SUMMARY

The mammalian pancreas is constructed during embryogenesis by multipotent progenitors, the identity and function of which remain poorly understood. We performed genome-wide transcription factor expression analysis of the developing pancreas to identify gene expression domains that may represent distinct progenitor cell populations. Five discrete domains were discovered. Genetic lineage-tracing experiments demonstrate that one specific domain, located at the tip of the branching pancreatic tree, contains multipotent progenitors that produce exocrine, endocrine, and duct cells in vivo. These multipotent progenitors are Pdx1\(^+\)Ptf1a\(^+\)cMyc\(^{\text{high}}\)Cpa1\(^+\) and negative for differentiated lineage markers. The outgrowth of multipotent tip cells leaves behind differentiated progeny that form the trunk of the branches. These findings define a multipotent compartment within the developing pancreas and suggest a model of how branching is coordinated with cell type specification. In addition, this comprehensive analysis of >1,100 transcription factors identified genes that are likely to control critical decisions in pancreas development and disease.

INTRODUCTION

Mammalian organs contain millions of cells and many different cell types, all of which arise from a small number of embryonic cells, the multipotent progenitors. Despite their importance, rather little is known about when multipotent progenitors arise or how they guide organ construction. The present study is concerned with multipotent progenitors in the pancreas. We identify genetic markers for these cells and show where they are positioned and how their progeny give rise to the branching organ and its exocrine and endocrine components.
of the pancreas? In the central nervous system, multipotent cells reside within a specific domain, the ventricular zone, with differentiated progeny migrating outward and organizing into different structures (Jessell, 2000). Similarly, in the gut, stem cells are localized at the base of the villi, sending differentiated progeny toward the tips (Fuchs et al., 2004). It is conceivable that a similar “multipotent domain” exists in the developing pancreas and that it serves as an organizing center for pancreatic organogenesis.

To identify previously unappreciated compartments in the pancreas, we performed genome-wide expression analysis of the developing pancreas with >1,100 TFs. Several discrete domains were recognized, representing the mesenchyme, epithelium, and vasculature of the developing pancreas. The epithelium domain can be further divided into two separate domains delineating the distal tip and trunk of the branches. Molecular marker analysis and genetic lineage tracing experiments suggest that cells of the distal tip domain, which coexpress Pdx1, Ptf1a, c-Myc, and Carboxypeptidase A1 (Cpa1), are multipotent progenitors that are capable of generating all pancreatic cell types, including exocrine, endocrine, and duct cells. The outgrowth of tip progenitors leaves behind endocrine and duct progeny that comprise the trunk of the branches. The multipotent tip progenitors undergo a developmental switch around midgestation (E14) and become exocrine cells. Our findings reveal the existence of a distinct domain of multipotent progenitors in the developing pancreas and, combined with earlier studies, suggest a model of how pancreatic branching is coordinated with cell-type specification.

RESULTS

Genome-Wide TF Expression Analysis Defines Five Domains of Gene Expression within the Developing Pancreas

A genome-scale, whole-mount in situ hybridization screen was performed with a recently developed mouse TF library that covers about 80% (~1,100) of all predicted mouse TFs (total, ~1,400) (Gray et al., 2004). E9.5 embryos and E14.5 dorsal pancreata were chosen for the screen. E9.5 represents the earliest stage of pancreatic organogenesis and E14.5 represents a stage at which the developing pancreas is undergoing active growth, branching, and cellular differentiation (Murtaugh and Melton, 2003).

Of the 1,100 TF genes analyzed, we detected the expression of 8 TF genes at E9.5 (see Figure 1 in the Supplemental Data available with this article online) and 94 TF genes at E14.5 (Table S1). The eight TF genes expressed at E9.5 are also present in E14.5 pancreas (Figure S1 and S2). Among TFs identified in the screen are many well-studied pancreatic genes, including Pdx1, Ptf1a, Nkx2.2, and Ngn3 (reviewed by Edlund, 2002 and Wilson et al., 2003). In addition, we identified genes, the expression of which, in the pancreas, has not been described previously, including Lisch7, Wbscr14, and Tbx3 (Table S1).

