The Methanol Dehydrogenase Gene, \textit{mxaF}, as a Functional and Phylogenetic Marker for Proteobacterial Methanotrophs in Natural Environments

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Abstract

The \textit{mxaF} gene, coding for the large ($\alpha$) subunit of methanol dehydrogenase, is highly conserved among distantly related methylotrophic species in the Alpha-, Beta- and Gammaproteobacteria. It is ubiquitous in methanotrophs, in contrast to other methanotroph-specific genes such as the \textit{pmoA} and \textit{mmoX} genes, which are absent in some methanotrophic proteobacterial genera. This study examined the potential for using the \textit{mxaF} gene as a functional and phylogenetic marker for methanotrophs. \textit{mxaF} and 16S rDNA gene phylogenies were constructed based on over 100 database sequences of known proteobacterial methanotrophs and other methylotrophs to assess their evolutionary histories. Topology tests revealed that \textit{mxaF} and 16S rDNA genes of methanotrophs do not show congruent evolutionary histories, with incongruencies in methanotrophic taxa in the Methylococcaceae, Methyloidybacaceae, and Beijerinckiaceae. However, known methanotrophs generally formed coherent clades based on \textit{mxaF} gene sequences, allowing for phylogenetic discrimination of major taxa. This feature highlights the \textit{mxaF} gene’s usefulness as a biomarker in studying the molecular diversity of proteobacterial methanotrophs in nature. To verify this, PCR-directed assays targeting this gene were used to detect novel methanotrophs from diverse environments including soil, peatland, hydrothermal vent mussel tissues, and methanotroph isolates. The placement of the majority of environmental \textit{mxaF} gene sequences in distinct methanotroph-specific clades (Methyloidybacaceae and Methylococcaceae) detected in this study supports the use of \textit{mxaF} as a biomarker for methanotrophic proteobacteria.

Introduction

Atmospheric methane contributes to $\sim$20% of the total radiative forcing by long-lived greenhouse gases globally. Due to the relatively short lifetime of atmospheric methane ($\sim$9 yrs) compared to CO$_2$, reductions in atmospheric methane flux would have an immediate impact on global warming [1]. Microbial methane oxidation is the only major biological sink of methane [2]. With the exception of members of the phylum Verrucomicrobia [3], and the yet to be cultured anaerobic methanotrophs (ANME) [4,5], and denitrifying methanotrophs of the ‘NC10’ phylum, aerobic proteobacterial methanotrophs are the only currently known groups of microorganisms capable of oxidizing methane as their sole carbon source [6,7,8], thus reducing atmospheric methane flux.

Aerobic proteobacterial methanotrophs are unique among the larger group of methylotrophic bacteria (which are able to utilize C$_1$ or one-carbon compounds) in that they oxidize methane to methanol, before subsequent metabolic reactions that are shared with other methylotrophs. In contrast, non-methanotrophic methylotrophs are unable to utilize methane, but can grow on other C$_1$ compounds (e.g., methanol, methylated amines, formate, or formamide) [9,10]. The vast majority of known aerobic methanotrophs belong to the Proteobacteria. Known exceptions include the phylum Verrucomicrobia, whose pathways for methanotrophy are still poorly understood [3,11,12], and enrichments of the uncultured methanotrophs such as \textit{Candidatus ‘Methylomirabilis oxyfera’} affiliated with the ‘NC10’ phylum, which appear to be incapable of oxidizing methane at low ($\approx$3%) oxygen levels in the laboratory [7,8,13]. Proteobacterial methanotrophs are placed in the families Methyloidybacaceae and Beijerinckiaceae in the Alphaproteobacteria [14,15].

The similarities in physiological characteristics, and the generally highly conserved nature of the 16S rRNA and functional gene sequences of proteobacterial methanotrophs, have enabled the use of targeted PCR primers for gene amplification to describe methanotroph diversity [15,16]. However, the 16S rRNA gene is non-protein coding (i.e., not linked directly to methanotroph physiology), and therefore does not directly determine function. For example, it cannot be determined whether 16S rDNA environmental sequences placed close to, but outside of, known
monophyletic methanotroph clades are indeed methanotrophic [17,19]. In comparison, functional genes for enzymes found in methanotroph metabolic pathways are directly related to physiology [19]. The initial step of methane oxidation to methanol by proteobacterial methanotrophs is mediated by particulate and/or soluble methane monoxygenases (MMOs) [15,20]. Genes in this step, such as pmOA [encoding a subunit of the particulate methane monoxygenase (pMMO)] and mmaX [encoding a subunit of the soluble methane monoxygenase (sMMO)] have been used for describing methanotroph diversity [17]. However, the pmOA gene is not present in the genera Methylocella and Methylophaga [21,22,23], and the mmaX gene is present only in a few strains of methanotrophs [15,24]. Hence, the use of both pmOA and mmaX genes in PCR-based studies results in the underestimation of methanotroph diversity and suggests that an alternative is needed.

