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The Role of *Candida albicans* SPT20 in Filamentation, Biofilm Formation and Pathogenesis

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Abstract

*Candida albicans* is a ubiquitous fungus, which can cause very serious and sometimes life-threatening infections in susceptible patients. We used *Caenorhabditis elegans* as a model host to screen a library of *C. albicans* mutants for decreased virulence and identified SPT20 as important for virulence. The transcription co-activator SPT20 was identified originally as a suppressor of Ty and solo δ insertion mutations, which can cause transcription defects in *Saccharomyces cerevisiae*. It is resistant to the toxicity caused by overexpression of GAL4-VP16. We constructed a *C. albicans* spt20Δ/Δ mutant and found the spt20Δ/Δ strain was significantly less virulent than the wild-type strain SC5314 in *C. elegans* (*p* < 0.0001), *Galleria mellonella* (*p* < 0.01) and mice (*p* < 0.001). Morphologically, spt20Δ/Δ mutant cells demonstrated a “snow-flake” shape and clustered together; prolonged culture times resulted in increased size of the cluster. The clustered morphology was associated with defects in nuclear distribution, as the nuclei were not observed in many cellular compartments. In addition, the *C. albicans* spt20Δ/Δ mutant resulted in defects in hyphae and biofilm formation (compared to the wild-type strain, *p* < 0.05), and sensitivity to cell wall and osmotic stressors, and to antifungal agents. Thus our study demonstrated a role of *C. albicans* SPT20 in overall morphology and distribution of nuclear material, which may cause the defects in filamentation and biofilm formation directly when this gene is deleted.

Introduction

*Candida albicans* is part of the human flora that can be isolated from the gastrointestinal tract, vagina, and mouth in healthy individuals. Nevertheless, when the anatomical barrier is damaged or when host immunity declines then an invasive fungal infection can follow [1]. *C. albicans* is the fourth most common pathogen isolated from blood cultures and candidemia is associated with mortality rates as high as 35% [2] or even higher [3,4]. A characteristic of *C. albicans* is that it can make the switch between several morphological forms: budding yeast, pseudohyphae, and true hyphae, according to the environmental conditions [5,6]. The pathogenic ability of *C. albicans* is closely related to the change between these morphological forms [7,8]. Additionally, the formation of hyphae [9] and biofilms [10], the integrity of the cell wall [11], the rapid adaptive capacity to external environmental condition [12] all contribute to virulence and are all related to morphogenesis.

Here we report SPT20 is involved in *C. albicans* virulence and is essential for hyphal and biofilm formation. The transcription co-activator SPT20 encodes a 604 amino acid nuclear protein that is rich in glutamine and asparagines [13] and was identified originally as a suppressor of Ty and solo δ insertion mutations, which can cause transcription defects in *Saccharomyces cerevisiae* [14]. There is ample evidence that *S. cerevisiae* SPT20 mutants have some broader transcriptional defects when compared with the other SPT genes [13–15]. Genome-wide expression analysis showed that SPT20 controls about 10% of normal transcription in *S. cerevisiae* [16]. Of note is that SPT20 has also been identified by another name, *ADA5*, as it is a member of the *ADA* group of genes, and it is resistant to the toxicity caused by overexpression of GAL4-VP16 which blocks the transcription factor combined with transcription complexes, thus killing the yeast cells [13]. Interestingly, we found that in *C. albicans*, SPT20 is associated with cell morphology; spt20Δ/Δ cells cluster together and demonstrate a “snow-flake” shape. Deformity in cell morphology is accompanied by defects in nuclear localization.
Materials and Methods

Strains and growth conditions

*C. albicans* strains were grown in YPD (1% yeast extract, 2% peptone, and 2% dextrose) at 30 °C. Nourseothricin-sensitive clones were identified by their small colony size and confirmed by re-streaking on YPD agar plates containing 200 µg/mL nourseothricin (Werner Bioagents, Jena, Germany) as described previously [17].

