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The Evolutionarily Conserved Mediator Subunit MDT-15/MED15 Links Protective Innate Immune Responses and Xenobiotic Detoxification

Read Pukkila-Worley¹²³, Rhonda L. Feinbaum²³, Deborah L. McEwan²³, Annie L. Conery²³, Frederick M. Ausubel²³

¹ Division of Infectious Diseases; Massachusetts General Hospital; Harvard Medical School; Boston, Massachusetts, United States of America, ² Department of Molecular Biology; Massachusetts General Hospital; Harvard Medical School; Boston, Massachusetts, United States of America, ³ Department of Genetics; Harvard Medical School; Boston, Massachusetts, United States of America

Abstract

Metazoans protect themselves from environmental toxins and virulent pathogens through detoxification and immune responses. We previously identified a small molecule xenobiotic toxin that extends survival of Caenorhabditis elegans infected with human bacterial pathogens by activating the conserved p38 MAP kinase PMK-1 host defense pathway. Here we investigate the cellular mechanisms that couple activation of a detoxification response to innate immunity. From an RNAi screen of 1,420 genes expressed in the C. elegans intestine, we identified the conserved Mediator subunit MDT-15/MED15 and 28 other gene inactivations that abrogate the induction of PMK-1-dependent immune effectors by this small molecule. We demonstrate that MDT-15/MED15 is required for the xenobiotic-induced expression of p38 MAP kinase PMK-1-dependent immune genes and protection from Pseudomonas aeruginosa infection. We also show that MDT-15 controls the induction of detoxification genes and functions to protect the host from bacteria-derived phenazine toxins. These data define a central role for MDT-15/MED15 in the coordination of xenobiotic detoxification and innate immune responses.

Introduction

In nature, organisms encounter environmental insults, such as chemical toxins, secreted microbial virulence factors and invasive pathogens, that threaten their ability to survive and reproduce. As a result, metazoans have evolved protective pathways to counter these challenges. For example, gene families such as cytochrome P450s (CYPs), glutathione-s-transferases (GSTs), and UDP-glucuronosyltransferases (UDPs) detoxify xenobiotic small molecule toxins and are conserved from nematodes to humans [1]. Likewise, innate immune defenses provide protection from invasive pathogens [2]. Recent publications have suggested that recognition of xenobiotic toxins is involved in the activation of immune response pathways [3,4]. From an evolutionary perspective, it is logical that hosts respond to threats encountered in the wild at least in part through surveillance pathways that monitor the integrity of core cellular machinery, which are often the targets of xenobiotic small molecules or microbe-generated toxins. These studies predict that organisms may integrate detoxification and immune responses as a means to respond rapidly to such challenges, but the mechanisms underlying this coordinated host response have not been reported.

Our research group and others use bacterial and fungal pathogenesis assays in the nematode Caenorhabditis elegans to investigate mechanisms of immune pathway activation in intestinal epithelial cells [2]. Genetic analyses of C. elegans that are hypersusceptible to bacterial infection have revealed that the nematode mounts defense responses through evolutionarily conserved innate immune pathways. For example, the C. elegans NSY-1/SEK-1/PMK-1 Mitogen Activated Protein (MAP) kinase pathway, orthologous to the ASK1 (MAP kinase kinase kinase)/MKK3/6 (MAP kinase kinase)/p38 (MAP kinase) pathway in mammals, is required for protection against pathogens [5]. C. elegans animals carrying loss-of-function mutations in this pathway have defects in the basal and pathogen-induced expression of immune effectors and are hypersusceptible to killing by bacterial and fungal pathogens [5–7].

We previously used a C. elegans pathogenesis assay as a means to identify small molecules that protect the host during bacterial infection [8]. One of the compounds identified in this screen, a small molecule called RPW-24, extended the survival of nematodes infected with the human bacterial pathogen Pseudomonas aeruginosa by stimulating the host immune response via the p38 MAP kinase PMK-1 pathway [9]. A genome-wide microarray analysis of animals exposed to RPW-24 revealed that, in addition
to inducing the transcription of putative immune effectors, this molecule also strongly upregulated Phase I and Phase II detoxification enzymes (CYPs, GSTs and UDPs), suggesting that RPW-24 is a xenobiotic toxin to C. elegans. Consistent with this hypothesis, RPW-24 caused a dose dependent reduction of nematode lifespan on nonpathogenic food and delayed development of animals that were exposed starting at the first larval stage.

Here we sought to use RPW-24 as a tool to characterize mechanisms of p38 MAP kinase PMK-1 pathway activation in C. elegans. We found that activation of PMK-1-regulated pathogen response genes is genetically linked to the induction of genes involved in the detoxification of small molecule toxins. We show that the evolutionarily conserved Mediator subunit MDT-15 function has important physiological effects on the ability of an animal to mount protective immune responses, resist bacterial infection and survive challenge from lethal bacterial toxins.

Results

RNAi Screen Identifies Regulators of p38 MAP Kinase PMK-1-Dependent Genes

To investigate mechanisms of immune activation in C. elegans, we generated a GFP transcriptional reporter for the immune response gene F08G5.6. F08G5.6 is a putative immune effector that contains a CUB-like domain [6] and is transcriptionally induced by exposure of C. elegans to several bacterial pathogens, including P. aeruginosa [6,10]. We chose F08G5.6 for these studies because it is upregulated more than 100-fold by RPW-24 in a manner that requires the p38 MAP kinase PMK-1 [9].

pF08G5.6::GFP was induced in the C. elegans intestine during P. aeruginosa infection and GFP expression was also robustly upregulated in pF08G5.6::GFP animals following exposure to RPW-24 when animals were feeding on nonpathogenic E. coli (Figure 1A). When three components of the p38 MAP kinase PMK-1 signaling cassette were individually knocked down by RNAi [tir-1 [11], pmk-1 [6] and af-7 [12]], pF08G5.6::GFP induction by RPW-24 was entirely abrogated (Figure 1A).

