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(Article begins on next page)
Effects of feminine hygiene products on the vaginal mucosal biome

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Background: Over-the-counter (OTC) feminine hygiene products come with little warning about possible side effects. This study evaluates in-vitro their effects on Lactobacillus crispatus, which is dominant in the normal vaginal microbiota and helps maintain a healthy mucosal barrier essential for normal reproductive function and prevention of sexually transmitted infections and gynecologic cancer.

Methods: A feminine moisturizer (Vagisil), personal lubricant, and douche were purchased OTC. A topical spermicide (nonoxynol-9) known to alter the vaginal immune barrier was used as a control. L. crispatus was incubated with each product for 2 and 24h and then seeded on agar for colony forming units (CFU). Human vaginal epithelial cells were exposed to products in the presence or absence of L. crispatus for 24h, followed by epithelium-associated CFU enumeration. Interleukin-8 was immunoassayed and ANOVA was used for statistical evaluation.

Results: Nonoxynol-9 and Vagisil suppressed Lactobacillus growth at 2h and killed all bacteria at 24h. The lubricant decreased bacterial growth insignificantly at 2h but killed all at 24h. The douche did not have a significant effect. At full strength, all products suppressed epithelial viability and all, except the douche, suppressed epithelial-associated CFU. When applied at non-toxic dose in the absence of bacteria, the douche and moisturizer induced an increase of IL-8, suggesting a potential to initiate inflammatory reaction. In the presence of L. crispatus, the proinflammatory effects of the douche and moisturizer were countered, and IL-8 production was inhibited in the presence of the other products.

Conclusion: Some OTC vaginal products may be harmful to L. crispatus and alter the vaginal immune environment. Illustrated through these results, L. crispatus is essential in the preservation of the function of vaginal epithelial cells in the presence of some feminine hygiene products. More research should be invested toward these products before they are placed on the market.

Keywords: Lactobacillus; L. crispatus; cytokines; Interleukin-8; Nonoxynol-9; vaginal microbiota; vaginal epithelial colonization; mucosal immunity
The presence of *Lactobacillus* has been associated with lower risk of inflammation (2). Lactobacilli and, especially, *L. crispatus* are among the bacteria that are most common in healthy women and characteristic for the healthy vaginal environment (4, 5). Lactobacilli are anaerobic bacteria that convert lactose and other sugars into lactic acid, which has a role in preventing infections (4). *Lactobacillus* is found in the gut and vagina, where it has anti-inflammatory and anti-cancer activities. In the industry, it is used to make yogurt and cheese, and it can also be used as a biotherapeutic. Because of its beneficial properties to the host, some *Lactobacillus* species have been used as probiotics available over the counter to aid mucosal health, for example, in the digestive tract. *Lactobacillus* products are also under investigation to restore vaginal health after prolonged use of antibiotics or to cure BV and prevent urinary tract infections (6, 7). *L. crispatus* has been found to effectively help inhibit the growth of harmful pathogenic microorganisms such as *Neisseria gonorrhoeae* (8). This study applied an experimental system for safety evaluation of vaginal products by testing their effects on *L. crispatus* growth and survival *in vitro*. The evaluation was carried out by comparing products representing over-the-counter (OTC) douche kits, moisturizers, and lubricants to cell culture medium that maintains the growth of Lactobacilli *in vitro* and nonoxynol-9 (N-9), a spermicide known to upset the vaginal immune barrier (9–11).

**Materials and methods**

**Test products**

The test articles listed in Table 1 (Vagisil Feminine Moisturizer, personal lubricant and moisturizer, and CareOne Douche) were purchased over the counter and were stored completely sealed, in their original box, at room temperature. The selection of products was made based on availability and public knowledge on each of the products. Once purchased, each product was screened to check for sterility. This was done by seeding each product undiluted over agar plates. None of the products showed any bacterial growth in this test. Phosphate-buffered saline (PBS) (Invitrogen by Life Technologies, Carlsbad, CA) and keratinocyte serum-free medium (KSFM) (Invitrogen, Carlsbad, CA) supplemented with epidermal growth factor, bovine pituitary extract, and CaCl₂ as described (12) were obtained from Invitrogen, Carlsbad, CA. Nonoxynol-9 (N-9) was obtained from Personal Products Company (Skillman, NJ) (13) and diluted to 2% with KSFM. For assessing their effect on bacterial growth, all products were applied at 1:1 ratio over the bacterial suspensions.

