Molecular Imaging of Inflammation in Atherosclerosis

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Molecular Imaging of Inflammation in Atherosclerosis

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Abstract

Acute rupture of vulnerable plaques frequently leads to myocardial infarction and stroke. Within the last decades, several cellular and molecular players have been identified that promote atherosclerotic lesion formation, maturation and plaque rupture. It is now widely recognized that inflammation of the vessel wall and distinct leukocyte subsets are involved throughout all phases of atherosclerotic lesion development. The mechanisms that render a stable plaque unstable and prone to rupture, however, remain unknown and the identification of the vulnerable plaque remains a major challenge in cardiovascular medicine. Imaging technologies used in the clinic offer minimal information about the underlying biology and potential risk for rupture. New imaging technologies are therefore being developed, and in the preclinical setting have enabled new and dynamic insights into the vessel wall for a better understanding of this complex disease. Molecular imaging has the potential to track biological processes, such as the activity of cellular and molecular biomarkers in vivo and over time. Similarly, novel imaging technologies specifically detect effects of therapies that aim to stabilize vulnerable plaques and silence vascular inflammation. Here we will review the potential of established and new molecular imaging technologies in the setting of atherosclerosis, and discuss the cumbersome steps required for translating molecular imaging approaches into the clinic.

Key words: Molecular imaging, Inflammation, Atherosclerosis

Inflammation in Atherosclerosis

Atherosclerotic cardiovascular disease is an increasingly common disease and contributes considerably to mortality and morbidity worldwide. Atherogenesis is driven by a combination of disturbed equilibrium of lipid accumulation and maladaptive immune responses. The disease entails chronic inflammation of the arterial wall and cross-talk with procoagulant pathways, culminating in plaque rupture and atherothrombosis. Atherosclerosis is characterized by arterial lesions that progress from an initial fatty streak towards an unstable (vulnerable) plaque in the arterial vessel wall. Early atherosclerotic lesion development is triggered by endothelial dysfunction and the local deposition of lipids (e.g. low density lipoprotein), particularly at sites of hemodynamic strain. In the intima, lipoproteins are prone to oxidative modifications and subsequently activate endothelial cells and intimal resident or infiltrated immune cells, causing a local inflammatory response that sustains leukocyte recruitment to the vessel wall. Monocytes that ingest excess lipids differentiate into macrophages and foam cells. The more advanced stable plaque consists of a thick fibrous cap with high collagen and smooth muscle cell content and a lipid core containing foam cells, debris and lipid droplets. The presence of an intact advanced plaque may lead to a
stenotic obstruction of the blood vessel (e.g. the coronary artery), a phenomenon which may clinically manifest as angina pectoris. Mechanisms that still largely remain unknown can render a stable plaque unstable and prone to rupture. Plaque rupture results in exposure of the plaque's prothrombotic core contents and leads to massive local blood coagulation and formation of a thrombus. Such thrombosis may lead to local and/or distal obstruction of blood vessels and gives rise to the major part of acute myocardial infarctions and stroke. Notably, plaque rupture is the most common type of plaque complication, accounting for ≈70% of fatal acute myocardial infarctions and/or sudden coronary deaths [1-4].

While animal models of disease have greatly advanced our understanding of the molecular mechanisms and cellular players underlying atherogenesis, atheroprogession and atherothrombosis as the pathogenetic sequence of CAD, the greatest challenge in cardiovascular medicine remains the identification of unstable or vulnerable (but often non-obstructive) arterial plaques that may be prone rupture. Although sensitivity, specificity, and overall predictive value of potential factors remain to be conclusively defined, criteria have been defined that make a plaque more likely to rupture. These include active inflammation, often defined as extensive macrophage accumulations, a thin fibrous cap with a large lipid core, superficial erosion and platelet aggregation or fibrin deposition, a fissured plaque cap and severe stenosis [1].

The development and refinement of non-invasive imaging therefore aims at providing reliable tools for the identification of preclinical disease and unstable lesions that reach beyond identification of flow-limiting stenosis. We here review molecular imaging modalities and discuss the cellular and molecular targets for imaging in the clinic.

**Modalities for non-invasive Molecular Imaging**

Imaging has become an indispensable tool both in cardiovascular research and clinical care within the last decades. Various imaging technologies are now available that each have their strengths and weaknesses (Table 1). Imaging in the clinical theatre is mostly restricted to depicting anatomy and quantifying the degree of vessel stenosis. However, more and more approaches aiming at the detection and characterization of vulnerable plaques are being translated into patient care [5, 6]. Molecular imaging of atherosclerosis is challenging as the vessels move rapidly with heart beat and respiration and most vessels of interest are in close proximity to tissue interfaces such as lung, blood or myocardium which can cause disturbing artifacts or strong background signal. ECG and respiratory triggering have facilitated data acquisition and lead to a significant improvement in image quality. When evaluating and comparing modalities for Molecular imaging both spatial resolution and temporal resolution are considered key properties of an imaging system. Spatial resolution describes a system’s ability to separate two closely spaced objects, or with respect to molecular imaging two closely spaced molecular probe concentrations. The higher the spatial resolution is, the higher the possibility to detect subtle molecular signals emitted from a cell or a molecular probe. Similarly, temporal resolution describes the ability to discriminate between two points in time. The temporal resolution is especially important when dynamic imaging is performed to track the kinetics of probe accumulation over time as well as when CINE imaging of moving/pulsating structures is performed. Sometimes there is a tradeoff between temporal and spatial resolution, and the temporal resolution theoretically achievable by a certain modality may be hampered by the special resolution and vice versa.

**Nuclear Imaging Techniques and Computed Tomography**

As nuclear imaging approaches are covered elsewhere in this issue only few important points relevant to multimodality approaches shall be mentioned here.

The most important disadvantage of both PET and SPECT is their limited spatial resolution. Small animal PET can resolve structures at ~1-2mm resolution, whereas clinical PET is limited to 4-5mm maximum spatial resolution. Such physical limitations make it unlikely that PET’s spatial resolution will improve significantly[5], however the limited spatial resolution may be compensated by hybrid imaging. PET-CT combines the excellent spatial resolution of CT with the high sensitivity in probe detection of the PET. A relevant concern using PET-CT is the radiation dose of ~10mSv a patient is exposed during imaging[7]. While CT offers only limited capabilities to differentiate various plaque components, recent integration of PET and MRI promises a significant advance for non-invasive characterization of vulnerable plaques. Combination of PET with MRI instead of CT significantly reduces radiation exposure due to replacement of the x-ray radiation, which is especially important for whole-body imaging. In comparison, average radiation exposure of coronary artery catheterization is ~7mSv, a SPECT based sestamibi stress test ~9mSv and coronary CT ~4-15mSv (depending on the technique used and the patient’s size). When comparing nuclear imaging techniques, cost and availability have to be taken into account. Short-lived
PET tracers require a cyclotron facility nearby the imaging site, while longer-lived SPECT tracers can be provided from outside sources. The minor costs of SPECT compared to PET have led to a broad availability in clinical medicine.