For the nematode studies, the *C. elegans* *gfp-4; sek-1* strain was propagated on nematode growth medium on lawns of *Escherichia coli* OP50 using standard methods [18].

Generation of a homozygous mutant and re-integrated strain

The flipper cassette from pSFS2 containing the dominant nourseothricin resistance marker *CaSAT1* was used to generate the *spt20Δ/A* mutant [17]. This *SAT1* flipping strategy was used to avoid any interference from the auxotrophic selectable marker genes in our virulence studies. The DNA fragment for homologous recombination was generated as follows: PCR was used to amplify a 924 bp *Apa*I-*Xho*I DNA fragment containing an upstream region and the 5’ end of *SPT20* gene with primers *SPT20-up-FWD* and *SPT20-up-RV* (Table S1). The fragment was then cloned into the pSFS2 vector upstream of the *MAL2p-CaFLP-CaSAT1* cassette, yielding plasmid pSFS2-SPT20hp. A 726 bp *Sac*I-*Sac*I fragment containing a downstream region and the 3’ end of the *SPT20* gene was amplified by PCR with primers *SPT20-down-FWD* and *SPT20-down-RV* (Table S1) and cloned into the pSFS2-SPT20hp vector downstream of the *MAL2p-CaFLP-CaSAT1* cassette using the *Sac*I and *Sac*I restriction sites, yielding the final plasmid pSFS2-SPT20hp-down. The pSFS2-SPT20hp-down plasmid was digested with *Apa*I and *Sac*I, and the 5856 bp fragment was incorporated into *C. albicans* SC5314 using the lithium acetate method [19]. Nourseothricin-resistant transformants were evaluated for the appropriate insert location by PCR (Table S2), and the correct transformants were used for further work. After excising the nourseothricin resistance marker (*caSAT1*), the same method was used to delete the remaining functional allele of *SPT20*. Another round of integration/excision yielded SPT20 homozygous mutant *spt20Δ/A*. The primers used to identify all the strains are listed in Table S2.

To re-integrate *SPT20* into the *SPT20* null mutant, *spt20Δ/A*, a 3640 bp *Apa*I-*Xho*I fragment containing the complete open reading frame as well as 790 bp of upstream and 594 bp of downstream flanking sequences was amplified with the primers *SPT20-FWD* and *SPT20-RV* (Table S1) and substituted for upstream and 5’ end of *SPT20* gene sequence in pSFS2-SPT20hp-down at the *Apa*I and *Xho*I restriction site to generate the plasmid pSFS2-SPT20Rec. The pSFS2-SPT20Rec plasmid was digested with *Apa*I and *Sac*I, and the 7848 bp fragment was used to reintroduce the *SPT20* sequence into the *C. albicans* mutant *spt20Δ/A*. Nourseothricin-sensitive derivatives were obtained in the following process, yielding the *SPT20* reconstituted strain *spt20/A*SPT20 for further investigations. The primers used to identify the strains are listed in Table S2. We were unable to produce a strain that re-integrated the *SPT20* sequence at the second allele site. However, testing of the single loci re-integrated strain exhibited morphology and virulence comparable to the wild-type strain.

