing. ^dOne subject missing.**Table 2.** Repeated measures generalized estimating equation models predicting the mean levels of serum concentrations of IGF-1 (ng/mL) in relation to blood lead levels and relevant covariates.

Covariate	Full multivariable model (n = 385 boys, 767 visits)		Final reduced model (n = 389 boys, 775 visits)	
	Adjusted mean change (95% CI)	p-Value	Adjusted mean change (95% CI)	p-Value
Lead (μg/dL)				
< 5	Reference		Reference	
≥ 5	−28.0 (−43.1, −12.9)	< 0.001	−29.2 (−43.8, −14.5)	< 0.001
Age (years)	51.9 (47.2, 56.6)	< 0.001	52.1 (42.4, 56.8)	< 0.001
Birth weight (kg)	−17.4 (−33.2, −1.5)	0.03	−17.5 (−31.5, −3.5)	0.01
BMI z-score (percentile)				
≤ 10	−58.6 (−73.9, −43.3)	< 0.001	−61.3 (−76.7, −45.8)	< 0.001
> 10–85	Reference		Reference	
> 85	13.8 (−4.4, 31.9)	0.14	12.9 (−5.2, 30.9)	0.16
Nutritional intake				
Total calories ^a	−2.6 (−10.7, 5.6)	0.54	−2.6 (−10.6, 5.5)	0.54
Fat (percent)	1.4 (0.2, 2.6)	0.02	1.3 (0.1–2.5)	0.03
Protein (percent)	3.0 (−1.6, 7.6)	0.21	3.1 (−1.4, 7.6)	0.18
Parental education				
Secondary education or less	−22.5 (−45.8, 0.8)	0.06	−24.9 (−47.7, −2.0)	0.03
Junior college/technical training	−3.4 (−19.3, 12.5)	0.69	−2.7 (−18.4, 13.0)	0.74
University graduate	Reference		Reference	
Monthly household income (US\$)				
< 175	−3.3 (−20.3, 13.8)	0.71		
175–250	−6.4 (−23.7, 10.9)	0.47		
> 250	Reference			
Gestational age (weeks)	0.4 (−4.2, 5.1)	0.85		
Breastfeeding (weeks)				
< 12	Reference			
12–24	0.7 (−18.2, 19.7)	0.94		
> 24	1.1 (−14.1, 16.4)	0.88		

^aPer 1,000 calories.

(Table 2). When pubertal status was added to the final reduced model, the association of high BLL with IGF-1 concentration was modestly attenuated (adjusted mean difference = −24.4 ng/mL; 95% CI: −37.7, −11.1).

The association of BLL with IGF-1 concentration differed according to pubertal status. In particular, the reduction in adjusted mean IGF-1 concentrations between high versus low BLL groups was greater among boys in mid-puberty than for prepubertal boys (−41.9 ng/mL; 95% CI: −15.1, −68.7 vs. −14.1 ng/mL; 95% CI: −0.9, −27.2; interaction *p*-value = 0.04). The reduction in adjusted mean IGF-1 concentrations between high versus low BLL groups was slightly larger for boys in early puberty (−18.0 ng/mL; 95% CI: −3.5, −32.5) than for prepubertal boys (interaction *p*-value = 0.64) (Figure 1). Adjusted mean percent decreases in IGF-1 concentrations between high versus low BLL groups were 9.3%, 12.2%, and 19.5% for prepubertal boys, boys in early puberty, and boys in mid-puberty, respectively.

In sensitivity analyses, further adjustment for parental heights among the subset of boys (*n* = 337) with these measures available had no appreciable impact on the estimated difference in IGF-1 concentrations for boys with high versus low BLL (adjusted mean difference = −31.6 ng/mL; 95% CI: −48.2, −15.0). The estimated difference in IGF-1 was also similar based on a model that included follow-up IGF-1 concentrations for 438 boys with at least one IGF-1 measurement (adjusted mean difference = −28.8 ng/mL; 95% CI: −42.5, −15.1).