**Preparation of bacterial suspension and treatment**

*L. crispatus* was originally isolated from vaginal swab samples from healthy women participating in a vaginal microflora research study and then characterized and expanded in the laboratory of Dr. A. Onderdonk, Brigham and Women’s Hospital (14). Once the volume needed was calculated, the frozen bacterial stocks were thawed in a room temperature water bath. The desired volume was transferred into a 50 mL tube and centrifuged down at 5000g for 10 min at 25°C. After discarding the supernant, the bacterial pellet was resuspended in KSFM cell culture medium to the concentration of 7 × 10⁵ CFU/mL. Hundred microliter of the bacterial suspension was then added over 100 microliter undiluted product in 96-well tissue culture plates (Becton Dickson and Company, Franklin Lakes, NJ). The designated plates were then incubated on an orbital shaker in an anaerobic chamber (AnaeroPack System, PML Microbiologicals, Wilsonville, OR) at 35°C for 24 h or 2 h. From the 200 µl of test products along with bacterial suspension, three replicates within the same tissue culture plate were made. Each replicate contained 50 µl of the mixture of product and bacteria, along with 50µl of 50% glycerol, and was cryopreserved at −80°C.

**Dilution of test plates**

To assess colony forming units, the bacterial suspensions in KSFM or in test product collected at each time point were serially diluted using 1 × PBS. The first row of the 96-well plate contained *L. crispatus* along with the product. The following rows contained the desired amount of dilution in PBS achieved by transferring desired volume from one row to another using multichannel pipettes, and when the product had high viscosity, we used positive displacement pipettes (Gilson Medical Electronics through Fisher Scientific, Pittsburgh, PA).

<table>
<thead>
<tr>
<th>Products</th>
<th>Ingredients (from label)</th>
<th>Approximate pH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vagisil Feminine Moisturizer</td>
<td>Water, glycerin, propylene glycol, hydroxyethylcellulose, DMDM hydantoin, diazolidinyl urea, disodium EDTA, polysorbate 20, methylparaben, propylparaben, <em>Anthemis noilis</em> (camomille) flower extract, polyquaternium-5, citric acid, <em>Aloe barbadensis</em>, tocopheryl acetate</td>
<td>4</td>
</tr>
<tr>
<td>CareOne Douche</td>
<td>Purified water, benzoic acid, and vinegar</td>
<td>3</td>
</tr>
<tr>
<td>Personal lubricant</td>
<td>Propylene glycol, glycerin, <em>Trifolium pratense</em> flower extract (clover), methylparaben</td>
<td>5</td>
</tr>
</tbody>
</table>
Seeding bacteria over agar plates
Brucella Agar Plates with 5% sheep blood, hemin, and vitamin K1 (BD, Franklin Lakes, NJ) were brought out from refrigerator to adjust to room temperature for 10 minutes. A pipette tip was used to mark either halves or thirds or the agar surface. After the serial dilutions of *L. crispatus* plus product were made, 30 μl was taken from a single well and was spread out over the agar, using a Sterilloop (Fisher Scientific, Pittsburgh, PA). The plates were then incubated in an anaerobic chamber for 48–72 h, or until there were visible colonies.

Colony forming units counts
Bacterial colonies were counted in the agar plates. The data was then entered into Excel, multiplied by the dilution factor, and transformed logarithmically (log10). The average log10 data from all dilutions that generated countable colony-forming units (CFU) for each culture in each experiment were used for statistical analysis of the triplicate cultures conducted in at least two independent seeding experiments for each time point and product.

Preparation of vaginal epithelial cells and seeding over 96-well plates
We utilized for this study an immortalized vaginal keratinocyte cell line (Vk2/E6E7), which maintains the differentiation patterns and phenotypic characteristics of its normal human tissue of origin (12, 15, 16). Vaginal cells were grown in a T75 flask using KSFM medium. Once grown to confluence, the cells were trypsinized. After decanting the old medium, 5 mL of warm trypsin was added and the flask was tipped back and forth to wash away any dead cells. After removing the trypsin, 10 mL of fresh trypsin was added and incubated for 10 min at 37°C. After seven minutes, detachment of cells was checked under a microscope. Once enough cells were detached, equal volume of neutralization medium was added on top of the trypsin. The cell suspensions were then spun down at 500 g for 10 min. Next, the supernatant was decanted, and the cells were resuspended in 2 mL of antibiotic-free medium (AB-) and counted. The cell density was adjusted to 3 in 2 mL of antibiotic-free medium (AB-) and counted.

