Introduction

CD4+ regulatory T cells (Tregs) have been shown to be essential for the development and the maintenance of peripheral tolerance and immune homeostasis [1]. Indeed, Treg dysfunction is associated with allergy, autoimmunity, cancer or early graft rejection [2]. In the context of infectious diseases, Tregs have the potential to limit excessive inflammatory immune responses, thereby reducing tissue damage, but can also suppress antimicrobial immune responses and promote pathogen persistence [3].

The role of Tregs during HIV-1 infection remains controversial [4,3,6]. During the course of HIV-1 disease progression, microbial translocation from the gut, viral factors and co-infections such as human cytomegalovirus (hCMV) have emerged as the major causes of persistent immune activation and have been associated with mortality and non-AIDS morbidity [7]. In this context, Treg activity could have a beneficial effect through suppression of generalized chronic immune activation, but also through inhibition of activated CD4+ T cells and subsequent control of viral replication, as demonstrated by Moreno-Fernandez et al. [8].

Regulatory T Cells Expanded from HIV-1-Infected Individuals Maintain Phenotype, TCR Repertoire and Suppressive Capacity

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Abstract

While modulation of regulatory T cell (Treg) function and adoptive Treg transfer are being explored as therapeutic modalities in the context of autoimmune diseases, transplantation and cancer, their role in HIV-1 pathogenesis remains less well defined. Controversy persists regarding their beneficial or detrimental effects in HIV-1 disease, which warrants further detailed exploration. Our objectives were to investigate if functional CD4+ Tregs can be isolated and expanded from HIV-1-infected individuals for experimental or potential future therapeutic use and to determine phenotype and suppressive capacity of expanded Tregs from HIV-1 positive blood and tissue. Tregs and conventional T cell controls were isolated from blood and gut-associated lymphoid tissue of individuals with HIV-1 infection and healthy donors using flow-based cell-sorting. The phenotype of expanded Tregs was assessed by flow-cytometry and quantitative PCR. T-cell receptor ß-chain (TCR-ß) repertoire diversity was investigated by deep sequencing. Flow-based T-cell proliferation and chromium release cytotoxicity assays were used to determine Treg suppressive function. Tregs from HIV-1 positive individuals, including infants, were successfully expanded from PBMC and GALT. Expanded Tregs expressed high levels of FOXP3, CTLA4, CD39 and HELIOS and exhibited a highly demethylated TSDR (Treg-specific demethylated region), characteristic of Treg lineage. The TCRß repertoire was maintained following Treg expansion and expanded Tregs remained highly suppressive in vitro. Our data demonstrate that Tregs can be expanded from blood and tissue compartments of HIV-1+ donors with preservation of Treg phenotype, function and TCR repertoire. These results are highly relevant for the investigation of potential future therapeutic use, as currently investigated for other disease states and hold great promise for detailed studies on the role of Tregs in HIV-1 infection.


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contrast, Tregs may play a detrimental role through inhibition of anti-HIV-1 immune responses [9,10,11,12], thus promoting HIV-1 persistence at the host’s expense. HIV-1 infection appears to directly and indirectly modulate Tregs in vivo, as suggested by data demonstrating that individuals with chronic HIV-1 infection have higher Treg frequencies than individuals who control HIV-1 infection and healthy control subjects [13,14]. This observation has not been fully elucidated to date, but could be explained by preferential survival, tissue redistribution, increased proliferation, or conversion of non-regulatory T cells into Tregs in chronic HIV-1 infection [4,15].

One of the main challenges for detailed functional analyses of Tregs in HIV-1 disease and their potential for future clinical application is the paucity of the natural Treg population in human peripheral blood, where thymus-derived Treg represent roughly 1–10% of the mature CD4+ T cell pool [16]. This poses an even greater challenge in progressive HIV-1 infection, where chronic viral replication and immune activation contribute to profound CD4+ T cell loss [17]. The functional characterization of Tregs in individuals with advanced HIV-1 disease, HIV-1-infected infants, for which only very small volume samples can be obtained, or from lymphoid or mucosal tissue sites where sample size is often limited, is therefore difficult. Based on our previous data demonstrating that ex vivo suppressive function of freshly isolated Tregs was preserved in HIV-1 positive individuals [13], we hypothesized that functional Tregs can be expanded in vitro from HIV-1-infected blood and tissue with preservation of phenotype and suppressive capacity.