The whole-mount in situ samples provide a clear view of overall expression patterns, but lack cellular resolution for structures within the pancreas. In situ hybridizations on tissue sections were performed with all 94 TFs identified from the initial whole-mount screen (Figures 1F–1J). Of the 94 genes, 60 yielded a clear, regionalized signal at E14.5, and these could be classified into five general patterns (Figures 1K–1O and Table S1). For simplicity, we referred to them as “pan-epithelium,” “tip,” “trunk,” “mesenchyme,” and “vascular.” Pan-epithelium genes, such as Hex, are expressed specifically in pancreatic epithelial but not mesenchymal cells (Figures 1A, 1F, and 1K). Tip genes are confined to the distal tips of the pancreatic epithelial tree (Figures 1B, 1G, and 1L, arrowheads), while trunk genes appear in cells scattered within the trunk but not the tips of the branching epithelium (Figures 1C, 1H, and 1M). In addition to these three epithelial domains, the mesenchyme and vascular classes of TFs have mesenchymal- and vascular-specific expression patterns, respectively (Figures 1D, 1E, 1I, 1J, 1N, and 1O).

Different Pancreatic Precursors Are Localized in Discrete Epithelial Domains

Given that all three major pancreatic cell types, including exocrine, endocrine, and duct cells, derive from the epithelium (Gu et al., 2002), we used specific markers to analyze whether the tip and trunk epithelial domains contain separate precursor populations.

Several tip genes, including Ptf1a, Mist1, and RBP-J (Table S1), have been implicated in the development of exocrine tissues (Beres et al., 2006; Lin et al., 2004; Pin et al., 2001; Zecchin et al., 2004). We tested the exocrine nature of the tip domain at E14.5 by double-labeled in situ hybridization of selected tip genes with Carboxypeptidase A1 (Cpa1) and Amylase, two markers of fully differentiated exocrine cells. Complete overlap was observed (Figures 2A and 2B), suggesting that the tip domain is occupied by exocrine precursors at E14.5. These data are consistent with published results (Pictet and Rutter, 1972).

In contrast to tip genes, most trunk TFs identified are well-established markers of endocrine precursors, with the exception of vitamin D receptor (VDR) and Wbscr14, the expression of which in the embryonic pancreas had not been described before (Table S1). Double-labeled in situ hybridization revealed that VDR and Wbscr14 partially overlap with a number of endocrine genes (not shown), suggesting that they are also expressed in endocrine cells. To visualize the extent of the pancreatic trunk that is occupied by endocrine precursors, a mixture of cRNA probes, including most of the endocrine precursor markers (Ngn3, NeuroD, Pax4, Pax6, Is11, Brm4, Myt1, MatB, Arx, Wbscr14, and VDR) was used for in situ hybridization on E14.5 pancreas (Figure 2C). While the majority of trunk cells were labeled (Figure 2C), some appear to have been negative (Figure 2C, arrows). It is noteworthy that the expression of endocrine genes did not extend inside the tips (Figure 2C, arrowheads). These data indicate that endocrine precursor cells reside exclusively within the
trunk of the pancreatic branches. Due to lack of definitive precursor markers for the pancreatic duct lineage, we could not assess where duct precursors are localized within the branching structure.

To further confirm that the tip and the trunk of the pancreatic branches at E14.5 represent discrete domains, we performed double-labeled in situ hybridization of Cpa1 (tip) and the endocrine progenitor marker Ngn3 (trunk) (Gradwohl et al., 2000; Gu et al., 2002). No overlap was observed (Figure 2D). Together, our analysis suggests that the tip and trunk of the branching pancreatic tree at E14.5 are separate domains and contain different precursor populations.

We further analyzed the expression of tip genes at E12.5, when pancreatic branching morphogenesis has just begun. Although most tip genes yielded no staining at this early stage, signals were detected for C-myc, Ptf1a, and Cpa1 in the newly formed branches (Figures 2E–2G, arrowheads). Interestingly, when one tip divides into two or more tips, the tip markers, Cpa1, Ptf1a, and c-Myc, are downregulated in the cleft region before overt morphological changes occur (Figure 2G, hollow arrowheads, and Figures S5A and S5B). Double-labeled in situ hybridization of Ptf1a, c-Myc, and Cpa1 shows that they are coexpressed in the same population of tip cells at E12.5 (Figures S5A and S5B). In addition, Ngn3+ endocrine cells and Cpa1+ tip cells occupy distinct domains (Figure 2H). Unlike at E14.5, however, the tip cells at E12.5 do not express the exocrine markers Amylase and Elastase (data not shown). These data raised the question of whether the Ptf1a+cMyc+Cpa1+ tip cells that exist before E14.5 are multipotent progenitors or committed exocrine precursors.

