The TAL1 complex targets the FBXW7 tumor suppressor by activating miR-223 in human T cell acute lymphoblastic leukemia

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Human T cell acute lymphoblastic leukaemia
(T-ALL) arises in thymocyte precursors through stepwise alterations in molecular pathways that often include the aberrant expression of intact master developmental regulatory transcription factors (Look, 1997; Ferrando et al., 2002; Armstrong and Look, 2005). One such transcription factor is TAL1/SCL, which is required for definitive haematopoiesis in early haematopoietic stem cells (HSCs), and is expressed by immature thymocytes after they migrate to the thymus, continuing until they reach the double negative 2 (DN2) stage of maturation, after which its levels are progressively down-regulated (Herblot et al., 2000; Lacombe et al., 2010; Lécuyer and Hoang, 2004; Porcher et al., 2010).

The oncogenic transcription factor TAL1/SCL is aberrantly expressed in 60% of cases of human T cell acute lymphoblastic leukaemia (T-ALL) and initiates T-ALL in mouse models. By performing global microRNA (miRNA) expression profiling after depletion of TAL1, together with genome-wide analysis of TAL1 occupancy by chromatin immunoprecipitation coupled to massively parallel DNA sequencing, we identified the miRNA genes directly controlled by TAL1 and its regulatory partners HEB, E2A, LMO1/2, GATA3, and RUNX1. The most dynamically regulated miRNA was miR-223, which is bound at its promoter and up-regulated by the TAL1 complex. miR-223 expression mirrors TAL1 levels during thymic development, with high expression in early thymocytes and marked down-regulation after the double-negative-2 stage of maturation. We demonstrate that aberrant miR-223 up-regulation by TAL1 is important for optimal growth of TAL1-positive T-ALL cells and that sustained expression of miR-223 partially rescues T-ALL cells after TAL1 knockdown. Overexpression of miR-223 also leads to marked down-regulation of FBXW7 protein expression, whereas knockdown of TAL1 leads to up-regulation of FBXW7 protein levels, with a marked reduction of its substrates MYC, MYB, NOTCH1, and CYCLIN E. We conclude that TAL1-mediated up-regulation of miR-223 promotes the malignant phenotype in T-ALL through repression of the FBXW7 tumor suppressor.
Activation of TAL1 expression through intrachromosomal deletion (SIL-TAL) or chromosomal translocation occurs in ~30% of patients, yet up to 60% of cases of T-ALL overexpress TAL1 through unknown mechanisms (Ferrando et al., 2002; Aifantis et al., 2008). TAL1-positive T-ALL is characterized by an arrest in differentiation at the CD4+CD8+ double-positive (DP) stage of thymocyte development, when the TAL1 gene is normally silent (Larson et al., 1996; Ferrando et al., 2002; Aifantis et al., 2008; Tremblay et al., 2010). Thus, transcriptional activity of TAL1 is clearly aberrant in T-ALL lymphoblasts compared with their normal DP counterparts.

The TAL1 gene encodes a class II basic helix–loop–helix (bHLH) transcription factor that binds E-box motifs only after heterodimerization with one of the class I bHLH E-proteins, such as E2A or HEB (Hsu et al., 1991; Hsu et al., 1994). Several of the core components of the transcriptional complex have now been elucidated and include GATA3, LIM domain only 1/2 (LMO1/2), and runt-related transcription factor 1 (RUNX1; Wadman et al., 1997; Lécuyer et al., 2002; Xu et al., 2003; Pali et al., 2011). Together, members of the core complex differentially interact with co-activators, such as the histone acetyltransferases p300 and p300/CBP-associated factor (PCAF), or with co-repressors such as SIN3A and histone deacetylase 1/2 (HDAC1/2; Huang et al., 1999; Huang and Brandt, 2000). Accordingly, binding of the TAL1 complex to promoter/enhancer regions can exert either a positive or negative influence on target gene expression.

We recently generated high-resolution maps of the genome-wide occupancy of the TAL1 complex in human T-ALL, which includes E2A, HEB, LMO1/2, GATA3, and RUNX1, using chromatin immunoprecipitation coupled to massively parallel DNA sequencing (ChiP-seq; Sanda et al., 2012). This analysis established that TAL1 acts predominantly as a positive regulator of the expression of its direct target genes, forms a positive interconnected auto–regulatory loop involving other members of the TAL1 complex, and activates several important target genes, including the MYB and TRIB2 oncogenes. Despite the emerging regulatory roles of miRNAs in normal and malignant hematopoiesis (Chen et al., 2004; Calin and Croce, 2006; Mavrakis et al., 2011), the underlying transcriptional mechanisms leading to dysregulated miRNA expression in cancer and T-ALL in particular remain poorly understood.