The level of F08G5.6 induction during bacterial infection is dependent upon the virulence of the invading pathogen [Figure S1]. We exposed C. elegans to several P. aeruginosa strains, each of which was previously shown to have a different pathogenic potential toward nematodes [13] and used qRT-PCR to determine the expression levels of F08G5.6 in these animals. In general, more pathogenic P. aeruginosa strains caused significantly greater induction of F08G5.6, suggesting that some aspect of P. aeruginosa virulence, rather than a structural feature of the bacteria itself, causes the activation of F08G5.6. Consistent with these data, it was previously shown that C. elegans primarily responds to virulence-related cues to mount its innate immune defenses towards P. aeruginosa [14–16].

To identify genes that regulate the p38 MAP kinase PMK-1 pathway in response to RPW-24, we screened a library of RNAi clones corresponding to 1,420 genes expressed in C. elegans intestinal epithelium (approximately 9% of the genome, Table S1B) for gene inactivations that abrogated the RPW-24-mediated induction of pF08G5.6::GFP. We specifically focused on intestinally expressed genes because of the recognized role for intestinal cells in coordinating the host response to ingested pathogens [2,17], and because P. aeruginosa and RPW-24 induce F08G5.6 expression in the intestine (Figure 1A). Our initial screening effort identified 153 genes that, when inactivated by RNAi, diminished or abrogated the induction of pF08G5.6::GFP by RPW-24.

We took several steps to identify specific regulators of p38 MAP kinase PMK-1-dependent immune effectors among these 153 gene inactivations. First, we noticed that knockdown of many of these genes markedly slowed nematode growth. To eliminate genes that simply reduced GFP reporter expression as a consequence of pleiotropic effects on worm growth and development, we determined if these 153 gene inactivations also affected induction of the C. elegans immune reporter irg-1::GFP [14]. irg-1 is strongly upregulated in intestinal epithelial cells during P. aeruginosa infection or by an E. coli strain that expresses the bacterial virulence factor Exotoxin A (ToxA), but via a pathway independent of p38 MAP kinase PMK-1 signaling [14–16]. Moreover, RPW-24 does not cause the induction of irg-1::GFP and mutation of the zip-2 gene, which encodes the transcription factor that regulates irg-1 expression, does not affect the RPW-24-mediated induction of F08G5.6(data not shown) or alter the ability of RPW-24 to extend the survival time of nematodes infected with P. aeruginosa [9]. We therefore discarded the genes that, when inactivated, reduced the induction of irg-1::GFP by E. coli expressing ToxA, reasoning that they were unlikely to be specific regulators of the p38 MAP kinase PMK-1-dependent pathogen response genes. Using this approach, we selected 56 of the 153 genes for further study.

In a tertiary screen, we determined the effects of these 56 gene inactivations on pF35E12.5::GFP, a second immune reporter that is also strongly induced in the intestine by RPW-24 in a PMK-1-dependent manner [9,10]. 29 of the 56 RNAi clones reduced or eliminated the induction of both the pF35E12.5::GFP and pF08G5.6::GFP reporters by RPW-24 (Table S1A). Validating the screen, the 29 clones we identified as putative regulators of the p38 MAP kinase PMK-1-dependent genes included the three known components of the p38 MAP kinase PMK-1 pathway that were present in the screening library, which suggested that the screen could identify additional, unrecognized components of this signaling pathway.

To confirm further the results of the screen, we used RNAi to knockdown the expression of a representative sample of the 29 genes identified, and tested the induction levels of F08G5.6 and...
confirmed by the pmyo-3::mCherry agEx114 pF08G5.6::GFP transgene. In the co-injection marker, which confirms the presence of the animals shown in this figure, red pharyngeal expression is the (pF08G5.6::GFP carrying the conditions. (B) 18 hours. Photographs were acquired using the same imaging food, RNAi strain, and then transferred at the L4 stage to either E. coli OP50 or P. aeruginosa as E. coli OP50 supplemented with RPW-24 for 18 hours. Photographs were acquired using the same imaging conditions. (B) C. elegans mdt-15(RNAi) and mdt-15(tm2182) animals carrying the pF08G5.6::GFP immune reporter with or without agEx114 (mdt-15::mdt-15) were exposed to RPW-24 as described above. In all animals shown in this figure, red pharyngeal expression is the pmyo-2::mCherry co-injection marker, which confirms the presence of the pF08G5.6::GFP transgene. In the mdt-15(tm2182) animals carrying the agEx114 array, the presence of the pmtd-15::mdt-15 transgene is confirmed by the pmyo-2::mCherry co-injection marker which is expressed in the body wall muscle. doi:10.1371/journal.ppat.1004143.g001

F35E12.5 by RPW-24 with qRT-PCR (Table S1A). For all six genes tested, we verified that inactivation of the gene by RNAi dramatically reduced the induction levels of the p38 MAP kinase PMK-1-regulated genes F08G5.6 and F35E12.5.

Mediator Subunit MDT-15/MED15 Regulates p38 MAP Kinase PMK-1 Pathway-Dependent Genes

One of the strongest hits from our RNAi screen was mdt-15, which encodes a subunit of the Mediator complex homologous to mammalian MED15 (78% sequence identity) [18–20]. Knockdown of mdt-15 eliminated all visible expression of the pF08G5.6::GFP and pF35E12.5::GFP reporters, and reduced expression of these genes by at least two orders of magnitude in response to RPW-24 (Figure 1B and Table S1). MDT-15 was previously found to regulate the transcription of detoxification genes, including cytochrome P450s, glutathione-s-transferases, and UDP-glucuronosyltransferases [19], gene classes that are strongly induced by RPW-24 [9]. To study the role of mdt-15 in the regulation of p38 MAP kinase PMK-1 gene activation, we crossed the pF08G5.6::GFP reporter into the hypomorphic mdt-15(tm2182) allele, which was previously shown to recapitulate many of the phenotypes observed in mdt-15(RNAi) animals [19,21]. As in mdt-15(RNAi) animals, we observed no induction of GFP when mdt-15(tm2182); pF08G5.6::GFP animals were exposed to RPW-24 (Figure 1B). An extrachromosomal array containing wild-type mdt-15 under its own promoter partially restored RPW-24-induced GFP expression in mdt-15(tm2182); pF08G5.6::GFP animals (Figure 1B).