In vitro hyphal formation and biofilm growth assays

For hyphal formation, *C. albicans* strains were incubated with 1 mL serum for 60 min at 37 °C with agitation in the dark, then collected by centrifugation and washed twice with phosphate buffered saline (PBS). Cells were suspended in Spider media and incubated at 37 °C for 20 h [18], then photographed using an Olympus BX51 series upright microscope. For biofilm growth evaluation [20], strains of *C. albicans* were grown in YPD medium overnight at 30 °C, diluted to an OD 0.00 of 0.5 in 2 mL Spider medium, and added to a sterile 12-well plate containing a prepared silicone square measuring 1.5 × 1.5 cm (cut from silicone sheets Bentec Medical Inc., Woodland, CA) that had been pretreated overnight with bovine serum (Sigma-Aldrich). The inoculated 12-well plate was incubated with gentle agitation (150 rpm) for 90 min at 37 °C for adhesion to occur. The squares were washed with 2 mL of PBS to remove any un-adhered cells, and moved to a fresh 12-well plate containing 2 mL of fresh Spider medium. This plate was incubated at 37 °C for an additional 60 h with agitation (150 rpm) to allow for biofilm formation. The silicone platform and attached biofilm were removed from the wells, dried overnight, and weighed the following day. The total biomass of each biofilm was calculated by subtracting the weight of the silicone platform material prior to biofilm growth from the weight after the drying period and adjusting for the weight of a control pad exposed to no cells. The average total biomass for each strain was calculated from four independent samples. Statistical significance was determined by the analyses of variance (ANOVA). For comparison between two *C. albicans* isolates, the Student *t* test was used. A *p*-value of less than 0.05 was considered significant.

Sensitivity assays

The sensitivity to cell stressors was evaluated using the cell wall stress agent sodium dodecyl sulfate (SDS) and osmotic stress agent sodium chloride (NaCl). *C. albicans* strains were grown overnight and then suspended in YPD at 2 × 10⁹ cells/mL. Ten fold serial dilutions from 10⁴ to 10⁰ of the wild-type, *spt20Δ/A* and *spt20Δ/A-SPT20* re-integrated strains were spotted in a volume of 5 µL on YPD agar plates with the indicated chemical agent added. Cells were grown under 25 °C, 30 °C, and 37 °C conditions until colonies appeared.

Virulence assay using the *C. elegans* infection model

The assay was carried out using an established protocol [18]. In brief, freshly grown *C. albicans* cells were inoculated into 2 mL of YPD broth and allowed to grow overnight at 30 °C. The following day, 100 µL of yeast was spread into a square lawn on a 10-cm plate containing brain heart infusion (BHI) agar and kanamycin (45 µg/mL), followed by incubation for approximately 20 h at 30 °C. Synchronized adult *C. elegans* *gfp-4; sek-1* nematodes grown at 25 °C were carefully washed from plates containing their normal food source (*E. coli* OP50 strain) using sterile M9 buffer. Approximately 400 to 500 washed worms were then added to the center of the *C. albicans* lawns. The plates were incubated at 25 °C for 4 h. Worms were then carefully moved into a 15 mL conical tube and washed four times with sterile M9. Sixty worms were then transferred into a single well of a six-well tissue culture plate (Corning, Inc.) containing 2 mL of liquid medium (80% M9, 20% BHI) and kanamycin (45 µg/mL). Worms were scored daily and dead worms were removed from the assay. Microscopy of nematodes was performed by using Nomarski optics on a Zeiss AxioImager microscope.

To screen for mutations that affect *C. albicans* virulence, we evaluated a miscellaneous homozygous mutant library from the Fungal Genetics Stock Center (Kansas City, MO), deposited by Dr. Aaron Mitchell. Worms were infected as described above and scored for visibility of fungal filaments protruding through the worms cuticle and survival at 72 h post infection. Survival was
determined by the ability to respond to touch with a platinum wire. The control group was infected with strain DAY286, the parent of the mutant strains in the library collection.

**Virulence assay using the Galleria mellonella infection model**

Inocula were prepared by growing *C. albicans* strains at 30°C overnight with agitation. Cells were collected with centrifugation and washed 3 times with PBS. Yeast cells were counted using a hemocytometer. *G. mellonella* larvae (Vanderhorst Wholesale Inc., St. Marys, OH) at the final instar stage were inoculated with 10³ cfu of *C. albicans* suspended in PBS. Each infection group contained 16 randomly chosen larvae of the appropriate weight (330 ± 25 mg). The inoculum was injected in a 10 μL volume directly to the last pro-leg using a Hamilton syringe [21]. After inoculation, larvae were incubated at 37°C. A mock inoculation with PBS was included as a control for each experiment to observe for killing due to physical trauma or infection by pathogenic contaminants. The number of dead larvae was scored daily.