CT is able to detect and accurately quantify vessel calcification, and coronary calcium scoring has been used as a risk predictor for future cardiovascular events. However, three-quarters of all coronary lesions are non-calcified plaques and high-risk vulnerable plaques prone to rupture usually do not contain significant calcifications [8-11]. With the help of iodinated contrast agents such non-calcified plaques can be detected. Improvements in multislice detector technology now enable the visualization of coronary anatomy in addition to large vessels with rapid (seconds) acquisition and minimal motion artifact. Similarly, novel reconstruction algorithms enable a significant dose reduction, making coronary CT attractive for wide applications. Moreover, detection of plaque inflammation using CT has been achieved by using N1177, an iodine containing agent taken up by macrophages [12]. Gold nanorods may become an attractive alternative as they are able to yield good contrast on CT imaging and can be coupled to gadolinium (Gd) compounds, NIR fluorochromes and nuclear tracers for multimodal imaging and be additionally used as drug carriers for various theranostic applications [13-15].

Table 1: Non-invasive Modalities for Molecular Imaging of Atherosclerosis

<table>
<thead>
<tr>
<th>Technique</th>
<th>Spacial Resolution</th>
<th>Depth</th>
<th>Acquis-</th>
<th>Quantitative</th>
<th>Imaging Agents</th>
<th>Molecular Targets</th>
<th>Clinical application</th>
<th>Specific Features</th>
</tr>
</thead>
<tbody>
<tr>
<td>MRI</td>
<td>10-100μm</td>
<td>No limit</td>
<td>min-h</td>
<td>Yes</td>
<td>Gd-Chelates, superparamagnetic nanoparticles (SPION, USPIO, VSOP)</td>
<td>Adhesion Molecules, Macrophages, pro-inflammatory enzymes (myeloperoxidase), Lipoproteins, Apoptosis/Necrosis, Integrins, Fibrin, Ca2+hydroxyapatite</td>
<td>Quantification of stenosis, Plaque morphology, Flow measurements</td>
<td>+ No radiation + high-soft-tissue contrast</td>
</tr>
<tr>
<td>CT</td>
<td>50μm</td>
<td>No limit</td>
<td>sec-min</td>
<td>Yes</td>
<td>Iodinated molecules</td>
<td>Plaque morphology, Coronary Plaque burden, Intima-Media Thickness, Flow velocities</td>
<td>- Radiation - high spatial and temporal resolution</td>
<td></td>
</tr>
<tr>
<td>Ultrasound</td>
<td>50μm</td>
<td>cm</td>
<td>sec-min</td>
<td>Yes</td>
<td>Microbubbles</td>
<td>Adhesion molecules, Integrins, Gycloproteins</td>
<td>- Operator-dependent + high temporal resolution</td>
<td></td>
</tr>
<tr>
<td>PET</td>
<td>~ 2mm</td>
<td>No limit</td>
<td>min-h</td>
<td>Yes*</td>
<td>111In, 123/124/125I, 58Fe, 64Cu Tracers</td>
<td>FDG, Adhesion Molecules, Integrins, Fibrin, Ca2+hydroxyapatite</td>
<td>Plaque inflammation</td>
<td>- Radiation - Cyclotron required - limited spatial resolution - stand alone PET not quantitative (overcome by hybrid PET/CT)</td>
</tr>
<tr>
<td>SPECT</td>
<td>~ 2mm</td>
<td>No limit</td>
<td>min-h</td>
<td>Yes</td>
<td>99mTc, 111In Tracers</td>
<td>Adhesion molecules, Lipoproteins, Macrophages, Proteases, Glycoproteins, Apoptosis</td>
<td>Myocardial Perfusion Imaging</td>
<td>- Radiation - limited spatial resolution - full quantification only by hybrid SPECT/CT - lower costs and higher availability compared to PET</td>
</tr>
<tr>
<td>Bioluminescence Imaging</td>
<td>2-5mm</td>
<td>Few cm</td>
<td>min</td>
<td>No</td>
<td>Luciferins</td>
<td>Gene expression, Cell tracking</td>
<td>No</td>
<td>- Experimental only - limited spatial resolution</td>
</tr>
<tr>
<td>Fluorescence Molecular Tomography</td>
<td>1mm</td>
<td>Few cm</td>
<td>min</td>
<td>Yes*</td>
<td>NIR fluorochromes</td>
<td>Adhesion molecules, Monocyte/Macrophages, Proteases, Collagen, Ca2+hydroxyapatite, Thrombosis</td>
<td>Not yet - semi-quantitative only (improved by hybrid SPECT/CT) - experimental only (Clinical applications under development) - interference with hemoglobin - experimental only</td>
<td></td>
</tr>
<tr>
<td>Optoacoustic Imaging</td>
<td>&lt;50μm</td>
<td>Few cm</td>
<td>min-h</td>
<td>Yes</td>
<td>NIR Fluorochromes</td>
<td>Catheter applications (see Table 2)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*when hybridized with CT/MRI for proper attenuation correction
**Magnetic Resonance Imaging**

MRI provides high spatial and temporal resolution and has the advantage of combining anatomic, functional and molecular imaging in an all-in-one approach. Its high soft tissue contrast enables to distinguish the major components of atherosclerotic plaques and allows risk stratifications based on plaque composition irrespective of the degree of stenosis [16, 17]. Contrast enhanced MRI enables highly sensitive molecular imaging of vulnerable plaques. Paramagnetic gadolinium based agents provide enhancement contrast by shortening the spin-lattice relaxation time (T1). Superparamagnetic iron-oxide nanoparticles, which target the phagocytic capacity of plaque-infiltrating leukocytes, consist of an iron-oxide core and a surrounding shell made of dextran, synthetic polymers or polyvinyl alcohol. Depending on their size these nanoparticles are classified as SPIO (small superparamagnetic iron oxide, >60nm), USPIO (ultra-small superparamagnetic iron oxide, <60nm) and VSOP (very small superparamagnetic iron oxide, <10nm) particles. The size of the particles affects their extravasation and biodistribution, as well as the mode of elimination from the body. Upon contact, these particles are rapidly taken up by phagocytic cells in vitro and in vivo. Superparamagnetic iron-oxide particles influence signal intensity mainly by shortening T2* and T2, which at a given echo time produces darkening of the contrast-enhanced tissue. Conversely, they can act as positive agents (T1 shortening and image brightening) when appropriate imaging sequences are used [18]. The USPIO/SPIO derived MRI signal correlates highly with macrophage presence in murine, rabbit and human atheroma [19-23]. The dextran or alcohol shell of the nanoparticles can be used to conjugate antibodies or other peptides for targeted imaging. In contrast, conjugation of high-affinity ligands to Gd-based paramagnetic agents is more difficult. A new and elegant tool for targeting mononuclear phagocytes is to pack micelles or liposomes with Gd agents, thereby reducing toxicity [24, 25]. A more sophisticated approach is the synthesis of activatable MR contrast agents that lead to signal amplification upon specific binding[26, 27].