We here describe the successful isolation and in vitro expansion of functional CD4+ Tregs from HIV-1-infected individuals, including HIV-1 controllers, individuals with progressive untreated HIV-1 infection, small volume specimen from HIV-1-infected infants and biopsies of gut-associated lymphoid tissue (GALT). Expanded Tregs were highly suppressive and exhibited an activated Treg phenotype with high expression of Treg markers and a demethylated TSDR, suggesting functional Treg lineage as opposed to activation-induced FOXP3 expression. We believe that our findings are of high relevance for potential future therapeutic exploration of Tregs and in addition will allow for more detailed investigations into the role and function of Tregs in HIV-1 disease.

**Methods**

**Study subjects**

The study was approved by the Institutional Review Board of the Massachusetts General Hospital (MGH, Boston, MA) and was conducted in accordance with the MGH human experimentation...
Written informed consent was obtained for all study participants.

Blood samples were drawn from 10 HIV-1 controllers with asymptomatic HIV-1 infection who maintained a plasma viremia below 300 copies/ml (median CD4 count: 779 cells/µl, interquartile range [IQR]: 526–1,084) in the absence of antiretroviral therapy, 13 individuals with chronic untreated HIV-1 infection (median viral load: 41,800 RNA copies/ml, IQR: 6,030–118,730 and median CD4 count: 362 cells/µl, IQR: 283–616) (Table 1) and a vertically HIV-1-infected infant (age: 511 days, viral load: 1,977,540 RNA copies/ml, CD4 count: 757). Blood samples from 5 HIV-1 uninfected individuals were studied as control specimen. Gut biopsies from 1 HIV-1-negative and 3 HIV-1-infected individuals (1 on antiretroviral therapy, 2 elite controllers) were also used in this study.

**Isolation of T cell subsets from peripheral blood and GALT**

CD4⁺ T Cell-enriched PBMC were isolated from peripheral blood by density centrifugation using the RosetteSep enrichment kit (Ficoll-Histopaque; Sigma-Aldrich and STEMCELL Technologies) and labeled with anti-CD3-PE-Cy7 (BD Pharmingen), CD4-FITC (eBioscience), CD25-APC (eBioscience), CD127-PE (BD Pharmingen).

Cryopreserved PBMC samples were stained using the same panel as described above except for the addition of an exclusion channel to select for viable cells (Invitrogen).

Pinch biopsies from HIV-positive individuals were obtained by endoscopy and an ileum biopsy from an HIV-negative individual was obtained from a laparoscopic small bowel resection. All gut samples were provided by the Ragon Institute tissue platform. After collection, biopsies underwent two rounds of collagenase type II (Sigma-Aldrich) digestions followed by filtration [18] and were stained with the same panel as the fresh PBMC described above.

CD3⁺CD4⁺CD25⁺CD127low Treg and CD3⁺CD4⁺CD25⁻CD127⁺ Tconv controls subsets were sorted on a FACS Aria cell sorter (BD Biosciences) equipped for handling biohazardous material.

**Expansion of CD4⁺ Tregs and conventional T cells (Tconv)**

Tregs and Tconvs were activated with anti-CD3/anti-CD28-coated microbeads (Invitrogen) at a 1:1 bead-to-cell-ratio. On day 2, media volume was doubled and exogenous IL-2 was added (300 U/ml, NIH Aids Research & Reference Reagent Program)