**Distal Tip Cells Represent a Novel, Fast-Proliferating Cell Type**

Among the three early tip genes, c-Myc expression, though concentrated in the tips, is also detected at lower levels in other epithelial cells (data not shown). Ptf1a expression is rather weak before E12.5, and cannot be easily or reliably detected by antibody staining or in situ hybridization. In contrast, Cpa1 expression is specific to the tip cells and readily detectable by antibody staining and in situ hybridization. We therefore used Cpa1 as a marker to further characterize the tip cells during early pancreatic branching morphogenesis.

Cpa1 mRNA is first detected in E9.5 and protein expression starts around E10.5 in the pancreatic buds (data not shown). At E11.5, Cpa1 is expressed in a scattered population of epithelial cells (Figures 3A, 3E, and 3I). Shortly
after E11.5, branching of the pancreas begins. By E12.5, there are well-formed primary branches, and Cpa1 expression is now largely restricted to the branching tips (Figures 3B, 3F, and 3J, arrows). The tip-restricted expression pattern of Cpa1 persists through successive branching and growth of the pancreatic tree (Figures 3C, 3D, 3G, 3H, 3K, and 3L). Note that Cpa1 protein levels vary among tip cells, perhaps reflecting their different mitotic stages.

At all embryonic stages examined, Cpa1+ cells do not express mature endocrine hormones (Figures 3A–3D) or the early endocrine progenitor marker, Ngn3 (Figures 3E–3H). In contrast, all Cpa1+ cells do express Pdx1 and constitute a subset of Pdx1+ epithelial cells (Figures 3I–3L). Note that Cpa1 protein levels vary among tip cells, perhaps reflecting their different mitotic stages.

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Genetic Lineage Tracing of the Tip Progenitors
To determine whether the tip cells constitute multipotent progenitors or committed cells, we generated a Cpa1CreERT2 mouse line that expresses an inducible form of Cre recombinase (CreERT2) from the Cpa1 genomic locus (Figures S3) (Metzger and Chambon, 2001). By crossing this mouse line with the reporter line, R26R, administration of tamoxifen (TM) allows permanent marking of Cpa1+ cells at designated developmental stages and allows us to follow the fate of Cpa1 progeny through subsequent development (Figure 4A).

To validate the Cpa1CreERT2 line, we first examined whether CreERT2 expression is restricted to Cpa1+ cells and whether CreERT2 activation is strictly dependent upon TM. Cpa1CreERT2;R26R animals were labeled with one TM injection at E13.5 (Figure 4C) and analyzed 20 hr later. Uninjected animals served as controls (Figure 4B). Many βgal+ cells were present in the pancreas of TM-injected embryos (Figure 4C), which all coexpress Cpa1 (Figure 4C, arrowheads). In contrast, no βgal+ cells were detectable in the absence of TM (Figure 4B). These data show that CreERT2 expression is restricted to Cpa1+ cells and, importantly, activation of CreERT2 is strictly TM dependent.

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A similar experiment on adult animals confirmed that CreERT2 expression is restricted to adult exocrine acini (Figure 4E, arrowhead), but not islets (Figure 4E, arrow) or ducts. Again, CreERT2 activation is TM dependent (Figure 4D). A maximum of ~20% labeling efficiency is observed in the adult (4 mg TM x 3).
TM has been shown to induce nuclear translocation of CreERT2 within 6 hr and to remain in the nucleus for approximately 36 hr (Ahn and Joyner, 2004; Danielian et al., 1998). Consistent with these reports, TM injection at E8.0, ~36 hr before the onset of Cpa1 expression, yielded no βgal+ cells in Cpa1CreERT2;R26R embryos, whereas many βgal+ cells were present if TM was given at E9.5 (data not shown), suggesting that CreERT2 is active from 12 to 36 hr after injection.