**Virulence assay using a murine candidiasis model**

A disseminated murine candidiasis model [22] was selected for the assay. Strains SC5314, spt20Δ/A, spt20Δ/SPT20 were used to infect mice. Cultures were grown overnight then washed 3 times with PBS. CD-1, six-week-old, female mice were infected with 1.5 × 10⁶ CFUs suspended in PBS via a tail vein injection in a 100 μL volume. Twelve mice were inoculated per strain tested and observed daily. Animal survival was evaluated by using the Kaplan-Meier method and differences were determined by using the log-rank test (STATA 6; STATA, College Station, TX). A p value of < 0.05 was considered statistically significant.

Fungal burden in the kidneys was assessed using four mice per infection strain [23]. We harvested the kidneys from mice asexptically 2 days post infection. Tissues were weighed and homogenized in sterile PBS by use of a Tissuemizer (model 398; Biospec Products Inc., Racine, WI). Serial dilutions were plated on YPD agar plates containing 100 μg/mL ampicillin, 100 μg/mL streptomycin, and 45 μg/mL kanamycin. The cfu/g kidney were counted after growth at 30°C for 48 h. Statistical analyses were performed using ANOVA and post hoc (Bonferroni and Student–Newman–Keuls) tests.

**Fungal cells staining with periodic acid Schiff (PAS)**

Histopathological analysis was performed to assess kidney infestation. The murine candidiasis model was prepared as described above. Two days after inoculation, kidneys were removed asexptically from each mouse before being fixed in 10% neutral buffered formalin. Kidneys were embedded in paraffin and removed aseptically from each mouse before being fixed in 10% neutral buffered formalin. Kidneys were embedded in paraffin and sections were stained with Periodic Acid Schiff (PAS) to reveal the hyphal structure of the fungal pathogens [23].

**Staining of nuclei**

Cells were stained for nuclei according to the protocol by Kopecka et al. and Fuchs et al. [24,25]. The strains were grown in YPD overnight at 30°C. Nuclei were identified by staining with 0.1 μg/mL DAPI for 15 min. The stained cells were collected then washed twice with 1% BSA in PBS for 5 min followed by a five minute rinse in 0.1% BSA in PBS. Cells were suspended in Vectashield (Vector Laboratories, Inc.) and visualized with an Olympus microscope. All observations were confirmed with 3 independent cell cultures.

**Fixation and staining of Calcofluor White**

Cells were fixed by adding 40% Formaldehyde (50 μL) to 450 μL cell cultures for 30 min and washed 2 times with PBS. Subsequently, cells were re-suspended in Calcofluor solution (Sigma) for 5 min at room temperature and washed 5 times with PBS. Then the stained cells were re-suspended in 10 μL mounting medium and observed using an Olympus confocal microscope.

**Drug susceptibility tests**

For the susceptibility assays, we grew cultures in YPD at 30°C with agitation. Cells were diluted in PBS to a density of 2.0×10⁹ cells/mL. The strains were plated as ten fold serial dilutions, spotting 5 μL of each dilution onto YPD agar plates supplemented with fluconazole (FLC; 2.5 μg/mL), amphotericin B (AMB; 1 μg/mL), 5-fluorocytosine (5FC; 4 μg/mL) or caspofungin (CAS; 0.075 μg/mL) and onto drug-free YPD agar plates as a control. Plates were incubated for 48 h at 35°C.

**Ethics statement**

All experiments were approved by Massachusetts General Hospital Research Animal Care Subcommittee and as outlined in the Guide for the Care and Use of Laboratory Animals of National Institutes of Health.