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**Figure 1. Cell sorting, FOXP3 TSDR and gene expression.** A. Flow-cytometry gating strategy used to isolate CD25⁺CD127low regulatory T cells (Tregs) and CD25⁺CD127⁺ conventional T cells (Tconv) from CD4⁺ T cells (Left Panel). Expansion fold change of Tregs isolated from HIV-1-infected (dark grey) (n = 8 controllers + 13 chronic untreated) and healthy (light grey) (n = 4) individuals during 7 days of cell culture (right Panel). B. Relative mRNA expression in arbitrary units (A.U.) of FOXP3 and IL-10 quantified by real time PCR in expanded Tregs (n = 3 controllers+2 chronic untreated) and Tconvs (n = 2 controllers+2 chronic untreated) isolated from HIV-1 infected individuals after 7 days of culture. C. Frequency of demethylation of the Treg Specific Demethylation (TSDR) region of the FOXP3 gene in expanded Tregs and Tconvs after 7 days of culture as assayed by real time PCR. Empty symbols represent HIV-1 controllers and solid symbols HIV-1 chronic untreated individuals.

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Figure 2. Phenotyping of expanded Tregs by flow cytometry. A. Representative examples of gating strategy used for CD25⁺FOXP3⁺ staining by flow-cytometry of ex vivo PBMC (upper panel) isolated from a HIV-1 controller and matched expanded Tregs (lower panel) at day 7 of expansion. B. Expression of different Tregs markers quantified by flow-cytometry of expanded (day 7) and ex vivo unexpanded Tregs and Tconvs. MFI = Mean Fluorescence intensity. Empty symbols represent HIV-1 controllers and solid symbols HIV-1 chronic untreated individuals. C. Representative example of flow-cytometry gating strategy used to phenotype Tregs, Tconvs (n = 3 controllers+9 chronic untreated) and ex vivo CD4 T cells (n = 3 controllers+3 chronic untreated) isolated from HIV-1 positive individuals based on their CD45RA and FOXP3 expression profiles [41]. The left dot plot shows ex vivo
CD4⁺ T cells from PBMC, the middle dot plot represents an example of expanded Tregs (black dots) and Tconvs (light grey dots). The right histogram graph quantifies the different Treg subsets in HIV-1 positive individuals. Gate 1 and white columns represent "resting" CD45RA⁺FOXP3⁺ Tregs, gate 2 and grey columns represent "non-suppressive cytokine-secreting" CD45RA⁻ FOXP3⁺ T cells and gate 3 and black columns represent "activated" CD45RA⁻ FOXP3⁺ Tregs.

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[19]. On day 5, cells were counted and fresh media added. On day 7 expanded T cells were assayed for their suppressive function and the remaining cells were cryopreserved for further analysis.

Immunophenotyping of T cell subsets by flow-cytometry

Cryopreserved expanded Tregs and Tconvs were thawed and immunostained with anti-CD3-PE-Cy7, anti-CD4-pqlot655 (Invitrogen), anti-CD25-PE-Cy5 (Biosciences), anti-CD39-FITC (Biosciences), anti-CD45RA-horizon v450 (BD Pharmingen), anti-FOXP3-PE (clone PCH101, Biosciences), anti-CTLA4-APC (BD Pharmingen), anti-HELIOS-FITC (Biolegend). For intracellular staining, the eBioscence FOXP3 staining buffer kit was used. Dead cells were eliminated using the LIVE/DEAD® Fixable Blue Dead Cell Stain Kit (Invitrogen). Flow-cytometry data were acquired on a LSR Fortessa (BD Biosciences).

RNA isolation and real-time RT-PCR

RNA was isolated using the RNeasy Plus Kit (Qiangen) and retro-transcribed using the SuperScript III Reverse Transcriptase (Invitrogen). Primers for FOXP3 (forward: 5'-CACGCCAGTCTCAGATGGCTC-3' and reverse: 5'-GGGTGTTAAACCAGGGTGGATAC-3') and IL10 (forward: 5'-GGCGCTGTCATCGATTTCCTC-3' and reverse: 5'-GATAGTGCCACCCTGATGAG-3') were designed using Primer3 [20] and chosen to span an exon-exon junction. Real-time PCR was performed in a Roche Applied Science LightCycler 480 using the SYBR Green I dye (Roche) and the SYBR Green I Master Kit (Roche). RNA polymerase II (forward: 5'-GGATCAGTTCTTTGGTTCAGCA-3' and reverse: 5'-GTTCAITTCACCTGCACACAG-3') gene expression was used to normalize the data by the Pfaffl method [21].