Note that, in addition to CreERT2, an IRES-H2BYFP element was knocked into the Cpa1 locus (Figure S2). The presence of the IRES element should allow coexpression of H2BYFP with CreERT2 from the Cpa1 locus. However, we only detect YFP autofluorescence in the adult, but not the embryonic pancreas of Cpa1CreERT2 animals. This is likely due to the fact that the expression level behind the IRES element is generally much reduced, and that Cpa1 expression in the embryos is weaker than that in the adults.
Cpa1+ Tip Progenitors are Multipotent and Give Rise to Endocrine, Exocrine, and Duct Cells before E14

Cpa1CreERT2;R26R embryos were labeled with a single pulse of TM at different developmental stages and analyzed at E18.5 (Figure 5 and Figures S3A). Cpa1+ cells marked at E9.5, E10.5, E11.5, and E12.5 gave rise to all three major cell types of the pancreas (Figures 5A–5C, 5E–5G, 5I–5K, and data not shown)—i.e., endocrine cells (Figures 5A–5C, arrows), exocrine cells (Figures 5E–5G, hollow arrowheads), and duct cells (Figures 5E–5G, white arrowheads). Notably, TM injections at both E11.5 and E12.5 should label Cpa1+ cells when they reside within the branching tips (labeling occurs around E12–E13 and E13–E14, respectively), suggesting that the early branching tip cells are multipotent progenitors. In sharp contrast, Cpa1+ cells marked at E13.5 or after generate only exocrine progeny (Figures 5D, 5H, and 5L, arrowheads).

The observation that Cpa1+ tip cells produce multiple pancreatic cell types before E14 does not necessarily mean that Cpa1+ cells are a homogeneous population of multipotent progenitor cells. It is possible that they are composed of a mixture of separate progenitors. To distinguish between these two possibilities, we examined clones of cells that likely derive from single Cpa1+ cells (Figures S4). By varying TM doses, we first determined that, at 0.5 mg/animal, only a small number of discrete clusters of cells (0–10 clusters) are present (Figures S4D and S4E, arrows), likely representing clones. In contrast, at 2 mg/animal, around 5%–20% of the pancreatic cells are labeled (Figures S4F). No lacZ-positive cells were ever observed in the absence of TM (Figures S4A). Clones of cells labeled at E11.5 and harvested at E13.5 typically contain just a few cells (Figures S4B and S4C), whereas clones harvested at E18.5 generally contain dozens of cells (Figures S4G and S4H).

For clonal analysis, pancreata with a single lacZ+ clone or two well-separated clones were selected. Individual clones were sectioned through their entirety and stained with insulin to visualize the primitive islets. The three major pancreatic cell types can be easily distinguished based on insulin and lacZ signals: endocrine cells reside within the primitive islets (Figures S4I, arrows); duct cells exhibit an elongated shape and reside within ducts (Figures S4I, black arrowheads); and exocrine cells appear as large round cells that are part of a rosette structure (Figures S4I, white arrowheads). Out of 43 clones examined, 32 (74%) are composed of exocrine, endocrine, and duct cells (Figures S4). In addition, a small number of clones were found to contain either a mixture of two cell types or only exocrine cells (Figures S4). Although the number of clones examined is rather small, these results strongly suggest that individual Cpa1+ tip progenitors labeled between E12 and E13 are tripotent.

Multipotent Tip Progenitors Give Rise to Mature Endocrine Cells in a Stepwise Fashion

We next sought to visualize the various differentiation steps that lead from the Cpa1+ multipotent tip progenitors to mature pancreatic cells. Among the three major pancreatic cell types, the differentiation steps of the endocrine cells are best understood (Wilson et al., 2003). The earliest recognizable endocrine progenitors are Ngn3+ (Gradwohl et al., 2000; Schwitzgebel et al., 2000; Wilson et al., 2003) and subsequently give rise to a number of late progenitor cell types expressing markers such as NeuroD, Pax4, Pax6, Arx, etc. (Wilson et al., 2003). Mature endocrine cells derive from these late progenitor cells.

To visualize each step of this differentiation process, we labeled Cpa1CreERT2;R26R embryos with a single dose of TM at E12 (labeling E12.5–E13.5) and harvested them on...
three successive days for analysis (Figure 6 and Figures S3B). One day after TM injection, the initial population of TM-responsive cells expresses both βgal and Cpa1 as expected (Figure 6A, arrows). Consistent with the labeling of an early multipotent progenitor cell type, these cells were negative for the early endocrine marker, Ngn3 (Figure 6D), the late endocrine marker, Pax6 (Figure 6G), and all endocrine hormones (Figure 6J). Starting from 2 days after labeling, we observed that Cpa1+/βgal+ cells appear in the trunk region of the branches (Figures 6B and 6C, arrowheads) in addition to Cpa1+βgal+ cells residing in the tips (Figures 6B and 6C, arrows). This observation, together with the data that most Cpa1+ cells are multipotent before E14 (Figure 5 and Figures S4), suggests that some Cpa1+ are capable of limited self-renewal (i.e., producing more Cpa1+ multipotent tip cells as well as Cpa1+/βgal+ differentiated progenies).

