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Lubiprostone Increases Small Intestinal Smooth Muscle Contractions Through a Prostaglandin E Receptor 1 (EP₁)-mediated Pathway

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Background/Aims
Lubiprostone, a chloride channel type 2 (ClC-2) activator, was thought to treat constipation by enhancing intestinal secretion. It has been associated with increased intestinal transit and delayed gastric emptying. Structurally similar to prostones with up to 54% prostaglandin E₂ activity on prostaglandin E receptor 1 (EP₁), lubiprostone may also exert EP₁-mediated procontractile effect on intestinal smooth muscles. We investigated lubiprostone’s effects on intestinal smooth muscle contractions and pyloric sphincter tone.

Methods
Isolated murine small intestinal (longitudinal and circular) and pyloric tissues were mounted in organ baths with modified Krebs solution for isometric recording. Basal muscle tension and response to electrical field stimulation (EFS; 2 ms pulses/10 V/6 Hz/30 sec train) were measured with lubiprostone (10⁻¹⁰⁻¹⁰⁻⁵ M) ± EP₁ antagonist. Significance was established using Student t test and P < 0.05.

Results
Lubiprostone had no effect on the basal tension or EFS-induced contractions of longitudinal muscles. With circular muscles, lubiprostone caused a dose-dependent increase in EFS-induced contractions (2.11 ± 0.88 to 4.43 ± 1.38 N/g, P = 0.020) that was inhibited by pretreatment with EP₁ antagonist (1.69 ± 0.70 vs. 4.43 ± 1.38 N/g, P = 0.030). Lubiprostone had no effect on circular muscle basal tension, but it induced a dose-dependent increase in pyloric basal tone (1.07 ± 0.10 to 1.97 ± 0.86 fold increase, P < 0.05) that was inhibited by EP₁ antagonist.

Conclusions
In mice, lubiprostone caused a dose-dependent and EP₁-mediated increase in contractility of circular but not longitudinal small intestinal smooth muscles, and in basal tone of the pylorus. These findings suggest another mechanism for lubiprostone’s observed clinical effects on gastrointestinal motility.

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Key Words
Gastrointestinal motility; Intestine, small; Lubiprostone; Receptors, prostaglandin E
Introduction

Lubiprostone, a chloride channel type 2 (ClC-2) activator, has been approved for the treatment of chronic constipation and constipation-predominant irritable bowel. Prior clinical study performed in healthy volunteers showed that lubiprostone is associated with increased intestinal transit, increased fasting gastric volume and delayed gastric emptying. The proposed primary mechanism of action of lubiprostone in the gastrointestinal tract is increased chloride ion transport into the intestinal lumen caused by opening of ClC-2, leading to increased intestinal secretion and enhanced mass transit. Nausea is the most common side effect of lubiprostone, reported in up to 31% of patients in one study. Several possible explanations for nausea have been suggested, including delayed gastric emptying, small intestinal distention secondary to increased gastric secretion, change in gastrointestinal sensation, or additional actions of lubiprostone on gastrointestinal motility. However, the underlying mechanism of nausea associated with lubiprostone remains unclear. In addition, lubiprostone has been shown to accelerate overall colonic transit without significantly changing the rate of ascending colon emptying. With the proximal colon likely reabsorbing the increased fluid load from the small intestine, it was postulated that a primary motor effect on the colon beyond the ascending portion may be responsible for this finding. Similarly, the presence of any possible direct smooth muscle effect of lubiprostone on the rest of the gastrointestinal tract has been suggested. In fact, a recent study has shown that lubiprostone increased contraction of rat and human stomach longitudinal muscle, an action that was inhibited by prostaglandin E receptor 1 (EP1) antagonist. Such effects may also be responsible for the clinical effect of lubiprostone such as nausea seen in humans.

Although lubiprostone is structurally similar to prostone, a bicyclic fatty acid compound derived from prostaglandin E1 (PGE1), its level of interaction with E-type prostanoid receptors and the possible resultant clinical effect are not completely clear. While prior studies have suggested that lubiprostone only has very weak activity on prostaglandin receptors, it has also been reported to have up to 54% PGE2 activity on EP1. Recent studies have suggested that lubiprostone may also exert prostaglandin effects on non-gastrointestinal tissues. Given lubiprostone’s structural similarity to prostone, reported activity on EP1, and effect on intestinal transit, we postulated that it may have direct procontractile effects on both phasic and tonic smooth muscles in the small intestine, possibly involving EP1 activation. In this study, we investigated the effect of lubiprostone on the basal tone and neuronally-stimulated contraction of the phasic intestinal smooth muscle and the tonic pyloric sphincter muscle in mice. We also studied the possible interaction of lubiprostone on EP1 using a specific antagonist.