Epigenetic analysis and TCR sequencing

Genomic DNA was isolated from Tregs and Tconvs using the DNeasy Blood & Tissue Kit (Qiangen). Quantification of TSDR demethylation by real-time PCR was performed by Epiontis (Berlin, Germany) as previously described [22].

The TCR diversity of ex vivo unexpanded and in vitro expanded Tregs was analyzed using a next generation sequencing protocol (NGS) [23,24]. Briefly, RNA was isolated using the RNeasy plus kit (Qiangen) and cDNA was synthesized with SuperScript IV Reverse Transcriptase and oligo-dT primers (Invitrogen) [25]. Linear amplification of the cDNA was performed on a T3000 thermocycler (Biometra). The amplified samples were analyzed by NGS on the Genome Sequencer FLX (Roche) using the titanium platform. After TCR sequencing, the VB8-, JB6 variants and the CDR3 were identified.

Assessment of Treg suppressive function using CFSE proliferation assays

Cryopreserved PBMC were labeled with CFSE (Invitrogen) and cultured in the presence or absence of autologous CD4⁺ Tregs or Tconvs at day 7 of expansion with anti-CD2/anti-CD3/anti-CD28 microbeads (Miltenyi Biotec) at a 1:1 bead-to-cell-ratio. After 4 days of co-culture, cells were stained with anti-CD3-PECy7, anti-CD4-APC (BD Pharmingen) and anti-CD8-AF700 (BD Pharmingen).
the FOXP3 TSDR (Treg-specific demethylated region) using a real-time PCR assay has recently been described as a reliable method to quantify and distinguish regulatory T cells from conventional activated T cells [22]. The DNA of this region is found to be methylated in activated and resting non-regulatory T cells, while the FOXP3 TSDR of T cells from the regulatory

**Figure 3. The TCR repertoire is not altered after in vitro expansion of Tregs.** A. Degree of expansion of the TCRβ repertoire (i.e. number of TCRs in a sample that belongs to an individual clone and expressed as percentage of total reads) from $2 \times 10^6$ ex vivo sorted unexpanded (light grey) and $2 \times 10^6$ in vitro expanded (Day 14; dark grey) Tregs isolated from the same original PBMC specimen. B. Distribution of variable-gene (Vβ-gene) variants from $2 \times 10^6$ ex vivo sorted unexpanded (light grey) and $2 \times 10^6$ in vitro expanded (Day 14; dark grey) Treg TCR-β clones isolated from the same PBMC specimen. C. Distribution of joining-gene (Jβ-gene) variants from $2 \times 10^4$ ex vivo sorted unexpanded (light grey) and $2 \times 10^4$ in vitro expanded (Day 14; dark grey) Treg TCR-β clones isolated from the same PBMC specimen.

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Tregs Expanded from HIV-1+ Donors Are Functional

A

\[ \text{% CD}8^+ \text{T cell suppression} \]

\[ \text{HIV-} \quad \text{HIV+} \]

\[ \text{ns} \quad \text{**} \]

\[ \text{ns} \quad **** \]

B

CD8^+ T cells

PBMC + Tregs + Tconvs

CD4^+ T cells

CFSE

C

% lysis

Target alone

Target + CTL

Target + Tregs

Target + CTL + Tregs

D

CD25

CD127

3.66%

E

CD25

CD127

7%

300

Treg cell numbers (x10^3)

days of expansion

PBMC + Tregs + Tconvs

% CD4 T cell suppression

ns

*
lineage is constitutively demethylated in this region [35]. In our study the FOXP3 TSDR of expanded Tregs was highly demethylated, while it was found to be methylated in expanded Tconvs (Figure 1B, left). Expanded Tregs therefore revealed high expression levels of stable FOXP3, suggesting their origin derived from true functional regulatory T cell lineage.

