The Villain Team-Up or how Trichomonas vaginalis and bacterial vaginosis alter innate immunity in concert

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INTRODUCTION

Homeostasis of the genital tract mucosa in reproductive-age women is challenged by resident microorganisms that vary depending on menstrual cycle, contraceptive and vaginal practices, and sexually transmitted pathogens.1–3 Culture techniques as well as modern genetic and metagenomic methods support the paradigm that both type and relative abundance of different bacterial taxa are indicative of vaginal health.4–7 A deviation from the norm, including increased numbers of certain species (eg, Gardnerella vaginalis, Atopobium vaginae and Prevotella bivia) and decreased abundance of Lactobacillus spp., has been coined as the syndrome of disturbed vaginal microbiota or bacterial vaginosis (BV).8–10 BV is among the most common conditions seen by primary care physicians in reproductive-age women and is often combined with infection by the flagellated protozoan *Trichomonas vaginalis* (TV).10 The occurrence rates and prevalence of BV and TV are believed to be underestimated because both are commonly asymptomatic and rather requires mandatory testing.

Common features of BV and TV are lack of effective adaptive immunity, high recurrence rate, high risk of reproductive problems (eg, chorioamnionitis, preterm birth, premature rupture of membranes, low birth weight, pelvic inflammatory disease) and failure of antibiotic treatment to break the inflammatory sequelae that complicate maternal–fetal interactions.11 Another common adverse outcome of both BV and TV is increased risk of HIV and other sexually transmitted infections.11 Both BV and TV are associated with vaginal discharge and abnormal levels of cytokines in cervicovaginal secretions.11 Although epidemiological findings link trichomoni asis to low abundance of lactobacillus, causative relationships between TV and BV, and their combined impact on vaginal immunity, have not been elucidated.10

We hypothesised that the TV parasite selectively interferes with patterns of vaginal colonisation and that vaginal bacteria reciprocally modify the mucosal immune balance and responses to TV virulence determinants, including its predominant surface lipophosphoglycan (LPG),12 13 also named lipoglycan,14 its ceramide-phosphoinositol-glycan core (CPI-GC), responsible for parasite adherence and chemokine response,12 13 and the endosymbiotic dsRNA viruses carried by TV (*Trichomonas vaginalis virus* (TVV)) that are sensed by the human host, causing inflammatory responses, magnified by antibiotic treatment.15 Here we newly applied a bacterial colonisation model16 17 to test this hypothesis.

METHODS

Epithelial cells

Epithelial cell lines Vtk2/E6E7, Ect1/E6E7 and End1/E6E7, representing normal human vagina,
ectocervix and endocervix, were maintained in keratinocyte serum-free medium (KSFM) supplemented with 50 μg/mL bovine pituitary extract, 0.1 ng/mL epidermal growth factor, penicillin/streptomycin (all from Invitrogen) and CaCl₂ (Fisher). Antibiotics were omitted in all experiments. Ect1/E6E7 and End1/E6E7 were derived from the same woman to allow an isogenic comparison of simple versus stratified non-keratinising cell types. All three cell lines, previously compared with their progenitors and a non-transformed organotypic ectocervical-vaginal tissue model, have shown immune responses to bacteria and a variety of pathogenic determinants including toll-like receptor (TLR) ligands, LPG and TVV similar to those by their primary counterparts.  