Starting from E14, some progeny of Cpa1+ cells labeled at E12 begin to express Ngn3 (Figures 6E and 6F, arrowheads), indicating that they have become committed endocrine progenitors. One day later, at E15, Pax6+βgal+ late endocrine progenitors, as well as endo+βgal+ mature endocrine progenies, were detected (Figures 6I and 6L). These data suggest that Cpa1+ cells appear to self-renew (making more Cpa1+ cells) and lay down daughters that go through a series of differentiation steps to generate mature pancreatic cells in vivo (Figure 6M).

DISCUSSION

Formation of the mammalian pancreas as a complex three-dimensional organ requires timely generation, migration, and differentiation of different cell types from multipotent progenitors. We performed genome-wide TF expression analysis of the developing pancreas and discovered multiple distinct gene expression domains that indicate the presence of specific pancreatic progenitor domains. Using a combination of serial immunofluorescence and genetic lineage tracing experiments, we propose that one type of multipotent pancreatic progenitor can be recognized by a combination of markers (Pdx1+Ptf1a+CMyc+H+/Cpa1+), resides specifically at the
branching tips of the growing pancreatic tree, and that its proliferation and differentiation play major roles in the branching morphogenesis of the pancreas. In addition, our analysis of TFs has uncovered pancreatic genes that are likely to play important roles in pancreas development and disease.

**Domain Organization of the Developing Pancreas**

Our data show that regionalized gene expression patterns are created very early and are maintained through successive branching and growth of the pancreatic tree. The embryonic pancreas contains relatively few gene expression domains, in contrast to the developing nervous system and kidney, where many more expression patterns have been discovered in similar screen efforts (Gray et al., 2004; A.P. McMahon, personal communication). Given the large number of cell types that exist in the adult brain and kidney, the simple domain organization of the developing pancreas may reflect the relatively small number of cell types that are produced in the developing pancreas. What causes the tip and trunk of the pancreatic branches to have different gene expression patterns? In other developing organ systems, such as the lung, nervous system, and limb bud, focal sources of morphogens are responsible for creating different domains of gene expression in an otherwise homogeneous tissue (Hogan, 1999; Jessell, 2000). Such focal sources of morphogens have not been identified in the developing pancreas. In fact, although many signaling molecules are expressed in the developing pancreas, few exhibit regionalized expression. One notable exception is the Notch pathway. Notch receptors and Hes1, an effector of Notch pathway, have elevated...
expression in distal tips of the embryonic pancreatic endoderm (Apelqvist et al., 1999; Wang et al., 2005). Similarly to other developing organ systems, the Notch pathway functions in pancreas to suppress differentiation and maintain progenitors in an undifferentiated state (Apelqvist et al., 1999; Esni et al., 2004; Hald et al., 2003; Jensen et al., 2000; Murtaugh et al., 2003). These studies suggest that tip cells of the branching pancreas are actively maintained in a progenitor state by elevated Notch signaling. It is unclear, however, as to what causes the preferential activation of Notch pathway in the early distal tips. Although Fgf10 signaling has been reported to promote or maintain Notch expression in embryonic pancreas (Hart et al., 2003; Norgaard et al., 2003), pancreatic expression of Fgf10 is diffuse and rather transient (Bhushan et al., 2001).

In other tissue and organ systems, it has been shown that stem cells reside in specific “niches,” or microenvironments (Fuchs et al., 2004). Similarly, the multipotent tip progenitors of the embryonic pancreas may also have a supporting niche, suggesting that mesenchymal cells surrounding the branching tips may exhibit different properties. However, no differential gene expression has been observed in the developing pancreatic mesenchyme.