To determine if MDT-15 is required for the induction of other RPW-24-induced genes, we used NanoString nCounter gene expression analysis to generate transcription profiles of 118 C. elegans genes with known involvement in immune, stress and detoxification responses (Table S2). As in our microarray analysis [9], we found that RPW-24 caused robust transcriptional changes in wild-type nematodes. 40 of the 118 genes in the NanoString codeset were induced at least 4-fold or greater, 25 of these 40 genes are putative immune effectors upregulated during pathogen infection (shown in Figure 2) and 13 are genes putatively involved in the detoxification of small molecule toxins (discussed below). Of note, we had previously observed that 31 of these 40 genes, including 28 of the 31 most strongly induced, were also upregulated in whole genome Affymetrix GeneChip microarray analysis of wild-type animals exposed to RPW-24 versus DMSO [9]. Of the 25 pathogen-induced genes upregulated by RPW-24, 21 have been shown to be induced during P. aeruginosa infection [6,7,22,23]. The RPW-24-induced expression of 13 of the 25 putative immune effectors was significantly reduced in pmk-1(km25) loss-of-function mutants compared to wild-type controls (Figure 2 top panel) in accord with the previously determined role for p38 MAP kinase PMK-1 in regulating both pathogen-induced [6] and RPW-24-induced [9] expression of putative immune effector genes.

Consistent with a role for MDT-15 in the expression of PMK-1-activated genes, the NanoString analysis revealed that the RPW-24-dependent induction of 10 of the 13 PMK-1-dependent genes was abrogated in mdt-15(RNAi) animals compared to controls (Figure 2 bottom panel). We used qRT-PCR to show that mdt-15 was knocked down by RNAi in this experiment and to confirm that mdt-15 depletion caused a dramatic reduction in the RPW-24-induced expression of three pmk-1-dependent immune genes (Figure S2A). Further, we found that the RPW-24-mediated induction levels of these three immune genes was reduced in the mdt-15(tm2182) mutant, which recapitulated our findings in mdt-15(RNAi) animals (Figure S2B).

Knockdown of mdt-15 reduced the expression of the top five most strongly upregulated p38 MAP kinase PMK-1-dependent immune effectors by several orders of magnitude (C32H11.1, F33E12.5, F08G5.6, F49F1.7, and F49F1.1) (Figure 2 top panel). These five genes were still induced by RPW-24 in pmk-1(km25) loss-of-function animals, albeit to levels markedly lower than in wild-type animals (Figure 2 top panel), but their induction was entirely abrogated by knockdown of mdt-15 (Figure 2 bottom panel). These data suggest that MDT-15 coordinates inputs from PMK-1 and other immune signaling pathway(s) to modulate the expression of these p38 MAP kinase PMK-1-dependent putative immune effectors. We also identified a requirement for mdt-15 in the induction of five putative immune effectors (elec-206, c29f7.2, K03B2.4, cdr-1, T28C12.4) that are not transcriptional targets of the PMK-1 pathway (Figure 2, compare top and bottom panels). Thus, MDT-15 is required for the induction of p38 MAP kinase PMK-1-dependent immune genes and a second group of defense effectors that are independent of the p38 MAP kinase PMK-1 signaling pathway.

The p38 MAP kinase PMK-1 pathway plays an important role in the regulation of putative immune effectors during P. aeruginosa infection [6]. To determine whether MDT-15/MED15 is also involved in the regulation of these genes during bacterial infection, we infected C. elegans with P. aeruginosa PA14 for 8 hours and used qRT-PCR to compare the induction levels of several immune response genes in mdt-15(RNAi) and control animals. The basal and pathogen-induced expression of three p38 MAP kinase PMK-1-dependent immune effectors (C32H11.1,
One possible explanation for these observations is that the Mediator subunit MDT-15 is a general regulator of transcription that non-specifically affects the expression of a large number of genes. In the NanoString experiment, however, we identified 10 genes that were induced 4-fold or greater in wild-type N2 animals by RPW-24 are presented in Figures 2 and 6. The 13 genes that were dependent on the p38 MAP kinase PMK-1 for their induction in the top panel are grouped and indicated in the bottom panel. Data are the average of two replicates each of which was normalized to three control genes with error bars representing standard deviation and are presented as the value relative to the average expression from the replicates of the indicated gene in the baseline condition (N2 animals (top) or vector control (L4440) animals (bottom) exposed to DMSO). We confirmed that mdt-15 expression was significantly knocked down by mdt-15(RNAi) in each of these experiments (p<0.001)(see Figure S2A). * p<0.05 for the comparison of the RPW-24-induced conditions.

Figure 2. The Mediator subunit MDT-15 regulates p38 MAP kinase PMK-1-dependent and independent immune genes in response to RPW-24. The expression of 118 C. elegans genes was analyzed using NanoString nCounter gene expression analysis in wild-type N2 and pmk-1(km25) animals (top) and in vector control (L4440) and mdt-15(RNAi) animals (bottom) exposed to either 70 μM RPW-24 or the solvent control DMSO. The 40 genes that were induced 4-fold or greater in wild-type N2 animals by RPW-24 are presented in Figures 2 and 6. The 13 genes that were dependent on the p38 MAP kinase PMK-1 for their induction in the top panel are grouped and indicated in the bottom panel. Data are the average of two replicates each of which was normalized to three control genes with error bars representing standard deviation and are presented as the value relative to the average expression from the replicates of the indicated gene in the baseline condition (N2 animals (top) or vector control (L4440) animals (bottom) exposed to DMSO). We confirmed that mdt-15 expression was significantly knocked down by mdt-15(RNAi) in each of these experiments (p<0.001)(see Figure S2A). * p<0.05 for the comparison of the RPW-24-induced conditions.