When BLL was divided into finer categories, adjusted mean IGF-1 concentrations decreased monotonically relative to

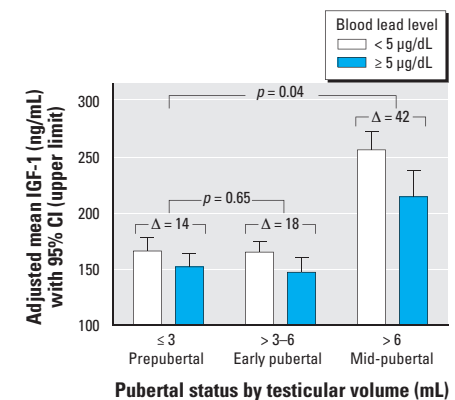


Figure 1. Adjusted mean IGF-1 concentrations for high versus low BLL by pubertal status. The adjusted mean IGF-1 difference for high versus low BLL was 14 ng/mL in prepubertal boys, 18 ng/mL in early-pubertal boys, and 42 ng/mL in mid-pubertal boys. Compared with prepubertal boys, the lead-associated IGF-1 difference was larger in mid-pubertal boys (*p* = 0.04) and larger, but not significantly larger, in early-pubertal boys (*p* = 0.65).

the reference group: -12.8 ng/mL (95% CI: $-29.9, 4.4$; $n = 176$) for BLL = $3-4$ $\mu\text{g/dL}$; -34.5 ng/mL (95% CI: $-53.1, -16.0$; $n = 97$) for BLL $5-9$ $\mu\text{g/dL}$; and -60.4 ng/mL (95% CI: $-90.9, -29.9$; $n = 12$) for BLL ≥ 10 $\mu\text{g/dL}$, compared with BLL ≤ 2 $\mu\text{g/dL}$ ($n = 109$) (Figure 2). Finally, each unit increase in natural log-transformed BLL was associated with a 22.2 -ng/mL decrease in mean serum IGF-1 (95% CI: $-33.9, -10.6$).

Discussion

We observed a negative association between BLLs measured at 8–9 years of age and serum IGF-1 concentrations at 10–11 and 12–13 years of age that was stronger among boys in mid-puberty than in prepubertal boys. This finding suggests one possible explanation for our previous finding of lower mean height z -scores for boys in the same cohort with BLL ≥ 5 $\mu\text{g/dL}$ compared with BLL < 5 $\mu\text{g/dL}$ (Burns et al. 2012). Results from the present analysis suggest a monotonic dose–response relationship between BLL and serum IGF-1 that adds further support to existing evidence of physiological effects of BLLs < 10 $\mu\text{g/dL}$ (Ballew et al. 1999; Bellinger et al. 1992; Gump et al. 2005; Kafourou et al. 1997; Karmaus et al. 2005; Lanphear et al. 2005; Little et al. 2009; Menke et al. 2006; Naicker et al. 2010; Selevan et al. 2003; Shukla et al. 1991; Williams et al. 2010).

A negative association between BLL and serum IGF-1 concentrations is consistent with lead-induced inhibition of the hypothalamic–pituitary–growth axis. An *in vitro* study in rat pituitary demonstrated that lead blocked the binding of growth hormone releasing hormone to its receptor, suggesting axis inhibition at the level of the pituitary (Lau et al. 1991). Lead could also affect the growth axis at the level of the pituitary through interference with calcium-dependent GH release. In *in vitro* studies of bovine and rat pituitary, other divalent cations such as zinc, nickel, cadmium, and magnesium blocked calcium-dependent

GH release (Carlson 1984; Lorenson et al. 1983), although additional studies are needed to determine whether this effect persists in *in vivo* and whether lead elicits similar actions. Consistent with these potential pituitary-mediated effects, a study of rodent pups (mean BLL, 18 $\mu\text{g/dL}$) demonstrated suppressed growth hormone releasing hormone-stimulated GH release (Camoratto et al. 1993), and in a separate rodent model, high lead exposure (up to mean BLL of 263 $\mu\text{g/dL}$) resulted in lower serum IGF-1 concentrations (Ronis et al. 1998a). Highly lead-exposed prepubertal children ($n = 6$) had decreased mean 24-hr serum GH concentrations and lower serum IGF-1 concentrations before chelation (BLLs > 40 $\mu\text{g/dL}$) compared with after chelation (BLLs ≤ 30 $\mu\text{g/dL}$) (Huseman et al. 1992). In contrast to these studies, the present analysis explored BLLs within currently acceptable ranges and still found a negative association between BLL and IGF-1.