Multipotent Progenitors of the Pancreas
Our genetic lineage tracing studies show that one type of multipotent pancreatic cell is localized specifically to the branching tips. Beside the tip cells, most of the trunk cells appear to be committed endocrine progenitors (Figure 2C). In addition, there is a population of trunk epithelial cells that do not express any of the known endocrine progenitor markers (Figure 2C). Since the trunk of the developing pancreatic tree is the predecessor of the adult duct system, we favor the idea that these unidentified cells represent duct progenitors. Nevertheless, we cannot rule out the possibility that some of these Cpa1− cells are also multipotent. Further genetic studies with additional molecular markers should help resolve this important issue.

Integrating Pancreatic Branching and Cell Type Specification
Our data suggest a simple model of how pancreatic branching is integrated with cell type specification. The early pancreatic bud consists primarily of multipotent progenitors (Figure 7A). When branching begins, Pdx1+/Ptf1a+/CMyC high Cpa1+ multipotent cells divide rapidly, and possibly in a directional fashion, such that they are propelled outward from the epithelial plane, thus initiating branches. The proliferation and outgrowth of the multipotent tip cells leave behind their more differentiated progeny (i.e., the endocrine and duct cells), which make up the trunk of the branches (Figure 7B). Eventually, endocrine cells differentiate and migrate out of the epithelium, leaving the trunk to be made entirely of duct cells. At
midgestation during the so-called “secondary transition” (Pictet and Rutter, 1972), tip progenitors undergo a developmental switch that converts them into exocrine cells (Figure 7B).

Pancreatic branching continues after the secondary transition. The mechanism for continued branching, however, remains unknown. In addition, we have observed that the timing of conversion from multipotent to acinar fate is not synchronized, with Amylase expression appearing in scattered tip cells from E13.5. Reduced production of endocrine and duct cells at these later stages may explain why an earlier study has suggested that duct cells are only specified before E12.5 (Gu et al., 2002). The methodology we used to detect duct cells (whole-mount X-gal staining of E18.5 pancreas) may be more sensitive than that employed in the previous study (section staining of alkaline phosphatase [AP] on 2 month adult pancreas).

Many mammalian organs, including the lung, kidney, mammary gland, and prostate gland, develop via branching morphogenesis (Davies, 2002; Hogan and Kolodziej, 2002). The branching tips of many of these organs express regionalized markers and exhibit increased proliferation (Fisher et al., 2001; Mollard and Dziadek, 1998; Xue et al., 2001). A recent study of developing kidney provided evidence that tips of the ureteric bud are bipotential and contribute to both tips and trunks (Shakya et al., 2005). Together with our analysis, these studies suggest that it may be a common theme for all branching organs to locate their stem/multipotent cells at tips.

Cpa1 as a Multipotent Progenitor Marker

The current consensus view is that Pdx1 marks multipotent pancreatic progenitor cells. Although this is likely to be the case at the pancreatic bud stage, Pdx1 continues to be expressed broadly throughout early embryogenesis, and cannot be used to distinguish between different progenitor pools. Cpa1, on the contrary, marks only a subset of Pdx1+ cells and these cells are multipotent in vivo before E14. Although our data show that these multipotent tip cells are positive for Pdx1, Ptf1a, c-Myc, and Cpa1 and negative for differentiated lineage markers, we propose that a simpler combination of markers (Pdx1+Cpa1+Amylase) is sufficient to unambiguously identify multipotent pancreatic progenitor cells in vivo and perhaps in vitro as well.

It is noteworthy that the expression pattern of Cpa1 is highly reminiscent of that of the pancreatic TF Ptf1a (Krap et al., 1996). Expression of Ptf1 first appears around E10 and later restricts to exocrine tissues (Krap et al., 1996). Due to weak expression of Ptf1a before E12.5, we were not able to assess whether Cpa1 and Ptf1a are always coexpressed in individual cells before E12.5. Nevertheless, it is likely that Ptf1a controls Cpa1 expression. The mutant phenotype of Ptf1a suggests that it plays a critical role in specifying the early pancreatic multipotent progenitors (Kawaguchi et al., 2002). This is consistent with the idea that Cpa1 marks multipotent cells.

