Inactivation of the RB family prevents thymus involution and promotes thymic function by direct control of Foxn1 expression

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The thymus plays a critical role in adaptive immunity as the site of mature T cell production (Miller, 2011). The thymus grows rapidly during early life, generating a large T cell repertoire. It then enters an involution process, leading to decreased production of naive T cells, resulting in impaired immune function in the elderly and preventing complete reconstitution of the immune system in various pathologies (Boehm, 2008; Rodewald, 2008; Carpenter and Bosselut, 2010). The mechanisms controlling thymic involution are poorly understood, hampering the development of therapeutic strategies to enhance immune function in a wide variety of patients (Napolitano et al., 2008; Sauce and Appay, 2011).

Although the thymus is composed mostly of T lymphocytes, T cell development requires a complex microenvironment including endothelial, dendritic, and thymic epithelial cells (TECs). TECs are highly proliferative during thymic expansion, and then their cell cycle slows considerably (Gray et al., 2007; Aw and Palmer, 2011; Manley et al., 2011). Recent data show that keratinocyte growth factor (KGF; or FGF-7) and growth hormone treatment in aged rodents and humans leads to transient thymic expansion and increased production of naive T cells (Min et al., 2007; Napolitano et al., 2008). In particular, increased TEC numbers were observed upon KGF treatment (Min et al., 2007; Rossi et al., 2007). IL-22 also can support thymic regeneration in mice after radiation treatment (Dudakov et al., 2012). These data suggest that increased numbers of TECs...
can enhance, at least temporarily, thymic function. Nevertheless, the molecular mechanisms regulating cell cycle activity in TECs are still poorly characterized, and no strategies have been devised yet for long-term thymic growth.

Through its ability to bind the E2F transcription factors, the RB family of proteins (RB, p107, and p130) plays a major role in the control of cell cycle progression. Growth factors and external signals activate Cyclin and Cyclin-dependent kinase (CDK) protein complexes. Upon activation, Cyclin-CDK complexes phosphorylate RB family proteins, resulting in their inactivation. Inactivation of RB family proteins by phosphorylation activates E2F, thereby promoting transcription of genes involved in the G1/S transition of the cell cycle (Iaquinta and Lees, 2007; Chinnam and Goodrich, 2011).

There is no reported thymic phenotype in mice in which any one of the RB family gene is inactivated, possibly because of the strong functional overlap between the three proteins (Dannenberg and te Riele, 2006). Nevertheless, emerging evidence suggests that some members of the RB pathway may play a role in thymic biology, including E2F2, Cyclin D1 (CCND1), p18
textsuperscript{Ink4c}, and p27
textsuperscript{Kip1} (Robles et al., 1996; Franklin et al., 1998; Pierce et al., 1998; Klug et al., 2000; Rodriguez-Puebla et al., 2000; Iglesias et al., 2004; Scheijen et al., 2004; Chien et al., 2006). However, the mechanisms underlying how the cell cycle machinery affects thymus development and involution are still unknown.

Here we report that deletion of Rb family genes in the thymus of mice leads to increased proliferation in TEC populations and prevents thymic involution. Furthermore, we found that the RB family regulates the transcription of Foxn1, an important regulator of TEC differentiation and function. Finally, we demonstrate that the increased expression of Foxn1 is required for the thymus expansion observed in Rb family mutant mice. These data identify a new RB-E2F-Foxn1 module as a critical regulator of thymic involution and function.
hyperproliferation in multiple organs (Viatour et al., 2008; Chen et al., 2011). In contrast, reintroduction of one copy of p107 (Mx1-Cre Rb+/lox/; p130lox/lox; p107+/−/ or Mx1-Cre p107-Single mice) rescues the lethality of the triple knockout mice and significantly extends their lifespan up to 9–12 mo of age (Viatour et al., 2008, 2011). At that age, although p107-Single mice that lack Cre-recombinase are still healthy, Mx1-Cre p107-Single mice exhibit weight loss and respiratory distress. Upon autopsy, we found that the thymus of Mx1-Cre p107-Single mice was significantly increased in size (Fig. 1 A, left), compressing the lungs. This thymic growth correlated with increased cellularity (Fig. 1 B). The plateau observed after several months of continuous growth in mutant mice may be the result of mechanical or vascular constraints, limits of the BM to generate T cell precursors, or a thymus-intrinsic effect.