We first sought to determine which miRNAs were either primary or secondary targets of the TAL1 oncoprotein by assessing changes in miRNA expression after knockdown of TAL1 in Jurkat cells, a T-ALL cell line that aberrantly expresses TAL1. By depleting TAL1 using two independent short-hairpin RNAs (shRNAs1 and 2; Fig. 1A), and assessing global changes in miRNA expression using a locked nucleic acid (LNA)–based profiling platform, we identified significant changes in the expression of 22 miRNAs (Fig. 1B). Of these were down-regulated on TAL1 knockdown (hence positively regulated by TAL1), whereas 13 were up-regulated (hence negatively regulated by TAL1). Because many of these changes were likely to be secondary, we used ChiP-seq analysis of TAL1 binding with its binding partners HEB, E2A, GATA3, and RUNX1 (Sanda et al., 2012) to address this issue. Of the 22 regulated miRNAs, five (miR-223, miR-181a*, miR-29c, miR-26a and miR-620) were judged to be direct targets of the TAL1 complex based on our ChiP-seq data, with evidence of TAL1 occupancy within 12 kb of the candidate miRNA gene or binding a nearby enhancer (Fig. 1B, red bars).

miR-223 is directly regulated by the TAL1 complex in TAL1-positive T-ALL

Of the five miRNAs directly regulated by TAL1, we chose to focus on miR-223, because it exhibited the most robust down-regulation after TAL1 knockdown. After using qRT-PCR to confirm the changes we had identified by miRNA expression profiling (Fig. 1C), we asked whether miR-223 was regulated by TAL1 in TAL1-positive T-ALL cell lines other than Jurkat cells. Knockdown with both TAL1 shRNAs1 and 2 in the TAL1-positive T-ALL cell lines RPMI-8402 and CCRF-CEM (each harboring an activating SIL-TAL1 deletion) resulted in down-regulation of miR-223 as determined by qRT-PCR, similar to our results in Jurkat cells (Fig. 1, D and E). We also examined the changes in miR-223 expression in an inducible TAL1 knockdown model previously established in Jurkat cells that expresses a TAL1 shRNA upon addition of doxycycline (termed Jurkat S1C1 cells; Pali et al., 2011). In agreement with our results from lentiviral knockdown, we found marked down-regulation of miR-223 after TAL1 depletion (Fig. 1F). Furthermore, transduction of the TAL1-negative cell line KOPT-K1 with TAL1 shRNAs1 and 2 did not affect miR-223 expression by qRT-PCR (unpublished data), consistent with the interpretation that TAL1 depletion mediates the reduction in miR-223 in TAL1-positive cell lines. Thus, the expression of miR-223 is consistently and directly activated by TAL1 in T-ALL.

Next, we examined the miR-223 locus in detail in TAL1-positive T-ALLs, including two cell lines (Jurkat and CCRF-CEM) and two “primagraft” samples (designated “Prima 2” and “Prima 5”), which were derived from primary T-ALL cells expanded in immunocompromised mice without prior exposure to in vitro culture. In all four T-ALL samples, a region...
within 4 kb of the miR-223 transcriptional start site was occupied by TAL1, HEB, E2A, LMO1/2, GATA3 and RUNX1 (Fig. 2 A), a result validated by ChIP-qPCR (Fig. 2 B). Furthermore, this putative regulatory element was co-occupied by the transcriptional co-activator CREB-binding protein (CBP). The promoter and body of the miR-223 gene had high levels of trimethyl histone H3 on lysine 4 (H3K4me3) and dimethyl histone H3 on lysine 79 (H3K79me2) and broad enrichment within 4 kb of the miR-223 transcriptional start site was occupied by TAL1, HEB, E2A, LMO1/2, GATA3 and RUNX1 (Fig. 2 A), a result validated by ChIP-qPCR (Fig. 2 B). Furthermore, this putative regulatory element was co-occupied by the transcriptional co-activator CREB-binding protein (CBP).
miRNA is selectively up-regulated in the subset of T-ALLs transformed by the TAL1 complex. Of the 19 T-ALL cell lines interrogated, 10 overexpressed TAL1, of which 4 harbored the SLY-TAL1 deletion (Fig. 3 A). Expression of miR-223 was significantly higher in the TAL1-positive cell lines as compared with the TAL1-negative cells (Fig. 3 B; P < 0.0001).