**Ultrasound**

Molecular imaging using ultrasound-targeted microbubbles to enhance ultrasound contrast may potentially be translated to imaging in patients [28]. A significant obstacle is the strong background signal that decreases detection sensitivity especially in deep vessels such as the coronaries. Yet, targeted microbubbles may not only be used for specific molecular imaging of inflammation and the detection of adhesion molecules, selectins or von-Willebrand Factor [29-32], they can additionally be used as drug carriers for theranostic applications[33]. A detailed overview of molecular imaging of cardiovascular disorders with ultrasound is provided in reference [34].

**Fluorescence imaging**

Fluorescence techniques have significantly enhanced our understanding of cardiovascular disease. Their high sensitivity, cost-effectiveness and the large portfolio of targeted agents have led to wide applications in preclinical research investigating pathophysiology at the microscopic, mesoscopic and macroscopic scale [26, 27, 35-39]. Fluorescence Molecular Tomography (FMT) technology permits a highly sensitive 3-D detection of enzyme activities [39-41], tissue calcification[42, 43] and integrin expression[44]. The use of fluorescent probes emitting light in the near-infrared (NIR) spectrum allows the detection of molecular signals in living animals in depths of several centimeters [39, 45]. Also, imaging in the NIR region minimizes tissue autofluorescence, thereby improving target to noise ratio.

Near-infrared fluorescence (NIRF) sensors can be constructed in three different ways: 1) Non-specific fluorescent blood-pool agents injected in the circulation of the animal that inform on perfusion or permeability of the tissue, 2) targeted probes that bind via specific ligands to protein structures on the cell surface or 3) ‘smart probes’, that are activated in situ by enzymatic conversion or enzymatic cleavage [35, 46]. These probes have been used to detect enzyme activity in live mice by FMT and with cellular resolution by fluorescence microscopy [43, 47, 48]. Currently, FMT allows the non-invasive detection of fluorochrome concentrations in the pico-molar range in deep tissues with a submillimeter resolution in vivo.

Clinical translation of optical imaging is limited by the restricted tissue penetration of NIR light. While non-invasive imaging may be applicable to superficial carotid arteries, deeper vessels would require more invasive assessment. Recent advances in miniaturization of fluorescence light source and detector technology have led to first catheter-based applications.

One challenge of Fluorescence Imaging is the proper signal quantification. As the fluorescent light emitted from the molecular target experiences diffuse scattering and attenuation on its way through complex tissues, full signal quantification is obtained by methods of attenuation correction using similarly acquired CT data [49]. These additional CT data sets can be obtained sequentially after FMT by using multimodal animal holders with fiducial markers [41]. However, the first prototypes of full hybrid FMT-XCT (Fluorescence Molecular Tomography- X-ray Computed Tomography) systems have shown improved...
performance by using a dual prior inversion method [50-52].

One advantage of fluorescence molecular imaging is the possibility to subsequently perform traditional ex vivo laboratory techniques. After in vivo fluorescence imaging the animal can be sacrificed and the biodistribution of the molecular sensor can be detected and quantified by cryoslicing[53]. Similarly, fluorescence microscopy can be performed on tissue sections after in vivo biodistribution of the injected probe; the sections can then be further evaluated ex vivo with classical tools such as immunofluorescence, immunohistochemistry, and in situ hybridization. Multicolor flow cytometry is an elegant tool to determine the cellular source of a fluorescence imaging signal obtained non-invasively in vivo [40, 47].

**Photoacoustic imaging**

Photoacoustic imaging is based on the photoacoustic effect, where ultra-short pulses of light that are absorbed in tissue create broadband ultrasound waves, which can be detected non-invasively outside the body. Because ultrasound scatters orders of magnitude less compared to fluorescence light in tissue it can provide optical images with a spatial resolution of ~50µm. By using multiple excitation wavelengths and tomographic signal detection, MultispectralOptoacoustic Imaging (MSOT) can provide images of specific chromophores including organic dyes as well as nanoparticles based on their unique spectra[37, 54]. First preclinical applications of MSOT in cardiovascular research have achieved successful anatomic visualization of the heart and major blood vessels as well as high-resolution imaging of inflammation in a model of acute myocardial infarction. First ex vivo studies with specimens from patients with symptomatic carotid artery disease have demonstrated that MSOT is capable of rendering detailed biodistribution of MMP activity together with high-resolution images of vulnerable plaques [55].

**Intravascular Molecular Imaging**

Intravascular catheter-based imaging techniques have contributed significantly to the understanding of human atherosclerosis. An overview of the specific features of existing techniques and their advantages or disadvantages is provided in Table 2. Intravascular Ultrasound (IVUS) is able to provide high-resolution images of the vessel wall and plaque morphology at an axial resolution of ~100µm using high-frequency detectors (up to 45 MHz). Spectral analysis of the IVUS backscatter radiofrequency signal enables color-coded images of certain plaque characteristics, referred to as virtual histology (IVUS-VH) [56]. With high spatial resolution this technique is able to differentiate lipid-rich areas, the necrotic core, fibrous tissue and calcifications on the basis of different echolucent structures [57]. With increased spatial resolution of 10-15µm optical coherence tomography (OCT) has facilitated the characterization of neointimal hyperplasia following percutaneous coronary intervention (PCI), and has allowed for depiction of the internal and external elastic lamina as well as a thin fibrous cap (<65µm)[58-60]. The drawback of OCT is the inability to image in the blood stream because the small wave lengths reflect very small objects such as blood cells. Further improvement of conventional time-domain OCT has been achieved by introducing frequency domain OCT (also referred to as OFDI, Optical frequency Domain Imaging), where a Fourier transformation of the acquired spectra enables to image at much higher frame rates of >100 frames/s. This allows fast 3D image acquisition of long vessel segments under a single non-occlusive saline flush without occluding the vessel [61].