Histopathological analysis showed no gross alteration of the thymic architecture in mutant mice compared with controls (Fig. 1 A, right). As expected (Yang et al., 2009; Aw and Palmer, 2011), the thymus from 6–9-mo-old mice accumulated lipids, as indicated by Oil Red O staining. In contrast, the thymus of young control (2–3 mo of age) and old mutant mice (6–9 mo of age) showed little lipid accumulation (Fig. 1 D). The only defect we detected in mutant mice outside of the thymus was epidermal hyperplasia (not depicted; previously described in Ruiz et al., 2004). Thymic size in Mx1-Cre Rb/p130 double mutant mice was indistinguishable from that of control mice (Fig. 1 D). Overall, these data show that loss of RB family function in Mx1-Cre p107-Single mice results in the expansion of the thymus, while preserving overall thymic architecture.

Flow cytometry analysis of progenitor thymocyte populations (Fig. 1 E and Fig. S1 A) revealed no significant differences in the frequencies of these cell populations in the thymus of control and mutant mice. There were no significant changes in the number of double-negative (DN) cells between control and mutant mice. Double-positive (DP) mature intrathy mic T cells were slightly more numerous in the mutant mice, whereas CD4+ and CD8+ populations were smaller (Fig. 1 F and Fig. S1 B). These changes may reflect a higher percentage of DP thymocytes failing positive selection in the mutant animals (Morris and Allen, 2012). As expected (Pénit and Vasquez, 1997), BrdU/propidium iodide (PI) cell cycle analysis showed some BrdU incorporation in CD4+ and CD8+ populations in control mice, but proliferation rates were higher in mutant CD4+ and CD8+ cells (Fig. 1 G). These observations show that loss of RB family function results in a general expansion of thymocytes rather than expansion of a specific subpopulation as would be seen in a T cell lymphoma. Notably, this increase in the number of intrathymic T cells was accompanied by an increase in the number of peripheral CD62L+ CD44− naive T cells in Rb family mutant mice (Fig. 1 H and Fig. S1 C). This last finding is consistent with a recent observation that forced expression of thymic Foxn1 leads to a decrease in peripheral memory T cells (Zook et al., 2011). Thus, inactivation of the Rb gene family results in a thymic expansion that correlates with increased production of naive T cells in mice.

