T cell–derived inducible nitric oxide synthase switches off TH17 cell differentiation

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T cell–derived inducible nitric oxide synthase switches off $T_{H17}$ cell differentiation

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RORγt is necessary for the generation of $T_{H17}$ cells but the molecular mechanisms for the regulation of $T_{H17}$ cells are still not fully understood. We show that activation of CD4+ T cells results in the expression of inducible nitric oxide synthase (iNOS). iNOS–deficient mice displayed enhanced $T_{H17}$ cell differentiation but without major effects on either $T_{H1}$ or $T_{H2}$ cell lineages, whereas endothelial NOS (eNOS) or neuronal NOS (nNOS) mutant mice showed comparable $T_{H17}$ cell differentiation compared with wild-type control mice. The addition of $N6$-(1-iminoethyl)-L-lysine dihydrochloride ($L$-NIL), the iNOS inhibitor, significantly enhanced $T_{H17}$ cell differentiation, and $S$-nitroso-$N$-acetylpenicillamine (SNAP), the NO donor, dose-dependently reduced the percentage of IL-17–producing CD4+ T cells. NO mediates nitration of tyrosine residues in RORγt, leading to the suppression of RORγt–induced IL-17 promoter activation, indicating that NO regulates IL-17 expression at the transcriptional level. Finally, studies of an experimental model of colitis showed that iNOS deficiency results in more severe inflammation with an enhanced $T_{H17}$ phenotype. These results suggest that NO derived from iNOS in activated T cells plays a negative role in the regulation of $T_{H17}$ cell differentiation and highlight the importance of intrinsic programs for the control of $T_{H17}$ immune responses.

Abbreviations used: eNOS, endothelial NOS; iNOS, inducible NOS; L-NIL, N6-(1-iminoethyl)-L-lysine dihydrochloride; MS, multiple sclerosis; NOS, nitric oxide synthase; nNOS, neuronal NOS; RA, rheumatoid arthritis.

IL-17–producing T helper cells ($T_{H17}$) are a recently identified T helper cell subset, which is clearly distinct from $T_{H1}$ and $T_{H2}$ cells. $T_{H17}$ cells mediate proinflammatory and autoimmune responses through the production of $T_{H17}$ signature cytokines including IL-17A, IL-17F, and IL-22 (Liang et al., 2006; Weaver et al., 2006; Bettelli et al., 2007; Zheng et al., 2007). Synergy between the cytokines TGF-β and IL-6 induces in vitro development of $T_{H17}$ cells (Veldhoen et al., 2006; Korn et al., 2007; Nurieva et al., 2007; Zhou et al., 2007), whereas IL-23 promotes the survival and expansion of $T_{H17}$ cell populations (Bettelli et al., 2006; Mangan et al., 2006; Veldhoen et al., 2006; Weaver et al., 2006). RORγt, a member of the orphan nuclear receptor family, has been identified as the master transcription factor for $T_{H17}$ cell development (Ivanov et al., 2006). Other transcription factors, including RORα, STAT3, IRF4, and IRF8, are also involved in the control of $T_{H17}$ cell differentiation (Ouyang et al., 2011). In addition, the differentiation of $T_{H17}$ cells is also regulated...
Nitric oxide (NO) is one of the smallest known bioactive products of mammalian cells and it can be produced by many mammalian cell types (Moncada et al., 1991). Three distinct isoforms of NO synthase (NOS) have been identified: neuronal NOS (nNOS), inducible NOS (iNOS), and endothelial NOS (eNOS; Griffith and Stuehr, 1995). It has been demonstrated that NO plays many important roles in neurotransmission, vascular functions, host defense, and immune regulation (Bogdan, 2001; Calabrese et al., 2007). These enzymes are products of different genes, with different regulation, localizations, and catalytic properties. nNOS and eNOS are primarily expressed in neurons and endothelial cells, and they are calcium dependent. iNOS can be induced by cytokines and other stimuli in many cell types and it is calcium independent. It is clear that NO is an important proinflammatory cytotoxic mediator that defends the host against various pathogens by inactivating and destroying infectious agents (Bogdan et al., 2000). Interestingly, NO also plays critical roles in immune suppression (Xiong et al., 1996; Niedbala et al., 2006). Previously, we and other groups reported that NO suppresses IL-12 production from dendritic cells and macrophages (Xiong et al., 2004), suggesting that NO may control the generation of TH1 immune responses by regulating IL-12 expression.

There is increasing evidence that TH17 cells are involved in the pathogenesis of various autoimmune/inflammatory diseases, including multiple sclerosis (MS), rheumatoid arthritis (RA), inflammatory bowel diseases (IBDs), and asthma (Korn et al., 2009). Thus, a more complete understanding of the molecular mechanisms involved in the regulation of TH17 immune responses should provide insights into the pathogenesis and treatment of these and possibly other inflammatory diseases. Although the activation program for TH17 cell differentiation has been well established, the intrinsic down-regulation for TH17 cell differentiation has not been fully understood.

Nitric oxide (NO) is one of the smallest known bioactive products of mammalian cells and it can be produced by many mammalian cell types (Moncada et al., 1991). Three distinct isoforms of NO synthase (NOS) have been identified: neuronal NOS (nNOS), inducible NOS (iNOS), and endothelial NOS (eNOS; Griffith and Stuehr, 1995). It has been demonstrated that NO plays many important roles in neurotransmission, vascular functions, host defense, and immune regulation (Bogdan, 2001; Calabrese et al., 2007). These enzymes are products of different genes, with different regulation, localizations, and catalytic properties. nNOS and eNOS are primarily expressed in neurons and endothelial cells, and they are calcium dependent. iNOS can be induced by cytokines and other stimuli in many cell types and it is calcium independent. It is clear that NO is an important proinflammatory cytotoxic mediator that defends the host against various pathogens by inactivating and destroying infectious agents (Bogdan et al., 2000). Interestingly, NO also plays critical roles in immune suppression (Xiong et al., 1996; Niedbala et al., 2006). Previously, we and other groups reported that NO suppresses IL-12 production from dendritic cells and macrophages (Xiong et al., 2004), suggesting that NO may control the generation of TH17 immune responses by regulating IL-12 expression.