Lead may also inhibit the reproductive axis at the level of the pituitary. Six men with occupational lead exposure (mean BLL, 38.7 $\mu\text{g/dL}$) had blunted LH response to gonadotropin-releasing hormone (GnRH) compared with nine men without occupational exposure (mean BLL, 16 $\mu\text{g/dL}$) (Braunstein et al. 1978). In a study of 77 lead smelter workers (mean BLL, 33 $\mu\text{g/dL}$) and 26 nonworkers (mean BLL, 4.1 $\mu\text{g/dL}$), a subset analysis demonstrated lower GnRH-stimulated follicle-stimulating hormone in 9 workers compared to 11 nonworkers (Erfurth et al. 2001). Also, lead exposure in rodents resulted in decreased serum LH concentration (Ronis et al. 1998b) and increased pituitary LH stores (Klein et al. 1994; Sokol et al. 1998), further suggesting possible pituitary hyporesponsiveness to GnRH. Consistent with these studies of gonadotropin inhibition, lead exposure has been associated with later onset of puberty in our cohort (Williams et al. 2010) and in other adolescent cohorts (Naicker et al. 2010; Selevan et al. 2003).

A lead-induced decrement in IGF-1 may contribute to gonadotropin inhibition and pubertal delay. Specifically, IGF-1 has been shown to activate GnRH *in vitro* (Zhen et al. 1997) and in rodent models (Hiney et al. 1991, 1996), and puberty is delayed in GnRH-specific IGF-1 receptor knock-out mice (Divall et al. 2010). Furthermore, IGF-1 administration to lead-exposed mice with delayed puberty restored pubertal timing (Pine et al. 2006), providing additional evidence for a potential mediating role of IGF-1 in the association between lead exposure and delayed puberty.

In addition to inhibition of GH and gonadotropin release, in animal studies, high lead exposure has been associated with other processes that could lead to growth delay such

as decreased food consumption (Hammond et al. 1989, 1990) and reduced formation of new bone (Hass et al. 1967; Hicks et al. 1996). Future studies should explore whether these effects can be observed in humans with low-level lead exposures.

As far as we are aware, our study is the first to identify puberty as a particularly vulnerable period in which to assess lead's effect on IGF-1. Both the absolute and percent decrease in IGF-1 in association with lead exposure was larger in mid-pubertal boys than in prepubertal or early-pubertal boys. Thus, in our cohort, puberty seemed to be a key time period in which to detect an effect of lead, and this may be generalizable to other environmental epidemiologic studies examining outcomes of growth and associated hormones.

The present study is limited by availability of the BLL measurement at only one time point, leading to an inability to explore other vulnerable windows of exposure, such as exposures during infancy that may have a stronger association with childhood height (Afeiche et al. 2012). Also, none of the participants had a baseline IGF-1 measurement. However, we believe that a prospective evaluation of BLL on subsequent IGF-1 values made for a stronger study design.

Future studies of lead and growth would benefit from measurement of serum insulin-like growth factor-binding protein 3, a less nutritionally dependent measure of GH activity. Inclusion of girls in future studies will also be important, because rodent models suggest that lead's effect on pubertal growth may be more pronounced in males than in females (Ronis et al. 1998a). Furthermore, the net effect of lead on growth in humans cannot be completely understood without information on the association between childhood lead exposures and adult height, so continued longitudinal follow-up through adulthood is warranted for this and other cohorts.

Conclusion

In the present study we found a negative monotonic dose–response association between blood lead levels in boys at 8–9 years of age and their serum IGF-1 concentrations at 10–11 and 12–13 years of age. With increasing attention to environmental exposures and potential health risks, it is essential to better understand effects of low-level lead exposure on key developmental processes such as growth and reproductive development.

REFERENCES

- Afeiche M, Peterson KE, Sanchez BN, Schnaas L, Cantonwine D, Ettinger AS, et al. 2012. Windows of lead exposure sensitivity, attained height, and body mass index at 48 months. *J Pediatr* 160(6):1044–1049.
- Ballew C, Khan LK, Kaufmann R, Mokdad A, Miller DT, Gunter EW. 1999. Blood lead concentration and children's anthropometric dimensions in the Third National Health

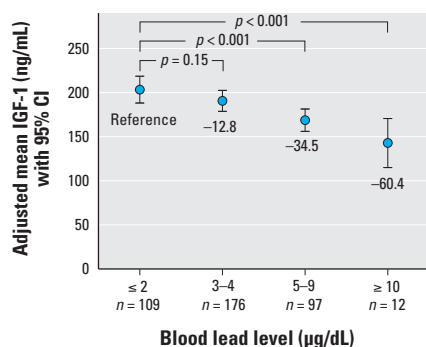


Figure 2. Adjusted mean IGF-1 concentrations by BLL category. Compared with BLL ≤ 2 $\mu\text{g/dL}$, the higher BLL levels of 5–9 $\mu\text{g/dL}$ and ≥ 10 $\mu\text{g/dL}$ were associated with significantly lower mean IGF-1 concentrations ($p < 0.001$ for both comparisons).