To investigate the origin of thymic expansion in Mx1-Cre p107-Single mice, we generated chimeric mice by transplanting Ly5.1+ / Ly5.2+ wild-type BM cells into 3–mo-old Ly5.1+ lethally irradiated mutant or control mice (Fig. 2 A). Upon reconstitution of hematopoiesis, chimeric mice were injected with polyinosine-polycytosine (pI:pC). 3 mo later, the thymi of Mx1-Cre p107-Single mice reconstituted with wild-type hematopoietic cells were enlarged significantly compared
Figure 3. Expansion of functional TECs in the thymus of Rb family mutant mice. (A) Immunoblot analysis (one of two replicates shown) of protein lysates from pooled CD45− EPCAM+ MHCII+ TECs from young (2 wk; n = 10), control (Ctrl; 15 wk injected; n = 10), and Mx1-Cre p107-single (15 wk; n = 3 injected, 2 noninjected) thymi against RB, phospho-RB, CCND1, CDK4, and α-Tubulin (loading control). (D) PCR analysis of genomic DNA isolated from CD45− EPCAM+ MHCII+ TECs for the lox-flanked or Cre-deleted exons of Rb and p130 in 15-wk-old control, non–pI:pC-injected Mx1-Cre p107-Single, and pI:pC-injected Mx1-Cre p107-Single mice (one of four shown). (C) RT-qPCR evaluation of expression of E2F target and S-phase genes in flow-purified TECs from 2-wk-old (n = 2), 3-wk-old (n = 5), and 15-wk-old control mice (n = 3), including E2f1 (p-values for all genes presented as 2–3 wk and 3–15 wk, P = 0.044 and 0.031, respectively), p107 (P = 0.001 and 0.04), PCNA (P = 0.026 and 0.018), bMyb (P = 0.026 and 0.041), Ezh2 (P = 0.0001 and 0.40), and Mcm3 (P = 0.018 and 0.073). (D) RT-qPCR evaluation of expression of E2F target and S-phase genes in flow-purified TECs from control and mutant TECs (15 wk; n = 3 each). Genes analyzed included E2f1 (P = 0.017), p107 (P = 0.004), PCNA (P = 0.033), bMyb (P = 0.026), Ezh2 (P = 0.059), and Mcm3 (P = 0.022). (E) PI analysis of cell cycle activity in TECs from 15-wk-old control and Mx1-Cre p107-Single mice (G1: Ctrl, 98.08 ± 1.18%; Mut, 69.17 ± 9.49%; P = 0.0003; S: Ctrl, 1.24 ± 0.78%; Mut, 18.86 ± 6.32%; P = 0.0002; G2: Ctrl, 1.24 ± 0.85%; Mut, 12.13 ± 3.68%; P = 0.0004; n = 10 control and 4 mutant). (F) BrdU incorporation.
with the thymi from chimeric wild-type/control mice (Fig. 2 B). In both cases, as expected, BM cells and thymocytes from chimeric mutant and control mice expressed both Ly5.1 and Ly5.2 isoforms, confirming their wild-type origin (Fig. 2 C). Although all T cell populations were expanded in chimeric mutant mice (Fig. 2 D), there was, overall, no significant difference in the proportional representation of DN, DP, CD4+, and CD8+ populations between wild-type and mutant chimeric mice (Fig. 2 E). Nevertheless, the proliferative index of T cell populations in mutant mice was increased compared with controls (Fig. 2 E). Mutant T cells failed to exhibit increased proliferation upon transplantation of mutant BM cells in recipient mice (not depicted). Thus, although there may be a contribution of increased T cell proliferation to the thymic expansion observed in mutant mice, the thymus expansion observed in Rb family mutant mice does not depend on the deletion of Rb family genes in BM-derived cells.

Because thymic stroma-derived cytokines and growth factors are important for thymus growth and T cell proliferation and differentiation, we assessed mRNA expression levels for several cytokines and receptors produced in the thymus (IL7, Sef/kitl, Fgfl2b, Wnt4, and Dll4). After normalization to Eva1 levels (Guttinger et al., 1998; DeMonte et al., 2007; Min et al., 2007), we found no significant difference in per-cell levels of these mRNAs between chimeric mutant and chimeric control groups (Fig. 2 F). These data provide more evidence that the thymic milieu is essentially normal in the enlarged thymus of Mx1-Cre p107-Single mice, further suggesting that the thymic microenvironment is expanded and maintained in Rb family mutant mice.

TECs are a critical component of the thymic microenvironment, providing many of the functions needed to control T cell differentiation and selection. Thus, we asked whether TECs may be responsible for the thymus phenotype observed in Mx1-Cre p107-Single mutant mice. We found that the pool of phosphorylated RB decreased with age, correlating with higher levels of CCND1 in TECs from younger mice; Cdk4 levels did not change with age (Fig. 3 A). Accordingly, transcription of canonical E2F targets in TECs significantly decreased with age (Fig. 3 C), further correlating thymic involution with increased RB activity. We also confirmed loss of RB by immunoblot analysis (Fig. 3 A) and deletion of Rb and p130 by PCR in injected and noninjected Mx1-Cre p107-Single TECs (Fig. 3 B). Deletion of Rb and p130 without pl:PC injections confirms that the Mx1 promoter can be induced by endogenous signals (Khün et al., 1995); this deletion in noninduced mice also led to a significant increase in thymus size (thymic cellularity in control, 9.8 ± 0.2 × 10^9; versus noninjected Mx1-Cre p107-Single, 1.2 ± 0.7 × 10^9; P = 0.0001). The extent of deletion of the Rb family seen in the thymus of these noninduced mice suggests that the mutant cells may outcompete wild-type or partially deleted cells as the thymus becomes enlarged. Notably, inactivation of the Rb gene family in Mx1-Cre p107-Single mice prevented the downregulation of E2F targets normally observed during aging (Fig. 3 D). Because CCND1 production is induced by mitogenic signals (Klein and Assoian, 2008), the lack of detectable CCND1 in 15-wk-old control and mutant mice argues that, at this age, TECs do not receive extracellular mitogenic signals, further suggesting that the thymic growth observed in Mx1-Cre p107-Single mice is caused by a TEC-intrinsic effect.