Figure 1. Enhanced TH17 cell differentiation in iNOS-deficient mice. (A) Naive CD4+ T cells from WT or iNOS−/− mice were differentiated under TH0 and TH17 polarizing conditions for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h, stained for intracellular IL-17 and analyzed by flow cytometry. Representative FACS dot plots gated on CD4+ T cells and the percentages of IL-17–producing CD4+ T cells are shown. Each bar represents mean ± SD from three independent experiments. *, P < 0.05 versus iNOS−/− cells. (B) The cells prepared in A were restimulated with PMA/ionomycin for 12 h and the supernatants were analyzed for IL-17 and IL-22 by ELISA. Each bar represents mean ± SD of at least three independent measurements. (C) The cells prepared in A were restimulated with PMA/ionomycin for 5 h and mRNA expression of indicated genes was determined by qPCR. Data present mean ± SD of measurements from two independent experiments, performed in triplicate. The data shown were normalized to levels of ubiquitin expression as analyzed by qPCR. *, P < 0.05 versus iNOS−/− cells. (D) Thymus and spleen cells from iNOS−/− and WT controls were prepared and the cells were stained for surface CD4 and CD8 and analyzed by flow cytometry. The data shown were normalized to levels of ubiquitin expression as analyzed by qPCR. *, P < 0.05 versus iNOS−/− cells. The results are representative of three independent experiments.
Transfer of naive T cells from iNOS-deficient mice induced more severe colitis in \( \text{Rag1}^-\) mice than T cells from normal controls. These findings suggest that NO plays a critical suppressive role in the control of T\(_{H17}\) differentiation and highlight the importance of T cell–derived iNOS in switching off T\(_{H17}\)-dependent immune responses.

**RESULTS**

iNOS deficiency enhances T\(_{H17}\) cell differentiation

To investigate the function of NO in T\(_{H17}\) cell differentiation, we first assessed the characteristics of CD4\(^+\) T cells from WT or iNOS\(^-\) mice were differentiated under T\(_{H17}\) conditions for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h and stained for intracellular IFN-\(\gamma\) or IL-4 by flow cytometry. Each bar represents mean ± SD from three independent experiments. **A** Naive CD4\(^+\) T cells from WT or iNOS\(^-\) mice were differentiated under T\(_{H17}\) conditions for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h and stained for intracellular IFN-\(\gamma\) or IL-4 by flow cytometry. Each bar represents mean ± SD from three independent experiments. **B** Naive CD4\(^+\) T cells from spleens and lymph nodes of WT and iNOS\(^-\) mice were prepared and the cells were activated with anti-CD3 and anti-CD28 antibodies for 3 d. \([\text{H}]\)-Thymidine was added during the last 8 h of culture. Then the cells were collected and counted with a \(\beta\)-counter. Alternatively, naive CD4\(^+\) T cells were labeled with CFSE and the cells stimulated with plate bound anti-CD3 and anti-CD28 antibodies for 3 d. T cell proliferation was analyzed by flow cytometry. Data represent mean ± SD from two independent experiments, performed in triplicate.

In addition, iNOS-deficient mice are more susceptible than WT mice to the development of inflammatory diseases such as EAE (Bogdan., 1998; Niedbala et al., 2011). More and more evidence indicates that NO affects T helper cell differentiation (Nath et al., 2010; Lee et al., 2011; Niedbala et al., 2011), suggesting that NO may dictate T cell immune responses. However, it is still not clear whether T cells express NOS, and if so, what functions T cell–derived NO might serve.

In the present study, we show that mice deficient in iNOS exhibited enhanced T\(_{H17}\) cell differentiation while exhibiting no significant effects on T\(_{H1}\) or T\(_{H2}\) cells. We demonstrated that iNOS protein was induced in activated CD4\(^+\) T cells and that use of an iNOS-selective inhibitor, N6-(1-iminoethyl)-L-lysine dihydrochloride (L-NIL), significantly increased the percentage of IL-17–producing CD4\(^+\) T cells in cell cultures from WT mice, whereas an NO donor, S-nitroso-N-acetylpenicillamine (SNAP), dose-dependently suppressed IL-17 production in WT and iNOS\(^-\) T cell cultures. In addition, the tyrosine residues of ROR\(\gamma\)t were nitrated resulting in the inhibition of ROR\(\gamma\)t-mediated IL-17 promoter activation.

Transfer of naive T cells from iNOS-deficient mice induced more severe colitis in \( \text{Rag1}^-\) mice than T cells from normal controls. These findings suggest that NO plays a critical suppressive role in the control of T\(_{H17}\) differentiation and highlight the importance of T cell–derived iNOS in switching off T\(_{H17}\)-dependent immune responses.