The mxaF gene was first proposed as a functional gene probe for methyloths by McDonald and Marrell [19]. Methane oxidation gene cluster (mxaF) encodes the α-subunit of the enzyme methanol dehydrogenase (MDH), an enzyme containing a pyrroloquinoline quinone (PQQ) cofactor that oxidizes methanol to formaldehyde in the second step of the methane oxidation pathway, following oxidation of methane to methanol [20,25,26,27,28]. Methanol dehydrogenase is common to all methanotrophs in the Proteobacteria [15,19], but is absent in members of the phylum Verrucomicrobia, which possess a homolog of the mxaF gene, called xoxF gene, a gene of unknown function [29,30,31,32]. Sequences of the mxaF gene are highly conserved in methylotrophic species in the Alpha-, Beta- and Gammaproteobacteria, reflecting 16S rDNA phylogeny conserved in methylotrophic species in the Alpha-, Beta- and Gammaproteobacteria. Analysis of mxaF gene phylogenies of methanotrophs from the GenBank [35] database indicates the extent of horizontal gene transfer in other proteobacterial methanotrophic species, and which taxa were involved [17]. Indications of horizontal gene transfer have also been conducted. In the only study of methylotroph diversity and suggests that an alternative is needed.

This study addresses the following questions, (a) Does the extensive mxaF gene phylogeny discriminate between methanotrophic and other methylotrophic sequences available in genetic databases? (b) Are the mxaF and 16S rRNA gene phylogenies congruent for methanotrophs in the families Methylocystaceae, Methylococcaceae, and Beijerinkiaeae? (c) Can we tell whether mxaF gene sequences from published databases, as well as mxaF sequences obtained in this study from surveys of diverse environments (forest soils, peat and Sphagnum moss, and symbiont-hosting mussel gills) and from methanotroph cultures. Overall, these analyses advance the study of methanotrophy diversity by showing that the mxaF gene consistently places methanotroph sequences in resolved phylogenetic clusters for all known members of the families Methylocystaceae and Methylococcaceae, and has the potential to elucidate the roles methanotrophs play in natural environments.

Materials and Methods

Cultures

Two mxaF gene sequences were determined in this study for cultured species of the Methylococcaceae, Methylophilonas rubra and Methylobacter latus, provided by J. Semrau and R. Knowles, respectively. Positive control cultures used in this study included two members of the Methylocystaceae and a member of the Methylococcaceae for which mxaF gene sequences are already available; Methylosinus trichosporium OB3b, Methylocystis parvus OBBP, Methylophilonas rubra, respectively.

Collections and Site Descriptions

Samples for DNA extraction or bacterial isolation were collected from four distinct habitats for DNA extraction or bacterial isolation: (a) Four soil samples, weighing ~15 g, from the organic horizon of long-term nitrogen-amended and control pine and hardwood forest soils of the Harvard Forest Long-Term Ecological Research (LTER) site, previously shown to oxidize methane at atmospheric levels, in Petersham, Massachusetts (42°30’N, 72°10’W) in 2004 [18,36], (b) Sphagnum recurvum moss and peat from Crystal Bog (Vilas county, Wisconsin), a 7 ha poor fen enclosing a 2.5 m deep, 0.5 ha, dystrophic lake on the North Temperate Lakes LTER site in 2005 [46’00’30”N 89°36’30”W] [37], (c) two species of methanotroph-hosting mussels (n = 3 per site) from the Mid-Atlantic Ridge (MAR) deep-sea vent sites sampled in 2003 using DSV Alvin: Bathymodiolus azoricus from Lucky Strike (LS; 37°17’N, 32°16’W; 1693 m deep) and Rainbow (RB; 36°13’N, 33°54’W, 2300 m deep) and Bathymodiolus puteoserpentis from Logatchev (LO; 14°45’N, 44°58’W; 3027 m deep) [38,39], and (d) the Halls Brook Holding Area (HBHA), an artificial lake in the Aberjona Watershed, sampled in 2004, near Boston, MA, which becomes stratified during the summer months, whereby the bottom depths become anoxic [40]. No specific permissions were required for collecting samples in these locations because samples did not involve endangered or protected species, Harvard Forest is owned by Harvard University and permission is granted to research employees, and Crystal Bog and Halls Brook Holding Area are located on public land.

Methanotroph Isolation, DNA Extraction, and Purification

Methanotrophic HBHA isolate 1 and HBHA isolate 2 were isolated from water collected at the oxic-anoxic interface at HBHA (~1 m depth) using sterile Tygon® tubing connected to a peristaltic pump, and injected into sterile flasks containing Nitrato
Mineral Salts (NMS) minimal medium [41], under 90:10 air-methane headspace, and incubated with shaking at room temperature. Both isolates failed to grow in the absence of methane and oxygen. Purified DNA from methanotrophic cultures was obtained using the Wizard® Genomic DNA Purification Kit (Promega Inc.). DNA from endosymbiont-containing mussel gill tissue and forest soils was obtained as previously described [18,30]. DNA was extracted from HBHA isolates, peat and Sphagnum moss following the method as previously described [10].