To assess whether TAL1 regulates miR-223 in primary T-ALL, we also analyzed RNA from primary pediatric T-ALL samples collected at presentation (Ferrando et al., 2002). Consistent with our findings in T-ALL cell lines, TAL1-positive patients exhibited significantly higher expression of miR-223 than TAL1-negative patients (P < 0.001), indicating that TAL1 regulates the expression level of miR-223 in T-ALL cells as they exist in patients and not only in T-ALL cell lines growing in tissue culture (Fig. 3 C).

In normal murine thymocytes, Tal1 expression is highest in early thymic development and then is rapidly silenced as the cells differentiate beyond the DN2 stage (Herblot et al., 2000). To explore whether miR-223 expression is a physiological or promiscuous target of TAL1, we isolated thymic subsets from mice and analyzed them for Tal1 and miR-223 expression by qRT-PCR. In accord with published studies, Tal1 expression was highest in early T cell progenitor and DN2 cells and was dramatically down-regulated in more differentiated cells (Fig. 3 D). Interestingly, miR-223 expression closely mirrored Tal1 expression (Pearson’s correlation r = 0.89; P < 0.001), suggesting that TAL1 regulates miR-223 physiologically during normal thymocyte development (Fig. 3 E).

Consistent with this finding, TAL1 and its partners HEB, E2A, LMO1/2, GATA3, and RUNX1 bind a region 4-kb upstream of the miR-223 transcriptional start site. (A) ChIP coupled to massively parallel DNA sequencing (ChIP-seq) was performed for TAL1, HEB, E2A, and LMO1 or LMO2, GATA3, and RUNX1 (Sanda et al., 2012). Gene tracks represent binding at the miR-223 locus in a human primagraft sample (Prima 5) and the T-ALL cell lines CCRF-CEM (top) and Jurkat (bottom), with the miR-223 locus shown below. Y-axis values represent the number of sequence reads. (B) Enrichment of miR-223 regions indicated in A in four T-ALL cell samples (Jurkat, CCRF-CEM, Prima 2, and Prima 5) was analyzed by ChIP-qPCR. Mean ± SD fold enrichment as normalized to binding at the NANGO promoter (negative control) is shown from two independent experiments performed in triplicate.

Figure 2. TAL1 and its partners HEB, E2A, LMO1/2, GATA3, and RUNX1 bind a region 4-kb upstream of the miR-223 transcriptional start site. (A) ChIP coupled to massively parallel DNA sequencing (ChIP-seq) was performed for TAL1, HEB, E2A, and LMO1 or LMO2, GATA3, and RUNX1 (Sanda et al., 2012). Gene tracks represent binding at the miR-223 locus in a human primagraft sample (Prima 5) and the T-ALL cell lines CCRF-CEM (top) and Jurkat (bottom), with the miR-223 locus shown below. Y-axis values represent the number of sequence reads. (B) Enrichment of miR-223 regions indicated in A in four T-ALL cell samples (Jurkat, CCRF-CEM, Prima 2, and Prima 5) was analyzed by ChIP-qPCR. Mean ± SD fold enrichment as normalized to binding at the NANGO promoter (negative control) is shown from two independent experiments performed in triplicate.

miR-223 and TAL1 expression are closely correlated during normal thymic development and in T-ALL
Based on our evidence that the TAL1 complex directly and positively regulates miR-223, we investigated whether this
Figure 3. TAL1 expression correlates with miR-223 expression in human T-ALL and during normal thymic development. (A) TAL1 protein expression as determined by Western blot in T-ALL cell lines. The double band is likely to reflect differing TAL1 isoforms. A representative blot from two independent experiments is shown. (B) Comparison of miR-223 expression as determined by qRT-PCR in T-ALL cells positive or negative for TAL1. Red bars indicate presence of an activating SIL-TAL1 deletion. Bars represent mean ± SEM from two independent experiments performed in triplicate. miR-223 expression is given as the percentage of RNU5A control. ***, P < 0.001 by two-tailed Student’s t test. (C) miR-223 expression as determined by qRT-PCR in 44 pediatric T-ALL patients according to TAL1 expression (n = 22 TAL1-positive; n = 22 TAL1-negative). Horizontal bars represent mean with SEM. TAL1 status was determined by qRT-PCR. miR-223 expression is given as the percentage of RNU5A control. ***, P < 0.001 by Mann-Whitney-Wilcoxon test. Expression levels of TAL1 (D) and miR-223 (E) as determined by qRT-PCR from thymic subsets sorted from mice (Pearson’s correlation, r = 0.89;
through an increase in apoptotic cells as determined by Annexin V staining (Fig. 4 E). The most dramatic inhibition of cell growth was observed in MOLT-16 cells that express the highest levels of miR-223 and are FBXW7 wild-type, whereas miR-223 inhibition had no effect on the cell growth or survival of Be-13 cells that harbor a biallelic deletion of FBXW7 (Fig. 4, D and E), indicating that miR-Zip-223 does not have antiproliferative effects other than those caused by FBXW7 reexpression (O’Neil et al., 2007). When we retrovirally overexpressed miR-223 in Jurkat cells, it rescued most of the phenotype induced by miR-ZIP-223, arguing against an off-target effect by the construct (Fig. 4 F).