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F08G5.6 and F35E12.5) was reduced in mdt-15(RNAi) animals by one to three orders of magnitude (Figure 3). We also tested the induction levels of three genes whose transcription is activated during P. aeruginosa infection in a manner independent of PMK-1 (irg-1, irg-2 and F01D5.5) and found that PMK-1-independent immune effectors were induced in mdt-15(RNAi) animals during P. aeruginosa infection to levels comparable to that observed in wild-type animals (Figure 3). The expression levels of irg-1 and irg-2 were higher under basal conditions in mdt-15(RNAi) animals compared to L4440 controls (Figure 3). Together, these data support the observations from our NanoString experiments that MDT-15 is required for the expression of some, but not all, immune genes that are activated in response to an environmental insult.

One possible explanation for these observations is that the Mediator subunit MDT-15 is a general regulator of transcription that non-specifically affects the expression of a large number of genes. In the NanoString experiment, however, we identified 10 genes that were induced by RPW-24 in mdt-15(RNAi) animals to levels similar to the wild-type controls (Figure 2 bottom panel). Also of note, our secondary screen demonstrated that mdt-15(RNAi) had no effect on the induction of irg-1::GFP by ToxA. We further wondered if the defects in expression of the PMK-1 targets in mdt-15(RNAi) animals were due to direct transcriptional regulation of the p38 MAP kinase PMK-1 pathway components by MDT-15. However, we found that the mRNA levels of pmk-1, tir-1 and sek-1 in mdt-15(RNAi) animals were not different from the wild-type control (Figure S2C).
MDT-15/MED15 Links Immunity and Detoxification

Mediator Subunit MDT-15/MED15 Is Required for Defense against *P. aeruginosa* Infection

The experiments in the preceding sections show that the Mediator subunit MDT-15/MED15 is necessary for the induction of the p38 MAP kinase PMK-1-regulated genes, both in response to RPW-24 and during *P. aeruginosa* infection. We therefore reasoned that mutation or RNAi-mediated knockdown of mdt-15 would result in enhanced susceptibility to *P. aeruginosa* infection. Initial *P. aeruginosa* pathogenesis assays showed a modest, but significant and reproducible, enhanced susceptibility to infection in mdt-15(RNAi) (Figure S3A) and mdt-15(tm2182) (Figure S3B) animals compared to controls. However, both mdt-15(tm2182) and mdt-15(RNAi) animals have reduced brood sizes and varying degrees of sterility. Sterile animals are more resistant to *P. aeruginosa* infection than wild-type animals, due in part to daf-16-dependent induction of stress response genes [25]. To eliminate this potentially confounding effect, we made all animals in the *P. aeruginosa* pathogenesis assay sterile by knocking down *cdc-25.1*, a technique that has been used previously in *C. elegans* bacterial pathogenesis assays [26]. Under these conditions, we found that mdt-15(tm2182) animals were markedly hypersusceptible to *P. aeruginosa* infection compared to control animals (Figure 5). Moreover, injection of the mdt-15 gene under control of its own promoter partially rescued the enhanced susceptibility to *P. aeruginosa* phenotype of mdt-15(tm2182) animals (Figure 5). Knockdown of mdt-15 by RNAi also caused a hypersusceptibility to *P. aeruginosa* phenotype in *C. elegans fer-15(hb26);fem-1(hc17) sterile animals [6,27], and as predicted, to a greater degree than wild-type animals that were not made sterile by *cdc-25.1(RNAi)* (Figure S3C).

We also found that the ability of RPW-24 to extend survival of *P. aeruginosa*-infected wild-type and sterile nematodes was significantly attenuated in mdt-15(RNAi) animals compared to controls, suggesting that MDT-15 is required for the immunostimulatory activity of RPW-24 (Figure S3A and S3C). The degree of lifespan extension by RPW-24 during *P. aeruginosa* infection was reduced in mdt-15(tm2182) compared to controls, but this difference did not reach statistical significance (*p* = 0.09) (Figure S3B). As discussed above, the gene expression defects of mdt-15 targets were more severe in mdt-15(RNAi) animals than in the hypomorphic mdt-15(tm2182) allele [19,21], which may account for this observation.

One caveat concerning the observation that mdt-15 depleted animals are hypersusceptible to *P. aeruginosa* infection is that mdt-15(RNAi) and mdt-15(tm2182) animals have a reduced lifespan due in part to RPW-24-mediated induction of RPW-24 during *P. aeruginosa* infection [26]. Under these conditions, we found that mdt-15(tm2182) animals were markedly hypersusceptible to *P. aeruginosa* infection compared to control animals (Figure 5). Moreover, injection of the mdt-15 gene under control of its own promoter partially rescued the enhanced susceptibility to *P. aeruginosa* phenotype of mdt-15(tm2182) animals (Figure 5). Knockdown of mdt-15 by RNAi also caused a hypersusceptibility to *P. aeruginosa* phenotype in *C. elegans fer-15(hb26);fem-1(hc17) sterile animals [6,27], and as predicted, to a greater degree than wild-type animals that were not made sterile by *cdc-25.1(RNAi)* (Figure S3C).

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One caveat concerning the observation that mdt-15 depleted animals are hypersusceptible to *P. aeruginosa* infection is that mdt-15(RNAi) and mdt-15(tm2182) animals have a reduced lifespan when grown on the normal laboratory food source *E. coli* OP50 (Figure 4B). During *P. aeruginosa* infection, we also observed a requirement for MDT-15 in the *csp-1(RNAi)*-mediated induction of F008G5.6 and F35E12.5, but not C32H11.1. The significance of this discrepancy is unclear, but may be explained by the observations of Taubert et al. who found that only 52% of mdt-15 targets were misregulated in the hypomorphic mdt-15(tm2182) allele, which encodes a truncated form of the MDT-15 protein [19,21]. Indeed, we also observed that mdt-15(RNAi) was more effective than mdt-15(tm2182) at reducing the expression of the six mdt-15 targets we studied following exposure to RPW-24, including C32H11.1 (Figures S2A, S2B and S2D). In any case, these data suggest that MDT-15 acts downstream of the p38 MAP kinase PMK-1 to coordinate the induction of at least some immune effector genes.