We next sought to carefully characterize the phenotype of expanded Tregs by comparing ex vivo unexpanded and expanded Tregs and Tconvs using flow cytometry. Examples of CD25/FOXP3 coexpression in expanded and ex vivo unexpanded Tregs from HIV controllers are shown in Figure 2A. While mean fluorescence intensity of CD25 expression was similar, CD25 expression levels of FOXP3 were significantly higher in expanded Tregs and Tconvs compared to conventional T cells [38] and in HIV-1 infection, CD39 expression on Tregs was reported to correlate with disease progression [14]. Similarly to FOXP3, our data show higher CTLA4 and CD39 expression in expanded Tregs compared to conventional T cells (Figure 2B), suggesting that these differences of CTLA4, CD39 and FOXP3 expression levels between Tconvs and Tregs were maintained after expansion.

The Treg marker HELIOS has recently been suggested as a more specific marker of thymic-derived Tregs [39]. As observed in our study, the frequency of cells expressing this molecule was high in both ex vivo unexpanded and in vitro expanded Tregs (median 72.5%, IQR: 70.1-100.6%, and median 64.8% IQR: 55.2-77.8%, respectively). HELIOS was not expressed in expanded Tconvs (Figure 2B) and the decrease of HELIOS expression found after stimulation in our culture is in line with the decreased of HELIOS-expressing FOXP3+ Tregs described after in vitro stimulation in a previous study, suggesting that obtaining large numbers of FOXP3+HELIOS+ Tregs after several rounds of stimulation may require use of a stabilizing reagent [40].

In 2009, Miyara et al. proposed an elegant classification scheme for the functional delineation of human CD4+ T cells based on the expression of FOXP3 and CD45RA [41]. Using this classification and based on ex vivo CD4+ T cell comparison as a reference, expanded Tregs showed a high amount of CD45RA+ FOXP3high activated Tregs, while expanded Tconvs were mostly constituted of CD45RA- FOXP3low cytokine-secretting non-suppressive T cells (Figure 2C).

In summary, the high expression of FOXP3 bearing a demethylated TSDR, high CTLA4, CD39 and HELIOS as well as the CD45RA+ FOXP3high phenotype suggest that after 7 days of expansion the CD4+CD25+CD127low T cells represent Tregs of an activated phenotype.

Treg expansion did not significantly alter the TCR repertoire

After determination of the phenotype of expanded Tregs, we next investigated if in vitro expansion would alter T cell receptor diversity and selectively expand specific Treg clones. The next generation sequencing analysis of the Vß-CDR3-Jß region allows for identification of unique T cell clones [23]. We sequenced the Vß, Jß variants and the CDR3 regions of the TCR of ex vivo unexpanded and in vitro expanded Tregs in a subset of HIV-1-infected individuals. No specific individual clones were preferentially expanded in our study sample (Figure 3A) and the TCR-β V- (Figure 3B) and J-usage (Figure 3C) did not appear to significantly differ after expansion, suggesting that the use of anti-CD3/anti-CD28-coated beads did not significantly alter the breadth of the TCR-ß repertoire. These results support work by Hoffmann et al. who found that the TCR Vß-chain of Tregs in vitro stimulated with artificial antigen-presenting cells proliferated polyclonally and did not lose clonotypes [42].

Expanded Tregs from HIV-1-infected individuals potently suppress T cell proliferation and HIV-1-specific cytotoxicity

Tregs are ultimately defined through their suppressive capacity. We therefore next explored if expanded Tregs isolated from HIV-1-infected individuals remained suppressive after expansion using standardized flow-based proliferation assays [13], where CFSE-labeled activated responder cells were cultured in the presence or absence of expanded Tregs (or Tconvs controls). Our data demonstrate that expanded Tregs isolated from HIV-1-positive individuals have preserved potent suppressive capacity. In contrast, no significant suppression of proliferation was observed in the presence of expanded conventional T cells (Figure 4A, B). Expanded CD4+ Tregs isolated from HIV-1-positive and negative individuals did not show significantly different suppressive capacities (Figure 4A, B). Expanded Tregs isolated from controllers and chronic untreated HIV-1 infected individuals were also equal in their ability to suppress T cell proliferation in this experimental system (data not shown), in line with preserved ex vivo Treg function in these two patient populations, as previously described [13]. Moreover when compared to our previous study [13], the suppressive function of expanded Tregs and ex vivo unexpanded Tregs isolated from HIV-1 positive individuals were not significantly different.