We and others have shown that knockdown of TAL1 slows the growth of TAL1-positive T-ALL cell lines through the induction of apoptosis without affecting the cell cycle (Pali et al., 2011; Sada et al., 2012). We thus tested whether the decrease in miR-223 expression observed after TAL1 knockdown was responsible, at least in part, for growth inhibition and apoptosis. We therefore infected the doxycycline-inducible TAL1 knockdown Jurkat cells with a retroviral vector expressing miR-223, such that miR-223 expression was no longer under the control of TAL1 in these cells (termed Jurkat-223 cells). Remarkably, retroviral expression of miR-223 partially rescued the growth inhibitory effects induced by doxycycline-induced TAL1 knockdown, whereas a control miR-223 construct in which four bases of the miR-223 seed sequence had been mutated (termed Jurkat-mut223) was unable to rescue cell growth (Fig. 5 A). Accordingly, levels of Annexin V staining and activation of caspases 3 and 7 were significantly lower in Jurkat-223 cells after TAL1 knockdown as compared with control cells (Fig. 5, B and C). Importantly, the degree of TAL1 knockdown was equivalent between Jurkat-223 and Jurkat-mut223 cells (Fig. 5 D). Thus, as a key downstream target of TAL1, miR-223 is essential for the survival and optimal growth of TAL1-positive T-ALL cells.

TAL1 regulates the FBXW7 tumor suppressor through miR-223
To identify candidate mRNA targets that could mediate the effects of miR-223 on cell survival and growth, we interrogated computational databases (TargetScan and microRNA.org; John et al., 2004; Lewis et al., 2005) and published studies (Xu et al., 2010; Lerner et al., 2011; Mavrakis et al., 2011). The highest ranked target of miR-223 predicted by TargetScan is the FBXW7 tumor suppressor, by virtue of three highly conserved and one poorly conserved binding site in its 3′UTR (Fig. 6 A). Notably, the colorectal carcinoma line HCT-116ΔOcer1−/− shows marked up-regulation of FBXW7 protein expression, consistent with its regulation by miRNAs (Xu et al., 2010; Lerner et al., 2011). Furthermore, in a miRNA library screen to identify miRNAs that can regulate a luciferase reporter containing the FBXW7 3′UTR, miR-223 exhibited the most potent suppression of luciferase activity, a finding consistent with other studies (Lerner et al., 2011; Xu et al., 2010). To demonstrate that this interaction is functional in human T-ALL, we stably overexpressed miR-223 in SUP-T1 and DND41 T-ALL cells, which lack TAL1 and have low miR-223 levels, and then analyzed endogenous FBXW7 protein expression by immunoprecipitation/immunoblot. As expected, FBXW7 expression was markedly down-regulated by miR-223 overexpression in both cell lines, confirming FBXW7 as a target of miR-223 in human T-ALL cells (Fig. 6 B). Additionally, inhibition of miR-223 in TAL1-positive cells with the miR-Zip-223 lentivirus resulted in reexpression of FBXW7, indicating its protein levels are repressed by miR-223 in these cells (Fig. 6 C).

FBXW7, an E3 ubiquitin ligase, targets a host of oncoproteins, including MYC, MYB, CYCLIN E, MCL-1, MTOR, and NOTCH-1 for proteasomal degradation, and is an important tumor-suppressor in T-ALL, with loss-of-function mutations present in 20% of patients (Maser et al., 2007; O’Neil et al., 2007; Thompson et al., 2007; Welcker and Clurman, 2008). Furthermore, Fbxw7 knockout mice develop T-ALL (Onoyama et al., 2007). Given the dynamic regulation of miR-223 by TAL1, we hypothesized that TAL1 represses FBXW7 by controlling miR-223 expression. Indeed, knockdown of TAL1 with two independent shRNAs in two TAL1-positive cell lines led to a significant increase in FBXW7 protein expression (Fig. 6 D). This effect was not mediated by direct regulation of FBXW7 transcription by TAL1, as we did not detect changes in FBXW7 mRNA levels after TAL1 knockdown (Sada et al., 2012).