The Mediator Subunit MDT-15 Acts Downstream of the p38 MAP Kinase PMK-1 to Regulate the Induction of F008G5.6 and F35E12.5

To determine if MDT-15 acts genetically upstream or downstream of the p38 MAP kinase PMK-1 in coordinating the induction of immune effectors during *P. aeruginosa* infection, we utilized the Map kinase phosphatase VHP-1, which is a negative regulator of the p38 MAP kinase PMK-1 [24]. Knockdown of *csp-1* in animals carrying the F008G5.6::GFP p38 MAP kinase PMK-1 immune reporter caused increased induction of GFP expression during *P. aeruginosa* infection in a manner dependent on pmk-1 expression (Figure 4A). This induction of F008G5.6::GFP by *csp-1(RNAi)* was entirely suppressed in the mdt-15(tm2182) partial loss-of-function allele (Figure 4A). We used qRT-PCR to confirm this observation (Figure 4B). We found that RNAi knockdown of *csp-1* in wild-type *C. elegans* caused constitutive activation of F008G5.6, F35E12.5 and C32H11.1 in a manner that required mdt-15 when nematodes were growing on their normal food source *E. coli* OP50 (Figure 4B). During *P. aeruginosa* infection, we also observed a requirement for MDT-15 in the *csp-1(RNAi)*-mediated induction of F008G5.6 and F35E12.5, but not C32H11.1. The significance of this discrepancy is unclear, but may be explained by the observations of Taubert et al. who found that only 52% of mdt-15 targets were misregulated in the hypomorphic mdt-15(tm2182) allele, which encodes a truncated form of the MDT-15 protein [19,21]. Indeed, we also observed that mdt-15(RNAi) was more effective than mdt-15(tm2182) at reducing the expression of the six mdt-15 targets we studied following exposure to RPW-24, including C32H11.1 (Figures S2A, S2B and S2D). In any case, these data suggest that MDT-15 acts downstream of the p38 MAP kinase PMK-1 to coordinate the induction of at least some immune effector genes.

Taken together, the data in this section indicate that MDT-15 is required for the transcriptional activation of immune effectors controlled by the p38 MAP kinase PMK-1, and at least one other pathway. However, MDT-15 is not required for the induction of all defense-related genes.

The Mediator Subunit MDT-15 Acts Downstream of the p38 MAP Kinase PMK-1 to Regulate the Induction of F008G5.6 and F35E12.5

To determine if MDT-15 acts genetically upstream or downstream of the p38 MAP kinase PMK-1 in coordinating the induction of immune effectors during *P. aeruginosa* infection, we utilized the Map kinase phosphatase VHP-1, which is a negative regulator of the p38 MAP kinase PMK-1 [24]. Knockdown of *csp-1* in animals carrying the F008G5.6::GFP p38 MAP kinase PMK-1 immune reporter caused increased induction of GFP expression during *P. aeruginosa* infection in a manner dependent on pmk-1 expression (Figure 4A). This induction of F008G5.6::GFP by *csp-1(RNAi)* was entirely suppressed in the mdt-15(tm2182) partial loss-of-function allele (Figure 4A). We used qRT-PCR to confirm this observation (Figure 4B). We found that RNAi knockdown of *csp-1* in wild-type *C. elegans* caused constitutive activation of F008G5.6, F35E12.5 and C32H11.1 in a manner that required mdt-15 when...
above that mdt-15(RNAi) animals fail to upregulate p38 MAP kinase PMK-1-dependent immune effectors both in response to a xenobiotic toxin and during *P. aeruginosa* infection, but retain the ability to induce other immune genes in response to pathogens and following exposure to the bacterial toxin ToxA. In addition, mdt-15(RNAi) animals do not respond to the immunostimulatory effects of RPW-24. We have shown previously that *C. elegans* with mutations in the ZIP-1 and FSHR-1 immune pathways, which act in parallel to the p38 MAP kinase PMK-1 cassette, are hypersusceptible to *P. aeruginosa* infection, but retain the ability to respond to RPW-24 [9]. That mdt-15(RNAi) animals are blind to the immunostimulatory effects of RPW-24 suggests a specific role of MDT-15 in regulating p38 MAP kinase PMK-1 pathway activity. It is also important to note that despite their reduced lifespan, mdt-15(RNAi) animals are not sensitive to all environmental insults. For example, animals deficient in mdt-15 are sensitive to the toxin fluoranthene, but not β-naphthoflavone, and are not more sensitive to high temperatures than wild-type animals [19].

Mediator Subunit MDT-15/MED15 Regulates the Induction of p38 MAP Kinase PMK-1-Independent Detoxification Genes and Is Required to Resist the Toxic Effects of the Xenobiotic RPW-24

We previously demonstrated that RPW-24 is a xenobiotic toxin [9]. MDT-15 is known to coordinate protection from the toxin fluoranthene and regulate the transcriptional induction of CYPs [19]. We found that the RPW-24-mediated induction of the 13 detoxification genes in the NanoString codeset was nearly entirely abrogated by RNAi knockdown of mdt-15 (Figure 6A top panel and Table S2). This result was confirmed for three detoxification genes by qRT-PCR (Figures S2B and S2D). To determine if MDT-15 is required to protect *C. elegans* from the toxic effects of RPW-24, we studied the development of wild-type, pmk-1(RNAi) and mdt-15(RNAi) in the presence of the xenobiotic RPW-24, an
assay that has been used previously to assess the toxicity of small molecules [19]. We found that RPW-24 slowed the development of control and pmk-1(RNAi) animals to similar degree, and that mdt-15(RNAi) animals were markedly delayed in the presence of RPW-24 (Figures 6B and S4). These data show that MDT-15 controls the induction of detoxification genes following exposure to RPW-24 and is required to resist the toxic effects of this xenobiotic.

Given that MDT-15 is required for the regulation of detoxification genes and the p38 MAP kinase PMK-1 pathway, we wondered if the detoxification machinery in C. elegans is regulated by the p38 MAP kinase PMK-1. We found, however, that 12 of the 13 RPW-24-induced detoxification genes were upregulated in pmk-1(km25) null mutants to levels comparable to that in wild-type animals exposed to RPW-24 (Figure 6A bottom panel). We used qRT-PCR to confirm this observation for three cytochrome P450 genes (Figure S2D). Thus, the MDT-15-dependent xenobiotic detoxification program is induced in a manner independent of the p38 MAP kinase PMK-1.