**Results**

Using a *C. elegans* infection model, we screened a *C. albicans* mutant library that consisted of 86 strains, for mutants that exhibited decreased virulence compared to strain DAY286. Through the course of our screening assay, we identified the SPT20 mutant strain as causing significantly less death in the *C. elegans* worms than the control strain DAY286. More specifically, the control infection produced an average of 47.6% dead worms with visible filaments protruding through the worm cuticle with a SD of 10.3%. However, the worms infected with the sp20 mutant strain did not produce any dead worms with protruding filaments. SPT20 deletion strains were developed for further analysis in order to confirm the role of this gene and the contributions of SPT20 in *C. albicans* virulence.

The *spt20Δ/A* mutant has reduced virulence in the *C. elegans* and the *G. mellonella* infection models

We confirmed the impact of SPT20 deletion in virulence using the *C. elegans* infection model (Figure 1). In this series of experiments, we performed the nematode assays in more detail with daily scoring of dead/alive nematodes. We found that more than half of the worms infected with the wild-type SC5314 (or the re-integrated strain *spt20Δ/A SPT20*) died within the first 48 h after infection (Figure 1A) and almost half the dead worms had visible hyphae piercing the cuticle (Figure 1B). At 120 h, >80% of the worms infected with the wild-type strain SC5314 or the re-integrated strain *spt20Δ/A SPT20* were dead, while only 30% of the worms were dead in the *spt20Δ/A* infection group (Figure 1A; p < 0.0001 compared to the wild-type strain or the reintegrated strain). Furthermore, no hyphae were observed in the *spt20Δ/A* infection group (Figure 1B).

We confirmed the decreased pathogenicity of the *spt20Δ/A* strain in an insect model. In the *G. mellonella* infection model, the *spt20Δ/A* was significantly less virulent than the wild-type strain SC5314 at 37°C (Figure 2; p < 0.01). All the larvae in the wild-type strain died within 4 days and the re-integrated strain group died within 5 days while the *spt20Δ/A* infection group died within 9 days (Figure 2; p < 0.01).
Figure 1. **SPT20 is essential for *C. albicans* virulence in the *C. elegans* infection model.** (A) Survival over 120 h (n = 60 worms infected per strain of *C. albicans*). Compared to infection with *C. albicans* SC5314, the spt20Δ/Δ mutant was avirulent to *C. elegans* (p < 0.0001). The reintegration of SPT20 restored the virulence of *C. albicans* to the wild-type level. (B) The images show *C. elegans* glp-4;sek-1 nematodes infected by *C. albicans* strains SC5314, spt20Δ/Δ and spt20Δ/SPT20 for 4 h and then moved into pathogen-free liquid media. On day 4 of the assay, the wild-type and spt20Δ/SPT20 infected worms exhibited a lethal infection with hyphae penetrating the worm cuticle. The hyphae were absent in the spt20Δ/Δ infected worms. The scale bar in Fig.1B represents 20 µm.

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Figure 2. **SPT20 influences virulence of *C. albicans* in the *G. mellonella* infection model.** Survival over 216 h (9 days) at 37°C (n = 16 larvae per strain of *C. albicans*). p < 0.01 compared to the groups infected with the wild-type or the spt20Δ/SPT20 reintegrated strain.

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The spt20Δ/Δ mutant has attenuated virulence in the murine candidiasis model

The spt20Δ/Δ strain was significantly less virulent than the wild-type strain SC5314 (p < 0.001) (Figure 3A). More specifically, all 12 mice inoculated with the wild-type strains within 7 days post infection by tail vein injection, while only 4 out of 12 mice in the spt20Δ/Δ group were dead at the same period.