Using expanded Tregs isolated from HIV-1 positive donors, we next tested their capacity to suppress the cytolytic function of HIV-1-specific cytotoxic T lymphocyte (CTL) clones in a 51Chromium release assay. Figure 4C shows a representative example of potent suppression by expanded Tregs of the cytotoxic activity of an
HIV-1-specific CTL clone after 4 h of co-culture at a ratio of 1:1 CTL/Treg/BCL target.

We here demonstrate that Tregs expanded from HIV-1-positive individuals retain their suppressive function in vitro, as shown by their capacity to suppress the proliferation of activated T cells and the cytolytic activity of HIV-1-specific CTL clones.

Expansion of Tregs from HIV-1-infected infant and gut-associated lymphoid tissue (GALT)

One of the major limitations in studying Treg biology in the context of HIV-1 infection is the limited amount of Tregs present in small volume samples. We therefore next investigated if functional Tregs can be expanded from tissue and small volume samples. Figures 4D and E show examples of Tregs isolated from the peripheral blood of an HIV-1-infected infant and from the colon of an HIV-1-infected adult. Using a flow-cytometry cell sorter we isolated $18 \times 10^6$ and $3.5 \times 10^3$ viable Tregs from $15 \times 10^6$ frozen PBMC and $110 \times 10^6$ cells isolated from fresh colonic tissue, respectively (gating is shown on Figure 4D and E, left). After 7 days, the cell number reached $2.9 \times 10^9$ (i.e. 161 fold-change) for the Tregs isolated from the infant specimen, whereas it reached $2.4 \times 10^6$ (i.e. 69 fold-change) after 9 days of culture of Tregs isolated from the adult GALT (Figure 4D and E, middle).

Suppressive function was quantified by flow-cytometry proliferation assays (Figure 4D and E, right) and showed that Tregs isolated from the peripheral blood of HIV-1-infected infants and the GALT of HIV-1-infected adults were functional and highly suppressive. In total Tregs from 5 gut samples (1 from a HIV-1-negative and 4 from HIV-1-positive individuals) were expanded and yielded similar results.

Discussion

Many unanswered questions related to Tregs in the context of HIV-1 immunopathogenesis remain and it is yet incompletely understood if this cell population contributes to promotion or prevention of disease progression. Studying Tregs in CD4+ T cell-depleted individuals has proven to be difficult in the context of limiting cell numbers and it is unknown to date, if Tregs can be expanded from HIV-1-positive individuals for experimental or potential future therapeutic use.

In the present study we describe for the first time the successful in vitro expansion of CD4+ regulatory T cells from HIV-1 positive individuals. Expanded Tregs from HIV-1-infected donors displayed the phenotype and function of genuine regulatory T cells with a preserved TCR repertoire. Expansion of functional Tregs isolated from different blood and tissue compartments of HIV-1 patients with preserved suppressive capacity suggests that these cells are not intrinsically defective in the context of HIV-1 infection. Indeed, when comparing the expanded Tregs isolated from HIV-1-positive individuals (HIV controllers and individuals with chronic HIV-1 infection) and healthy control subjects, no differences in their capacity to inhibit proliferation of activated lymphocytes were observed after in vitro expansion. These results support our previous studies demonstrating conserved suppressive function between Tregs isolated ex vivo from HIV-1 positive and negative individuals [13]. However, like these previously reported ex vivo functional data, our results do not exclude the possibility of impairment of in vivo Treg function during HIV infection, e.g. in a pathologically impaired tissue microenvironment, through disregulated interplay with antigen presenting cells such as dendritic cells [43] or loss of function as a result of direct HIV-1 infection [unpublished data], [44].