**Primer Design, PCR Amplification, Cloning, and Sequencing**

Methylotroph *mxaF*-specific PCR primers F1003 and R1561 ([33]; Table 1) sequences were verified through BLAST [42] to determine their specificity to proteobacterial methanotrophs in the GenBank [35] database, and used to amplify partial *mxaF* gene sequences from soils, peat, *Sphagnum* moss, methanotrophic HBHA isolates 1 and 2, and control methanotroph cultures. The amplified region encompasses three amino acid residues (out of eight) in the MDH active sites – Asn-261, Asp-303 and Arg-331 – based on the amino acid sequence of *Methylobacterium extorquens* MDH [26]. BLAST searches of the primer pair F1003 and R1561 retrieved only proteobacterial *mxaF* genes, but not that of *Candidateus* ‘Methylomirabilis oxyfera.’

Additionally, degenerate primers F1003degen and R1561degen (Table 1) were created after comparing amino acid sequences of MDH retrieved from the GenBank database and identifying degenerate base positions where more than one nucleotide codon specified the same amino acid residue. The degenerate primers were used to PCR amplify *mxaF* genes from DNA extracts of mussel tissues, as primers F1003 and R1561 failed to PCR amplify *mxaF* genes from the same DNA extracts. The same primers were also used to PCR amplify *mxaF* genes from *Sphagnum* moss in order to verify these primers also PCR amplify *mxaF* sequences outside of the Methyllococaceae. All PCR reaction mixtures contained 1 × PCR buffer (Qiagen Inc.), 2.5 mM MgCl$_2$, 300 mM final concentration of each dNTP, a 1.0 μM final concentration of each primer, 1.0 U of Taq polymerase, and approximately 300–800 ng of template DNA in a final volume of 25 μL. PCR conditions were: denaturation at 94°C for 45 sec, primer annealing at 60°C for 1 min, extension at 72°C for 1.5 min for 30 cycles, and a final 10 min extension at 72°C. PCR products of the expected size (~550 bp for *mxaF* gene) were purified, cloned, and sequenced (≥3 clones per sample).

**Cloning and Sequencing**

PCR products of the expected size (~550 bp for *mxaF*) were purified (QiAquick PCR Purification Kit, Qiagen Inc.), cloned (pCR®2.1-TOPO vector from the TOPO TA Cloning Kit, Invitrogen Corp.), using chemical method on TOP 10 competent cells, and plated on LB agar plates containing 50 μg ml$^{-1}$ kanamycin, and 40 mg ml$^{-1}$ X-Gal (5-bromo-4-chloro-3-indolyl-β-D-Galactopyranoside). Colonies were screened for inserts using the respective primers via PCR. Plasmid DNA was isolated from positive clones using the QIAprep® Spin Miniprep Kit (Qiagen Inc.). Sequencing reactions were performed using the ABI PRISM® Big Dye Terminator Cycle Sequencing kit (version 3.1, Applied Biosystems®) and an ABI model 3100 automated sequencer (Applied Biosystems®) according to the manufacturer’s instructions. Resequencing were performed in both directions using M13 forward and reverse primers. For environmental DNA extracts, at least five clones were sequenced from each PCR reaction. For *Bathydendrion* mussels, three clones each were sequenced from frozen gill tissue of three *B. azoricus* mussels from LS, three *B. azoricus* mussels from RB, and three *B. putorosetensis* mussels from LO, for a total of 27 sequences.

**Phylogenetic Analyses of Methylotrophic Bacterial Sequences from GenBank Database**

The ADH gene and/or 16S rRNA gene sequences of *Solibacter undistatus* Ellin 6076 (GenBank Accession no. NC_005536/CP000473, Pflum Acidobacteria) were used as outgroup in all phylogenetic analyses. *mxaF* gene sequences were aligned using ClustalW2 ([http://www.ebi.ac.uk/Tools/msa/clustalw2/](http://www.ebi.ac.uk/Tools/msa/clustalw2/)) and manually inspected in MacClade 4.08 (Sinauer Associates Inc., Sunderland, MA). Phylogenetic reconstruction was implemented using PAUP 4.0B10 [43] and Geneious 4.85 ([http://www.geneious.com/](http://www.geneious.com/)) with PAUP plug-in. Unless stated otherwise, statistical support for all trees were obtained from 1,000 bootstrap replicates under the same initial settings (only bootstrap values >50% are reported). Pairwise base comparisons of *mxaF* nucleotide sequences within and between phylogenetic groups were determined using ClustalW2 and reported as % identity values.