We also evaluated the fungal burden in the mouse kidneys as part of the pathogenicity assessment. The fungal burden in the spt20Δ/Δ group was significantly lower than that in the wild-type SC5314 infection group (p, 0.01; Figure 3B). Evaluation of the kidney tissue by histopathology illustrated the differences in the burden and morphology of the infecting fungi. PAS staining revealed a large number of fungi in the kidneys of mice infected with the wild-type or the re-integrated strains while rare fungal cells were observed in the kidneys from the mice infected with the spt20Δ/Δ strain (Figure 3C). We also observed differences in the morphological state of the infecting fungi. Hyphae were visible in the tissues of the wild-type and re-integrated strains infected mice but not in the kidney of mice infected with the spt20Δ/Δ strain (Figure 3C). Of note is that the fungal burden of the kidneys of mice infected with spt20Δ/SPT20 re-integrated strain was restored, and the P value was not significantly different compared to the wild type.

The role of SPT20 in C. albicans cell wall integrity

To examine the cell wall integrity, we tested the susceptibility of wild-type strain SC5314, the spt20Δ/Δ mutant and the re-integrated strain spt20Δ/SPT20 to the cell wall stress agent SDS and osmotic stress agent NaCl. Interestingly, the spt20Δ/Δ mutant was susceptible to 0.01%, 0.03% SDS and 1 M, 1.5 M, 2 M NaCl at all the temperatures tested, 25°C, 30°C and 37°C (Figure 4A, 4B and 4C). Thus, the disruption of SPT20 affected C. albicans stress responses and suggests that SPT20 plays a role in maintaining cell wall integrity.

Sensitivity to antifungal agents

In our drug susceptibility testing, the wild-type, spt20Δ/Δ and spt20Δ/SPT20 re-integrated cells grew similarly on YPD agar plates and on YPD agar plates supplemented with 5FC, whereas the spt20Δ/Δ mutant was hypersensitive to amphotericin B, fluconazole and caspofungin compared to the wild-type and spt20Δ/SPT20 re-integrated strains (Figure 4D).

SPT20 is involved in C. albicans morphology

An examination of the cell morphology at 30°C growth temperature was established. C. albicans spt20Δ/Δ has a morphological defect (Figure 5). Micrographs of C. albicans morphology after 2, 8, and 24 h of growth at 30°C are presented. We found that the spt20Δ/Δ cells cluster together and demonstrate a “snow-
flake” shape rather than the normal separated and round shape of the wild-type and spt20Δ/Δ strain. The longer the cells are cultured, the larger the cell cluster.

To further validate the clustering morphology, we evaluated the cells for defect in chitin. We analyzed chitin distribution in wild-type strain and spt20Δ/Δ strain using Calcofluor white and tested the expression of CHT3 by RT-PCR. The results showed that spt20Δ/Δ did not exhibit defects in chitin distribution (Figure 6) and the expression of CHT3 between wild-type strain and spt20Δ/Δ strain have no statistical differences (Figure S1).

Figure 4. The disruption of SPT20 affected C. albicans stress responses and antifungal susceptibility. Ten-fold serial dilutions of the wild-type, spt20Δ/Δ, and spt20Δ/SPT20 were evaluated for susceptibility to cell wall stressors and antifungal agents. The spt20Δ/Δ mutant was specifically susceptible to SDS and NaCl at all the temperatures tested, 25°C (A), 30°C (B) and 37°C (C). The spt20Δ/Δ mutant was hypersensitive to amphotericin B, fluconazole and caspofungin, but was not sensitive to 5-fluorocytosine (D).

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SPT20 is associated with hyphal and biofilm formation

The visible difference in hyphal formation observed in the C. elegans infection assay and the lack of filaments in the infected kidney tissue, suggests that SPT20 is needed for hyphal formation, and plays an important role in biofilm formation. We confirmed this role by studying hyphal development and biofilm formation in vitro. The results showed that the spt20Δ/Δ mutant was defective in hyphal (Figure 7A) and biofilm formation (p < 0.05; Figure 7B and 7C) compared to either the wild-type strain or the re-integrated strain.
Defects in nuclear localization