Bacterial-epithelial co-culture and epithelial-associated CFU counts
Details on the bacterial colonization model used in this study are published elsewhere (17). In brief, once the vaginal keratinocytes reached 100% confluence, medium was removed and replaced by 50 μL of bacterial suspension and 50 μL of test product. The plate was then placed in an anaerobic jar with one AnaeroPack inside and incubated for 24 h. After 24 h incubation, supernatants were collected and frozen for cytokine assays. Using 200 μL of (1 ×) PBS, cell layers were washed in original plate twice with a multichannel pipette. Cells were then lysed by adding 100 μl of Hypure water and incubated at room temperature for 15 minutes followed by adding 100μl of (2 ×) PBS over the Hypure water and mixing well. Lysates were serially diluted in PBS and seeded over agar plates with a sterile loop. Agar plates were incubated under anaerobic conditions for 48–72 h, followed by CFU counting.

Statistical analysis
Two-tail ANOVA analysis of variance and Prism GraphPad Software Inc, San Diego, CA) were used to compare the CFU values, viability, and IL-8 levels obtained for test product and the control medium.

Results
Effect of products on planktonic *L. crispatus* culture
To simulate a naturally thriving environment of the natural microbiota of the vagina, *L. crispatus* was incubated at 2 and 24 h with KSFM medium, which supports the growth of human vaginal epithelial cells (12). Acting as a control, it was expected that under these conditions the bacteria would grow to a sufficient number of colonies. These plates were used as a maximum threshold to compare the serial dilutions done in the absence or presence of OTC vaginal products. The results from the series of incubations and dilutions concurred with the expectations for good support of bacterial growth demonstrating recovery of CFU input counts (Fig. 1 columns one). Acting as a bactericidal control, N-9 was incubated along with *L. crispatus* for both 2 and 24 h. As expected, N-9 had a negative effect on the bacteria. At 24 h N-9 completely killed the bacteria, while at 2 h, N-9 significantly (p <0.01) decreased the number of bacterial colonies (Fig. 1). The personal lubricant explored during this study decreased number of bacterial colonies by less than 1 log at 2 h incubation period (p <0.05) (Fig. 1A); however, at 24 h, it completely prevented bacterial growth and no colonies were visible from seeding of either diluted or undiluted bacterial suspensions (Fig. 1B). Vagisil significantly suppressed *Lactobacillus* growth even at a short 2 h incubation time (Fig. 1A) and at 24 h completely...
douche
production (Fig. 2D).

difference from control.
differences from the medium control. n.s.
to bacteria twice. 

at (A) 2 h and (B) 24 h for each vaginal product exposed
standard errors of the mean from triplicate CFU measurements
CFU. Bars represent logarithmically transformed means and

bacteria the douche triggered increased IL-8 production
surface in response to infection (18, 19). In the absence of
vaginal epithelium to attract leukocytes to the mucosal

inflammatory innate immunity mediator produced by the

the production of IL-8, which is an important proin-
flammatory innate immunity mediator produced by the
vaginal epithelium to attract leukocytes to the mucosal
surface in response to infection (18, 19). In the absence of
bacteria the douche triggered increased IL-8 production
(Fig. 2C). L. crispatus counteracted this effect. In the
presence of L. crispatus, all products suppressed IL-8 production (Fig. 2D).

Effect of products on bacterial-epithelial interactions

All products tested were toxic to the vaginal keratinocytes when applied undiluted (at 100% dose) (Fig. 2A). The
douche was the least toxic and the moisturizer most toxic over a set of dilutions (Fig. 2A). While the undiluted
douche stimulated bacterial growth (increasing recovery of epithelial cell-associated CFU to \(\sim 200\%\)), the other
products suppressed bacterial growth, with the moisturizer being suppressive up to a 4-fold dilution (at 25% dose) (Fig. 2B). The products had differential effects on the
production of IL-8, which is an important proinflammatory mediator produced by the vaginal epithelium to attract leukocytes to the mucosal

Fig. 1. Composite effect of vaginal products on L. crispatus
cFU. Bars represent logarithmically transformed means and
standard errors of the mean from triplicate CFU measurements
at (A) 2 h and (B) 24 h for each vaginal product exposed
to bacteria twice. \(P\) values \(<0.05\) and \(<0.01\) show significant
differences from the medium control. n.s. = no significant
difference from control.

abolished bacterial colonies (Fig. 1B). The vaginal
douche tested in this study did not show any significant
effect on L. crispatus as compared to the KSFM control (Fig. 1A and B).