Intravascular radiation detector systems have been tested for endovascular sensing of 18F-FDG in atherosclerotic lesions. Various systems were able to detect 18F point sources in the canine femoral as well as in the coronary arteries using an open chest model [62]. Using a model of femoral artery denudation in hypercholesterolemic rabbit’s intravascular 18F detection in injured artery segments correlated with increased macrophage content [63]. Miniaturized catheters systems were even able to detect increased 18F accumulation in murine aortas of apoE−/− mice [64]. The catheter based 18F detection may be superior to regular PET/SPECT as spatial mapping of the probe distribution within the lesion may be improved.

Intravascular NIRF Imaging is an attractive approach for human coronary artery imaging and has the potential to identify vulnerable plaques upon the high macrophage load as well as increased protease activity. A first study in hypercholesterolemic rabbits has demonstrated that this technique can detect the fluorescent signal emitted from a previously injected protease sensor activated in plaques within the iliac artery [65]. Further improvement of the technique has led to engineering of new catheter capable of 2D fluorescence imaging in a 360° view during automated pullback. This improvement enabled a submillimeter axial resolution, nanomolar sensitivity to the injected NIR probe and only moderate light attenuation by the circulating blood [66]. The whole system was based on a 2.9F angiography catheter and thus potentially applicable to human coronary arteries. In a recent report intravascular NIRF imaging has been combined with OFDI technology for molecular and microstructural imaging of atherosclerosis [67]. Intravascular MSOT approaches similarly aim to provide
high-resolution anatomical images of the atherosclerotic plaque and to report on the biological activity by using targeted molecular probes [68]. The successful preclinical application of these intravascular molecular imaging approaches in various animal models will pave the route for first human trials with the aim to detect vulnerable plaques prior to percutaneous coronary intervention via a transarterial approach.

Table 2: Invasive Modalities for Molecular Imaging of Atherosclerosis

<table>
<thead>
<tr>
<th>Technique</th>
<th>Spacial Resolution</th>
<th>Depth</th>
<th>Acquisition Time</th>
<th>Quantitative Imaging</th>
<th>Molecular Targets</th>
<th>Clinical application</th>
<th>Specific Features</th>
</tr>
</thead>
<tbody>
<tr>
<td>OCT</td>
<td>~10 µm</td>
<td>2-3mm</td>
<td>min</td>
<td>Yes</td>
<td>-</td>
<td>Plaque and thrombus characterization</td>
<td>- Blood-free field required</td>
</tr>
<tr>
<td>OFDI</td>
<td>~10 µm</td>
<td>2-3mm</td>
<td>min</td>
<td>Yes</td>
<td>-</td>
<td>Plaque and thrombus characterization</td>
<td>+ Faster image acquisition compared to OCT</td>
</tr>
<tr>
<td>IVUS-VH</td>
<td>~100µm</td>
<td>10mm</td>
<td>min</td>
<td>Yes</td>
<td>-</td>
<td>Plaque characterization</td>
<td>- Interference with blood at frequencies&gt;40MHz</td>
</tr>
<tr>
<td>Angioscopy</td>
<td>10-50µm</td>
<td>Surface Imaging</td>
<td>Min</td>
<td>No</td>
<td>-</td>
<td>Imaging of plaque surface, thrombus characterisation</td>
<td>- Blood-free field required</td>
</tr>
<tr>
<td>Intravascular Fluorescence Imaging</td>
<td>1mm</td>
<td>Few cm</td>
<td>min</td>
<td>Yes</td>
<td>NIR Fluorochromes</td>
<td>Macrophages, Proteases, Apoptosis</td>
<td>+ Imaging of arterial inflammation</td>
</tr>
<tr>
<td>Intravascular Optoacoustic Imaging</td>
<td>&lt;50µm</td>
<td>Few cm</td>
<td>min</td>
<td>Yes</td>
<td>NIR Fluorochromes</td>
<td>targets comparable to FMT</td>
<td>- Interference with hemoglobin</td>
</tr>
</tbody>
</table>

(+ ) advantage, (-) disadvantage
OCT=Optical Coherence Tomography, IVUS-VH=Intravascular Ultrasound-Virtual Histology, OFDI=Optical Frequency Domain Imaging

Cellular and Molecular Targets within the inflamed vessel

Initiating steps of inflammation: shear stress and lipid accumulation

A schematic overview of plaque formation and its consequences are shown in Figure 1. Atherosclerotic lesions develop in particular predisposed regions of the vasculature. While flow with physiological levels of laminar shear stress in straight segments is regarded as atheroprotective, areas experiencing low and especially oscillatory shear stress or turbulent blood flow such as inner curvatures as well as vessel bifurcations are prone to develop atheroma [69]. Oscillatory shear stress in particular leads to increased activation of the endothelium [70], and it is therefore not surprising that these regions are characterized by a higher inflammatory load and an increase of reactive oxygen species, and accumulate lipids within the intima [71-73]. Simulations based on computational mechanics will help to better understand the effects of wall shear stress on the arterial wall and the associated cellular and molecular adaptations to altered mechanical forces. High-resolution MR Imaging can aid in identifying atherosclerosis-prone areas with altered shear stress [74].

Paving the route for immune cell influx: Endothelial activation and permeability

Endothelial activation one of the initiating steps of vessel inflammation and atherosclerotic lesion development. Upon activation endothelial cells express various adhesion molecules triggering leukocyte recruitment across the blood-vessel wall [75]. Selectins (e-selectin, p-selectin and l-selectin), adhesion molecules such as intercellular adhesion molecule (ICAM)-1 and vascular cell adhesion molecule (VCAM)-1, and chemokines control leukocyte rolling, adhesion and migration into the vessel wall. The adhesion molecule is expressed on activated endothelial cells, smooth muscle cells but also macrophages, and recruits leukocytes via binding of the integrin very late antigen (VLA)-4. Thus, imaging approaches targeting VCAM-1 use VLA-4 peptides, anti-VCAM-1 antibodies and peptides of the MHC-I molecule interacting with VCAM-1. These targeting moieties can be conjugated to various nuclear tracers (123I, 99mTc, 18F) for PET and SPECT imaging, NIR fluorochromes for intravital microscopy and non-invasive tomo-
graphic optical imaging, iron-oxide based nanoparticles (SPIO) for MRI or microbubbles for CEU [76-84]. These agents have been reported to successfully detect vascular inflammation and atherosclerotic lesions and to correlate with intraplaque VCAM-1 mRNA levels. However, VCAM-1-targeted probes do not exclusively detect activated endothelium, as also activated macrophages express VCAM-1 and other integrins [30, 80, 85]. To enhance uptake and probe accumulation, targeting multiple moieties represents an interesting and promising approach. It has been shown that combined VCAM-1 and selectin targeting leads to > 5-fold binding to activated endothelium compared to single-targeted probes [29, 86, 87]. ICAM-1 targeted microbubbles as well as a liposomal Gd-based MRI agent are similarly able to visualize activated endothelium and inflammatory atherosclerotic lesions in vivo [88, 89]. Selectin-target probes have similarly been engineered for targeting inflammation. A dendritic polyglycerol sulfate based fluorescent probe has been applied to target inflammation within the injured myocardium [90]. The inhibitory properties of this agent impair leukocyte recruitment to sites of inflammation, rendering an agent for both imaging and treatment of atherosclerosis [91].