(a) **Congruency tests between *mxaF* and 16S rRNA gene tree topologies.** The program Modeltest [44] was used to determine the best fit substitution model under Maximum Likelihood (ML) analysis (from PAUP software) for each dataset, and searched heuristically for the best model of nucleic acid sequence evolution that best fits our data. The topology of the best tree from ML analyses of each dataset was saved and then enforced as a topological constraint during subsequent paired ML phylogenetic analyses for each dataset. One-tailed Templeton and Shimodaira-Hasegawa (SH) tests were used to compare the constrained and unconstrained topologies using reestimated log likelihoods (RELL) simulation [45,46] and full optimization distributions using 10,000 bootstrap replicates under the likelihood tree scores menu. Uncorrected and Bonferroni-corrected *P*-values were reported for one-tailed Templeton and SH tests [45]. Confidence intervals were determined for the null hypotheses: the unconstrained and constrained ML tree of each dataset tested have significantly different likelihood scores. The data consisted of nineteen methanotroph taxa spanning three families (Methyllocystaceae, Beijerinckiaceae, and Methylcocaceae) and one outgroup for which both genes were available: (a) seven members of the Methyllocystaceae – *Methylosinus trichosphorum* BF1, *M. trichosphorum* O19/1, *M. trichosphorum* KS21, *Methylosinus sporium* F10/1b, *Methylocystis echinoids* IMET10491, *Methylocystis* sp. IMET10490, and *Methylcystis* sp. IMET10494, (b) four members of the methanotrophic Beijerinckiaceae – *Methylcella silvestris*, *M. tundra*, *M. palustria*, and *Methylcystis acidiphila*, and (c) eight members of the Methylcocaceae – *Methylcoccus capsulatus* Bath, *Methylcococcus* sp. E10a, *Methyloccomas methanica*, *M. album*, *M. rubra*, *Methylbacterium luteus*, *Methylbacterium* sp. 5FB, and *Methylimonium* sp. For the Methylcocaceae, there are insufficient *mxaF* sequences in GenBank.

### Table 1. PCR Primers sets used in this study.

<table>
<thead>
<tr>
<th>Name</th>
<th>Sequence (5′ → 3′)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>F1003</td>
<td>GCGGCACCAACTGGGGCTGGST</td>
<td>33</td>
</tr>
<tr>
<td>R1561</td>
<td>GGGAGCCCTCCATGCTGCCC</td>
<td>33</td>
</tr>
<tr>
<td>F1003degen</td>
<td>GGNCANACTCYGGGGNTGTT</td>
<td>This study</td>
</tr>
<tr>
<td>R1561degen</td>
<td>GGGARCCNYATGYGNCTCNC</td>
<td>This study</td>
</tr>
</tbody>
</table>

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hence the mxaF of *Methylomonas rubra* str. BG8 (GenBank Accession no. L33682) was matched to the 16S rRNA gene sequence of *Methylomonas sp.* (GenBank Accession no. D09279). Separate ML phylogenetic analyses were conducted on three datasets: (a) mxaF gene sequences (~513 bp) of these methanotrophs, (b) the corresponding 16S rRNA gene sequences (~1471 bp) of the same taxa and (c) the concatenated mxaF and 16S rRNA gene sequences (~1984 bp).

(b) Phylogenetic analyses of cultured methanotrophs and other related methylotrophs. Maximum parsimony (MP) phylogenies of known methylotrophs (including methanotrophs), and related mxaF POQ-linked dehydrogenase genes, which have recently been described [32] and were included to verify their phylogenetic placement relative to mxaF genes, were constructed based on nucleotide and on inferred amino acid sequences (n = 145) available in Genbank [35]. Alignments for mxaF nucleotide sequences included sequences with and without third nucleotide positions. Phylogenetic analyses (MP) implemented amino acid sequences (513, 342, and 171 characters, respectively) for 145 taxa. The resulting tree was obtained via random stepwise addition of sequences, as consensus of 9 saved, respectively.) for 145 taxa. The resulting tree was obtained via random stepwise addition of sequences, as consensus of 9 saved, respectively.

Phylogenetic Analyses of Environmental mxaF Gene Sequences

All environmental mxaF nucleotide sequences detected in this study were verified through BLAST searches, and mxaF nucleotide sequences of their closest relatives (identified through homology searches) were included in the assembly. The final alignments for mxaF nucleotide sequences, mxaF sequences without third nucleotides and inferred mxaF amino acid sequences comprised 513, 342, and 171 characters, respectively, for 85 taxa. Maximum parsimony (MP) methods (implemented as described above) were used to generate phylogenetic trees from the nucleotide alignments. The resulting tree was obtained as consensus of 4 saved, respectively, for most parsimonious trees obtained from a heuristic search.

Nucleotide Sequence Accession Numbers

The mxaF gene nucleotide and inferred amino acid sequences of *Methylophilus rubra* and *Methyllobacter luteus, HBJA isolates 1 and 2, and uncultured clones have been deposited in GenBank, under accession numbers JX312966–JX313018.

Results

Congruency Tests

Congruency tests showed that the mxaF and 16S rRNA genes of representative methanotrophic species do not share congruent tree topologies. Topology tests failed to reject the null hypothesis that mxaF and 16S rRNA gene trees, when each was constrained to match the topology of the other gene, have significantly different likelihood scores. The SH test examines the topology, but does not indicate specific nodes or taxa causing incongruence [47]. Trees for mxaF gene, 16S rRNA gene, and combined mxaF+16S rRNA sequences were observed to be incongruent, with incongruences for representative taxa from the Methylocystaceae and Methyllococcaceae, and the methanotrophic Beijerinckiacaeae (Fig. 1).