The ability to identify multipotent pancreatic progenitors has practical implications. Islet transplantation has recently been shown to be an effective method to treat type I diabetes patients (Lakey et al., 2006). To generate large amounts of beta cells for transplants, many ongoing efforts are focused on the directed differentiation of beta cells from embryonic stem cells. As an important intermediate cell type in these cultures, it is critical to recognize the early multipotent pancreatic precursors so that their survival and in vitro expansion can be optimized. Additionally, it would also be interesting to see whether such Pdx1+Cpa1+Amylase cells reappear in the adult pancreas after injury and serve as adult stem cells for tissue regeneration.

Experimental Procedures

In Situ Hybridization

Whole-mount in situ hybridization screen with the TF library on E9.5 mouse embryos and E14.5 pancreata was performed essentially as described previously (Gray et al., 2004). Briefly, a plasmid library that represents ~1,100 independent mouse TFs served as template for PCR amplification of the inserts. Digoxigenin-labeled cRNA probes were made directly from the PCR products using T7, T3, or SP6 polymerases (Roche) and purified through Micro Bio-spin columns (BioRad). E9.5 embryos and E14.5 pancreata were dissected and fixed with 4% paraformaldehyde overnight. After proteinase K treatment, the pancreata were hybridized with individual cRNA probes. Posthybridization washes and antibody incubation were performed with a BioLane in situ hybridization machine (Holle and Huttner, AG). Signals were developed with BM purple (Roche). Samples were cleared in 80% glycerol and photographed.

Single- and double-labeled section in situ hybridization on paraffin-embedded tissues was performed as previously described (Gray et al., 2004). For double-labeled in situ hybridization, two different probes were labeled with either digoxigenin or fluorescein. The first probe was detected with AP-conjugated anti-digoxigenin antibody, and developed with INT/BCIP (Roche), which yields a purple precipitate. After the NBT/BCIP reaction, the AP-conjugated anti-digoxigenin antibody was inactivated at 85°C and the slides were subsequently incubated with AP-conjugated anti-fluorescin antibody and detected with INT/BCIP (Roche), which yields a reddish brown precipitate.

Immunohistochemistry

Mouse embryos and pancreata were fixed by immersion in 4% paraformaldehyde from 1 to 6 hr depending on the age. Samples were subsequently incubated in 30% sucrose solution overnight and embedded with OCT compound (Vector). The following primary antibodies were used: rat anti-E-cadherin (Zymed), rabbit anti-carboxypeptidase A1 (Biogenesis), goat anti-Ngn3 (Santa Cruz), Guinea pig anti-insulin (Dako), guinea pig anti-glucagon (Linco), guinea pig anti-pancreatic polypeptide (Linco), goat anti-somatostatin (Santa Cruz), goat anti-Pdx1 (gift of Dr. Chris Wright), rabbit anti- Ins-1/2 (Bethyl), rabbit anti-Insulin (Dako), rabbit anti-Amylase (Sigma), sheep anti-Amylase (Abcam), mouse anti-Ki67 (BD PharMingen), rabbit anti-Pax6 (Chemicon), and FITC-conjugated mouse anti-phospho-histone H3 (Upstate). Rodamin-red-X, FITC, Cy5, and Alexa dye-conjugated donkey secondary antibodies were obtained from Jackson Immunoresearch Laboratories and Molecular Probes Inc. Biotin-labeled Dolichos biflorus agglutinin is from Vector Laboratories Inc.

Generation of Cpa1CreER2 Knockin Mice

The knockin vector for Carboxypeptidase A1 (Cpa1) was generated by inserting a CreER2–ires-H2B-YP–AC in cassette between a 2.7 kb 5’ arm and a 4.5 kb 3’ arm. Both arms were derived from PCR amplification of AV3 ES cell genomic DNA and confirmed by sequencing. ACN is a neoycin selection cassette that self-deletes during germ line
transmission (Bunting et al., 1999). The linearized construct was elec-
trotoped into AV3 ES cells. After neomycin selection, positive clones
were picked and screened by Southern blot with both a 5’
tm and a 3’ probe. Three recombined ES clones were injected into C57BL/6
blastocysts. Two gave germ line transmission. PCR analysis confirmed
that the ACN selection cassette was self-deleted from all germ line-
transmitted animals. Since Cpa1CreERT2 homozygous animals are
healthy and fertile, this mouse line was maintained as homozygous-
gotes. The Rosa-loxP-stop-loxP-lacZ reporter mice (R26R) were
purchased from the Jackson Laboratory and maintained as homozygotes.
All animal experiments described in this article have been approved
by Harvard University’s Institutional Animal Care and Use Committee.