**RESULTS**

iNOS deficiency enhances T\(_{H17}\) cell differentiation

To investigate the function of NO in T\(_{H17}\) cell differentiation, we first assessed the characteristics of CD4\(^+\) T cells from iNOS-deficient mice. Naive CD4\(^+\) T cells from iNOS\(^-\) or WT control mice were primed in vitro for 3 d under neutral (T\(_{H0}\)) or T\(_{H17}\) (IL-6 plus TGF-\(\beta\)) polarizing conditions. The cells were then restimulated with PMA/ionomycin and examined for the percentages of IL-17–producing cells by intracellular staining using flow cytometry. Notably, the frequency of IL-17–producing cells generated from iNOS\(^-\) T cell cultures was significantly greater than cells from WT cultures (Fig. 1 A). These observations correlated with enhanced
IL-17, IL-22, and IL-9 secretion by iNOS−/− T_{H}17 cells as determined by ELISA (Fig. 1 B). In addition, transcript levels of the signature T_{H}17 cytokines, IL-17 and IL-21, were significantly enhanced in iNOS−/− T_{H}17 cells (Fig. 1 C). To rule out the possibility that the enhanced T_{H}17 cell differentiation was a result of abnormal T cell development, we analyzed CD4+ T cells from spleens and lymph nodes of WT and iNOS−/− mice (Fig. 1 D). In contrast to the dramatic effect of iNOS deficiency on T_{H}17 cell differentiation, T_{H}11 and T_{H}12 differentiation were not noticeably affected in iNOS−/− T cell cultures (Fig. 2 A). Furthermore, when we polarized naive CD4+ T cells under conditions with TGF-β/IL-6 plus IL-23, we found that IL-17 single-positive cells were significantly increased in iNOS−/− T cell cultures, but there was no clear difference in the number of IFN-γ single-positive cells between WT and iNOS−/− T cell cultures, whereas IL-17/IFN-γ double-positive cells were just minimally increased (unpublished data). iNOS−/− mice had normal numbers of CD4+ T cells (unpublished data) and exhibited comparable expression of T cell activation markers CD62L, CD44, CD25, and CD69 to relative cells from WT mice (unpublished data). In addition, [3H]-thymidine incorporation assays and CFSE dilution showed that the proliferation of CD4+ T cells from iNOS−/− or WT control mice cultured under T_{H}17 conditions was comparable (Fig. 2 B). Collectively, these results indicate that T_{H}11 cell differentiation is enhanced in CD4+ T cells deficient in iNOS, suggesting that NO plays a negative role in T_{H}17 cell differentiation.

To investigate whether the enhancement of T_{H}17 cell differentiation was the result of T_{reg} dysfunction in iNOS-deficient mice, we examined FOXP3+CD4+ T cells in these mice. Naive CD4+ T cells from the spleens and lymph nodes of WT and iNOS−/− mice were activated in vitro for 3 d under neutral T_{H}10 or T_{reg} (TGF-β) polarizing conditions. There were no significant differences between the FOXP3+ CD4+ T cell populations of WT and iNOS−/− mice under T_{reg} inducing conditions (Fig. 3 A). In addition, production of IL-10, another inhibitory cytokine, was comparable in WT and iNOS−/− T_{H}17 cells (Fig. 3 B). Thus, the enhanced T_{H}17 cell differentiation in iNOS−/− mice was not a result of the alterations of either TGF-β-derived T_{reg} or IL-10 production.

**iNOS is induced in activated CD4+ T cells**

The enhanced T_{H}17 cell differentiation of iNOS−/− T cells cultured under T_{H}17 polarizing conditions prompted us to think that iNOS expression in T cells could be responsible for reduced T_{H}17 cell differentiation in WT mice. To address this question, naive CD4+ T cells from the spleens and lymph nodes of WT and iNOS−/− mice were activated in vitro for 3 d under T_{H}10 or T_{H}17 polarizing conditions. Western blotting showed that iNOS protein was indeed induced in WT CD4+ T cells but not in iNOS−/− T cells (Fig. 4 A). In addition, stimulation of OT-II cells with OVA peptide (323–339) resulted in significant induction of iNOS protein as determined by Western blotting (Fig. 4 A). Staining cultured CD4+ T cells with anti-CD4 and anti-iNOS antibodies demonstrated that CD4+ T cells expressed iNOS after stimulation with anti-CD3 and anti-CD28 antibodies (Fig. 4 B). Furthermore, real time RT-PCR experiments revealed the expression of iNOS mRNA at 6 and 12 h after TCR engagement (Fig. 4 C). In addition, single cell FACS analysis showed that iNOS was expressed in activated WT CD4+ T cells but not in iNOS−/− CD4+ T cells (Fig. 4 D), and iNOS/IL-17 double-positive cells were present in WT T cells but not in iNOS−/− T cells (unpublished data). To rule out the possibility that myeloid cells may have been contaminated in the CD4+ T cell cultures, we stimulated CD4+ T cells with 1 µg/ml LPS for 24 and 48 h and Western blotting was performed. The results

Figure 3. There are no changes of T_{reg} differentiation and IL-10 production in iNOS−/− mice. (A) Naive CD4+ T cells from WT or iNOS−/− mice were differentiated under T_{H}0 and T_{reg} polarizing conditions for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h, stained for intracellular FOXP3, and analyzed by flow cytometry. Representative FACS dot plots gated on CD4+ cells and the percentages of FOXP3-positive CD4+ cells are shown. Each represents mean ± SD from three independent experiments. (B) Naive CD4+ T cells from WT or iNOS−/− mice were differentiated under T_{H}0 and T_{H}17 polarizing conditions for 3 d. Cells were then restimulated with PMA/ionomycin for 24 h and the supernatants were analyzed for IL-10 by ELISA. Data represent mean ± SD from three independent experiments.
were primed in vitro for 3 d under T\(_0\) or T\(_{17}\) polarizing conditions. The cells were then restimulated with PMA/ionomycin and examined for the percentages of IL-17–producing cells by intracellular staining using flow cytometry. The percentages of IL-17–producing CD4\(^+\) T cells from eNOS\(^{-/-}\) and nNOS\(^{-/-}\) CD4\(^+\) T cell cultures were comparable to cells from WT cell cultures (unpublished data). CD4\(^+\) T cells developed normally in eNOS\(^{-/-}\) and nNOS\(^{-/-}\) mice (unpublished data). These results suggest that iNOS is expressed in activated T cells and may play a role in T\(_{17}\) cell differentiation.

showed that macrophage cell line 264.7 cells expressed high level of iNOS protein after LPS stimulation, whereas LPS-stimulated CD4\(^+\) T cells did not express iNOS protein at all (unpublished data). These results suggest that NO derived from iNOS expressed by activated CD4\(^+\) T cells plays a negative role in T\(_{17}\) cell differentiation.