Materials and Methods

Tissue Isolation and Preparation

Ten adult wild-type mice were sacrificed by carbon dioxide asphyxiation followed by cervical dislocation. All efforts were made to minimize the number of animals used. Immediately after death of the mice, a midline incision was made exposing the peritoneal cavity and contents. The stomach and entire small intestine were excised. The tissues were placed immediately in a pre-mixed Krebs solution (118 mM NaCl, 4.8 mM KCI, 1.2 mM KH2PO4, 1.2 mM MgSO4, 2.5 mM CaCl2, 25 mM NaHCO3 and 11 mM glucose), which was previously equilibrated with 5% CO2 in O2 at a temperature of 37°C. The pylorus was isolated and 1 cm transverse sections of the small intestine were obtained. The mucosa was left intact. The tissues were thoroughly flushed with the Krebs solution to remove any remaining food or bile.

For small intestine, longitudinal muscles were isolated by using 1 cm length segments of small intestinal tissues. Both ends of the longitudinal muscles were attached to ring clips that were mounted on the isometric force transducers. For circular smooth muscles, the small intestinal tissues were cut into smaller transverse segments of 0.2-0.3 cm width. The muscle ring prep was then mounted on the force transducers allowing measurement of circular muscle contraction. The pyloric tissues were similarly mounted in a ring prep fashion of approximately 0.5 cm in width for measurement of basal tone.

Tissue Bath Experiment

Isolated small intestinal (longitudinal and circular) and pyloric sphincter tissues were mounted in 5 mL organ baths with Krebs solution for isometric recording. The Krebs solution was maintained in the organ baths by bubbling with 5% CO2/95% O2 at 37°C. Isometric force transducers measured changes in muscle tension of the tissues. Tissue samples were suspended under 4 mN tension for isometric recording between 2 platinum ring electrodes. This preload was noted to be in the appropriate range of optimal electrical field stimulation (EFS)- and carbachol-in-
duced contractions. The EFS parameters were chosen from prior experiments that demonstrated tetrodotoxin-inhibitable neurogenic responses. For this experiment, peak contraction responses were achieved using 30 seconds train of biphasic square-wave pulses of 2 ms pulse width at 10 volts and frequency of 6 Hz. Baseline measurement was then taken with EFS stimulation in the absence of lubiprostone. Sequential addition of escalating doses of lubiprostone (0.0001-10 μM) was then performed to construct a cumulative concentration-response curve for neuronally-stimulated smooth muscle contraction. Similar lubiprostone concentration-response curve for EFS-stimulated smooth muscle contraction was constructed in the presence of EP1 antagonist (SC19220; Sigma, Saint Louis, MO, USA) to evaluate the involvement of the EP1 cascade in the lubiprostone-mediated smooth muscle contraction response.

In a separate set of experiments, the basal tone of the smooth muscle of small intestines and the pylorus were studied in the absence of EFS. By similarly suspending tissue samples under 4 mN for isometric recording in 5 mL wells of bubbling Krebs solution, the basal muscle tone of the muscles in response to escalating concentration of lubiprostone were recorded. At the end of the experiment, the tissues were placed on a slide and dried in an oven for obtaining dry weight. Tension measurements were normalized for the mass of tissue.

Data Acquisition and Analysis

Data acquisition and analysis was performed using the LabChart v. 7.0 software (ADInstruments, Colorado Springs, CO, USA). In the EFS experiments, the difference between the maximal muscle tension post-EFS stimulation and the minimal muscle tension pre-EFS stimulation was calculated for baseline and each lubiprostone concentration. This change in muscle tension pre- and post-EFS stimulation represents the EFS-mediated pro-contractile response. In experiments for basal tone without the use of EFS, the maximal muscle tension reached with each lubiprostone concentration was used. The muscle tension was normalized for the dry weight of the tissue used in each experiment. All data are expressed as means ± standard error of mean. Statistical significance was established using two-tailed Student’s t test and P < 0.05.

Chemical Reagents Used

All drugs were freshly prepared before use. Lubiprostone was extracted using a 24 gauge needle directly from commercially available 24 microgram capsules and suspended in 100% ethanol solution. Different concentrations of lubiprostone were obtained by further dilution with 100% ethanol. The EP1 antagonist (SC19220; Sigma) was dissolved in pre-synthesized Krebs solution.