In the “snow-flake” morphology, clustered cells appear to have defects in the ability of the daughter cells to separate from the mother cells, thus causing the clustering phenotype. We investigated whether the cell complexes all have nuclei using 6-diamidino-2-phenylindole dihydrochloride (DAPI) to evaluate the presence of nuclei in the complexes of the clustered cells. Interestingly, many gathered cells show defects in the location of nuclei (Figure 8). This indicates that the nucleus is not properly dividing or being distributed to the dividing cells. Overall, the lack of nuclei in some gathered compartments indicates a role for SPT20 in nuclear division. The lack of nuclear material in complexes of the clustered cells may also be associated with the cytokinesis between the cell complexes of the clustered SPT20 mutant cells.

Discussion

In this report, we show that in C. albicans, SPT20 plays a role in maintaining cell wall integrity. The defects in the cell wall are coupled with altered morphology and the inability of spt20Δ/Δ cells to form hyphae and generate a biofilm. Strikingly, these defects translate into reduced pathogenicity of the strain in multiple infection models.
The defects associated with cell morphology are characterized by a clustering effect, suggesting that *spt20*Δ/Δ cells are defective for cytokinesis, the mother and daughter cells do not separate properly. Moreover, some of the *spt20*Δ/Δ cells lacked nuclei therefore it is more accurate to describe some of the cells as “compartments” or “complexes”. These clusters are able to enlarge in the number of compartments that comprise the overall entity over time through the continuation of cell proliferation without complete separation. Interestingly, the *SPT20* mutant daughter cells of *S. cerevisiae* do not separate well from the mother cell [26], but the cells appears to have a nucleus [27], an aspect that differs in the *C. albicans* clustered complexes.

In *S. cerevisiae*, the growth rate of the deletion mutant is slow, and the *SPT20* mutant displays an elongated and irregularly bud morphology instead of the normal round morphology of wild-type cells [14,28]. Although both fungi display morphological defects they differ in the rate of growth. Our examination of growth on solid media indicated that the *SPT20* mutant strain grew at rates comparable to the wild-type strain.

Adapting to variations in the environment is a basic feature for microorganism survival and, through adaptive evolutionary mechanisms, this process is under strict regulation and control. In *C. albicans*, we found that *SPT20* is involved with the ability of the fungal cells to adapt to compounds such as SDS that are toxic to the cell wall as well as to high osmolarity caused by sodium chloride. Similarly, in *S. cerevisiae* *SPT20* is essential for cell survival during environmental stress conditions [26,29]. It’s worth noting that *SPT20* maintains the structural integrity and the function of SAGA complex[30–33]. *SPT20* and SAGA complex in *S. cerevisiae* and *C. albicans* are also highly conserved [29].

Figure 7. Hyphae and biofilm defects of the *spt20*Δ/Δ. (A) *C. albicans* strains SC5314, *spt20*Δ/Δ and *spt20*Δ/SPT20 were grown in Spider medium for 20 h at 37°C and examined at ×400 magnification. Compared to the *C. albicans* SC5314, *spt20*Δ/Δ showed attenuated hyphal formation in vivo (also showed attenuated hyphal formation in vitro (also showed attenuated hyphal formation in vivo, see Figure 1). This phenotype was restored in the *spt20*Δ/SPT20 reintegrated strain. The scale bar given in panel A represents 20 μm. (B) Strains were grown under in vitro biofilm assay conditions for 60 h. Biofilm formed on the silicone surface of the SC5314 and *spt20*Δ/SPT20 strains but failed to form on silicone sheets seeded with *spt20*Δ/Δ. (C) Dry weight of the biofilm biomass. Standard deviations are depicted and based on 6–8 silicone pad measurements. *p* < 0.05.