MDT-15/MED15 Is Not Required for the Avoidance Behavior Induced by RPW-24

Many xenobiotic toxins, including RPW-24, induce an avoidance response wherein C. elegans leave a lawn of bacterial food, to which they are otherwise attracted, if it contains a toxic compound [3,9]. We therefore wondered if MDT-15 is required for the avoidance behavior induced by RPW-24. However, pmk-1(km25) and mdt-15(tm2182) animals left the lawn of E. coli containing RPW-24 as readily as wild-type animals (Figure S5). We also observed a similar phenotype when we knocked down the expression of pmk-1 and mdt-15 in the neuronally-sensitive RNAi strain TU3311 (data not shown). Thus, animals lacking the function of MDT-15 and PMK-1 are still able to recognize RPW-24 as a toxin.

In summary, the data in this and the preceding section suggest that MDT-15, but not PMK-1, controls the induction of genes that are required to resist the toxic effects of RPW-24, and that neither are required for avoidance of RPW-24.

Mediator Subunit MDT-15/MED15 Promotes Resistance to Secreted P. aeruginosa Phenazine Toxins

We have shown that MDT-15 regulates the C. elegans detoxification response to the xenobiotic RPW-24. We therefore hypothesized that MDT-15 might also be required for protection from lethal secreted toxins produced by pathogenic bacteria. To address this question, we used an assay that allows the specific study of C. elegans killing by secreted low molecular weight toxins of P. aeruginosa. When P. aeruginosa is grown on high osmolarity media, phenazine toxins produced by the bacteria are lethal to nematodes [28,29]. In contrast to the “slow killing” infection assay, which was used in the assays described above, wild-type nematodes exposed to these “fast killing” conditions die within a few hours via a process that does not require live bacteria [28,29]. Wild-type C. elegans at the fourth larval stage of development, but not young adult animals, are exquisitely sensitive to the phenazine toxins produced by P. aeruginosa in this assay and are killed within 6 hours of exposure. We took advantage of the inherent resistance of young adult animals to test whether mdt-15(tm2182) animals are hypersusceptible to the phenazine toxins produced by P. aeruginosa. As expected, both wild-type and mdt-15(tm2182) L4 animals were rapidly killed by P. aeruginosa in the “fast kill” assay (Figure S6A); P. aeruginosa carrying deletions of both phenazine biosynthetic operons (ApzE2) does not make these toxins and accordingly had...
reduced ability to kill both wild-type and mdt-15(tm2182) animals (Figure S6A). In contrast to L4 adults, we observed that almost no young adult, wild-type animals were killed after six hours of exposure to the phenazine toxins compared with 98% death of L4 staged animals (Figures 7A and S6A), which reproduces the findings of others [28,29]. In contrast to wild-type animals, young adult mdt-15(tm2182) animals were dramatically susceptible to P. aeruginosa in this assay and this pathogenesis required the secretion of phenazine toxins, as P. aeruginosa Δphz was markedly less pathogenic toward mdt-15(tm2182) young adults (Figure 7A). Moreover, mdt-15(tm2182) animals were not simply hypersusceptible to the high osmolarity conditions of this assay because we observed no mortality over the course of the assay in mdt-15(tm2182) mutants exposed to “fast kill” media containing the normal nematode food source E. coli OP50 (Figure 7A).

We confirmed that phenazine toxins are lethal to mdt-15-depleted animals by supplementing “fast kill” growth media with both phenazine-1-carboxylic acid and 1-hydroxyphenazine in the absence of pathogen [28]. These two particular phenazines are toxic to wild-type nematodes [28] and, as expected, the mixture of phenazine-1-carboxylic acid and 1-hydroxyphenazine rapidly killed L4 animals (Figure S6B). Young adult wild-type animals were resistant to the lethal effects of these molecules (Figure 7B). However, mdt-15(RNAi) young adult animals were dramatically susceptible to phenazine-mediated killing (Figure 7B).

We also found that pmk-1(km25) loss-of-function mutants were more susceptible to P. aeruginosa in the “fast killing” assay than wild-type animals, although were less susceptible than mdt-15(tm2182) animals. This lethality, however, was mediated by factors other than phenazine toxins, since there was no difference in pathogenicity of the wild-type and Δphz P. aeruginosa toward pmk-1(km25) mutants (Figure 7A). Likewise, pmk-1(RNAi) animals were no more susceptible to phenazines-1-carboxylic acid and 1-hydroxyphenazine than L4440 RNAi control animals (Figure 7B).

Together, these data define a role for MDT-15, but not PMK-1, in the protection from bacterial-derived phenazine toxins.

Discussion

Detecting and countering environmental threats is central to the ability of organisms to survive and reproduce in the wild. We examined the C. elegans response to the xenobiotic RPW-24, which is able to induce a host immune response that is protective for animals infected with the lethal bacterial pathogen P. aeruginosa [9]. In an RNAi screen with RPW-24, we identified a number of genes, including mdt-15/MED15, which are required for induction of p38 MAP kinase PMK-1-dependent immune effectors. mdt-15 encodes a subunit of the highly conserved Mediator complex that controls the activation of a variety of genes involved in the response to external stress. We demonstrate that: (i) MDT-15 is required for the induction of p38 MAP kinase PMK-1-dependent immune effectors following exposure to a xenobiotic toxin, as well as during infection with P. aeruginosa, (ii) MDT-15 controls the expression of some p38 MAP kinase PMK-1-independent immune effectors, but not all defense genes, (iii), MDT-15 functions downstream of the p38 MAP kinase PMK-1 cascade to control the induction of at least two immune effectors, (iv) the induction of xenobiotic detoxification genes and protection from the toxic

![Figure 7. Resistance to P. aeruginosa phenazine toxins requires MDT-15.](image-url)