Ethical Approval

This study was approved by the VA Health System Institutional Animal Care and Use Committee prior to inception of this investigation.

Results

Circular Muscles

In the measurement of EFS-stimulated smooth muscle contractions, lubiprostone induces a dose-dependent increase in circular smooth muscle. The addition of the prostaglandin E receptor 1 (EP1) antagonist, SC19220, attenuated the lubiprostone-induced response in muscle contractions. EFS, electrical field stimulation.

![Figure 1](image1.png)

**Figure 1.** Sample tracing demonstrating increasing electrical field stimulation-induced contractions of intestinal circular smooth muscle with increasing concentrations of lubiprostone. EFS, electrical field stimulation.

![Figure 2](image2.png)

**Figure 2.** Electrical field stimulation induced contractions of the circular smooth muscle demonstrated a dose-response relationship with increasing concentrations of lubiprostone (no antagonist). The addition of the prostaglandin E receptor 1 (EP1) antagonist, SC19220, attenuated the lubiprostone-induced response in muscle contractions. EFS, electrical field stimulation.
Lubiprostone Induces EP1-mediated Contraction

Figure 3. Basal circular smooth muscle tone of the small intestine did not show any significant differences with increasing concentrations of lubiprostone.

Figure 4. Longitudinal smooth muscle tone of the small intestine demonstrated no significant changes with increasing concentrations of lubiprostone.

Figure 5. Neuronal contractions of the longitudinal smooth muscles as simulated by electrical field stimulation showed no response to increasing concentrations of lubiprostone.

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Figure 6. Neuronal contractions of the longitudinal smooth muscles as simulated by electrical field stimulation showed no response to increasing concentrations of lubiprostone.

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Figure 6. The pyloric sphincter tone revealed a mild, significant increase in basal tone with lubiprostone. This increase in tone was inhibited by the prostaglandin E receptor 1 (EP1) antagonist SC19220.

As lubiprostone treats constipation by reducing intestinal transit time, we hypothesized that it may also have a procontractile effect on the phasic intestinal smooth muscles. Using EFS to simulate neuronal stimulation, our study demonstrated that lubiprostone indeed increases the circular smooth muscle contractions in a dose-dependent manner, while it has no effect on the longitudinal muscles. The disparity in the response of these muscles is consistent with the known mechanism of propulsion of intestinal contents, which requires coordinated but reciprocal activities of longitudinal and circular muscles during peristalsis. With each peristalsis, circular muscles contract oral and relax caudal to the site of stimulation, and longitudinal muscles contract and relax in reverse to circular muscles to provide the coordinated forward propulsion. Therefore, longitudinal muscles exert ascending relaxation and descending contraction with each peristalsis. With these physiological properties of intestinal smooth muscles, our results can explain another mechanism for the accelerated intestinal transit caused by lubiprostone besides the volume transit through its effect on CIC-2. The enhancement of contractions in one type of muscles (circular) coupled with the lack of effect in the other (longitudinal) by lubiprostone could result in stronger forward propulsive force.

Our study has also shown that the procontractile effect on circular smooth muscle tension observed may have been achieved through an EP1-mediated pathway, as pretreatment with an EP1 antagonist attenuated the procontractile effect of lubiprostone. Previous studies have demonstrated that PGE2 and its different receptor subtypes exert varying effects on gastrointestinal motility. In particular, in a study of circular smooth muscles isolated from pig and guinea-pig ileum, EP1 agonist enhances contraction of the muscle cells. EP1 are G protein-coupled receptors that regulate Ca2+-channel gating and they have been shown to increase Ca2+ concentration in Chinese hamster ovary cells without detectable phosphatidylinositide response. Enhanced smooth muscle contractions, therefore, may be induced by EP1 via changes in intracellular Ca2+ concentrations of smooth muscle cells. While different subtypes of PGE2 receptors are present in varying concentrations throughout the gastrointestinal tract, EP1 have been detected in the muscularis mucosa, enteric glial cells, and smooth muscle layers of the small intestine. Therefore, the increased muscular contractions by lubiprostone could be a result of effects on muscle or glial cells. Prior studies have suggested roles for enteric glial cells in neurotransmission by mediating intercellular Ca2+ waves and muscular contraction by increasing intracellular Ca2+ concentration. Clinical studies of pa-
tients with severe, chronic, slow-transit constipation demonstrate a loss of enteric glial cells and interstitial cells of Cajal, but the specific roles of these cells in EP1-mediated contraction remains poorly characterized. The lubiprostone-mediated increase in circular muscle contractions during EFS observed in this study may have resulted from activation of EP1 located on either muscle or enteric glial cells.