Because T cells have also been reported to express eNOS and nNOS (Williams et al., 1998; Ibiza et al., 2006), we further wanted to explore whether eNOS or nNOS is involved in the regulation of T\(_{17}\) cell differentiation. Naive CD4\(^+\) T cells from eNOS\(^{-/-}\), nNOS\(^{-/-}\), and WT mice were primed in vitro for 3 d under T\(_{17}\) polarizing conditions. The cells were then restimulated with PMA/ionomycin and examined for the percentages of IL-17–producing cells by intracellular staining using flow cytometry. The percentages of IL-17–producing CD4\(^+\) T cells from eNOS\(^{-/-}\) and nNOS\(^{-/-}\) CD4\(^+\) T cell cultures were comparable to cells from WT cell cultures (unpublished data). CD4\(^+\) T cells developed normally in eNOS\(^{-/-}\) and nNOS\(^{-/-}\) mice (unpublished data). These results suggest that iNOS is expressed in activated T cells and may play a role in T\(_{17}\) cell differentiation.
Figure 5. NO suppresses Th17 cell differentiation. (A) Naive CD4+ T cells from B6 mice were differentiated under Th17 polarizing conditions in the presence of 0.5 mM L-NIL for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h, stained for intracellular IL-17, and analyzed by flow cytometry. Representative FACS dot plots gated on CD4+ cells and the percentages of IL-17–producing CD4+ cells are shown. Each bar represents mean ± SD from three independent experiments. *, P < 0.05 versus cells added with L-NIL. (B) The cells prepared in A were differentiated under Th17 polarizing conditions in the presence of SNAP (10, 100, or 200 µM) for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h, stained for intracellular IL-17, and analyzed by flow cytometry. Representative FACS dot plots gated on CD4+ cells and the percentages of IL-17–producing CD4+ cells are shown. (C) The cells prepared in A were restimulated with PMA/ionomycin for 12 h and the supernatants were analyzed for IL-17 by ELISA or the cells prepared in A were restimulated with PMA/ionomycin for 5 h and mRNA expression of indicated genes was determined by qPCR. The data shown were normalized to levels of ubiquitin expression as analyzed by qPCR. Each bar presents mean ± SD from three independent experiments. *, P < 0.05 versus Th17 cells only. The results are representative of three independent experiments. (D) Naive CD4+ T cells from iNOS−/− mice were differentiated under Th17 polarizing conditions in the presence of 0.5 mM L-NIL or 100 µM SNAP for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h, stained for intracellular IL-17, and analyzed by flow cytometry. Representative FACS
The effect of L-NIL, a pharmacologic iNOS inhibitor, on Th17 cell differentiation

Experiments with iNOS−/− mice indicated that NO may be involved in the negative regulation of Th17 cell differentiation. We next tested this by adding an iNOS inhibitor to WT T cell cultures. We used L-NIL, a selective iNOS inhibitor, to treat CD4+ T cells cultured under Th17 conditions. Naive CD4+ T cells from spleens and lymph nodes of WT mice were activated in vitro for 3 d under Th17 polarizing conditions in the presence of 0.5 mM L-NIL. Addition of L-NIL significantly enhanced the percentage of IL-17–producing cells and IL-17 protein release in WT CD4+ T cell cultures (Fig. 5 A), which mimicked the iNOS−/− CD4+ T cell cultures. We next used SNAP, which releases NO spontaneously and is widely used as an NO donor. SNAP dose-dependently reduced the percentage of IL-17–producing CD4+ T cells (Fig. 5 B). Similarly, IL-17 protein release and IL-17 mRNA expression were inhibited by SNAP in a dose-dependent manner (Fig. 5 C). In addition, SNAP suppressed Th17 cell differentiation in both WT and iNOS−/− cell cultures, whereas L-NIL had no significant effect on the percentage of IL-17–producing cells and IL-17 protein release in iNOS−/− CD4+ T cell culture (Fig. 5 D). Furthermore, we also used 2-phenyl-4,4,5,5-tetramethyl-imidazoline-1-oxyl-3-oxide (PTIO), an NO scavenger. We demonstrated that PTIO treatment significantly increased the percentage of IL-17–producing CD4+ T cells in WT T cell cultures but not in iNOS−/− T cell cultures (Fig. 6). To test whether L-NIL and SNAP affects Th1 or Th2 cell differentiation, naive CD4+ T cells from spleens and lymph nodes of WT mice were activated in vitro for 3 d under Th1 or Th2 conditions in the presence of 0.5 mM L-NIL or 50 µM SNAP. We did not find a significant effect of L-NIL or SNAP on Th1 or Th2 cell differentiation (unpublished data). In addition, the compound SNAP or L-NIL had no significant effect on cell viability and proliferation (unpublished data). Thus, these results suggest that NO derived from iNOS expressed in T cells selectively inhibits Th17 cell differentiation.

NO suppresses RORγt-mediated IL-17 transcription

The above findings prompted us to probe the molecular basis for NO control of Th17 cell differentiation. Because many studies have demonstrated a critical role for RORγt in Th17 cell differentiation both in vitro and in vivo (Ivanov et al., 2006), we asked if NO might affect RORγt expression, resulting in the control of Th17 cell differentiation. First we examined RORγt expression in iNOS-deficient CD4+ T cells. Naive CD4+ T cells from iNOS−/− or WT littermate mice were primed in vitro for 3 d under Th0 or Th17 polarizing conditions. The cells were then restimulated with PMA/ionomycin and examined for the percentages of RORγt-positive cells by intracellular staining using flow cytometry. Interestingly, the percentage of RORγt-positive cells in iNOS−/− CD4+ T cell cultures was comparable to that in WT cell cultures (Fig. 7 A). This was confirmed by Western blotting experiments showing that the levels of RORγt, STAT3, IRF4, and AHR protein were similar in iNOS−/− and WT T cells (Fig. 7 B). These results suggest that enhanced Th17 cell differentiation in iNOS−/− mice is not the result of a change in RORγt expression at the protein level. To further assess whether NO might directly affect RORγt protein expression, we transfected RORγt into 293T cells treated with different concentrations of SNAP for 40 h and then examined RORγt expression by Western blotting. The results showed that the treatment with SNAP did not affect the levels of RORγt expression (Fig. 7 C) or nuclear translocation (Fig. 7 C).