Genetic Lineage Tracing

TM (Sigma T-5648) was dissolved in corn oil (Sigma C-8267) at 10 mg/ml.
Cpa1CreERT2 homozygous males were mated with R26R homozygous
females to produce double-heterozygous Cpa1CreERT2/R26R
embryos. The noon of the day of a vaginal plug was designated as E0.5.
We have found that TM doses over 2 mg per pregnant female (weighing
30–40 g each) induced significant embryonic lethality. All embryonic
experiments were performed with less than 2 mg TM per animal given
intraperitoneally.

Supplemental Data

Supplemental Data, including five additional figures and one table, are
available online at http://www.developmentalcell.com/cgi/content/
full/13/1/103/DC1/.

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REFERENCES

cells to positive hedgehog signaling during mouse limb patterning. Cell
178, 505–516.

Apeltqvist, A., Li, H., Sommer, L., Beatus, P., Anderson, D.J., Honjo, T.,
Hrabe de Angelis, M., Lendahl, U., and Edlund, H. (1999). Notch signal-

Beres, T.M., Masui, T., Swift, G.H., Shi, L., Henke, R.M., and MacDon-
ard, R.J. (2006). PTF1 is an organ-specific and Notch-independent
basic helix-loop-helix complex containing the mammalian suppressor

Bhushan, A., Itoh, N., Kato, S., Thiery, J.P., Czernichow, P., Bellusci,
S., and Scharffmann, R. (2001). Fgfl0 is essential for maintaining the
proliferative capacity of epithelial progenitor cells during early pancre-

Brennand, K., Huangfu, D., and Melton, D. (2007). All beta cells con-
10.1371/journal.pbio.0050163.

Bunting, M., Bernstein, K.E., Greer, J.M., Capecchi, M.R., and

Danielian, P.S., Muccino, D., Rowitch, D.H., Michael, S.K., and McMa-
8, 1323–1326.

Davies, J.A. (2002). Do different branching epithelia use a conserved
developmental mechanism? Bioessays 24, 937–948.

Dor, Y., Brown, J., Martinez, O.I., and Melton, D.A. (2004). Adult pan-
creatic beta-cells are formed by self-duplication rather than stem-cell

Edlund, H. (2002). Pancreatic organogenesis—developmental mecha-


hibits Ptf1 function and acinar cell differentiation in developing mouse

MAP kinase regulates branching morphogenesis in the developing

Fuchs, E., Tumber, T., and Gعاش, G. (2004). Socializing with the

Neurogenin3 is required for the development of the four endocrine cell line-

Gray, P.A., Fu, H., Luo, P., Zhao, Q., Yu, J., Ferrari, A., Tenzen, T., Yuk,
D.I., Tsung, E.F., Cai, Z., et al. (2004). Mouse brain organization re-
vealed through direct genome-scale TF expression analysis. Science
306, 2255–2257.

Gu, G., Dubaikaitel, J., and Melton, D.A. (2002). Direct evidence for the
pancreatic lineage: NGN3+ cells are islet progenitors and are dist-

Hald, J., Hjorth, J.P., German, M.S., Madsen, O.D., Serup, P., and Jen-
sen, J. (2003). Activated Notch1 prevents differentiation of pancreatic
acinar cells and attenuate endocrine development. Dev. Biol. 260,
426–437.

Hale, M.A., Kagami, H., Shi, L., Holland, A.M., Elsasser, H.P., Hammer,
R.E., and MacDonald, R.J. (2005). The homeodomain protein PDX1 is
required at mid-pancreatic development for the formation of the exo-

Hart, A., Papadopoulou, S., and Edlund, H. (2003), Fgfl0 maintains
notch activation, stimulates proliferation, and blocks differentiation of
pancreatic epithelial cells. Dev. Dyn. 228, 185–193.


Jensen, J., Pedersen, E.E., Galante, P., Hald, J., Heller, R.S., Ishibashi,
Control of endobrachial endocrine development by Hes-1. Nat. Genet.
23, 36–44.


ector-factor 1 is required for pancreas development in mice. Nature
371, 606–609.

Kawaguchi, Y., Cooper, B., Gannon, M., Ray, M., MacDonald, R.J.,
and Wright, C.V. (2002). The role of the transcriptional regulator