Next, we sought to quantify the extent to which loss of RB family activity in TECs increased TEC numbers in the mutant thymus. CD45+, MHCII+ TECs increased 18-fold compared with CD45+ hematopoietic cells in the thymus of the mutant mice (control, 0.9 ± 0.49% TECs; mutant, 16.42 ± 6.9% TECs; n = 3 each; P = 0.018). Mutant TECs also displayed a higher proliferative index compared with their wild-type counterparts (Fig. 3 E). Additional separation of CD45+ MHCII+ TECs from mutant thymus based on the Ly51 surface marker (Williams et al., 2009) showed that mutant cortical TECs (cTECs; Ly51+) and medullary TECs (mTECs; Ly51-) displayed a high proliferative activity (Fig. 3, F and G). Compared with Ly51+ TECs, we noted an increase in the number of Ly51+ TECs in G2, which is sometimes seen in cells with loss of RB function (Stark and Taylor, 2006; Conklin et al., 2012).

To determine whether there was a change in proportional representation of particular TEC subsets, we quantified levels of the mature mTEC marker Aire (Gray et al., 2007; Dooley et al., 2008) in control and mutant thymi. We found no significant difference in Aire expression in Mx1-Cre p107-Single mice (Fig. 3 F), suggesting that there is no preferential expansion of either cTECs or mTECs in Rb family mutant mice. We also measured by flow cytometry the proportions of cTEC precursors as identified by CD40 and CD205 expression (Shakib et al., 2009; Nowell et al., 2011). There was...
To examine further the differentiation state of RB family-deficient TECs, we performed immunofluorescence analysis of markers for various cell populations in the thymic microenvironment, including Keratin 5 (K5), Keratin 8 (K8), Keratin 14 (K14), the proteasomal β-5t subunit (β5t), and CD205 (Rodewald, 2008; Shrimpton et al., 2009; Takahama et al., 2012). We found no evidence for alteration of thymus structure in mutant mice (Fig. 3 L). We occasionally observed subcapsular cysts in very large mutant thymi (not depicted), perhaps reflecting the physical limitations placed by the thoracic cavity upon the enlarged thymus. This observation highlights the fact that we performed our analyses of TEC subpopulations at a late time point, when the cells analyzed may represent no difference in CD40/CD205 subpopulations between control and mutant mice (Fig. 3 I and Fig. S1 D). Similarly, there were no differences in mTEC precursor populations as measured by CD80 and UEA1 expression (Fig. 3 J and Fig. S1 E; Akiyama et al., 2012). When we measured mRNA expression levels for two thymocyte growth cytokines that are expressed by differentiated TECs (Il7 and Scf), we found that the expression levels of these genes on a per-cell basis were not significantly different between control and mutant thymi (Fig. 3 K). These data support the idea that the major difference between control and Mx1-Cre p107-Single mice lies in the number of TECs and not in the differentiation state of these cells.

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These observations suggested that, in addition to its canonical role in regulating progression through the cell cycle, the RB family also controls TEC expression of Foxn1 (Balciunaite et al., 2002). Little is known about the transcriptional control of Foxn1. There are two alternate promoters and first exons in the Foxn1 gene (Schorpp et al., 1997; Cunliffe et al., 2002): the more distal promoter region (1a) is transcriptionally active in both the skin and the thymus; the second promoter region (1b) is active only in keratinocytes of the skin (Fig. 4 D). We found by computational analysis that region 1a harbors several conserved putative E2F binding sites. We labeled some of these putative sites “site 1” and “site 2–3”, with site 2–3 containing two independent but adjacent putative sites (Fig. 4 E).

To test the possibility that E2Fs might act at the Foxn1 promoter, we cotransfected an expression vector coding for E2F3, an activating member of the E2F family, with a reporter plasmid upstream of a Foxn1 promoter fragment (Zook et al., 2011). Thus, the thymic outgrowth in Rb family mutant mice is probably also dependent on the increased proliferation seen in mutant TECs, and this increased proliferation is likely to be greater than that with increased Foxn1 levels alone.