We then proceeded to analyze whether NO modulates posttranslational modification of RORγt protein. First, we considered if NO affects RORγt ubiquitination. To test this, we cotransfected T7-RORγt and HA-ubiquitin overexpression plasmids into 293T cells in the presence of the NO donor SNAP at different concentrations for 40 h. Cell lysates were analyzed by immunoprecipitation with anti-T7 antibody and immunoblotted with anti-HA mAb. A high molecular mass smear typical of ubiquitinated protein was detected only in samples cotransfected with T7-RORγt and HA-ubiquitin. SNAP treatment had no significant effect on

![Figure 6. NO suppresses Th17 cell differentiation.](Image)

Figure 6. NO suppresses Th17 cell differentiation. Naive CD4+ T cells from WT or iNOS−/− mice were differentiated under Th17 polarizing conditions in the presence of 200 µM PTIO for 3 d. Cells were then restimulated with PMA/ionomycin for 5 h, stained for intracellular IL-17, and analyzed by flow cytometry. Representative FACS dot plots gated on CD4+ cells and the percentages of IL-17–producing CD4+ cells are shown. The data are representative of three similar experiments.

![Image](Image)
the pattern of ubiquitination (unpublished data), suggesting that RORγt ubiquitination is not the target for the effect of NO on RORγt.

The mouse amino acid sequence of RORγt has 15 tyrosine residues (unpublished data), which may be subject to nitration induced by NO. To investigate this possibility, naive CD4+ T cells from spleens and lymph nodes of WT mice were activated in vitro for 3 d under Th17-polarizing conditions in the presence of L-NIL or SNAP. Flow cytometric analyses clearly revealed a population of RORγt-Nitrotyrosine+ cells in WT but not in iNOS−/− cells (Fig. 7 D). Treatment with SNAP resulted in an expanded cell population (Fig. 7 D), which was reversed in the presence of L-NIL (Fig. 7 D). ChIP analysis indicated that SNAP treatment suppressed RORγt binding to the promoter region of IL-17 gene (Fig. 7 E), whereas L-NIL treatment significantly enhanced the binding of RORγt there (Fig. 7 E). To further analyze the effect of NO on nitration of tyrosine residues in RORγt, we performed coimmunoprecipitation experiments using cell lysates from primary CD4+ T cells cultured under Th17-polarizing conditions. We found that anti-nitrotyrosine antibody coimmunoprecipitated RORγt from lysates of Th17 cells (Fig. 8 A), suggesting that tyrosine residues of RORγt were nitrated under Th17 conditions. To confirm these results, 293T cells were transfected with T7-RORγt in the presence of SNAP for 40 h. Cell lysates were immunoprecipitated with anti-nitrotyrosine antibody and immunoblotted with anti-T7 antibody. SNAP treatment clearly induced tyrosine nitrosylation...
of RORγt (Fig. 8B), suggesting that NO-induced alterations of tyrosine residues may affect RORγt activation. To analyze which part of RORγt is sensitive to NO, we generated three T7-tagged RORγt truncation mutants (Δ5-166, Δ169-491, and Δ342-491). The mutants were transfected into 293T cells in the presence of SNAP for 40 h and cell lysates were immunoprecipitated with anti-nitrotyrosine antibody and immunoblotted with anti-T7 antibody. SNAP treatment had no significant effect on RORγt mutant Δ5-166 (Fig. 8C) but clearly increased nitrotyrosine levels of RORγt truncation mutants Δ169-491 and Δ342-491 (Fig. 8C). Thus, tyrosine residues in the region from residue 169 to 491 of RORγt are sensitive to nitration by NO. Because NADPH oxidase is required for the formation of peroxynitrite, which is involved in the tyrosine nitration, we found that T cells expressed NADPH p47-phox (unpublished data), as reported previously (Jackson et al., 2004). To examine the effect of NO on RORγt at the functional level, we cotransfected an IL-17A promoter reporter construct containing the 6-kb promoter and a RORγt plasmid in the presence of SNAP (50, 100, or 200 µM) or 293T cells were transfected with an IL-17A promoter reporter construct and WT RORγt or different RORγt mutant plasmids for 30 h. Luciferase assays were performed and luciferase activities were normalized to β-galactosidase activity. Data indicate mean ± SD of triplicate cultures and are representative of three independent experiments.
Figure 9. iNOS deficiency promotes the T<sub>H</sub>17 immune responses in experimental colitis. CD4<sup>+</sup>CD45RB<sup>hi</sup> T cells were purified from spleens and lymph nodes of WT or iNOS<sup>−/−</sup> mice and 5 × 10<sup>5</sup> cells were injected (i.p.) into recipient Rag1<sup>−/−</sup> mice. Body weight change was monitored every week and mice were sacrificed 5 wk later. (A) Changes in body weight of Rag1<sup>−/−</sup> mice (n = 5–6 mice per group) after i.p. transfer of WT or iNOS<sup>−/−</sup>CD4<sup>+</sup>CD45RB<sup>hi</sup> T cells were recorded. Data are presented as the mean ± SD of the percentage of initial body weight and are representative of two similar experiments. *, P < 0.05 versus recipients of iNOS<sup>−/−</sup> cells. Morphology of intestines (B), disease scores (C; each bar represents mean ± SD of measurements made from five mice; *, P < 0.05 versus recipients of iNOS<sup>−/−</sup> cells), and sections of colons with colitis (D) from Rag1<sup>−/−</sup> mice (n = 5–6 mice in each group) on day 35 after naive T cell transfer was as described above. Bars, 100 µm. (E) The percentage of IL-17–producing cells from mesenteric lymph nodes of Rag1<sup>−/−</sup> mice in B, C, and D (white column, transfer with WT cells; black column, transfer with iNOS<sup>−/−</sup> cells). Each bar represents mean ± SD of measurements made from five mice. *, P < 0.05 versus recipients of iNOS<sup>−/−</sup> cells.
36 h and analyzed them for IL-17 promoter activation. The data showed that SNAP suppressed RORγt-mediated IL-17 promoter activation in a dose-dependent manner (Fig. 8 D). Furthermore, we have analyzed two tyrosine residues Tyr346 and Tyr359 of mouse RORγt, corresponding to Tyr369 and Tyr382 of human RORγt, that are important for ligand binding (Fig. S1, A and B). We found that mutating these two tyrosine residues significantly impaired RORγt-mediated IL-17 promoter activation, suggesting that Tyr346 and Tyr359 are critical for RORγt transcriptional function (Fig. 8 D). Thus, the results indicate that NO suppresses IL-17 expression at the transcriptional level by nitration of tyrosine residues in RORγt.