Discussion

The mucosal lining of the vagina provides essential
help in maintaining a healthy vaginal environment. The
results from this experimental study demonstrate that some vaginal products may be harmful to the Lactobacillus bacteria and therefore should be used with caution. Referring to Fig. 1, both the lubricant and N-9 showed
to have a significant effect on the growth of L. crispatus. At 24 h (Fig. 1), both the lubricant and N-9 completely
inhibited bacteria growth in our experimental system.

The results from this experimental study confirm the
hypothesis that some feminine hygiene products may
alter the vaginal immune barrier by having a negative
effect on the epithelial cell integrity (measured by MTT),
survival of beneficial Lactobacillus species in the vaginal
microenvironment (measured by epithelial-cell-associated
CFU), and by changing the ability of the vaginal epithelial
cells to produce protective or inflammatory immune
mediators, for example, IL-8. The results confirm clinical
findings of the harmful effects of certain vaginal hygiene
practices on susceptibility to infections. The moisturizer
and the lubricant were especially cytotoxic and negatively
affecting L. crispatus survival. The douche, on the other
hand, could promote a proinflammatory environment
in the absence of Lactobacillus or when the microbiome is
disturbed. More clinical studies are needed to test the
findings and conclusions from these experiments, which if
confirmed would argue for the benefit of using probiotics
for vaginal health.

Our results provide experimental warning that women
who have used nonoxynol-9, Vagisil, or lubricant may
have a weakened vaginal barrier due to destroyed
L. crispatus and perhaps other normal microflora species
not assessed in our study, thus becoming at risk for
the development of the syndrome of disturbed vaginal
microbiome (20). Women using such products on a regu-
lar basis should take measures such as regular use of
condoms to prevent sexually transmitted infections and
should be evaluated for BV, which can sometime be
asymptomatic and thus remain unnoticed by the vaginal
product user. Although the particular douche kit tested
did not show to have a negative effect on L. crispatus
growth in our experimental system, it must be noted that
some douching products have been associated with B
streptococci and different Candida species (21). Douching
has also been associated with a non-regression of low-
grade squamous intraepithelial lesions (21). The use of
vaginal spermicides and vaginal medications has been
known to onset an unstable microflora and BV (22). It has
to also be noted that the effects of vaginal products on
bacterial growth can be selective and can vary among
commercial brands, as suggested by another in-vitro study,
which demonstrated differences among seven commercial
douches tested against Lactobacillus isolates as well as
of biological triplicates. * = p < 0.05, ** = p < 0.01, *** = p < 0.001, values in product-treated cultures lower than medium control; + = p < 0.05, ++ = p < 0.01, +++ = p < 0.001, values in product-treated cultures higher than medium control.

Fig. 2. (A) Effect of product on vaginal keratinocyte (Vk) viability assessed by MTT assay, (B) L. crispatus colonization assessed by Vk-associated CFU, and Vk IL-8 production in (C) the absence or (D) the presence of L. crispatus. Data represent means and SEM of biological triplicates. * = p < 0.05, ** = p < 0.01, *** = p < 0.001, values in product-treated cultures lower than medium control; + = p < 0.05, ++ = p < 0.01, +++ = p < 0.001, values in product-treated cultures higher than medium control.

BV associated bacteria (23). All three products tested were estimated to be fairly acidic (Table 1), although the pH varied within values typical for normal vaginal environment, except for the douche, which was more acidic.

Using the data that was amassed from this project, more research can hopefully go into vaginal health and development of probiotics for restoring and maintenance of a healthy vaginal microflora. Conventional health care wisdom has warned consumers against the use of douches due to their stripping the vagina of its protective lining and altering its pH. Though the brand we tested did not show to inhibit the growth and altering its pH. Though the brand we tested did not show to inhibit the growth of L. crispatus in vitro of L. crispatus, frequent douching with this or other brands may throw the vagina’s internal protective system off balance and promote non-sexually and sexually transmitted infections. Regardless of this, millions of females still continue to use it for personal and cultural reasoning. Hopefully this research may blossom into a new field in the biopharmaceutical market. Just as vaginal suppositories exist containing estrogen replacement therapies and anti-fungal agents to combat non-sexually transmitted infections such as candida (yeast) and BV, we may envision vaginal probiotic replacement therapies in the future in order to promote vaginal health and curb the frequency of reoccurring yeast infections and as a supplement available to women who continue to use these products. Preventative therapies are the path of modern medicine. Finding treatments for tomorrow’s disease may be the cloud with a silver lining for patients who are afflicted, but preventing them is golden.

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References

2. Fichorova RN, Onderdonk AB, Yamamoto H, Delaney ML, Dubois AM, Allred E, et al. Maternal microbe-specific

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