Fig 1. Tools and targets for molecular imaging of atherosclerosis. Figure demonstrates schematic evolution of atherosclerotic plaques and potential targets for molecular imaging.
Concomitant to the expression of adhesion molecules, endothelial dysfunction and increased endothelial permeability are considered as initiating events in plaque formation. Progressive endothelial leakage permits low-density lipoproteins to diffuse across the endothelial cell lining and accumulate in the arterial intima. Gadofosveset is a clinically approved albumin-binding contrast agent. Binding of albumin leads to a prolonged half-life within the blood pool and a dramatically higher r1-relaxivity. The uptake of gadofosveset has been linked to mechanically damaged endothelium in a porcine model of coronary injury [92] as well as with atherosclerosis-induced endothelial damage in apoE−/− mice [93]. Figure 2 demonstrates gadofosveset accumulation in atherosclerotic plaques with underlying endothelial damage indicated by a bright contrast in T1-weighted MR images. Similarly, amphiphilic NIR fluorochromes can be used for optical imaging of vulnerable plaques. Indocyanine green (ICG), a clinical approved NIRF dye, also binds to albumin with high affinity and accumulates in atherosclerotic plaques after in vivo injection in hypercholesterolemic rabbits as well as after incubation of human carotid artery specimens[94].

Monocyte accumulation, phagocytosis and metabolism

During atherogenesis, monocytes accumulate in the arterial intima and differentiate into macrophages, which then ingest oxidized lipoproteins, secrete a diverse array of proinflammatory mediators, and mature towards foam cells [95, 96]. Monocyte heterogeneity has been conserved in various species [97]. At least two different subpopulations exist with divergent properties. In hypercholesterolemic mice, Ly6C hi (Gr-1+) monocytes preferentially accumulate in lesions [98]. Proinflammatory Ly6C hi monocytes dominate hypercholesterolemia-associated monocytes and give rise to macrophages in atheroma [99]. The role of Ly6C lo monocytes is less clear in atherosclerosis, but it appears that these cells exhibit an anti-inflammatory phenotype and play a major role in extracellular matrix formation and tissue remodeling. Also in humans, two major subpopulations have been described. CD14 hi/CD16 lo monocytes (corresponding to Ly6C hi monocytes in mice) display elevated serum levels in patients with coronary artery disease [100, 101] whereas levels of CD14 hi/CD16 lo monocytes (corresponding to Ly6C hi monocytes in mice) are predictive for myocardial salvage after myocardial infarction [102]. Targeting monocytes has been performed using various formulations of superparamagnetic nanoparticles.. An example of targeting macrophages in murine atheroma using VSOPs is shown in Figure 3. Interestingly, under anti-inflammatory therapy with atorvastatin leading to a decrease in macrophage activity, uptake of USPIOs was significantly reduced [103]. While murine Ly6C hi and Ly6C lo monocytes exhibit equal phagocytic capacity of cross-linked iron-oxide nanoparticles[104], human CD14 hi/CD16 lo monocytes display a higher phagocytic capacity in vitro compared to their CD14 hi/CD16 hi counterparts[101, 105]. This phagocytic phenotype may therefore be well suited for specific targeting of monocyte subsets in patients with atherosclerosis. Imaging of iron-oxide loaded inflammatory macrophages has been successfully performed in animals and humans and constitutes a promising approach for identifying vulnerable plaques[106-108]. While most studies report on the direct injection of the nanoparticles into the circulation, ex vivo labeling with subsequent reinjection and recruitment of labeled phagocytes to sites of inflammation has also been utilized [109]. The latter approach may enhance target-to-background signal by eliminating unspecific probe accumulation, yet it has to be considered that ex

Fig 2. MR Imaging of endothelial permeability. Uptake of gadofosveset in regions of the brachiocephalic artery of control (Panel A-E) and atherosclerotic mice after 4 (Panel F-J) and 12 weeks (Panel K-O) of high-fat diet is associated with endothelial permeability. After 12 weeks significant increase in R1 relaxivity is observed in the inflamed vessel wall following gadofosveset injection (Panel P). Image courtesy of Alkystis Phinikaridou and René M. Bonnar, King’s College London.
**vivo** labeling with iron-oxides affects the phenotype of the labeled cells and potentially their function [105]. *Ex vivo* labeling with $^{111}$In-oxine was similarly successful at visualizing macrophage accumulation in murine atherosclerotic lesions [110]. Also nanoparticles can be conjugated to radionuclides as well as NIR fluorescent dyes for hybrid nuclear and optical imaging in conjunction to MRI [47, 81]. Beside targeting the phagocytic capacity of monocytes/macrophages, receptor-based imaging of phagocytes has been reported by using Gd-loaded micelles targeting the macrophage scavenger receptor, the cannabinoid receptor and the neutrophil gelatinase-associated lipocalin 2 (NGAL) in murine plaques [24, 111-113] as well as targeting the MCSF-receptor on human monocyte subsets [101]. Furthermore, activated macrophages in atherosclerotic lesion express various integrins such as $\alpha_\text{v}\beta_3$. Targeting $\alpha_\text{v}\beta_3$ using RGD based nuclear tracers [114, 115] as well as MR contrast agents [116] allows identification of activated macrophages in atherosclerotic plaques in various animal models.

Aside from targeting immune cells via their phagocytic capacity or receptor profile, imaging of metabolic activity of the cellular infiltrate has been applied to assess inflammation in atherosclerotic plaques by PET/CT [117-120]. The glucose analogue $^{18}$F-FDG is taken up by metabolically active cells, especially by macrophages and foam cells, and thus reports on inflammatory activity in atherosclerotic lesions (Figure 4). First clinical trials of $^{18}$F-FDG PET/CT showed strong association between $^{18}$F-FDG uptake in carotid artery lesions with type 2 diabetes [121] and more importantly with the occurrence of ischemic stroke [122, 123]. Exciting are the results of the dal-PLAQUE study, one of the first multicenter clinical trials employing non-invasive multimodality imaging, MRI and PET-CT, to assess structural and inflammatory indices as primary endpoints of the effects of dalcetrapib on carotid disease [124]. In patients, the $^{18}$F-FDG signal correlates closely with the expression of biomarkers such as GLUT-1, HK2, cathepsin K and CD68 in carotid artery lesions [125]. Interestingly, in a similar approach, the investigators were not able to find significant correlations between $^{18}$F-FDG uptake and CD68 levels in patients with peripheral arterial occlusive disease [126]. A recent *ex vivo* study investigated the in vitro uptake of FDG in several cell types involved in ath-eroma formation, revealing that predominantly hy- poxia but not inflammatory cytokines stimulates cells to accumulate FDG. Therefore FDG uptake signals in atheroma may reflect hypoxia-stimulated macrophages rather than the mere inflammatory burden [127]. Yet these results await further evaluation *in vivo*.