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two sites in site 2–3 partially inhibited the ability of E2F3 to activate the Foxn1 promoter (Fig. 4 F). Coexpression of RB markedly inhibited E2F3-mediated activation of the Foxn1 promoter (Fig. 4 G). We further tested direct binding of E2Fs to the Foxn1 promoter by performing chromatin immunoprecipitation (ChIP) experiments in several TEC lines (TEC100, TEC71, and 210R; Friend et al., 1994; Wang et al., 1998). We found that E2F3 and E2F4, a repressor E2F, can bind to both site 1 and site 2–3 but do not bind significantly to regions of nonspecific DNA on the fifth chromosome of the mouse (Fig. 4 H for TEC100; TEC71 and 210R not depicted). We repeated similar experiments after the transient ectopic expression of RB in TEC100 cells and found a decrease in endogenous Foxn1 expression (fold change: Rb mRNA, 3.1 ± 0.11; Foxn1 mRNA, −1.8 ± 0.11; n = 3 each), as well as a switch between the E2F3 activator and the E2F4 repressor at the Foxn1 promoter (Fig. 4 I). Collectively, these data show that Foxn1 is a direct target of E2F regulation; E2F binds to and controls transcription at the Foxn1 promoter, and RB inhibits this activation.

To assess the in vivo functional role of increased Foxn1 levels in the thymus expansion observed in Mx1-Cre p107-Single mice, we studied thymic growth in Mx1-Cre p107-Single; Foxn1−/− mutant mice. Foxn1−/− is a hypomorphic allele of Foxn1, which causes down-regulation of Foxn1 protein expression from soon after birth; of interest, Foxn1−/− heterozygous mice have no appreciable thymic defects (Chen et al., 2009). Similar to Mx1-Cre p107-Single mice, Mx1-Cre p107-Single Foxn1−/−/+ mice displayed significant thymic expansion. In contrast, Mx1-Cre p107-Single Foxn1−/−/− mice had a thymus equivalent in size to thymus from control (Rbp7−/−, p130−/−, p107−/−; Foxn1+/− without Cre) mice (Fig. 5 A and B). As expected, Foxn1 levels were reduced in the thymus of Mx1-Cre p107-Single Foxn1−/−/+ mice compared with Mx1-Cre p107-Single Foxn1−/−/+ or Mx1-Cre p107-Single mice (Fig. 5 C and D). We also observed decreased expression of E2F targets in the thymus of Mx1-Cre p107-Single Foxn1−/−/+ mice (Fig. 5 E). This result suggests, as has been observed in zebrafish (Ma et al., 2012), that Foxn1 plays a role not only in differentiation of TECs but also in promoting proliferation through regulation of S phase genes. These observations also indicate that decreased expression of Foxn1 is sufficient to rescue the thymic phenotype observed in Rb family mutant mice, indicating that increased Foxn1 levels induced by unrestricted E2F activity upon inactivation of Rb family genes are a major cause of the expanding thymus in these mice.

The transcriptional activation of Foxn1 in Rb family mutant cells is reminiscent of the interactions between RB, E2F, and PPARγ (peroxisome proliferator-activated receptor gamma subunit): increased levels of PPARγ upon loss of RB function in mesenchymal cells changes the fate of these cells toward the adipocytic lineage and may affect the spectrum of tumors in mice (Cálo et al., 2010). However, a unique aspect of the RB-E2F-Foxn1 module is that it keeps in check the protumorigenic consequences of losing RB family function, at least in part by activating prodifferentiation functions of Foxn1.

Our data demonstrate that the RB pathway plays a key cell-autonomous role in TECs to control maximal thymic size and aging. This result is striking when one considers that thymic size likely is controlled by complex feedback interactions between stromal cells and maturing T cells. Perhaps surprising, given the well-characterized tumor suppressor role of RB family members, loss of RB family function in the thymus does not perturb the differentiation of TECs or T cells and simply increases the proliferation of some of these cells. These observations raise the possibility that loss of RB family function could be used to restore or promote thymic expansion and T cell production in patients (van den Brink et al., 2004; Wils and Cornelissen, 2005). Clearly, however, constant inactivation of RB family function in the thymus, although initially beneficial for the immune system of mice, eventually becomes detrimental to the animals when the mutant thymus has reached a large size in the chest. Thus, an important future step will be to identify novel means to transiently inactivate the RB family in a controlled fashion to minimize the potential negative effects of this inactivation while optimizing the benefits to the immune system of patients.