**iNOS regulates T_{h}17 cell differentiation in vivo**

Accumulating evidence indicates that T_{h}17 cells are involved in the pathogenesis of various autoimmune/inflammatory diseases, including MS, RA, IBD, and asthma (Weaver et al., 2006; Wilke et al., 2011). To further assess the effects of iNOS on T_{h}17 cell development in vivo, we performed adoptive transfer experiments using CD4^{+}CD45RB^{hi} cells from WT and iNOS−/− mice to induce colitis in Rag1^{−/−} mice. Rag1^{−/−} mice reconstituted with iNOS−/− T cells began losing weight earlier and lost significantly more weight than mice transferred with WT cells (Fig. 8 A). Parallel histological studies of colon sections from Rag1^{−/−} mice reconstituted with iNOS−/− T cells revealed more severe inflammatory cell infiltrates and significantly higher pathological scores than those observed in sections from mice reconstituted with T cells from WT mice (Fig. 9, B–D). In addition, mice reconstituted with iNOS−/− cells had significantly higher percentages of IL-17–producing cells than control mice (Fig. 8 E), whereas the percentage of IFN-γ–producing cells was comparable (Fig. 9 E). Thus, iNOS deficiency in T cells promotes intestinal inflammation in a T cell–mediated model of colitis, suggesting that iNOS expressed in T cells may play a negative role in the regulation of T_{h}17 immune response.

To further understand the role of iNOS in T_{h}17 cell function in vivo, we extended our studies to include experimental autoimmune encephalomyelitis (EAE), a mouse model of human MS. Previous studies showed that NO will suppress EAE (Zielasek et al., 1995; Okuda et al., 1997; Fenyk-Melody et al., 1998). To investigate the role of iNOS expressed by T cells on the development of EAE, we immunized WT and iNOS−/− mice with the myelin oligodendrocyte glycoprotein (MOG_{35-55}) peptide in complete Freund’s adjuvant. iNOS−/− mice developed EAE with an accelerated time course and greater severity than WT mice as indicated by disease scores (Fig. 10 A). As expected, iNOS mRNA expression was significantly induced in CD4^{+} T cells from WT but not from iNOS−/− mice in these experiments (Fig. 10 B). We then examined the expression of T_{h}17 and T_{h}1 cytokines in EAE mice. Infiltrating CD4^{+} T cells from the central nervous systems of iNOS-deficient mice with EAE expressed significantly higher percentage of IL-17–producing and IFN-γ–producing CD4^{+} T cells and expressed higher levels of transcripts for IL-17 and IFN-γ compared with cells from WT mice (Fig. 10, C and D). Thus, iNOS deficiency promotes inflammation in central nervous system in EAE, further confirming that NO negatively regulates T_{h}17 cell differentiation in vivo.

**DISCUSSION**

T_{h}17 cells are a new member of the still-growing family of T helper cell subsets, which play critical roles in the pathogenesis of autoimmune and inflammatory diseases. Therefore, understanding the intrinsic suppressing programs for T_{h}17 cells will help to dissect mechanisms for the control of T_{h}17 immune responses and elucidate the mechanism involved in the development of human inflammatory diseases including IBD, MS, and RA. In the present study, we demonstrated that iNOS-deficient naive CD4^{+} T cells polarized under T_{h}17 condition led to more efficient T_{h}17 cell differentiation without major effects on either the T_{h}1 or T_{h}2 cell lineages. In vivo, transfer of CD4^{+} CD45RB^{hi} cells into Rag1^{−/−} mice induced more severe colitis than transfer of control cells. In addition, mice reconstituted with iNOS−/− T cells had a significantly higher percentage of IL-17–producing CD4^{+} T cells than mice transferred with WT cells. These results suggest that iNOS derived from activated T cells selectively regulates T cell differentiation.