**TV LPG and CPI-GC**  
UR1, a TV isolate that carries three TVV species, was previously obtained from a symptomatic patient, cloned and cultured in Diamond’s modified media as described. We chose this isolate for our experiments because it represents the majority (>80%) of those in our collection in being infected with multiple TVV species. For LPG extraction, parasites were harvested in late log phase (24 h) by centrifugation, washed twice with phosphate-buffered saline (PBS, pH 7.4) (Invitrogen), consecutively extracted with methanol/chloroform (1:2) and solvent E and purified on an octyl-Sepharose column. The CPI-GC core was released by mild acid treatment (100 mM TFA) of LPG as described. LPG and CPI-GC purity and lack of endotoxin contamination were confirmed as described. Both LPG and CPI-GC were used at a dose of 240 μg/mL based on previously established lack of toxicity, peak of inflammatory response or used for viability assessment. As controls, epithelial cells were lysed for colony-forming units (CFU) counts, used for viability assessment or used for microscopy. For CFU counts, epithelial cells were washed twice with PBS and hypotonically lysed in HyPure water (Fisher) for 15 min. The lysates were seeded on agar following adjustment of osmolarity by adding equal volume of 2× PBS as described. Bacterial suspensions were also allowed to grow in parallel in the absence of epithelial cells in the presence or absence of TV to determine the initial multiplicity of infection for each microorganism, the degree of growth of each during the experiment and direct effects of TV. In some experiments, epithelia-free bacterial suspensions or colonised epithelial monolayers were exposed to a combination of 100 U/mL penicillin and 100 μg/mL streptomycin (both from Invitrogen) or 100 μM metronidazole (Acros Organics) previously shown to be non-toxic to epithelial cells and incubated for 4 h followed by plating on agar for relative assessment of extracellular and intracellular CFU.

**Cell viability**  
Epithelial cell viability in the presence of LPG, CPI-GC, MALP-2 or TV virions was assessed by the non-radioactive CellTiter96 MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) assay (Fisher). Cell viability in the presence of TV and bacteria was assessed microscopically and by the trypan blue exclusion assay (Fisher) because the microorganisms also convert the MTT dye and thus the MTT assay is not an optimal choice for coculture conditions.

**Inflammation-associated proteins**  
The following chemokines were measured in culture supernatants using electrochemiluminescence multiplex assays on a Sector Imager 2400 (Meso Scale Discovery): interleukin-8 (IL-8, CXCL8), chemoattractant primarily for neutrophils; regulated and normal T cell expressed and secreted protein (RANTES, CCL5), chemoattractant primarily for T cells; macrophage inflammatory protein-3α (MIP-3α, CCL20), chemoattractant primarily for dendritic cells; and interferon γ-induced protein 10 (IP-10, CXCL10), chemoattractant for T cells, monocytes, NK cells and dendritic cells. Secretory leucocyte protease inhibitor (SLPI) levels were measured by Quantikine ELISA (R&D Systems).

**Statistics**  
Data were analysed by analysis of variance (ANOVA) (GraphPad Prism, V5.5.0). p Values <0.05 were considered significant.

**RESULTS**  
Inoculation of live TV reduced the CFU numbers of epithelia-associated L.acidophilus and L.jensenii by more than two log₁₀ values (figure 1A). L crispatus and G vaginalis showed a trend of decrease, which however did not reach a log₁₀ difference. Live TV had no effect on P bivia or A vaginae. LPG, CPI-GC and TVV had no significant effects except that epithelia-associated G vaginalis was slightly increased by CPI-GC. These data suggested that the live TV was responsible for the observed reduction in epithelia-associated bacterial CFU numbers.