Novel hybrid imaging technologies will be helpful in correlating specific tracer accumulation within the vessel wall/atherosclerotic lesions and lesion morphology when imaged simultaneously by CT or MRI. Figure 5 shows successful $^{18}$F-FDG imaging of plaque inflammation in a hypercholesterolemic rabbit by full hybrid MR-PET (Panel A-C) as well as vessel wall inflammation in a patient with large-vessel vasculitis by both PET-CT and MR-PET (Panel D-L). PET-MR may in particular be promising for detection of vulnerable plaques as it combines the molecular information reported by various tracers from the PET with high-resolution and functional imaging by MRI.

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**Fig 3. MR Imaging of vascular inflammation using very small superparamagnetic nanoparticles (VSOP).** VSOPs target inflammatory macrophages in high-fat diet induced atherosclerosis in mice, inducing shortening of $T_2^*$ relaxation in the vessel wall in HFD fed mice (lower row) as compared to controls (upper row). Imaging findings are corroborated by histology (A4-A6 and B4-B6). EvG=Elastica van Gieson, HFD=high-fat diet, SGM=susceptibility gradient mapping. TOF=Time-of-Flight angiography. Image courtesy of René M. Botnar, King’s College London.
Fig 4. Imaging of the vulnerable plaques in human coronary atherosclerosis. Representative images of 18F-FDG PET (A), CT (B), PET/CT (C), and coronary angiography (D) from patient with good suppression with coronary 18F-FDG uptake (arrows). Reprinted with the permission of the Society of Nuclear Medicine from Wykrzykowska et al. [183]

Fig 5. Molecular Imaging of atherosclerosis by hybrid PET-CT and MR-PET. Inflammation in plaques of hypercholesterolemic rabbits can be assessed and quantified by 18F-FDG PET and co-localized to carotid artery by simultaneously acquired MRI. Panel A: TOF angiography, Panel B, contrast enhanced fat-suppressed T1 weighted MRI (delayed enhancement), Panel C: MR-PET fusion showing increase tracer accumulation around the left carotid artery. Images demonstrate good correlation of PET signal and contrast-enhanced MRI but also show limited spatial resolution of PET technology. Hybrid Molecular Imaging in a patient with large-vessel vasculitis (Panels D-L). Increased 18F-FDG uptake can be visualized by whole-body PET and correctly co-localized to the aortic arch by the subsequently performed contrasted enhanced CT (Panels D-G: PET-CT). Similar co-localization can be performed using hybrid MR-PET (Panels H-J). Whole body MRA (Panel K) and CTA (Panel L) can be routinely performed during hybrid image acquisition. Images courtesy of Isabel Dregely, Stefan Nekolla and Ambros J. Beer from the Munich PET/MR consortium of TUM and LMU (funded by DFG).
Lipids and Oxidative Stress

Oxidative stress promotes lipid oxidation and cell death and is one of the major triggers of lesion progression and destabilization. Oxidized lipids have been targeted in preclinical atherosclerosis models using MDA (malondialdehyde-lysine) MR contrast agents [128]. Oxidative stress is predominantly mediated through reactive oxygen species (ROS). In atherosclerotic plaques, ROS are produced by macrophages, neutrophils and smooth muscle cells. A signature enzyme of inflammation and ROS production is myeloperoxidase (MPO). Advanced human atheroma as well as ruptured plaques contains increased numbers of MPO expressing macrophages [129]. Targeting of MPO may therefore be useful for the identification and characterization of vulnerable plaques. A prototype of an amplifiable MRI contrast agent has been reported by Chen et al [26, 27]. This Gd-MPO [bis-5HT-DTPA (Gd)] increases its relaxivity upon contact with MPO; subsequent polymerization and protein binding leads to signal amplification in situ compared to its native state. This agent has been applied for molecular imaging of various inflammatory conditions [130, 131] and for targeting inflammation in a rabbit model of atherosclerosis (Figure 6)[132]. Similarly, MPO-sensing in cardiovascular diseases can be achieved by fluorescent nanoparticles detecting hypochlorous acid production in vivo [133].

Consequences of infiltrating immune cells: proteolysis and matrix degradation

In concert with the phagocytic activity of other infiltrating immune cells, macrophages in particular secrete proteolytic enzymes which can degrade extracellular matrix proteins promoting thinning and rupture of the fibrous cap [134]. Among the various existing proteases cathepsins and MMPs have most frequently been used for molecular imaging, and targeted by two major strategies. Nuclear tracers based on small molecule protease inhibitors bind to the active site of the enzyme of interest and enrich at sites with a high protease concentration. These MMP inhibitors (MMPI) can be coupled to various radiotracers such as $^{123}$I, $^{111}$In, $^{99m}$Tc or $^{18}$F for PET and SPECT imaging. SPECT imaging signal of MMPI has been

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**Fig 6. Myeloperoxidase (MPO) – targeted MRI of vascular inflammation.** MPO-Gd MR imaging of atherosclerosis in a rabbit model fed high cholesterol diet for 24 months. MPO-Gd imaging identifies areas of high MPO activity and content (red circles) that are corroborated by MPO immunostaining. Images courtesy of John W. Chen, Massachusetts General Hospital, Harvard Medical School.
shown to correlate to MMP-2/-9 activity in atherosclerotic plaques [135, 136].