- and Nutrition Examination Survey (NHANES III), 1988–1994. *J Pediatr* 134(5):623–630.
- Bellinger DC, Stiles KM, Needleman HL. 1992. Low-level lead exposure, intelligence and academic achievement: a long-term follow-up study. *Pediatrics* 90(6):855–861.
- Betts KS. 2012. CDC Updates Guidelines for Children's Lead Exposure. *Environ Health Perspect* 120:A268.
- Braunstein GD, Dahlgren J, Loriaux DL. 1978. Hypogonadism in chronically lead-poisoned men. *Infertility* 1(1):33–51.
- Burns JS, Williams PL, Sergeev O, Korrick SA, Lee MM, Revich B, et al. 2012. Serum concentrations of organochlorine pesticides and growth among Russian boys. *Environ Health Perspect* 120:303–308.
- Camoratto AM, White LM, Lau YS, Ware GO, Berry WD, Moriarty CM. 1993. Effect of exposure to low level lead on growth and growth hormone release in rats. *Toxicology* 83(1–3):101–114.
- Carlson HE. 1984. Inhibition of prolactin and growth hormone secretion by nickel. *Life sciences* 35(17):1747–1754.
- Divall SA, Williams TR, Carver SE, Koch L, Bruning JC, Kahn CR, et al. 2010. Divergent roles of growth factors in the GnRH regulation of puberty in mice. *J Clin Invest* 120(8):2900–2909.
- Erfurth EM, Gerhardsson L, Nilsson A, Rylander L, Schutz A, Skerfving S, et al. 2001. Effects of lead on the endocrine system in lead smelter workers. *Arch Environ Health* 56(5):449–455.
- Gump BB, Stewart P, Reihman J, Lonky E, Darvill T, Matthews KA, et al. 2005. Prenatal and early childhood blood lead levels and cardiovascular functioning in 9½ year old children. *Neurotoxicol Teratol* 27(4):655–665.
- Hammond PB, Chernausek SD, Succop PA, Shukla R, Bornschein RL. 1989. Mechanisms by which lead depresses linear and ponderal growth in weanling rats. *Toxicol Appl Pharmacol* 99(3):474–486.
- Hammond PB, Minnema DJ, Shulka R. 1990. Lead exposure lowers the set point for food consumption and growth in weanling rats. *Toxicol Appl Pharmacol* 106(1):80–87.
- Hass GM, Landerholm W, Hemmens A. 1967. Inhibition of intercellular matrix synthesis during ingestion of inorganic lead. *Am J Pathol* 50(5):815–847.
- Hicks DG, O'Keefe RJ, Reynolds KJ, Cory-Slechta DA, Puzas JE, Judkins A, et al. 1996. Effects of lead on growth plate chondrocyte phenotype. *Toxicol Appl Pharmacol* 140(1):164–172.
- Hiney JK, Ojeda SR, Dees WL. 1991. Insulin-like growth factor I: a possible metabolic signal involved in the regulation of female puberty. *Neuroendocrinology* 54(4):420–423.
- Hiney JK, Srivastava V, Nyberg CL, Ojeda SR, Dees WL. 1996. Insulin-like growth factor I of peripheral origin acts centrally to accelerate the initiation of female puberty. *Endocrinology* 137(9):3717–3728.
- Huseman CA, Varma MM, Angle CR. 1992. Neuroendocrine effects of toxic and low blood lead levels in children. *Pediatrics* 90(2 pt 1):186–189.
- Kafourou A, Touloumi G, Makropoulos V, Loutradi A, Papanagiotou A, Hatzakis A. 1997. Effects of lead on the somatic growth of children. *Arch Environ Health* 52(5):377–383.
- Karmaus W, Brooks KR, Nebe T, Witten J, Obi-Osius N, Kruse H. 2005. Immune function biomarkers in children exposed to lead and organochlorine compounds: a cross-sectional study. *Environ Health* 4(1): 5; doi:10.1186/1476-069X-4-5 [Online 14 April 2005].
- Klein D, Wan YJ, Kamyab S, Okuda H, Sokol RZ. 1994. Effects of toxic levels of lead on gene regulation in the male axis: increase in messenger ribonucleic acids and intracellular stores of gonadotrophs within the central nervous system. *Biol Reprod* 50(4):802–811.
- Lanphear BP, Hornung R, Khoury J, Yolton K, Baghurst P, Bellinger DC, et al. 2005. Low-level environmental lead exposure and children's intellectual function: an international pooled analysis. *Environ Health Perspect* 113:894–899.