The CFU reduction caused by TV was not due to epithelial destruction as confirmed by light and electron microscopy as well as immune responses were assessed after another 24 h. Synthetic TLR2/TLR6 ligand MALP-2 (25 nM; Alexis Biologicals) analogue of Mycoplasma fermentans lipopeptide macrophage activating lipopeptide was used as a pro-inflammatory control. At the end of each stimulation period, supernatants were unless specified collected for soluble mediators and epithelial cells were lysed for colony-forming units (CFU) counts, used for viability assessment or used for microscopy. For CFU counts, epithelial cells were washed twice with PBS and hypotonically lysed in HyPure water (Fisher) for 15 min. The lysates were seeded on agar following adjustment of osmolarity by adding equal volume of 2× PBS as described. Bacterial suspensions were also allowed to grow in parallel in the absence of epithelial cells in the presence or absence of TV to determine the initial multiplicity of infection for each microorganism, the degree of growth of each during the experiment and direct effects of TV. In some experiments, epithelia-free bacterial suspensions or colonised epithelial monolayers were exposed to a combination of 100 U/mL penicillin and 100 μg/mL streptomycin (both from Invitrogen) or 100 μM metronidazole (Acros Organics) previously shown to be non-toxic to epithelial cells and incubated for 4 h followed by plating on agar for relative assessment of extracellular and intracellular CFU.
Figure 1  Effect of *Trichomonas vaginalis* (TV) on bacterial colonisation in an in vitro cervicovaginal colonisation model. (A) Colony-forming units (CFU) associated with endocervical epithelial cells 48 h postcolonisation and 24 h after exposure to live TV or purified TV virulence factors. Bars represent means±SEM of duplicate measurements representing consistent results from five independent experiments. ***p<0.001, *p<0.01, *p<0.05, live TV, lipophosphoglycan, ceramide-phosphoinositol-glycan core and *Trichomonas vaginalis* virus virions different from medium control, two-way ANOVA, Bonferroni post-test. Similar results were obtained with vaginal and ectocervical cells (data not shown). (B) Electron micrographs from the vaginal colonisation model. Arrows in the higher magnification image (upper panel) indicate adherent or internalised *Atopobium vaginae* in vaginal epithelial cells. (C–H) CFU numbers assessed after 4 h antibiotic treatment of epithelia-free bacterial suspensions (C–E) or vaginal epithelial monolayers (F–H) precolonised with bacteria for 24 h. Data are means±SEM of duplicate cultures representing one of three experiments. med, antibiotic-free medium, p/s, combined penicillin–streptomycin; metro, metronidazole, Vk, vaginal epithelial cells. *p<0.05, **p<0.01, ***p<0.001, antibiotics different from no antibiotics, +p<0.05, ++p<0.01, +++p<0.001, supernatant different from lysate. (I) Reduction of planktonic or epithelia-associated CFU after 24 h infection with TV. Data are means and SEM from duplicate cultures of vaginal (Vk), ectocervical (Ect) and endocervical (End) epithelial cells representing two independent experiments. *p<0.05, **p<0.01, ***p<0.001, TV—infected compared with non-infected control; ++, p<0.01, +++p<0.001, planktonic compared with epithelial-associated CFU reduction (two-way ANOVA, Bonferroni post-test).
In the absence of bacteria, RANTES (figure 2B) was induced by TV and TVV but not LPG or CPI-GC, as expected from our previous work, in which RANTES upregulation was dependent on TVV dsRNA sensing in the context of multiple TV strains. RANTES was upregulated >fourfold by *A. vaginae* or *G. vaginalis*, less by *P. bivia* (∼2-fold) and not by lactobacilli. *A. vaginae*, *G. vaginalis* or MALP-2 significantly amplified the RANTES response to TV (∼2-fold) and even more to TVV: 30- to 40-fold over medium control; five- to eightfold over TVV alone, TV in the absence of bacteria or TV in the presence of lactobacilli; and >18-fold over lactobacilli alone. In contrast to *A. vaginae* and *G. vaginalis*, *P. bivia* suppressed TV- and TVV-induced RANTES.

SLPI (figure 2C) was significantly suppressed by live TV and even more by purified LPG or CPI-GC, both in sterile and bacteria-colonised epithelia. SLPI was slightly upregulated by *G. vaginalis*, MALP-2 or TVV, but was nevertheless suppressed below baseline by TV, LPG or CPI-GC, even in the presence of *G. vaginalis* or MALP-2.

To further investigate the dual (pro-inflammatory and immunosuppressive) effects of *P. bivia*, we performed additional experiments in which TV infection was conducted in cervical cells colonised with either *P. bivia* or *L. acidophilus*, which had invariably shown a non-inflammatory profile (figure 2A–C and previous investigations). Four chemokines were measured simultaneously by a Meso Scale Discovery multiplex. Again, similar to the results in figure 2A,B, *P. bivia* induced vigorous IL-8 production and amplified IL-8 production induced by TV (figure 2D) but caused only a low RANTES response and suppressed TV-upregulated RANTES (figure 2E). MIP-3α was upregulated by either *P. bivia* or TV alone in the presence or absence of bacteria, but together they yielded lower MIP-3α induction than *P. bivia* alone (figure 2F). Similarly, IP-10 was upregulated by either *P. bivia* or TV alone, but was completely suppressed by their combination (figure 2G). *L. acidophilus*, in contrast, neither induced chemokines nor altered responses to TV (figure 2D–G).