mxaF Gene Phylogeny

The simplified mxaF gene tree (based on all codon positions) for known proteobacterial methylotrophs is shown in Fig. 2. Trees resulting from analyses of 1st and 2nd codon positions and inferred mxaF amino acid sequences of methylotrophs were identical in branching order of major taxa (not shown). Figure 2 indicates five distinct clusters of methylotrophs: (a) cluster 1 encompassing the Methylocystaceae, (b) cluster 2 containing mostly methylophilic *Methyllobacterium* sp. and methanotrophic Beijerinckiacaeae (*Methylocapsa* sp. and *Methyllobcicola*), (c) cluster 3 containing mostly *Hyphomicrobium* sp., (d) cluster 4 consisting solely of the monophyletic family Methyllococcaceae, including sequences obtained in this study for the cultured members *Methylomonas rubra* and *Methyllobacter luteus*, and (e) cluster 5 containing betaproteobacterial methylotroph genera. In addition, a well-supported monophyletic cluster 6 falls outside of other mxaF clusters and consists of oxidF gene sequences, which are distantly related to methylotroph mxaF sequences [32]. The list of taxa comprising the non-methanotrophic methylophilic and oxidF/oxidF-like clusters in Fig. 2 is shown in Table 2. The complete tree with all taxa (n = 145) is shown in Supplement S1.

Cluster 1 includes all members of the Methylocystaceae, forming two separate sub-clusters. Both sub-clusters are polyphylectic for the two genera *Methylosinus* and *Methylocystis*. The methanotrophic Beijerinckiacaeae (genera *Methylocapsa*, *Methyllobcicola* and *Methylolfura*) are placed polyphyletically within cluster 2 with other methylotrophs (Fig. 2). There is a lack of distinction in the placement of species of methanotrophic Beijerinckiacaeae in cluster 2: *Methylacetos* is most closely related to the non-methanotrophic genus *Acidomonas* [48], while the close relatives *Methylocella palustris* strain K and *Methylocella tundræ* cluster with the methylophilic *Methylorosula polaris*, gen. nov. Yu-22 and V-22 [49], *Methylocella violacei* clusters with the non-methanotrophs, *Beijerinckia mobilis* DSM 2326, and *Methylorosula stellata* clusters with the methylophilic *Methylorosula ligni* strains BW363 and BW372 [50]. Based on the mxaF gene tree (Fig. 2), the alphaproteobacterial groups – the Methylocystaceae (cluster 1), alphaproteobacterial methylotrophs including the methanotrophic Beijerinckiacaeae (clusters 2 and 3) – are more closely related to one another than to betaproteobacterial methylotrophs (cluster 5) and gammaproteobacterial methanotrophs (cluster 4). This pattern is also reflected in the 16S rRNA phylogeny.

Pairwise nucleotide sequence variations within the families Methylocystaceae and Methyllococcaceae and between methanotrophic Beijerinckiacaeae were ≥87%, ≥75%, and ≥78%, respectively. In contrast, the oxidF cluster shared ≤69% identity with all mxaF genes, and *Solibacter usitatus* Ellin 6076 ADH (outgroup) shared ≥55% identity with all mxaF genes.

Phylogenetic Analyses of PCR-amplified Environmental mxaF Gene Sequences

All inferred amino acid sequences translated from mxaF genes amplified in this study possessed the three amino acid residues involved in the interactions of the active site. MP phylogenetic trees based on all nucleotides, 1st and 2nd codon positions, and inferred mxaF amino acid sequences were identical in topology; the tree based on all three nucleotides is shown in Fig. 3. Of the mxaF clones amplified from Harvard Forest soils (labeled P_C, P_F, and H_F), one was placed in the Methylocystaceae and five were placed in the Methyllococcaceae, with the remaining three (P_F#1; #2; #3; #6; #10, P_F#4, and P_F#5) closely related to environmental clones detected in other soils. Three sequences detected in *Sphagnum* moss (labeled Sphaq, Fig. 3) clustered with the methylotroph *Methylorhabdus multivorans*, although this clade lacked bootstrap support. Of the four sequences detected in peat (labeled Peat) in the northern WI bog, one (Peat#1) clustered with mxaF sequences from clones detected in acidic forest and acidic
Figure 1. Congruency tests between mxaF and 16S rRNA gene nucleotide sequences of methanotrophs from GenBank database. Phylogenetic trees for congruency tests based on maximum likelihood (ML) analysis of mxaF (~513 bp) and 16S rRNA gene nucleotide sequences (~1471 bp) from methanotrophs in GenBank, including the mxaF nucleotide sequences of Methylomonas rubra and Methylobacter luteus sequenced in this study. The ADH gene of Solibacter usitatus Ellin 6076 was used as outgroup. Methanotrophs (in the Methylococcaceae, Methylocystaceae and Beijerinckiaceae) are indicated by shaded clusters. Accession numbers of mxaF and 16S rRNA gene sequences downloaded from GenBank are indicated in parentheses. Bootstrap values from 1,000 replicates are indicated at the nodes of branches (if >50). The scale bar represents the number of nucleotide changes.