(A) Wild-type N2, mdt-15(tm2182) and pmk-1(km25) animals were tested in the toxic phenazine P. aeruginosa pathogenesis assay (also called “fast killing” assay). C. elegans young adult animals were exposed to wild-type (WT) P. aeruginosa, P. aeruginosa carrying a deletion in both of the phenazine biosynthetic operons (.Δphz) or E. coli OP50. The difference between mdt-15(tm2182) and N2 animals exposed to P. aeruginosa WT is significant (p<0.001), as is the difference in mdt-15(tm2182) animals exposed to P. aeruginosa WT and P. aeruginosa Δphz (p<0.001). There is no significant difference in pmk-1(km25) animals exposed to P. aeruginosa WT and P. aeruginosa Δphz (p>0.05). This assay is representative of two independent experiments. Figure S6A presents the control condition for this experiment showing that both C. elegans N2 and mdt-15(tm2182) at the L4 stage are sensitive to toxic phenazine-mediated killing (p=0.001). (B) Vector control (L4440), mdt-15(RNAi) and pmk-1(RNAi) animals were exposed to high osmolarity “fast kill” media (pH 5) with E. coli as the food source in the presence or absence of phenazine-1-carboxylic acid and 1-hydroxyphenazine (labeled “toxic phenazines” above) at the approximate concentrations produced by P. aeruginosa under standard assay conditions. The difference in killing between mdt-15(RNAi) animals exposed to toxic phenazines and the other conditions is significant (p<0.001).
MDT-15/MED15 Links Immunity and Detoxification

C. elegans and Bacterial Strains

C. elegans were grown on standard NGM plates with E. coli OP50 [37] unless otherwise noted. The previously published C. elegans strains used in this study were: N2 Bristol [37], pmk-1(lm62) [5], AY101 [nls101; pDB09.1[pF35E12.5::GFP]; prF4[rol-6(n2100)]) [10], XAX702 mdt-15(tm2182)[19,21], C512 fer-15(b26); fem-1(he17) [38], and AU0135 [qa17[pmyo-2::gfp]; pmyo-2::mCherry] [14]. The C. elegans strains created for this study were: AU0307 [pF65.5:GFP::unc-54:3×UTR; pmyo-2::mCherry], AU0316 [mdt-15(tm2182); qaf44], AU0325 [mdt-15(tm2182); aqEx116 (mdt-15[pmyo-3::mCherry]), AU0326 [mdt-15(tm2182); aqEx117 (mdt-15[pmyo-2::mCherry]), AU0327 [mdt-15(tm2182); aqEx118 (mdt-15[pmyo-3::mCherry])].

Materials and Methods

C. elegans and Bacterial Strains

C. elegans were grown on standard NGM plates with E. coli OP50 [37] unless otherwise noted. The previously published C. elegans strains used in this study were: N2 Bristol [37], pmk-1(lm62) [5], AY101 [nls101; pDB09.1[pF35E12.5::GFP]; prF4[rol-6(n2100)]) [10], XAX702 mdt-15(tm2182)[19,21], C512 fer-15(b26); fem-1(he17) [38], and AU0135 [qa17[pmyo-2::gfp]; pmyo-2::mCherry] [14]. The C. elegans strains created for this study were: AU0307 [pF65.5:GFP::unc-54:3×UTR; pmyo-2::mCherry], AU0316 [mdt-15(tm2182); qaf44], AU0325 [mdt-15(tm2182); aqEx116 (mdt-15[pmyo-3::mCherry]), AU0326 [mdt-15(tm2182); aqEx117 (mdt-15[pmyo-2::mCherry]), AU0327 [mdt-15(tm2182); aqEx118 (mdt-15[pmyo-3::mCherry])].

The data presented in this study imply the existence of surveillance mechanisms that monitor for xenobiotic toxins (or their effects) and signal through MDT-15/MED15 to simultaneously activate protective detoxification and innate immune responses. Whether such activation occurs in the context of xenobiotic-induced organellar dysfunction is not known. McEwan et al. [13] and Dunbar et al. [16] found that C. elegans monitor the integrity of its translational machinery as a means to detect pathogen invasion. Inhibition of translation by the secreted bacterial toxin ToxA leads to an antibacterial response via known immune pathways involving the transcription factor ZIP-2 and the p38 MAP kinase PMK-1. Several observations suggest, however, that MDT-15-mediated regulation of immune activation and detoxification gene induction by RPW-24 does not occur via a mechanism involving translation inhibition. First, we previously found that a loss-of-function mutation in the zip-2 gene did not affect the ability of RPW-24 to extend the survival of animals infected with P. aeruginosa and genome-wide transcriptional analysis of animals exposed to RPW-24 did not suggest that this compound is an inducer of translation [9]. Second, the transcriptome analysis of animals exposed to ToxA did not show an abundance of Phase I and II detoxification genes [15]. Finally, we show here that the ToxA-responsive, ZIP-2-regulated genes, ing-1 and ing-2, are induced in mdt-15(RNAi) animals during P. aeruginosa infection to levels comparable to that observed in wild-type animals.

Melo et al. recently studied the behavioral response of C. elegans to xenobiotic toxins known to disrupt the function of the mitochondria, the ribosome, and the endoplasmic reticulum [5]. Inhibiting these essential processes by the action of small molecules or through targeted gene disruptions triggered an avoidance response, which required serotonergic and JNK signaling pathways. They proposed that organisms monitor disruption of core metabolic processes as a means to detect pathogen invasion and challenges from xenobiotic toxins. Based on the data reported here, however, it is not clear, whether immune signaling is an integral part of xenobioteic-elicited avoidance behavior, at least with respect to RPW-24. We found that the function of neither MDT-15 nor PMK-1 was required for the avoidance of RPW-24, indicating that the behavioral component of this protective response occurs upstream of MDT-15, or via separate mechanism altogether.