MATERIALS AND METHODS

Animals. All mice where housed in the Stanford University School of Medicine Research Animal Facility in accordance with institutional and National Institutes of Health guidelines. All animal care and experiments were approved by the Stanford University Administrative Panel on Laboratory Animal Care and followed the guidelines of this animal use committee. Mice were of a mixed C57/129 background. Mx1-Cre Rbp7−/− p130−/− p107−/− mice were previously described (Viatour et al., 2008). Mx1-Cre p107-Single mice were generated by breeding Mx1-Cre Rbp7−/− p130−/− p107−/− mice with Mx1-Cre Rbp7−/+ p130−/+ p107−/− mice. The Foxn1-IRES-LacZ transgene was described previously (Chen et al., 2009). These mice were bred through successive generations with Mx1-Cre Rbp7−/− p130−/− p107−/− to generate Rbp7−/− p130−/− p107−/− Foxn1−/−/+ and Rbp7−/− p130−/− p107−/− Foxn1−/−/+ with and without Mx1-Cre. As appropriate, mice were injected intraperitoneally with plp/C at 3–8 wk of age. Thymi were removed from euthanized mice and processed as appropriate for the particular experiment.

BM transplantation. Chimeric mice were generated by lethal irradiation (9 Gy) of 3-mo-old Mx1-Cre p107-Single and control mice expressing only the Ly5.1 antigen. Irradiated mice were rescued by retroorbital transplantation of 2 × 106 wild-type BM cells expressing both the Ly5.1 and Ly5.2 antigens. plp/C was injected 12 wk after transplantation. Tissues were harvested 12–16 wk later.

Immunofluorescence. Immunofluorescence was performed on frozen sections or sections from paraffin-embedded tissues as described previously (Chen et al., 2009; Gordon et al., 2010; Bryson et al., 2011; Park et al., 2011). Images were obtained by confocal microscopy using a confocal LSM 510 Meta (Carl Zeiss) with a Plan-Apochromat 20×/0.8 objective and analyzed with AxioVision 4.8 software (Carl Zeiss). Primary antibodies were directed against Foxn1 (goat anti-mouse, G-20; Santa Cruz Biotechnology, Inc.), K5 (rabbit anti–mouse; Covance), K8 (Trömml-1: Developmental Studies Hybridoma Bank), K14 (rabbit anti–mouse; Covance), B6 (rabbit anti–mouse; MBL International Corporation), CD205 (rat biotinylated; UEA-1–biotin (Vector Laboratories), CD31 (goat anti–mouse; BD), and PDGFR-β (goat anti–mouse; R&D Systems). Secondary antibodies included donkey anti–goat Cy5, donkey anti–rabbit 594, donkey anti–rat FITC, donkey anti–rabbit Texas Red, and streptavidin–FITC (Jackson ImmunoResearch Laboratories, Inc.) and donkey anti–goat Alexa Fluor 594 (Invitrogen).
Histology. Hematoxylin and eosin (H&E) staining was performed on paraffin-embedded sections. For H&E staining, thymi were fixed with 4% paraformaldehyde overnight, dehydrated in gradient ethanol solutions, and embedded in paraffin. 5–10-µm sections were cut and stained with H&E. Oil Red O staining was performed on frozen sections as follows: sections were fixed in 4% formalin for 10 min, washed with running water, rinsed in 60% isopropanol, stained for 15 min in fresh Oil Red O solution, washed in 60% isopropanol, stained briefly with hematoxylin, and washed with running water for 10 min. Microscopy was performed on a DM2000 microscope (Leica) at room temperature using a DFC500 camera (Leica). Images were analyzed using LASV3.8 software.