**DISCUSSION**

Our findings indicate that TV infection significantly reduced epithelia-associated *Lactobacillus* spp. but not BV spp. The results thus provide evidence for a cause–effect relationship between TV and BV and support clinical observations that trichomoniasis is associated with a specific vaginal bacterial community in reproductive-age women, characterised by a lack of significant numbers of lactobacilli and a higher proportion of strictly anaerobic organisms such as *A. vaginae*, *P. bivia*, *G. vaginalis* and other BV spp.

Since we frequently observed *A. vaginae* and *P. bivia* within epithelial cells, we hypothesise that some types of vaginal bacteria, particularly BV spp. and especially *A. vaginae*, may escape the hostile vaginal environment and attacks by TV by localising inside epithelial cells. In our experiments, antibiotics at doses that killed planktonic or extracellular bacteria had less effect on *P. bivia* and did not reduce *A. vaginae* CFU obtained from epithelial lysates, suggesting these bacteria may remain viable at significant numbers within epithelial cells. It cannot be excluded, though, that these and other bacteria might form a surface biofilm that makes them more resistant to antibiotic treatment and perhaps also to TV or TV products. Our findings thus warrant further investigations that may suggest novel potential mechanisms for managing recurrent and antibiotic-resistant BV.

The combination of BV spp. with TV significantly affected the host immune response to the individual TV virulence...
factors LPG and TVV as well as to the live parasite as a whole (schematised in figure 3). Importantly, our experimental results show also a remarkable causative relationship between altered epithelial responses and the type of vaginal bacteria populating the epithelia at the time of TV infection, with *Lactobacillus* spp. lacking and BV spp. possessing the ability to alter different aspects of LPG- or TVV-driven responses. The fact that TV teams up with *A vaginae* and *G vaginalis* to induce much higher chemokine responses (represented here by IL-8 and RANTES) on the background of significantly reduced SLPI due to LPG/CPI-GC signalling suggests a mechanism for inflammatory damage accompanied by recruitment of CD4 cells and weakened antiviral barrier thus facilitating viral co-infections including herpes simplex virus, human papillomavirus and HIV. SLPI is an innate-immune mediator with direct virucidal effects, which is produced and stored at abundant levels in mucosal epithelial cells and keratinocytes. 16 17 SLPI is also capable of dampening inflammatory responses to LPS. 25 We have recently shown that vaginal SLPI levels decrease in women with trichomoniasis in a manner dependent on parasite load. 28 Higher

Figure 2  Effect of *Trichomonas vaginalis* (TV) and bacteria on chemokine levels in an in vitro cervicovaginal epithelial model. (A–C) Levels of IL-8 (A), RANTES (B) and secretory leucocyte protease inhibitor (C) measured in epithelial-cell supernatants collected 48 h after bacterial colonisation as indicated (1–7, bacteria identified below panel C) and 24 h after exposure to TV isolate UR1 (TV), lipophosphoglycan (LPG), ceramide-phosphoinositol-glycan core (CPI-GC), *Trichomonas vaginalis* virus (TVV) virions or MALP-2. Data are means±SEM from duplicate cultures in one of five independent experiments. ***p<0.001, **p<0.01, *p<0.05, TV, LPG, CPI-GC and TVV different from medium control within each group, two-way ANOVA, Bonferroni post-test; +++p<0.001, ++p<0.01, +p<0.05, medium+bacteria or medium+MALP-2 different from ‘no bacteria’ medium control, one-way ANOVA, Dunnett post-test. (D–G) Levels of IL-8 (D), RANTES (E), MIP3α (F) and IP-10 (G) assessed simultaneously by Meso Scale Discovery multiplex in epithelial-cell supernatants collected 48 h after colonisation with *Lacidophilus acidophilus* and *Prevotella bivia* and 24 h after exposure to TV (1–6, conditions identified below panel G). ***p<0.001, **p<0.01, different from medium control, +++p<0.001, ++p<0.01, *P bivia* different from *P bivia*+ TV, one-way ANOVA, Bonferroni post-test.
vaginal SLPI levels may have protective anti-HIV activity as suggested by studies in HIV controllers.10