Many studies have demonstrated that NO can play a dual role in the modulation of immune responses (Niedbala et al., 2006). NO derived from iNOS in macrophages and other innate immune cells is proinflammatory and an essential component of host defenses against various pathogens including bacteria, parasites, and viruses (Bogdan et al., 2000). However, mounting evidence indicates that NO can also contribute to immune suppression. We and other groups previously reported that IL-12 mRNA and protein expression were significantly increased in iNOS KO mice, suggesting that NO may suppress IL-12–mediated T_{h}1 immune responses (Xiong et al., 2004). Huang et al. (1998) suggested that the enhanced T_{h}1 immune responses in iNOS KO mice were a result of increased production of IL-12 by iNOS−/− macrophages after infection with *Leishmania major*. In addition, Giordano et al. (2011) reported that expression of inflammatory cytokines, including TNF, IL-6, IL-12p70, and IL-23, was up-regulated in iNOS−/− bone marrow–derived dendritic cells. Collectively, these results indicate that iNOS expressed in innate immune cells, including macrophages and dendritic cells, can modulate inflammatory cytokine production. Although the exact molecular mechanisms responsible for this regulation are not fully understood, NO–mediated control of NF-κB activation may be involved (Xiong et al., 2004).

It is well established that iNOS is expressed in different cell types including, macrophages, dendritic cells, NK cells, and by both primary tumor cells and tumor cell lines (Bogdan, 2001). In addition, there is a controversy for the expression of iNOS in T cells. Vig et al. (2004) reported that T cell blasts expressed iNOS, which plays an important role in immune memory, whereas Thürling et al. (1995) did not find iNOS expression in T cells clones or T cells from naive and *L. major*–infected mice. In the present study, we clearly demonstrated...
that iNOS expressed by activated CD4+ T cells negatively regulated T<sub>H</sub>17 cell differentiation. The results are consistent with recent studies showing that the NO donors NOC-18 and S-nitrosoglutathione suppressed T<sub>H</sub>17 cell differentiation (Nath et al., 2010; Niedbala et al., 2011). Thus, our observations support the notion that NO derived from iNOS in activated T cells controls T cell differentiation by selectively suppressing T<sub>H</sub>17 cell development.

The importance of ROR<sup>γt</sup> in T<sub>H</sub>17 cell development has been well studied in mice. ROR<sup>γt<sup>-/-</sup> mice fail to develop lymph nodes or Peyer’s patches and T<sub>H</sub>17 cell development is severely impaired, indicating that ROR<sup>γt</sup> is a master transcription factor in T<sub>H</sub>17 cell differentiation (Ivanov et al., 2006). Interestingly, ROR<sup>γt</sup> expression in iNOS<sup>-/-</sup> CD4<sup>+</sup> T cells cultured under T<sub>H</sub>17 conditions was comparable to WT CD4<sup>+</sup> T cell cultures, implying that the enhanced T<sub>H</sub>17 cell differentiation observed is not the result of increased ROR<sup>γt</sup> protein levels. Instead, we found that the NOS donor SNAP suppressed ROR<sup>γt</sup>-mediated IL-17 promoter activation in a dose-dependent manner, suggesting that NO

![Figure 10](image-url)

**Figure 10.** iNOS is expressed in CD4<sup>+</sup> T cells infiltrated in CNS of WT mice with EAE and iNOS<sup>-/-</sup> mice develop more severe EAE. (A) iNOS<sup>-/-</sup> or WT mice were induced for EAE, and disease development was scored. Data are mean ± SD (n = 15 mice per group) pooled from three experiments. The score was based on a five-point scale. (B) CD4<sup>+</sup> T cells were purified from CNS of WT and iNOS<sup>-/-</sup> mice with EAE. Total RNA was extracted and iNOS mRNA expression was determined by qPCR. Each bar represents mean ± SD from two independent experiments. (C) WT or iNOS<sup>-/-</sup> mice were induced for EAE and CD4<sup>+</sup> T cells were purified from CNS of iNOS<sup>-/-</sup> or WT mice with EAE. Cells were then restimulated with PMA/ionomycin for 5 h, stained for intracellular IL-17 or IFN-γ, and analyzed by flow cytometry. Representative FACS dot plots gated on CD4<sup>+</sup> cells and the percentages of IL-17–producing or IFN-γ–producing CD4<sup>+</sup> cells are shown. (D) WT or iNOS<sup>-/-</sup> mice were induced for EAE and CD4<sup>+</sup> T cells were purified from CNS of iNOS<sup>-/-</sup> or WT mice with EAE. Total RNA was extracted and qPCR was performed for the analysis of IL-17, IFN-γ, and CCR6 mRNA expression. Each bar represents mean ± SD from three independent experiments. *, P < 0.05 versus WT cells.
Cells were stimulated with PMA and ionomycin for 5 h in the presence of brefeldin A before intracellular cytokines were measured by flow cytometry. Cells stimulated under neutral conditions were defined as T<sup>+</sup> cells, and cells stimulated in the presence of TGF-<beta> and anti–IFN-<gamma> were defined as T<sup>-</sup> cells. Sorted cells were primed for 96 h with 1 µg/ml anti–CD3 (145-2C11; BD) and 2 µg/ml of soluble anti-CD28 (37.51; BD). The cells were rested for 48 h and were then restimulated for 5 h with PMA plus ionomycin, and intracellular cytokines were measured by flow cytometry. The following antibodies were purchased from BD, as conjugated to FITC, PE, PE-Cy5, perCP-Cy5.5, or APC: CD4 (L3T4), NOX2 (NOS type II), CD8 (53–6.7), CD3e (145-2C11), CD25 (PC61.5), IFN-<gamma> (1A6) and p47phox (mouse) were purchased from EMD Millipore. Antibodies for iNOS (NOS1), NOS2 (iNOS), and NOS3 (eNOS) were purchased from R&D Systems. Antibodies for AHR (BML-SA210) was purchased from Enzo Life Sciences. Antibodies for IL-2 (JES6-1A12), ROR<gamma> (B2D), IL-4 (11B11), IL-10 (JES-16E3), and Foxp3 (FK3-16S) were purchased from eBioscience. Antibody for AHR (BML-SA210) was purchased from Enzo Life Sciences. Antibodies for NOX2 (NOS2, NOS type II), and NOS3 (eNOS) were purchased from R&D Systems. Antibodies for AHR (BML-SA210) was purchased from Enzo Life Sciences. Antibodies for IL-2 (JES6-1A12), ROR<gamma> (B2D), IL-4 (11B11), IL-10 (JES-16E3), and Foxp3 (FK3-16S) were purchased from eBioscience. Antibodies for AHR (BML-SA210) was purchased from Enzo Life Sciences. Antibodies for NOX2 (NOS2, NOS type II), and NOS3 (eNOS) were purchased from R&D Systems. Antibodies for AHR (BML-SA210) was purchased from Enzo Life Sciences.
permeabilization buffer, and stained with PE–anti-mouse IL-17, APC–anti-IFN-γ, and PE-Cy 5.5 anti-mouse CD4 antibodies. Flow cytometry was performed on a FACSCalibur (BD).