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Figure 2. Simplified phylogenetic tree of methanotrophs and their close relatives based on mxaF nucleotide sequences from GenBank database. Unrooted phylogenetic tree based on maximum parsimony (MP) analysis of known proteobacterial partial mxaF and xoxF/xoxF-like nucleotide sequences (~513 bp) from GenBank and the mxaF nucleotide sequences (in bold) of Methylomonas rubra and Methylobacter luteus sequenced in this study. The ADH gene of Solibacter usitatus Ellin 6076 was used as outgroup. Accession numbers of sequences downloaded...
from GenBank are indicated in parentheses. Bootstrap values from 1,000 replicates are indicated at the nodes of branches (if $>50$). The three bacterial families containing methanotrophs (Methylcocaceae, Methyllyctaceae and methanotrophic members of the Beijerinckiaceae) are indicated by shaded clusters and the other alphaproteobacterial and betaproteobacterial methyloths are delineated by lines. The identity of mxaF and mxaF-like sequences from the “Methyllobacterium cluster (within cluster 2)”, “Mainly Hypomicrobiium (Cluster 3)”, “-I-proteobacterial methyloths (Cluster 5)”, and “mxaF/mxaF-like genes” is shown in Table 2. The scale bar represents the number of nucleotide changes. The complete phylogenetic tree of methanotrophs and their close relatives based on mxaF nucleotide sequences is shown in Supplement S1.

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Discussion

Establishing the link between microbial phylogeny and physiology is complicated by the high level of physiological diversity in most microbial taxa (e.g., many microbes utilizing several carbon sources), and the potential for horizontal gene transfers – the movement of microbial genes between divergent genomes. Consequently, it is necessary to evaluate the accuracy of candidate functional genes as a diagnostic of key metabolic processes, as well as being accurate markers of evolutionary history. Doing so requires evaluating the result of phylogenetic reconstruction of gene sequences from all cultured organisms possessing the gene, before analyses of sequences from uncultured environmental clones. Methanotrophs are unique due to their preference for methane as a metabolic substrate and their possession of functional genes involved in this process. Here, we investigated the phylogeny, based on the mxaF gene, of all known proteobacterial methanotrophs from GenBank database and highlighted the mxaF gene’s ability to detect most methanotrophic bacteria and describe their molecular diversity in natural environments.

mxaF and 16S rRNA Gene Phylogenies

In this study, the topological differences between 16S rRNA and mxaF gene trees, as seen in incongruencies between both trees (Fig. 1), suggest multiple occurrences of horizontal gene transfer in the mxaF genes of many methanotrophic taxa in the Methylcocaceae, Methylocystaceae, and Beijerinckiaceae. Phylogenetic analyses based on mxaF genes placed the vast majority of cultivated proteobacterial methanotrophs, excluding members of the genera Methylacopia, Methylocella, and Methylomonas, in distinct and coherent clades representing the Methylcocaceae and Methylocystaceae, with higher nucleotide identity between taxa within these clades than between members of different clades. These family-level clade distinctions are consistent with patterns evident in the 16S rDNA phylogeny. However, below the family level, mxaF-based analyses failed to differentiate between distinct subclades based on genera. The mxaF gene has poor resolving power for methanotrophs within the Beijerinckiaceae. Our mxaF gene tree (Fig. 2) indicates that these methanotrophs, composed of the genera Methylacopia, Methylocella, and Methylomonas, are polyphyletic, sharing common ancestry with other alphaproteobacterial methyloths. This pattern corroborates previous studies, which assessed fewer methylothrix taxa [23,49,53], and showed different topologies between mxaF and 16S rRNA genes, for example, in the genus Methylocella [22,50,53]. The more extensive mxaF gene phylogeny inferred here (compared to previous studies) suggests that
methanotrophy (a) arose once in the Beijerinckiaceae and was lost by some methylotrophic taxa, (b) arose separately in more than one taxon in the Beijerinckiaceae, and/or (c) multiple occurrences of horizontal gene transfers have occurred in the common ancestor.