Our data also suggest that expansion of functional Tregs from HIV-infected individuals theoretically raises the possibility to use these cells therapeutically, should an appropriate clinical indication outside of their HIV disease (transplantation, autoimmune disease) arise. However, immunotherapy targeting Tregs in the context of HIV-1 infection remains controversial [45,46] and will require further careful investigation into the role of Tregs in HIV disease.

The concept of immune silencing and potentially enhancing Treg function in vivo to control HIV-1-related immune activation and virus replication in conventional T cells is appealing, yet challenging to achieve. IL-2 cytokine therapy in humans promotes the generation and proliferation of effector T cells and has been shown to improve CD4 counts in HIV-1-positive individuals but not their clinical outcomes [47]. Interestingly, IL-2 treatment of HIV-infected patients on suppressive antiretroviral therapy resulted in the expansion of Tregs, which may have impaired the function of conventional CD4+ T cells [48] and could explain the overall disappointing results of this approach. Indeed, the suppressive capacity of Tregs critically depends on IL-2 [49]. In a SIV animal model, IL-2 treatment resulted again in Treg expansion but also promoted CD4+ T cell activation and spontaneous apoptosis [50], further highlighting the difficulties of using IL-2 in vivo to modulate the course of HIV-1 infection.

One alternative, but still highly experimental approach of enhancing Treg activity in vivo would be the transfer of autologous Tregs. In the transplantation setting, numerous animal studies described the use of polyclonally expanded autologous Tregs to induce allograft control [51,52,53,54] and control autoimmune diseases [55,56,57]. Indeed adoptive transfer of activated Tregs provided neuroprotection in an HIV-1 encephalitis mouse model [58] and this was linked to down-regulation of proinflammatory cytokines, oxidative stress, and viral replication. However, besides technical difficulties, a major risk and challenge of isolating and expanding Tregs from HIV-1 infected donors for potential cell therapy is the re-activation of replicating virus, which needs additional careful exploration, but could potentially be managed safely in the era of HAART. Future studies should aim to reach the highest degree of Treg purity (e.g. using rapamycin alone [19] or in combination with retinoic acid [43]) and stability possible (e.g. using Oligodeoxynucleotides [39]) as adoptive transfer of activated conventional CD4+ T cells in the context of HIV-1 may add “fuel to the fire” in the form of new targets for the virus.

The Treg expansion approach may also be used to enrich or detect small Treg subsets such as antigen-specific Tregs [59]. Moreover, expanding functional Tregs from different tissue compartments could prove to be a useful tool to study the biology and impact of Tregs on HIV-1 infection, as the Treg TCR repertoire varies by anatomic location, presumably due to antigen encounter [60]. Tregs are important for maintenance of intestinal immune homeostasis by controlling inflammatory responses triggered by continuous antigen challenge in healthy individuals [61]. During the earliest days of HIV-1 infection, increased inflammation and immune activation occur in the gut associated lymphoid tissue [62], however little is known about the role and specificity of Tregs present in GALT and other mucosal tissues during early HIV-1 events [63,64]. Despite access to mucosal samples and the scarcity of the Treg population, which contribute to the lack of data, are drawbacks that could be partially overcome by Treg expansion approaches such as outlined here.

We therefore believe that this study will greatly facilitate the investigation of the role of Tregs during HIV-1 infection. A more detailed understanding of this unique T cell subset and its influence on HIV pathogenesis, immune activation and HIV-1-specific immunity will be critical for the design of potential immunotherapeutic strategies targeting Tregs (both up regulation
and down regulation of Treg activity are under active investigation and consideration [46] and possibly in the context of HIV-1 eradication [65].

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Author Contributions

Conceived and designed the experiments: MA PLK MMA. Performed the experiments: MA PLK MK SMS AR APT. Analyzed the data: MA PLK MMA. Contributed reagents/materials/analysis tools: ESM APT IT ATC PJG TN DSK. Wrote the paper: MA MMA.

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