We also examined known oncogenic substrates of FBXW7, to further strengthen the evidence that FBXW7 is the primary mediator of cell growth inhibition downstream of TAL1 and its direct target miR-223. After the addition of doxycycline to Jurkat cells harboring a doxycycline-inducible TAL1-specific shRNA, the reduction in TAL1 was associated with re-expression of FBXW7 and a striking reduction in the protein levels of MYC, intracellular NOTCH1 (ICN-1), MYB, and CYCLIN E (CCNE1), consistent with reactivation of FBXW7 activity (Fig. 7 A). However, we observed only marginal reductions in MCL-1 protein and no changes in MTOR levels, which may relate to cell type-specific aspects of the control of degradation of these FBXW7 substrates in T-ALL.

Together, our results suggest a model in which the TAL1 complex directly regulates the expression of miR-223, leading to its aberrant expression in transformed DP thymocytes, where it represses the ubiquitin ligase FBXW7, a tumor suppressor that can degrade oncoproteins with critical functions in the pathogenesis of T-ALL (Fig. 7 B).
et al., 2010; Palii et al., 2011; Sanda et al., 2012). In the T cell lineage, TAL1 has preferential binding to E-boxes with adjacent GATA, ETS, and RUNX motifs and regulates genes involved in T cell differentiation (Palii et al., 2011). It also targets several downstream oncogenes, including MYB, TRIB2, and STAT5A (Sanda et al., 2012). Although several other genes have been identified as direct targets of TAL1 and its regulatory partners in human T-ALL, such as pre-T cell receptor

DISCUSSION
The t(1;14)(p34;q11) translocation involving TAL1 in T-ALL was first described over 20 yr ago (Finger et al., 1989; Bernard et al., 1990; Xia et al., 1992), yet the mechanisms through which TAL1 is able to transform thymocytes are only beginning to be dissected. Studies from our laboratory and others using ChIP-seq analysis have shed light on the regulatory networks activated during malignant transformation by TAL1 (Wilson et al., 2010; Palii et al., 2011; Sanda et al., 2012). In the T cell lineage, TAL1 has preferential binding to E-boxes with adjacent GATA, ETS, and RUNX motifs and regulates genes involved in T cell differentiation (Palii et al., 2011). It also targets several downstream oncogenes, including MYB, TRIB2, and STAT5A (Sanda et al., 2012). Although several other genes have been identified as direct targets of TAL1 and its regulatory partners in human T-ALL, such as pre-T cell receptor
α (p10x), aldehyde dehydrogenase 1 member A2 (ALDH1A2), NK3 homeobox 1 (NKX3-1), and cyclin-dependent kinase 6 (CDK6), the relative contributions of these targets to leukemogenesis remain unclear (Bernard et al., 1998; Ono et al., 1998; Herblot et al., 2000; Palomero et al., 2006; Kusy et al., 2010; Palii et al., 2011; Sanda et al., 2012).

Genome-wide analysis of transcription factor binding using ChIP-seq has made it possible to assess the direct transcriptional regulation of miRNAs within the genome as a contributing factor to carcinogenesis. Here, we used this approach to address the possibility that TAL1-regulated miRNAs contribute to the pathogenesis of T-ALL driven by this oncoprotein. Of 22 miRNAs that changed expression on TAL1 knockdown, we identified five as direct targets of the TAL1 complex, including miR-26a, miR-29c, miR-181a*, miR-620, and miR-223. Of these, miR-223 was selected for further study because it was most highly down-regulated upon TAL1 knockdown in several different T-ALL cell lines.

Our results are in accord with the notion that TAL1-mediated oncogenesis occurs in an anachronistic manner, i.e., the physiological target genes of TAL1 are activated at a stage of thymocyte differentiation in which they are normally silent. Thus, we found evidence of TAL1-binding at the miR-223 locus in HSCs, and there was a close correlation between TAL1 and miR-223 expression throughout normal thymocyte maturation, suggesting that the regulation of miR-223 by TAL1 is part of a conserved developmental pathway. These findings may explain the high levels of miR-223 that have been described in T-ALL cases with features characteristic of an immature cell of origin (Chiaretti et al., 2010; Van Vlierberghe et al., 2011). Considering that miRNA expression during T cell development is tightly regulated (Neilson et al., 2007; Ghisi et al., 2011; Kirigin et al., 2012), the expression of miR-223 in DP thymocytes would be expected to aberrantly down-regulate the expression of several key proteins, including FBXW7, that are not normally regulated by this miRNA at such a developmental time point.

miR-223 was one of the first miRNAs shown to be expressed by hematopoietic cells and until recently was considered a myeloid-specific miRNA (Chen et al., 2004). In acute promyelocytic leukemia, ATRA induces the expression of CCAAT/enhancer binding protein-α (C/EBPA), which up-regulates miR-223 through binding to its promoter (Fazi et al., 2005). miR-223 then acts to repress nuclear factor-IA (NFI-A), thereby contributing to myeloid differentiation (Fazi et al., 2005). Whether the regulation of miR-223 by TAL1 contributes to early myeloid lineage commitment would be an interesting area for future study because the depletion of TAL1 in HSCs leads to a reduction in myeloid cells through an unknown mechanism (Brunet de la Grange et al., 2006; Zardo et al., 2012).