![Figure 7](image-url)
signal transduction pathways can regulate intracellular synthesis and storage of glycerol, which resulted in increased intracellular osmotic pressure to improve the adaption to high osmolarity environments [34,35]. SAGA is necessary for cell survival at hyperosmotic conditions [36]. In the hyperosmotic environment, it activates the HOG1 MAP kinase and is transferred from the cytoplasm to the nucleus, phosphorylating Sko1, and forms the Sko1-Cyc8-Tup1-Hog complex that recruits SAGA to the modified cytoplasm to the nucleus, phosphorylating Sko1, and forms the osmotic pressure to improve the adaption to high osmolarity and storage of glycerol, which resulted in increased intracellular stress inducible [43]. Researches have shown that the SAGA complex of S. cerevisiae is no intact SAGA complex in spt20 mutant and the sensitivity to cell wall damage is similar to the SAGA complex of S. cerevisiae [43]. SAGA controls 10% of all adjustable genes, and is known to be stress inducible [43]. Researches have shown that the SAGA complex of C. albicans is similar to the SAGA complex of S. cerevisiae [26]. Environmental stress conditions that influence gene expression that tends to be SAGA dominated include: heat, oxidation, acidity and nitrogen starvation, conditions that also influence the cell wall integrity pathway. In S. cerevisiae, SAGA regulates expression of FKS1 and ERG11, involved in cell wall glucan synthase and ergosterol synthesis, respectively [43].

An important part of C. albicans’ pathogenicity is its ability to form hyphae and generate biofilm, which play important roles in the colonization of medical devices. The hyphal form belongs to the overall factor of C. albicans’ virulence by invading epithelial and endothelial cells and evading phagocytes that cause the release of hydrolytic enzymes, forming biofilms that colonize medical devices, and lead to tissue damage. This process requires active changes to the cell wall in order to accommodate expanding volume, relying on actin to maintain the cytoskeletal structure during the process of hyphae and pseudohyphae formation. Our investigation of Spt20 suggests that it plays a role in biofilm formation and filamentation. The spt20Δ/A showed attenuated hyphal formation within the clustered morphology. The defective filaments were unable to support the formation of a biofilm, as assessed by the reduced biofilm mass. Importantly, the inability to form hyphae was reflected in the pathogenicity studies by the inability to produce hyphae that could penetrate the skin cuticle in the C. elegans infection model or the lack of hyphae penetration the kidney tissue in the murine model. The aspect of pathogenicity is a differing attribute between S. cerevisiae and C. albicans, where in the former does not cause disease but the latter can be an opportunistic pathogen. Thus the role that SPT20 plays in C. albicans can contribute to virulence, a marked difference from the contribution made to S. cerevisiae.

Although S. cerevisiae do not form hyphae for the invasion of host tissues, filaments are formed during other biological functions. In S. cerevisiae, the ability of SPT20 mutant to form filaments is impaired [14,26,44–46]. Combined with our results, the data strongly indicates that SPT20 acts as an activator of filamentous and biofilm growth. It may affect the expression of hyphal-specific genes. In addition to SPT20, several SAGA related genes such as SPT6, ADA2, SPT3 have shown to be related for the filamentous growth of C. albicans. The genes SPT6 and ADA2 play a positive role in filamentous growth [18,47], while SPT3 plays a negative role [26]. Both ADA2 and SPT3 play an important role in pathogenicity [18,26].

In this work, we show that C. albicans SPT20 is associated with virulence. It is likely that the defects in hyphae and biofilm formation of the spt20Δ/A mutant and the sensitivity to cell wall and osmotic stresses may explain the loss of virulence in both invertebrates and mammals. Moreover, SPT20 plays a vital role in cell morphology and distribution of nuclear material, which may lead to the defects in filamentation and biofilm formation directly when this gene is deleted.

Supporting Information

Figure S1 The expression of CHT3 was not affected by spt20Δ/A. CHT3 mRNA levels were tested by RT-PCR in wild-type strain and spt20Δ/A strain. No significant difference between the two strains was found (P>0.05).

Table S1 Primers for the disruption and reconstitution of SPT20.

Table S2 Primers used to identify replacements of the SPT20 alleles.
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References