### Table 2. Taxa included (but not shown) in the phylogenetic analyses for Fig. 2.

<table>
<thead>
<tr>
<th>Cluster name</th>
<th>Taxa and GenBank Accession number (in parentheses)</th>
</tr>
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</table>
| Methylobacterium cluster (within cluster 2) | Methylobacterium nodulans strain ORS2060 (AF220764)  
Methylobacterium aquaticum DSM1371 (EF562464)  
Methylobacterium sp. MP3 (EF030549)  
Methylobacterium lusitanum strain MP2 (EF030548)  
Methylobacterium rhodesianum strain DSM5567 (EF562473)  
Methylobacterium hispanicum strain DSM16372 (EF562468)  
Methylobacterium jeotgali strain 52R03-9 (EF031552)  
Methylobacterium sp. MPS (EU047511)  
Methylobacterium mesophilicum strain DSM1708 (EF562470)  
Methylobacterium organophilum DSM760 (EF562471)  
Methylobacterium radiotolerans JCM2831 1819 (EF562472)  
Methylobacterium fujisawaense strain KACC10744 (EF562467)  
Methylobacterium oryzae strain CBMB20 (EF562478)  
Methylobacterium suomiense strain CBMB130 (EF562479)  
Methylobacterium suomiense strain CBMB120 (EF562477)  
Methylobacterium suomiense strain KCTC12963 (EF562474)  
Methylobacterium sp. MP3 (EF030547)  
Methylobacterium sp. MPS (EU047511)  
Methylobacterium suomiense strain CBMB130 (EF562479)  
Methylobacterium suomiense strain CBMB120 (EF562477)  
Methylobacterium suomiense strain KCTC12963 (EF562474)  
Methylobacterium sp. MP3 (EF030547)  
Methylobacterium sp. MPS (EU047511)  
Methylobacterium suomiense strain CBMB130 (EF562479)  
Methylobacterium suomiense strain CBMB120 (EF562477)  
Methylobacterium suomiense strain KCTC12963 (EF562474)  
Methylobacterium sp. MP3 (EF030547)  
Methylobacterium sp. MPS (EU047511)  
Methylobacterium suomiense strain CBMB130 (EF562479)  
Methylobacterium suomiense strain CBMB120 (EF562477)  
Methylobacterium suomiense strain KCTC12963 (EF562474)  
Methylobacterium sp. MP3 (EF030547)  
Methylobacterium sp. MPS (EU047511)  
Methylobacterium suomiense strain CBMB130 (EF562479)  
Methylobacterium suomiense strain CBMB120 (EF562477)  
Methylobacterium suomiense strain KCTC12963 (EF562474)  
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Methylobacterium suomiense strain CBMB130 (EF562479)  
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Methylobacterium suomiense strain CBMB130 (EF562479)  
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Methylobacterium sp. MPS (EU047511)  
Methylobacterium suomiense strain CBMB130 (EF562479)  
Methylobacterium suomiense strain CBMB120 (EF562477)  
Methylobacterium suomiense strain KCTC12963 (EF562474)  |
| Mainly Hyphomicrobium (Cluster 3) | Hyphomicrobium sp. strain B 314 (HSMXAF314)  
Hyphomicrobium sp. strain B 327 (HSMXAF327)  
Hyphomicrobium sp. strain DPB 2c (HSMXAF2C)  
Hyphomicrobium aestuarii (HAMXAF)  
Hyphomicrobium sp. strain B 583 (HSMXAF583)  
Hyphomicrobium sp. strain B 520 (HSMXAF520)  
Hyphomicrobium sp. strain B 294 (HSMXAF294)  
Hyphomicrobium sp. strain P 324 (HSMXAF324)  
Hyphomicrobium zavarzinii (HZMXAF)  
Hyphomicrobium sp. strain B 69 (HSMXAF69)  
Hyphomicrobium vulgaris (HVMXAF)  
Hyphomicrobium sp. strain P 768 (HSMXAF768)  
Hyphomicrobium sp. strain P 495 (HSMXAF495)  
Hyphomicrobium sp. P 495 (HSMXAF495)  
Hyphomicrobium sp. P 768 (HSMXAF768)  
Hansschlegelia plantiphila strain S1 (DQ525143)  
Hansschlegelia plantiphila strain S2 (DQ525144)  
Hansschlegelia plantiphila strain S4 (DQ525145)  
Paracoccus kordatiavæ NCIMB 13773 (AJ878072)  
Methylosulfonomonas methylovora (MMU70525)  
Ancylobacter aquaticus strain IAM 12364 (AB455976)  
Allobacter methylotrophs DSM 13819 (AJ878069)  
Methyllophilus capsulata ATCC 700716 (AJ878071)  |
| β-proteobacterial Methylotrophs (Cluster 5) | Flavisobacter sp. Vu-144 (EU912489)  
Duganella sp. B41 (EU439303)  
Methylophilus methylotrophus (MMU41040)  
Methylococcus glycogens ATCC29475 (AJ878073)  
Methylophilus sp. sp. 351 (AF184915)  |
| xoxF/xoxF-like genes | Paracoccus denitrificans (U34346)  
Rhodobacter sphaeroideas 2.4.1 (CP000143)  
Methylobacterium radiotolerans JCM 2831 Mrad 2831 1932 (NC_010505)  
Methyloversatilis universalis strain FAM5 (EUS45068)  
Burkholderiales bacterium RZ18-153 (EUS45065)  
Methylobacterium extorquens PA1 Mext_1809 (NC_012808)  
Methylophilus methylotrophus (MMU41040)  
Methylococcus glycogens ATCC29475 (AJ878073)  
Methylophilus sp. sp. 351 (AF184915)  |

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**Figure 3. Phylogenetic tree of environmental mxaF gene sequences detected in this study.** Phylagenetic tree based on MP analyses of environmental partial mxaF nucleotide sequences (~513 bp) detected in this study (in bold) in comparison with their close relatives, with Solibacter usitatus Ellin6076 as outgroup. All mxaF sequences were obtained using primer pair F1003 and R1561, except for the 13 putative symbiont mxaF genes from Bathymodiolus azoricus and B. puteoserpentis, which were obtained using primer pair F1003degen and R1561degen. Accession numbers of sequences downloaded from GenBank are indicated in parentheses. Bootstrap values from 1,000 replicates are indicated at the nodes of branches (if >50). Clone sequences are labeled P_C, pine soil (control); P_F, pine soil (fertilized); H_F, hardwood soil (fertilized); Sphag, Sphagnum moss; HBHA, Halls Brook Holding Area; RB, Rainbow; LS, Lucky Strike; LO, Logatchev, followed by clone number (#). Methanotrophs found in the three bacterial families (Methylcoccaceae, Methylocystaceae and Birgerinkciaceae) are shaded. Bootstrap values from 1,000 replicates are indicated at the nodes of branches (if >50). The scale bar represents the number of nucleotide changes. doi:10.1371/journal.pone.0056993.g003

of methanotrophic Beijerinckiaceae. It is therefore difficult to ascertain whether microorganisms with mxaF gene sequences placed near the polyphyletic Beijerinckiaceae genera Methylocapsa, Methylocella, and Methylofera are indeed methanotrophic.