Using a combination of miRNA expression profiling of primary T-ALL patient samples and an unbiased library screen...
Figure 6. TAL1 represses FBXW7 via miR-223 in TAL1-positive T-ALL. (A) Binding sites for miR-223 and the FBXW7 3′UTR are shown for three conserved and one poorly conserved site (bottom), as determined by the TargetScan algorithm. Position numbers refer to distance from the FBXW7 stop codon. (B) TAL1-negative miR-223 low T-ALL cell lines, SUP-T1 and DND-41, were retrovirally transduced to overexpress miR-223, and FBXW7 protein expression determined by immunoprecipitation/immunoblot. Loading controls are 10% of immunoprecipitation input. A representative blot from two independent experiments is shown. (C) FBXW7 protein expression was determined after transduction of TAL1-positive CCRF-CEM and Jurkat cells with miRZip-223 compared with a miRZip control. A representative blot from two independent experiments is shown. (D) TAL1 was depleted in TAL1-positive MOLT-16 and RPMI-8402 cells by lentiviral knockdown with two independent shRNAs, and FBXW7 protein expression was determined by immunoprecipitation/immunoblot. A representative blot from two independent experiments is shown.

of miRNAs that were able to rescue lymphocytes from cytokine withdrawal, Mavrakis et al. (2011) recently identified several oncogenic miRNAs in T-ALL. Of the miRNAs that scored on their screen, miR-223 was the most abundantly expressed in T-ALL patients and accelerated Notch-induced leukemia in a mouse model, although the mechanism leading to its dysregulation in human T-ALL was not established (Mavrakis et al., 2011). It is notable that in this study miR-223 had a pro-survival role in lymphocytes. Similarly, we found that inhibition of miR-223 in T-ALL induced apoptosis, suggesting T-ALL cells are dependent on its sustained expression. Moreover, in an inducible TAL1 knockdown system, we found that miR-223 undergoes significant down-regulation when TAL1 expression is blocked, and that retroviral transduction
several groups have shown that miR-223 regulates FBXW7 expression in a variety of cellular contexts (Lerner et al., 2011; Mavrakis et al., 2011; Xu et al., 2010). Here, we show miR-223 can also regulate FBXW7 protein expression in human T-ALL cells. The reactivation of FBXW7 protein expression observed after \emph{TAL1} knockdown indicates it is actively suppressed by \emph{TAL1} through its regulation of miR-223, although we cannot rule out the presence of other miR-223 targets important to the malignant phenotype. Regulation of FBXW7 positions \emph{TAL1} at the top of a genetic network that directly stabilizes oncoproteins such as c-MYC, MYB, and NOTCH that have vital roles in T-ALL pathogenesis. FBXW7 is known to play a prominent role as a tumor suppressor in T-ALL, and our paper firmly establishes a major mechanism leading to its down-regulation by \emph{TAL1} in the majority of T-ALL cases that lack gene-specific FBXW7 inactivating mutations or deletions. Our paper implicates a carcinogenic pathway involving over-expression of an oncogenic transcription factor in the coordinate down-regulation of a key tumor suppressor through a miRNA intermediate, and further study will establish whether this mechanism is more broadly used in cancer biology.

**MATERIALS AND METHODS**

**T-ALL cell samples.** Human T-ALL cell lines were maintained as described previously, and the identity of all cell lines was confirmed by short-tandem repeat analysis (Sanda et al., 2010). Diagnostic T-ALL samples from \emph{TAL1}-positive cases were obtained with informed consent and Institutional Review Board approval from children treated on Dana Farber Cancer Institute study 05–01. The development of primagraft samples for ChIP-seq analysis has been previously described (Sanda et al., 2012). RNA was obtained at presentation from 44 pediatric T-ALL patients as previously reported (Ferrando et al., 2002).