- Lau YS, Camoratto AM, White LM, Moriarty CM. 1991. Effect of lead on TRH and GRF binding in rat anterior pituitary membranes. *Toxicology* 68(2):169–179.
- Little BB, Spalding S, Walsh B, Keyes DC, Wainer J, Pickens S, et al. 2009. Blood lead levels and growth status among African-American and Hispanic children in Dallas, Texas—1980 and 2002: Dallas Lead Project II. *Ann Hum Biol* 36(3):331–341.
- Lorenson MY, Robson DL, Jacobs LS. 1983. Divalent cation inhibition of hormone release from isolated adeno-hypophysial secretory granules. *J Biol Chem* 258(14):8618–8622.
- Martinchik AN, Baturin AK, Baeva VS, Feoktistova AI, Piatnitskaia IN, Azizbekian GA, et al. 1998. Development of a method of studying actual nutrition according to analysis of the frequency of consumption of food products: creation of a questionnaire and general evaluation of the reliability of the method [in Russian]. *Vopr Pitan* 3:8–13.
- Menke A, Muntner P, Batuman V, Silbergeld EK, Guallar E. 2006. Blood lead below 0.48 µmol/L (10 µg/dL) and mortality among US adults. *Circulation* 114(13):1388–1394.
- Naicker N, Norris SA, Mathee A, Becker P, Richter L. 2010. Lead exposure is associated with a delay in the onset of puberty in South African adolescent females: findings from the Birth to Twenty cohort. *Sci Total Environ* 408(21):4949–4954.
- Pine MD, Hiney JK, Dearth RK, Bratton GR, Dees WL. 2006. IGF-1 administration to prepubertal female rats can overcome delayed puberty caused by maternal Pb exposure. *Reprod Toxicol* 21(1):104–109.
- Rockett HR, Breitenbach M, Frazier AL, Witschi J, Wolf AM, Field AE, et al. 1997. Validation of a youth/adolescent food frequency questionnaire. *Prev Med* 26(6):808–816.
- Ronis MJ, Badger TM, Shema SJ, Roberson PK, Templer L, Ringer D, et al. 1998a. Endocrine mechanisms underlying the growth effects of developmental lead exposure in the rat. *J Toxicol Environ Health A* 54(2):101–120.
- Ronis MJ, Gandy J, Badger T. 1998b. Endocrine mechanisms underlying reproductive toxicity in the developing rat chronically exposed to dietary lead. *J Toxicol Environ Health A* 54(2):77–99.
- Schwartz J, Angle C, Pitcher H. 1986. Relationship between childhood blood lead levels and stature. *Pediatrics* 77(3):281–288.
- Selevan SG, Rice DC, Hogan KA, Euling SY, Pfahles-Hutchens A, Bethel J. 2003. Blood lead concentration and delayed puberty in girls. *N Engl J Med* 348(16):1527–1536.
- Shukla R, Dietrich KN, Bornschein RL, Berger O, Hammond PB. 1991. Lead exposure and growth in the early preschool child: a follow-up report from the Cincinnati Lead Study. *Pediatrics* 88(5):886–892.
- Silbergeld A, Litwin A, Bruchis S, Varsano I, Laron Z. 1986. Insulin-like growth factor I (IGF-I) in healthy children, adolescents and adults as determined by a radioimmunoassay specific for the synthetic 53–70 peptide region. *Clin Endocrinol* 25(1):67–74.
- Sokol RZ, Berman N, Okuda H, Raum W. 1998. Effects of lead exposure on GnRH and LH secretion in male rats: response to castration and α -methyl-*p*-tyrosine (AMPT) challenge. *Reprod Toxicol* 12(3):347–355.
- Vandenberg LN, Colborn T, Hayes TB, Heindel JJ, Jacobs DR Jr, Lee DH, et al. 2012. Hormones and endocrine-disrupting chemicals: low-dose effects and nonmonotonic dose responses. *Endocr Rev* 33(3):378–455.
- WHO (World Health Organization). 2011. The WHO Child Growth Standards. Available: <http://www.who.int/childgrowth/en/> [accessed 12 June 2013].
- Williams PL, Sergeev O, Lee MM, Korrick SA, Burns JS, Humblet O, et al. 2010. Blood lead levels and delayed onset of puberty in a longitudinal study of Russian boys. *Pediatrics* 125(5):e1088–e1096.
- Zhen S, Zakaria M, Wolfe A, Radovick S. 1997. Regulation of gonadotropin-releasing hormone (GnRH) gene expression by insulin-like growth factor I in a cultured GnRH-expressing neuronal cell line. *Mol Endocrinol* 11(8):1145–1155.