Overall, we data suggest that the partial mxaF gene (~550 bp) amplified by the primer set mentioned is a useful phylogenetic marker and provides sufficient resolution to broadly discriminate between known proteobacterial methanotroph families via the Methylcoccaceae and Methylocystaceae clusters, which together encompass the vast majority of known methanotrophs. However, it has poor resolution at the sub-family level across the Methylcoccaceae (not shown in previous studies) and Methylocystaceae, and ambiguities exist between the methanotrophic Beijerinckiaceae and other methyloches in the Alphaproteobacteria, due possibly to horizontal transfers of the mxaF gene.

**mxaF Sequences from Diverse Environments, Isolates and Endosymbiotic Methanotrophs**

The mxaF gene was used here to examine the phylogenetic placement and diversity of over fifty novel sequences of putative methanotrophs from a range of natural environments and from cultured isolates in light of evolutionary information from the mxaF gene phylogeny determined above. The mxaF datasets from Harvard Forest soil, peat and Sphagnum moss from northern Wisconsin, and the HBHA water column contained diverse sequences that clustered primarily in the Methylcoccaceae and Methylocystaceae, as well as sequences related to methanotrophic Beijerinckiaceae and other alphaproteobacterial methyloches. Notably, the sample of Sphagnum moss contained three sequences (Sphag#1, Sphag#2, and Sphag#3), suggestive of three strains, most closely related to the methyloch Methylobacterium multivorans. However, these three sequences are also closely related to the methanotrophic Beijerinckiaceae genus Methylocella (Fig. 3). Given the uncertainty with which mxaF discriminates between methanotrophic Beijerinckiaceae and certain alphaproteobacterial methyloches discussed above, the sequences detected in peat, forest soils and Sphagnum moss could represent methanotrophic bacteria.

Our analyses also provide the first insights into the phylogenetic placement and biogeography of the previously unknown mxaF genes from putative methanotrophic endosymbions of deep-sea hydrothermal vent Bathymodiodid mussels. Bathymodiolus azoricus and B. puteoserpentis are the dominant species of mussels within the two spatially separate mussel hybrid zones of Lucky Strike and Rainbow, and Logatchev on the Mid-Atlantic Ridge [54]. Here, dual bacterial endosymbions provide nutrition to the mussel hosts through thiotrophy and methanotrophy, but the mode of methanotrophic symbiont transmission, whether vertical (symbi- ons are passed from parent to offspring) or horizontal (symbi- ons are taken up from the environment, or from co-occurring hosts) is not known [38,39,53,56]. Here, we show that these novel mxaF gene sequences of putative endosymbiotic methanotrophs belong to the Methylcoccaceae, in agreement with their placement in the Gammaproteobacteria with other methanotrophs based on 16S RNA phylogeny [6]. B. azoricus mussels at the adjacent Lucky Strike and Rainbow vent sites harbored closely related putative methanotrophic symbions, with most mussels possessing a heterogeneous population of putative endosymbions, based on the closely-related but unique mxaF gene sequences we detected. This result is consistent with the hypothesis of environmental acquisition of mussel symbions [38,55,57,58], where each mussel may be expected to contain multiple genetic variants.

In contrast, B. puteoserpentis mussels at Logatchev, a vent site over 2600 km away, harbored a separate monophyletic group of closely related putative methanotrophic symbions. The phylogenetic clustering of B. azoricus and B. puteoserpentis symbions in two distinct clades suggests either taxon-specific differences in the specificity of the symbiont-host relationship (i.e., each host species associates with a unique symbiont strain) or that B. azoricus mussels at Lucky Strike and Rainbow acquire putative symbions from an environmental pool that is genetically distinct from that available to B. puteoserpentis mussels at Logatchev vent site. Our phylogenetic analyses of mxaF gene sequences indicate that the putative endosymbiotic methanotrophs have diverged from an ancestral sequence into two monophyletic groups, either in response to host-symbiont co- speation or to geographic separation. Indeed, the fragmented distribution of deep-sea hydrothermal vents may promote spatial isolation that acts as a barrier to symbiont dispersal [59]. With more information, the extent to which symbiont diversification is driven by either geographic isolation and symbiont-host specificity and co-evolution, can be assessed.

Proteobacterial methanotrophs currently constitute the vast majority of known aerobic methane oxidizing bacteria. In this study, we demonstrate the usefulness of the mxaF gene in studying proteobacterial methanotroph diversity in non-anoxic environments. Studying the evolutionary history of this gene from known methanotrophs extensively may provide insights into the placement of novel taxa detected in different environments and avoid incorrect inferences from their phylogenetic placement. Our results indicate the mxaF gene can be a functional and phylogenetic marker for proteobacterial methanotrophs, providing more information about an important group of microorganisms involved in the global biogeochemical methane cycle.

**Supporting Information**

**Supplement S1** (DOCX)

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References