**Microarray expression analysis for mRNA or miRNA.** For global miRNA expression analysis, Jurkat cells were harvested at 48 h after lentiviral transduction with \emph{TAL1} shRNAs 1 and 2, or control shRNAs targeting GFP and luciferase, with each shRNA performed in duplicate. Efficient \emph{TAL1} knockdown was assessed by Western blotting. miRNAs were isolated and purified with the High Pure miRNA isolation kit (Roche). Global miRNA expression profiling was performed with the Exiqon miRCURY LNA fifth generation platform (miRBase version 14.0). The difference in log2 median ratios (Hy3/Hy5) between sample groups was calculated and significance was determined by two-tailed Student’s \emph{t} test. \emph{P} values of \leq 0.05 were considered to indicate statistical significance. For mRNA expression analysis, total RNA samples were extracted by TRIzol (Invitrogen) from T-ALL cell lines grown in parallel under identical conditions and purified by RNeasy Mini kit (QIAGEN). Gene expression was analyzed by Affymetrix HG U133 2.0 plus microarrays.

**ChIP.** ChIP was performed according to previously described methods (Lee et al., 2006). The antibodies and detailed ChIP conditions and analysis parameters have been described in detail (Sanda et al., 2012). Data showing \emph{TAL1} binding and histone modifications in human HSCs were from previous studies (Cui et al., 2009; Novershtern et al., 2011). Relative enrichment of the miR-223 promoter was determined by ChIP quantitative PCR as described Sanda et al. (2012) using the primers miR-223-F1 5’-CAGAAA-GCCCAATTTCTTCT-3’ and miR-223-R1 5’-GGGCAAATGGATACCAG-TAG-3’ and NANOGrp-R1 5’-CCACGAGACCGTTAAAAATCC-3’.
Isolation of murine T cell subsets. For RNA isolation from DP and SP thymocyte subpopulations, cells from 6-wk-old C57BL/6 mice were stained for CD4 (RM4-5), CD8 (53-67.7), and TCRβ (H57-597). Thymocytes with CD4+CD8− (DP), TCRβ+CD4+CD8− (CD4 single-positive, CD4SP) and TCRβ-CD4+CD8− (CD8 single-positive, CD8SP) phenotypes were sorted. For CD4/CD8 double-negative (DN) thymocyte subpopulation isolation, cells were first stained with biotinylated antibodies to CD4 and CD8, before CD4+CD8+ deletion using streptavidin-conjugated magnetic beads (Invitrogen). Cells were then stained with fluorochrome-conjugated antibodies against CD25 (PC61), CD44 (IM7), CD28 (E18), and cKit (2B8). Thymocytes with CD4+CD8− cKit+CD44+CD25+ (early T cell progenitor), CD4+CD8− cKit−CD44+CD25− (DN2), CD4−CD8+ cKit−CD44− cKit−CD28− (DN3a), CD4−CD8− cKit−CD44+ cKit−CD28− (DN3b) phenotype were sorted. All cell sorts were performed with a FACSAria instrument (BD).

Generation of plasmids. The pMX-miR-GFP/PURO retroviral expression vector was obtained from Cell Biologics Inc. (RTV-017). A 310-bp fragment including premiR-223 was amplified by PCR from MOLT-16 DNA using the primers Xhol-miR-223-F 5′-TCCAGCTAGCTACCCGGGCTA-AGGGTGTGA-3′ and Xhol-miR-223-R 5′-TCCAGCTAGCTACAAGTGCA-CATAAATACCTGTGGA-3′. PCR products were cloned into the Xhol site of the pMX-mir-GFP/PURO vector to generate pMX-miR-223-GFP/PURO. To construct a mutant miR−223 vector, the overlapping primer technique was used by first amplifying MOLT-16 DNA with Xhol-miR-223-F and MUT-miR-223-R 5′-ATTGCAGCAAGGGCCTGGTTGAAGAAGAT-3′, a separate PCR using MUT-miR223-F (5′-GATGCCTGGTTTGGTGTGAG-3′) and Xhol-miR-223-R, followed by a third PCR to anneal the first two fragments using primers Xhol-miR-223-F and Xhol-miR-223-R. The 310-bp fragment was then cloned into the Xhol site of pMX-miR-GFP/PURO vector to generate pMX-miR-MUT223-GFP/PURO. The mature miRNA of this construct (5′-TCCGCC- GGTTGCTAATCCGCCAACA-3′) has no predicted targets by TargetScan. The proof-reading enzyme Phusion (New England Biolabs) was used for all reactions and all plasmids were verified by sequencing.