Alternatively, optical reporters are being used that are activated upon contact with the target enzyme [137]. The sensor is injected in its inactive state in which fluorochromes are not excitable due to auto-quenching. Proteolytic cleavage of the scaffold releases the fluorochromes and results in extensive fluorescence generation (de-quenching). Amplification is achieved because one active enzyme moiety can activate multiple reporters [39]. Both FMT and Optoacoustic Tomography are able to resolve this process non-invasively in small animals as well as tissue specimens. In a pilot study an activatable NIRF sensor, activated by gelatinases A (MMP2) and B (MMP9), was able to detect MMP activity in aortas of apoE-/- deficient mice on high-cholesterol diet. Both ex vivo FRI and in vivo FMT yielded high MMP activity compared to injected apoE-/- mice and animals not injected with the probe [138]. Similar to the detection of MMP activity, cathepsin can be detected in inflammatory lesions. Using pan-cathepsin NIRF sensors, high protease activity was detected in atherosclerotic lesions of apoE-/- as well as double knock out apoE-/- eNOS-/- mice [139]. Strong co-localization of cathepsin B within the lesion pointed towards mononuclear phagocytes as the main source of secreted proteases. Using a combined FMT-CT imaging approach and a variety of nanosensors tracing cathepsin activity, the study localized the molecular fluorescence signal to the aortic root of mutant mice [41]. The combination of FMT and CT further allowed proper quantification of the pan-cathepsin signal and thus allowed to appropriately monitor atorvastatin therapy. The anti-inflammatory effect of statins in atherosclerosis was similarly reported using a cathepsin B activatable NIRF sensor [140]. While most of the protease sensors discussed report pan-MMP or pan-cathepsin activity, more selective probes become available. A cathepsin K selective optical reporter demonstrated high protease activity by intravital microscopy as well as ex vivo FRI [141]. Activity of cathepsin S was associated with vascular calcification and co-localized with an osteogenesis targeted imaging agent in mice with chronic renal disease [142]. With respect to monocyte heterogeneity, as discussed above, these sensors can distinguish increased protease contents in proinflammatory Ly6C+ monocytes as compared to lower activity in Ly6C- monocyte subsets [104].

The only MRI-based approach targeting proteases has been performed using P947. This gadolinium chelate is coupled to a peptide, which binds MMPs [143, 144]. These pilot results are encouraging; and suggest that a non-invasive imaging approach may be clinically applicable at identifying vulnerable plaques.

The dusk of inflammation: Apoptosis

The three major characteristics of vulnerable plaques are a high degree of inflammation, thinning of the protective fibrous cap and a lipid rich ‘necrotic’ core. Lipid-laden foam cells are prone to undergo apoptosis [145]. Cell debris further promotes inflammation, secretion of proteases, and thus progressive destabilization of the plaque, ultimately resulting in rupture and thrombosis. During apoptosis, activated flippases rapidly externalize phosphatidylserine (PS) chains from the inner membrane of the lipid double layer to the outer layer of the membrane. The externalized PS are recognized by the 35kD plasma protein Annexin V (A5) [146]. A5 has been used for various molecular imaging approaches both in preclinical atherosclerosis models as well as in human cardiovascular disease. It can be coupled to nuclear tracers, NIRF fluorochromes or iron-oxide nanoparticles for MR imaging. For plaque imaging A5 labeled with 99mTc proved most suitable both in experimental as well as in first human trials [147]. In atherosclerotic mice, rabbits and swine 99mTc labeled A5 showed focal tracer accumulation in inflammatory plaques in vivo, which correlated with immunohistochemistry of cell death and high macrophage load [148-150]. A pilot report of four patients with symptomatic carotid artery disease revealed high tracer accumulation at the affected carotid bifurcation, and post-endarterectomy histology evaluation confirmed high macrophage load as well as intraplaque hemorrhage [151]. Similarly, SPECT imaging of apoptosis can detect therapeutic effects of anti-inflammatory as well anti-apoptotic regimens in hypercholesterolemic rabbits [152].

An A5 conjugated micellar nanoparticle carrying multiple Gd-labeled lipids for MRI as well as fluorescent lipids for optical imaging was able report on apoptosis and macrophage accumulation in atherosclerotic aortas of apoE-/- mice [153]. Coupling of Annexin V to the dextran shell of superparamagnetic iron-oxide nanoparticles enabled apoptosis imaging by T2* weighted nanoparticles in various models of cardiovascular disease [154, 155].

Yet, PS externalization is not specific to apoptosis but also occurs in activated macrophages and stressed cells in general [146, 149]. This has to be taken into account when evaluating anti-apoptotic therapies by A5-based molecular imaging [152]. Nevertheless, cell stress and macrophage activation are likewise hallmarks of vulnerable plaques, so that cumulative targeting of these aspects of vascular inflammation may in fact enhance probe accumulation and increase contrast in biomedical imaging. Oligo-targeted mo-
molecular imaging that visualizes apoptosis and protease activity may thus enhance in vivo imaging signals and study interlinked processes of inflammation in cardiovascular disease [135].

**Extracellular Matrix and Vascular Remodeling following active inflammation**

During inflammation, increased proteolysis as described above is responsible for the degradation of ECM proteins and subsequent vascular remodeling. Molecular Imaging of ECM proteins has been achieved by MRI using Gadofluorine, a Gd-based contrast agent targeting collagen, proteoglycans and tenascin[156-158]. The ECM protein elastin has been shown to be crucial for arterial remodeling in atherosclerosis. Human atheroma and macrophage-rich plaque regions display a high content of immature disorganized tropoelastin but lack stabilizing cross-linked, mature elastin. Recently an elastin-targeted molecular MR agent (ESMA) has been introduced and showed promising results for non-invasive imaging of the vessel wall (Figure 7). ESMA was able to visualize and quantify plaque burden in atherosclerotic apoE−/− mice [159] and depict stent induced coronary injury in a swine model [160, 161]. Also paramagnetic micelles (CNA35) targeting collagen were able to detect murine atherosclerosis by molecular MRI [162].

The end stage of vascular remodeling is the formation of microcalcifications and sclerosis [163], and there is clear evidence that both inflammation and calcification are closely related to each other[42, 43]. Molecular targeting of vessel wall calcifications has been achieved by fluorescence imaging and hydroxylapatite-targeted PET [42, 43, 164, 165]. Together with high-resolution CT hydroxylapatite targeted PET may give rise to a more comprehensive evaluation of plaque phenotype, integrating morphology, plaque composition, degree of the resulting vessel obstruction, and stage of plaque progression. Already today, coronary CT is able to distinguish between stable and vulnerable plaques based on the existence and pattern of calcifications. While large calcifications are characteristic of stable plaques, both non-calcified plaques as well as spotty calcifications are more frequently observed in patients presenting with an acute coronary syndrome [166].

**Fig 7. Imaging of Vascular Remodeling.** Vascular remodeling can be assessed with an Elastin-targeted Magnetic Resonance Agent (=ESMA). Panels A and B show ESMA-enhanced MR images of the aortic arch and supraaortic vessels in swine with increased SNR and CNR compared to non-targeted Gd-DTPA (Panel C and D). A similar approach is able to detect vascular injury following coronary stent implantation. Magnetic Resonance Angiography (MRA, Panel E), delayed-enhancement MRI after injection of ESMA (Panel F) and fusion of E and F (Panel G). Quantification of Elastin by MRI yields good correlation with Histology (Panel H). Images courtesy of Marcus Makowski and René M. Botnar, Kings College London and Christian von Bary, Universität Regensburg.
Rupture of the inflamed atherosclerotic plaque and thrombus formation

Rupture of the inflamed atherosclerotic plaque leads to exposure of a variety of prothrombotic plaque constituents to the circulation, initiating atherothrombosis and sequelae such as the development of stroke or myocardial infarction. Selective targeting of clot-bound platelets has been achieved using peptides or antibodies with high affinity for the glycoprotein IIb/IIIa[167, 168].