In the study of pyloric muscles, lubiprostone increased the basal tone of the pyloric sphincter in a dose-dependent manner. This procontractile effect was attenuated by pretreatment with an EP1 antagonist, suggesting that the lubiprostone acted through an EP1-mediated pathway. Prior studies have demonstrated the presence of EP1 in the mucosal and smooth muscle layers of the gastric body and antrum. However, the existence of EP1 or other PGE2 receptors in the pylorus remains unclear. Our results demonstrated the likely presence of EP1 on pyloric tissues and the possible effect of activation of the EP1 pathway. These pharmacologic effects will need to be confirmed on human tissues.

One of the main side effects of lubiprostone is nausea, which may affect up to a third of the patients. The underlying mechanism of lubiprostone-induced nausea is unclear. However, several possible processes have been suggested, including delayed gastric emptying, small intestinal distention from increased gastric secretion, change in gastrointestinal sensation, or alteration of gastrointestinal motility. The elevation of basal pyloric sphincter tone caused by lubiprostone may contribute to the nausea symptoms by delaying gastric emptying or in the form of pylorospasm. The resultant nausea may be particularly pronounced as lubiprostone increases gastric secretion through the activation of chloride channels. Gastric distention from decreased pyloric outflow and increased gastric secretion may produce nausea symptoms, especially in those with decreased compliance of the gastric wall.

Our results are significant in demonstrating prostaglandin-like actions of lubiprostone in the gastrointestinal tract, in addition to its known specific chloride channel effects. It is now less clear whether the clinical effects observed in patients with constipation result predominantly from activation of CIC-2, as formerly believed, or from its prostaglandin-like effects on intestinal smooth muscles. Recent evidence suggests that lubiprostone acts predominantly on the classical cystic fibrosis transmembrane conductance regulator (CFTR), and no chloride secretion could be observed using rectal epithelial cells from a patient homozygous for defective (F580del) CFTR. In contrast, lubiprostone has been demonstrated in a recent case series to relieve constipation in patients with cystic fibrosis. Thus the clinical benefit seen in these patients with defective CFTR may be related to lubiprostone’s action on EP1, as demonstrated by our results. Therefore, the general mechanisms of lubiprostone in relieving constipation may be multimodal.

Through understanding the underlying mechanism of lubiprostone effects, the newly described pathway of action for lubiprostone may open potential new use for the medication. For example, patients with small intestinal pseudo-obstruction or ileus may respond to the procontractile effects of lubiprostone, while those with bile reflux may improve due to the increase in basal pyloric sphincter tone. This may also enhance bicarbonate secretion and have cytoprotective effects, akin to misoprostol. On the other hand, potential negative consequences of PGE2 receptor activation should also be considered. Previous studies have demonstrated that blockade of EP1 and EP3 inhibits the development of colon cancer in mouse models. Given lubiprostone’s demonstrated agonist effect on EP1, its chronic use may result in adverse consequence. Further studies are required to delineate lubiprostone’s other potential short- and long-term clinical effects resulting from its prostaglandin-like activities. Orally administered lubiprostone is rapidly metabolized, not only by the liver, but also immediately on the mucosal surface. Thus the intact compound is not detectable in the blood. Therefore, it is possible that the observed pro-contractile effects of lubiprostone are mediated by its metabolites. Also its primary site of action may be the mucosa, and not necessarily directly on the smooth muscles. To understand lubiprostone’s actual clinical effect, we chose to study intact muscle preparation without inhibiting adrenergic or cholinergic effects and without removing the mucosa. Further mechanistic studies discerning its site of action, metabolites, and pathways would be warranted. Its interactions with other PGE2 receptor subtypes should also be explored.

In conclusion, lubiprostone, previously described as a selective chloride channel activator with little prostaglandin effect, caused a dose-dependent and EP1-mediated increase in contractility of circular but not longitudinal small intestinal smooth muscles in mice. It had no effect on the basal tone of these phasic muscles. However, in the tonic pyloric sphincter, lubiprostone caused a dose-dependent increase in basal tone that was attenuated by an EP1 antagonist. These findings suggest potential prostaglandin-like activities of lubiprostone. They also provide another mechanism for the previously-reported accelerated intestinal transit and delayed gastric emptying in normal individuals given lubiprostone.
References