RNA isolation and quantitative real-time RT–PCR (qPCR). Total RNA was extracted using an RNeasy plus kit (QIAGEN) and cDNA was generated with an oligo (dT) primer and the SuperScript II system (Invitrogen), followed by analysis using iCycler PCR with SYBR Green PCR master Mix (Applied Biosystems). Results were normalized based on the expression of ubiquitin. The following primer sets were used: IL-17a sense, 5′-CTTCCAGAAGGCCTTACAGTAC-3′; IL-17a antisense, 5′-AGGTTCCTCCCTGCCATGACAG-3′; IL-21 sense, 5′-CGGCTTGGTATTAGCTTCCG-3′; IL-21 antisense, 5′-GAGCCCTTTACATTTGTTGAGA-3′; RORγt sense, 5′-CGGCTGAGAGGGCTTCAC-3′; RORγt antisense, 5′-TGCGAGGAGCAGCATAACA-3′; iNos sense, 5′-CCGAAGCAACATCACATTCA-3′; iNos antisense, 5′-GTTCTAAAGGCTCCGGGCT-3′; ubiquitin sense, 5′-TGCTTATTATTATGCGTCGCA-3′; and ubiquitin antisense, 5′-GCAAGTGCTGCTAGGCAAGTAA-3′.

Transfection and luciferase reporter assay. 293T cells were transiently transfected with an IL-17 promoter luciferase reporter plasmid together with RORγt in the presence of SNAP at different concentrations. For each transfection, 2.0 µg of plasmid was mixed with 100 µl DMEM (without serum and antibiotics) and 4.0 µl Lipofectamine 2000 reagent. The mixture was incubated at room temperature for 20 min and added to 12-well plates containing cells and complete medium. The cells were incubated for 30 h and harvested using reporter lysis buffer (Promega) for determination of luciferase activity. Cells were cotransfected with a β-galactosidase reporter plasmid to normalize experiments for transfection efficiency.

T cell proliferation assay. Naive CD4+ T cells were purified from spleens and lymph nodes of WT and iNos−/− mice. 105 cells/well were cultured in the absence or presence of 1 µg/ml anti-CD3 and 2 µg/ml anti-CD28 antibodies for 3 d in 96-well microplates. [3H]-Thymidine was added during the last 8 h of a 72-h culture. The cells were then harvested and counted with a β-counter.

Immunoblotting analysis. Cells were washed with cold phosphate-buffered saline and lysed for 15 min on ice in 0.5 ml lysis buffer (50 mM Tris-HCl, pH 8.0, 280 mM NaCl, 0.5% Nonidet P-40, 0.2 mM EDTA, 2 mM EGTA, 10% glycerol, and 1 mM dithiothreitol) containing protease inhibitors. Cell lysates were clarified by centrifugation (4°C, 15 min, 14,000 rpm), and proteins were subjected to 10% SDS-PAGE and immunoblotting was performed. Cell lysates were processed using an assay kit according to the manufacturer's instructions. The degree of inflammation in the epithelium, submucosa, and submuscularis propria was scored separately as described previously (Totsuka et al., 2007).

Chromatin immunoprecipitation (ChIP) assay. The ChIP procedure was performed using an assay kit according to the manufacturer's instructions (EMD Millipore). In brief, Tgfβ17 cells were cross-linked by 1% formalde-hyde for 10 min at 37°C. Nuclei were prepared and subjected to sonication to obtain DNA fragments. Chromatin fractions were preclreated with protein A–agarose beads followed by immunoprecipitation overnight at 4°C with 3 µg of anti-RORγt (Santa Cruz Biotechnology, Inc.) or control antibody. Cross-linking was reversed at 65°C for 4 h, followed by proteinase K digestion. DNA was purified and subjected to qPCR. The input DNA was diluted 200× before PCR amplification. The input and immunoprecipitated DNA were amplified by qPCR using primers encompassing the CNS2 and ~50 to ~250 regions of the mouse IL-17 promoter.

Induction of experimental autoimmune encephalomyelitis (EAE). iNos−/− and WT mice were immunized subcutaneously with 200 µl emulsion containing 100 µg myelin oligodendrocyte glycoprotein (MOG)35–55 peptide (MEGVYWRSPFSRVRHYLRNGK), complete Freund's adjuvant, and 0.4 mg Mycobacterium tuberculosis extract H37Ra (Difco). Mice were given 100 ng pertussis toxin (list Biological Laboratories) intraperitoneally on days 0 and 2. Mice were sacrificed ~3 wk later. EAE was scored as follows: 0, no diseases; 1, limp tail; 2, hind limb weakness; 3, partial paralysis and hind limb paralysis; 4, front and hind limb paralysis; 5, death.

Cytokine ELISA. Supernatants from cell cultures were collected after activation under various conditions and secreted cytokines in the supernatants were measured by ELISA kits with purified coating and biotinylated detection antibodies anti-IL-17 (R&D Systems) and anti-IL-10 (BD).

Statistical analysis. Statistical analysis was performed using Student's t test. P-values <0.05 were considered statistically significant.


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