Western blots and immunoprecipitation. Whole-cell lysates were prepared in RIPA buffer. Immunoblotting was performed with the following antibodies: mouse monoclonal anti–β-Tubulin (clone B3-TL73; Millipore), mouse monoclonal anti–α-tubulin antibody diluted 1 in 1,000 (clone B-5-1-2; Sigma-Aldrich), mouse monoclonal anti–GAPDH (Santa Cruz Biotechnology, Inc.), mouse anti-CCNE1 (HE12; Santa Cruz Biotechnology), rabbit anti-MCL-1 (S-19; Santa Cruz Biotechnology), mouse anti-MYB (05–175, Millipore), rabbit anti-cleaved NOTCH1 (Val1744; Cell Signaling Technology), rabbit anti-MTOR (C25; Cell Signaling Technology), rabbit anti-MCL-1 (S-19; Santa Cruz Biotechnology, Inc.), mouse anti-CCNE1 (HE12; Santa Cruz Biotechnology). For immunoprecipitation experiments, 750 ng rabbit FBXW7 antibody (Biotechnolog, Inc.) was incubated with 100 µg whole-cell lysate on a rotator overnight at 4°C. Beads were washed five times on a magnet and eluted according to the manufacturer’s instructions (Dynabeads Co-immunoprecipitation kit; Invitrogen). Proteins were immunoblotted with FBXW7 antibody (A301-720A; Bethyl Laboratories Inc.) covalently coupled to Dynabeads (Invitrogen) was incubated with 100 µg whole-cell lysate on a rotator overnight at 4°C. Beads were washed five times on a magnet and eluted according to the manufacturer’s instructions (Dynabeads Co-immunoprecipitation kit; Invitrogen). Proteins were immunoblotted with FBXW7 antibody (A301-720A; Bethyl Laboratories Inc.) diluted 1 in 1,000. As a loading control, 10% of immunoprecipitation input was immunoblotted with anti–β-actin or anti–α-tubulin antibodies. Secondary horseradish peroxidase-linked IgG antibodies to mouse or rabbit diluted 1 in 10,000 were obtained from Cell Signaling Technology.

shRNA knockdown and doxycycline-inducible TAL1 knockdown cells. The shRNA sequences directed against TAL1, expressed in the p.KO.1-puro vector, and production of lentivirus have been previously described (Sanda et al., 2012), as have the doxycycline-inducible TAL1 knockdown Jurkat cells (S1C1 clone; Pali et al., 2011). Jurkat S1C1 cells were maintained under selection in blasticidin and G418. After retroviral transduction with pMX-miR-GFP/PURO, cells were also maintained in puromycin. Doxycycline was used at 6.25 µg/ml. The level of knockdown was verified by qRT-PCR for RNA or by Western blotting.

RNA extraction, cDNA, and expression analysis. miRNAs were isolated and purified with the High Pure miRNA isolation kit (Roche) and reverse transcribed with the miScript kit (QIAGEN), and their expression was quantitated with the miScript SyberGreen kit using a miScript universal primer and miR-223-specific primer (MS00003871; QIAGEN). Expression was normalized to expression of RNU5A (MS00013993; QIAGEN). Both the miR-223 and RNU5A primers are compatible with human and mouse gene orthologues. To assess mRNA knockdown, a small fraction of cells was collected and mRNA was extracted by TRizol (Invitrogen), followed by column purification using the RNeasy Mini kit (QIAGEN). Purified RNA was reverse-transcribed with the miScript kit (QIAGEN). Quantitative real-time PCR was performed with the AB7300 Detection System (Applied Biosystems) using gene-specific primers and Power SYBR Green PCR Master Mix (Applied Biosystems). Primer sequences for qRT-PCR were as follows: human TAL1-F, 5′-AGGGCCTGTTGGAAGGATG-3′; human TAL1-R, 5′-AAGTAAGGGGCGCTGTTGGT-3′; murine Tel-1-F, 5′-CCCCCCATGTTCACCCAAACAC-3′; murine Tel-1-R, 5′-CCGCATACCTTTGTTGAG-3′; GAPDH-F, 5′-TGCAACACCACTGTTAGC-3′; GAPDH-R, 5′-GCCATGGACTGTGGTCAATGAC-3′. Among T-ALL cases, TAL1 expression exhibited a bimodal distribution, and patients were determined as TAL1-positive if TAL1 transcripts were >3.7% of GAPDH.

Apoptosis assays. Annexin V staining was assessed by flow cytometry using Annexin V-APC antibody (BD) according to the manufacturer’s recommendations. Cleaved caspase-3 and -7 activity was determined by luminescence using the Caspase-Glo 3/7 assay from Promega after normalization for cell number.

Accession nos. Expression data and ChIP-seq can be found in the Gene Expression Omnibus under superseries accession no. GSE29181, and chromatin marks and RNA Pol II ChIP-seq data from Jurkat are found under accession no. GSE33850. Chromatin marks and RNA Pol II in human hematopoietic stem and progenitor cells are found under accession no. GSE26015, mRNA expression data are available under accession no. GSE46957.

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