The immunosuppressive effects of *P. bivia* warrant further investigation. Suppression of IP-10, MIP-3α and RANTES may serve to evade the arm linking innate to adaptive immunity since those chemokines in combination are particularly important for recruitment of antigen-presenting dendritic and T cells to the site of microbial invasion. Our data also suggest that the effects of TV on the vaginal immunobiome depend on which BV spp. are prevalent at the time of TV inoculation. The fact that TVV and BV spp. but not bacterial vaginosis (BV) spp. cause a tremendous (in some cases over 30-fold) amplification of the inflammatory response to TV is a point of particular concern and suggests the need to include antiviral and anti-inflammatory components in the combined therapeutic–preventative approach to trichomoniasis leading to or occurring in concert with BV.

CONCLUSIONS

Our findings have important implications for understanding how TV and its major virulence factors LPG and TVV affect the mucosal immune/microbiome and for designing better therapeutic approaches for combining antiparasitic therapy with attempts to restore the normal microbiome by introducing *Lactobacillus*-based probiotics and preventing TV-attributable BV and inflammatory sequelae. Further experimental and clinical studies should follow for identifying the molecular targets for novel drug therapies preventing additive or synergistic pro-inflammatory and immunosuppressive effects of TV, TVV and BV in concert.

**Key messages**

- *Trichomonas vaginalis* reduced epithelia-associated *Lactobacillus* spp. but not bacterial vaginosis (BV) spp.
- T vaginalis and BV species in concert amplified pro-inflammatory and suppressed protective innate-immune responses.
- Virulence factors of *T vaginalis* that altered epithelial responses in concert with BV species include the surface lipophosphoglycan and endosymbiotic dsRNA viruses.
- Future therapeutic/preventative approaches should combine antiparasitic therapy with microbiome restoration and target the pro-inflammatory and immunosuppressive effects of *T vaginalis* and BV.

**Figure 3** Schematic presentation of chemokine response and suppressed secretory leukocyte protease inhibitor in response to *Trichomonas vaginalis* (TV) virulence factors and combined bacterial vaginosis (BV)-TV infection. Arrows show magnitude and direction of change. Arrows in parentheses indicate responses dependent on type of BV bacteria.

**REFERENCES**


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**Contributors** RNF conceived and directed the co-infection model and concept development, cloned the UR1 isolate, wrote the manuscript, and coordinated reviews and coauthors’ contributions. ABO, BNS and MLN contributed to concept development and provided critical review of results and interpretations, ABO provided microbiology expertise and characterised vaginal bacteria isolates; BNS provided the original UR1 isolate and expertise in *Trichomonas vaginalis* biochemistry; and MLN provided virology expertise and TVV virions. ORB, HSY and TF conducted the experiments, cell viability assessment and data analyses; MD characterised and expanded bacterial isolates; HYD, BF, OB and TF performed bacterial cultures; ORB and HYD performed immunoassays; DHB and TF cultured *T. vaginalis*, and GRH and BNS purified LPG and CPI-GC; and YT purified the TVV virions. All authors read, provided critical comments and agreed with the final manuscript content.

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**Competing interests** None.

**Ethics approval** The original protocol for collecting *Trichomonas vaginalis* isolates with patients’ informed consent was approved by the IRB boards at SUNY Upstate Medical University (Syracuse, New York, USA) and Brigham and Women’s Hospital (Boston, Massachusetts, USA).

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