Cytometry. FACS analysis sorting was performed on an LSRII, Vantage, or FACs ARIA LSRII (BD). Data were acquired on CellQuest or FACSDiva (BD) and analyzed with FlowJo software (Tree Star). Antibodies for Kit, CD25, CD3e, CD4, CD8, MHCCII, CD40, CD205, CD80, CD45, Ly5.1, Ly5.2, Ly51, Mac1, Ter119, B220, Gr1, and Sc1 were purchased from eBioscience. FITC–conjugated UEA1 was purchased from Vector Laboratories. Cell cycle analysis was performed by intraperitoneal injection of BrdU, followed by euthanasia 1–4 h after injection. Cells were FACS sorted and analyzed for BrdU content as described previously (Viour et al., 2008). TECs were isolated by enzymatic digestion. Thymin were cut into ~1-mm3 pieces, and fragments were digested by a mixture of DNase I, and collagenase A/D: Dispase for 60 min at 37°C. Cells were passed through a 45-µm filter and resuspended for analysis in PBS/serum.

Plasmids and sequence analysis. Promoter sequence identification and alignment are as described on the University of California, Santa Cruz (UCSC) genome browser. The Foxn1 promoter (from −1550 to 64) was cloned in pGL3 basic. Point mutations were introduced by subcloning a 129-bp fragment of the 2–3 E2F binding sites. E2F3- and RB-expressing plasmids have been described previously (Angus et al., 2003; Burkhart and Sage, 2008). Primer sequences are available upon request.

ChIP. ChIP analysis in TEC100, TEC71, and 210R cell lines (a gift from A. Farr, University of Washington, Seattle, WA) and RT–quantitative PCR (RT-qPCR) were performed as described in Burkhart et al. (2010). Chromatin was sonicated using a fisher probe sonicator 30% output power for eight cycles of 30 s. The chromatin was preclared before being diluted and bound by 4 µg of the primary antibody overnight at 4°C. Each ChIP was then incubated with 8 µg rabbit anti-mouse IgG (MP Biomedical) or anti-mouse p16 antibody (N-20; Santa Cruz Biotechnology, Inc.) as a secondary for 1 h. Nuclearprotein complexes were pulled down using Pansorbin cells (EMD Millipore). Complexes were digested with Proteinase K and RNase A and purified by a QIAquick PCR Purification kit (QIAGEN). Analysis was performed using qPCR with primers directed against Foxn1, Cyclin A, and GAPDH promoters (primer sequence available upon request) using SYBR–green reagents (Applied Biosystems). PCR was run on a C1000 thermal cycler (Bio-Rad Laboratories).

Immunoblot analysis. Protein lysates were prepared by Triton X-100–based lysis from TECs pooled according to age and genotype from 5–12 animals each. Equal amounts of protein were loaded onto either manually prepared 10% Bis-Tris gels or Novex NuPAGE 4–12% gradient gels (Invitrogen) and run until appropriate separation was achieved. Gels were transferred to polyvinylidene difluoride membrane (GE HealthCare). Blots were probed with primary antibodies anti-Tubulin (T9026; Sigma-Aldrich); anti-Foxn1 H-270, anti-Foxn1 G-20, anti-RB C-15, and anti-CCND1 H-295 (Santa Cruz Biotechnology, Inc.); p300 anti-phospho-RB S807/S811 and S608 (Cell Signaling Technology); and anti-CDK4 (DCS-31; Invitrogen). HRP-conjugated secondary antibodies were purchased from Jackson Immuno-Research Laboratories, Inc. Signal was detected using either ECL Western blotting reagent (Thermo Fisher Scientific) or ECL–Prime Western blot detection reagent (GE Healthcare).

Cell culture, transfections, and luciferase analysis. TEC lines (TEC100, TEC71, and 210R) were grown in RPMI 1640 or DMEM supplemented with 10% serum, glutamine, penicillin, and streptomycin. Transfections were performed using either Lipofectamine 2000 (Invitrogen) or Eugene (Promega) and included experimental plasmids with TK-Renilla (Promega) and pcDNA plasmid to ensure transfection of equivalent amounts of DNA. Luciferase assays were performed at least three times in triplicates using the Dual Luciferase Assay System (Promega) and analyzed using the Hybrid Synergy Reader and Gen5V2.00 software (BioTek).

Statistical analysis. For analysis of two populations, statistical analyses were performed using a two-tailed paired Student’s t test. A p-value of <0.05 was required for significance (actual p-values are listed in figure legends). For analyses for three or more populations, significance was determined using ANOVA analysis.

Online supplemental material. Fig. S1 describes complex gating strategies used in this study. Online supplemental material is available at http://www.jem.org/cgi/content/full/jem.20121716/DC1.

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