Fibrin-targeted MRI has vast potential in thrombus imaging and has already passed Phase I and II clinical studies after promising results in animal models [169-172]. EP-2104R is a Gd-based MR imaging agent functionalized with a short peptide with high affinity to fibrin [173-175]. A novel 64Cu labeled EP-2104R showed promising results in depicting arterial thrombi by hybrid MR-PET in a rat model [176]. Quantification of intraplaque and endothelial fibrin has been achieved by applying novel T1 mapping techniques of the injected EP2104R [177]. An example of this approach is shown in Figure 8.

Another preclinical approach uses peptide substrates for factor XIII, an important factor in cross-linking fibrin monomers for thrombus stabilization. Fluorescent nanosensors were able to reveal factor XIII activity in experimental models of thrombosis and may enable to differentiate acute versus chronic thrombi in vivo [178]. Similar approaches using MRI showed feasible factor XIII detection in preclinical animal models [179].

Target selection for clinical molecular imaging

As outlined above, a variety of molecular targets have so far been successfully employed in preclinical models of cardiovascular disease. However, the translation of imaging agents and technologies into the clinic has been much slower than anticipated. When selecting potential targets for molecular imaging of patients, the target has to fulfill a set of criteria.
to be suitable for non-invasive detection: The expression and kinetics of a potential molecular target should be known in the course of the disease, and the target density at the region of interest should be high enough to be captured with a high contrast-to-noise ratio. Above all, an imaging agent has to be available that is able to detect the target with high sensitivity and specificity, is non-toxic, and preferably available at low costs. Currently, there is no consensus on the best target for imaging in the clinical setting. In humans, our knowledge about the expression of molecular markers within the vasculature is mostly limited to one-time snapshot, for example when analyzing a carotid plaque after a patient had undergone endarterectomy. The kinetics of inflammatory processes in atherosclerosis and in particular the course of expression of potential molecular markers within the inflamed vessel, are yet to be defined. To date, targets that may be most promising for clinical imaging applications comprise adhesion molecules and monocytes/macrophages, as hallmarks of inflammations. Integrins and cell adhesion molecules are not only expressed on the activated endothelium, but also on inflammatory macrophages within the plaque, which should enable imaging with a good contrast-to-noise ratio. Similarly, macrophages are highly abundant within the vulnerable plaque and their presence is moreover clearly associated with the risk of plaque rupture. Macrophages are therefore attractive imaging targets.

**Perspectives on Clinical Translation**

Our understanding of the inflammatory nature and molecular processes involved in atherosclerosis has substantially advanced over the last 30 years, and it is now widely accepted that inflammation plays a key role in plaque development, maturation and rupture [134]. Molecular imaging, unlike anatomic imaging, is able to elucidate the immunobiological processes inside the vessel wall, otherwise invisible by x-ray, coronary angiography or established intravascular imaging methods. While traditional anatomic imaging modalities have failed to identify the vulnerable plaque that is prone to rupture, it is the aim of ongoing experimental efforts to define imaging modalities that enable the detection and quantification of parameters that can reliably be integrated into clinical diagnostic and therapeutic decision making. In particular, it remains a pressing question in the clinic, which lesion to treat in order to prevent potentially life-threatening complications, such as myocardial infarction and stroke [180, 181]. Molecular imaging is able to translate our advanced knowledge in vascular biology towards these applications. Yet, many hurdles remain to be solved before molecular imaging platforms can be implemented in routine clinical practice. The lack of probe specificity needs to be overcome by more specific sensors targeting distinct cellular or subcellular processes, and more sensitive detection technologies. Some obstacles, such as the low spatial resolution of nuclear imaging or the restricted penetration depths of near-infrared light, are rooted in physical principles that may not be significantly improved. Modification of the application of these technologies, e.g. imaging via the intravascular route, however, may have the potential to further improve these imaging technologies. First applications of fluorescence imaging in surgical oncology have proven promising when used in cytoreductive surgery [182]. As more and more selective molecular sensors will undergo FDA approval, molecular imaging approaches that have been successful in preclinical animal models will soon be tested in the clinical setting. When the discussed novel imaging platforms are considered for clinical translation, radiation exposure will become a key issue. Although costs of these new imaging technologies are still immense, these may still be modest when compared to the general healthcare costs that arise from the treatment of vascular complications such as myocardial infarction or stroke. Identifying these previously silent culprit lesions before myocardial infarction or stroke occur is a major task for molecular imaging.

When envisioning successful implementation of molecular imaging into daily patient care in the future, one could imagine the following scenario: A patient presents to the emergency room with acute chest pain. The laboratory values as well as the echocardiographic findings point toward an acute coronary syndrome. The patient is therefore brought to the catheterization laboratory for invasive angiography. In addition to the regular x-ray angiogram for detecting vessel obstructions, intravascular fluorescence imaging is performed to screen the entire coronary vascular tree for possible vulnerable plaques, which are not identified by conventional coronary angiography but may nevertheless cause subsequent infarctions within the near future. After angioplasty of an obstructed artery segment the patient is placed on an anti-inflammatory therapeutic regimen. The treatment success can be subsequently monitored non-invasively, either by hybrid PET-CT/PET-MRI or by molecular MRI alone and can guide further therapeutic triage. One could imagine that in parallel, patients would be subjected to genomic risk analysis to guide the aggressiveness of the anti-inflammatory regimen and determine the frequency of follow-up examinations by molecular imaging. Although such implementation of molecular imaging in the clinical routine is appealing, many hurdles, as outlined above,
still need to be overcome [45]. As a concluding remark, the cardiovascular field may benefit from the cancer community where imaging of biomarkers has found its ways into standard daily patient care and is utilized in large clinical trials [45]. In our pursuit of establishing personalized medicine for improving patient care, molecular imaging tools will be of great value for helping physicians to identify vulnerable plaques and triage tailored treatment approaches in individual patients with the aim to avert serious complications of atherosclerosis.

Competing Interests

The authors have declared that no competing interest exists.

References


145. Tabas I. Macrophage apoptosis in atherosclerosis: consequences on plaque
131: 15739-44. doi:10.1021/ja903922u.