It will be interesting to determine the mechanism by which MDT-15 activates immune and detoxification responses in C. elegans. In our genetic screen for regulators of the p38 MAP kinase PMK-1 pathway, we identified a number of genes that are involved in fatty acid biosynthesis, including mdt-15, fat-6, fat-7, elo-5,acs-19 and C25A1.5. Indeed, MDT-15 is known to control the expression of fat-6 and fat-7 [18–20]. It is therefore possible that a fatty acid signaling molecule or membrane component is required for p38 MAP kinase PMK-1 activity. We found, however, in epistasis analyses with the MAP kinase phosphatase VHP-1 that MDT-15 functions downstream of PMK-1 to coordinate the expression of F08G5.6 and F33E12.5. Thus, for at least a subset of immune genes, MDT-15 likely also physically interacts with sequence-specific regulators, such as ATF-7, a transcription factor that is the downstream signaling target of the p38 MAP kinase PMK-1 pathway [12], to coordinate protective host responses mounted following exposure to xenobiotic toxins.
C. elegans Bacterial Infection and Other Assays

“Slow killing” P. aeruginosa infection were performed as previously described [9,44]. In all of these assays, the final concentration of DMSO was 1% and RPW-24 was used at a concentration of 70 μM, unless otherwise indicated. The propensity of wild-type C. elegans to leave a lawn of bacteria supplemented with RPW-24 was assayed using a previously described protocol [39, 47] with minor modifications. Rather than adding the toxin on top of the small lawn of food, 20 μg of RPW-24 was mixed with E. coli OP50, which was spotted onto NGM plates. To assess the toxicity of RPW-24, we assayed the development of animals exposed to vector control (L4440), pmk-1(RNAi) and mdt-15(RNAi) in the presence of 70 μM RPW-24, as previously described [9]. P. aeruginosa “fast kill” pathogenesis assays were conducted with late L4 and early young adult animals (picked 1–3 hours after the L4 molt) obtained from timed egg lays as described [28, 29]. For the killing assay using toxic phenazines, 50 μg/ml phenazine-1-carboxylic acid (PCA) and 5 μg/ml 1-hydroxyphenazine in DMSO were added to modified “fast kill” media (1% bactopeptone, 1% glucose, 1% NaCl, 150 mM sorbitol, 1.7% bacter agar, 5 μg/ml cholesterol and 50 mM sodium citrate, pH 5) [28]. These phenazine concentrations correspond to the amount of PCA and 1-hydroxy-phenazine that are produced under “fast kill” conditions [28]. E. coli OP50 was used as the food source. The modified “fast kill” media pH 5.0 plus 1% DMSO was used as the control condition. These assays were incubated at 21–23°C.

Quantitative RT-PCR (qRT-PCR) and NanoString nCounter Gene Expression Analyses

Synchronized L1 staged C. elegans N2 animals were grown to L4/young adult stage on the indicated RNAi strain, transferred to assay plates and incubated at 25°C for 24 hours. To prepare the assay plates, 70 μM RPW-24 or DMSO was added to 20 mL “slow killing” media [44] in 10 cm petri dishes seeded with E. coli OP50. N2 and pmk-1(km25) animals were raised on E. coli OP50 and exposed to the above conditions for 18 hours at 20°C. For qRT-PCR studies of nematodes infected with P. aeruginosa PA14 or the indicated strain of P. aeruginosa, 20 mL of “slow killing” media containing either DMSO or 70 μM RPW-24 was added to 10 cm petri dishes. Plates were seeded with either 75 μL of E. coli OP50 or P. aeruginosa, each from cultures grown for 15 hours at 37°C. The plates were incubated for 24 hours at 37°C and 24 hours at 25°C. L4/young adult animals were added to the assay plates and incubated at 25°C for eight hours. RNA was isolated using TriReagent [14] and analyzed by NanoString nCounter Gene Expression Analysis (NanoString Technologies) using a “codeset” designed by NanoString that contained probes for 118 C. elegans genes (Table S2). Probe hybridization, data acquisition and analysis were performed according to instructions from NanoString with each RNA sample normalized to the control genes snb-1, ama-1 and act-1. For the qRT-PCR studies, RNA was reverse transcribed to cDNA using the Retroscript kit (Life Technologies) and analyzed using a CFX1000 machine (Bio-Rad) with previously published primers [6, 18]. The qRT-PCR primers for the mdt-15, pmk-1, irg-1 and sek-1 genes were designed for this study and are available upon request. All values were normalized against the control gene snb-1. Fold change was calculated using the Pfaffl method [45].

Microscopy

Nematodes were mounted onto agar pads, paralyzed with 10 mM levamisole (Sigma) and photographed using a Zeiss AXIO Imager Z1 microscope with a Zeiss AxioCam HRm camera and
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Axiovision 4.6 (Zeiss) software. For comparisons of GFP expression in the F08G5.6::GFP transgenic animals, photographs were acquired using the same imaging conditions.

Statistical Analyses

Differences in survival of C. elegans animals in the P. aeruginosa pathogenesis assays were determined with the log-rank test in each of two biological replicates. Differences were considered significant only if the p value was less than 0.05 for both replicates. In the manuscript, data from one experiment that is representative of both replicates is shown and the sample sizes for these experiments are given in Table S3. To determine if the increase in survival conferred by RPW-24 treatment was different in one population compared to another, we examined the difference in the effect of RPW-24 treatment on the hazard in each group using a Cox proportional hazard model (Stata13, Stata, College Station, TX) from two biological replicates, as previously described [9]. Fold changes in the qRT-PCR analyses were compared using unpaired, two-tailed student t-tests.

Accession Numbers

Accession numbers for genes and gene products are given for the publically available database Wormbase (http://www.wormbase.org). The accession numbers for the principal genes mentioned in this paper are: afi-7 (C07G2.2), C32H11.1, cyp-33A1 (C036G14.14), cyp-35B2 (K07G3.3), cyp-35C1 (C06G3.3), F35E12.5, F08G5.6, F01D5.5, fsh-1 (C50H2.1), ig-1 (C07G5.2), ig-2 (C49G7.5), mdt-15 (R12B2.5), nyc-1 (F59A5.1), pmk-1 (B0218.3), sek-1 (R03G5.2), skn-1 (T19E7.2), tir-1(F13B10.1), and zip-2 (K02F3.4). Other accession numbers are given in Figure 2, Figure 6, Table S1 and Table S2.

Supporting Information

Accession numbers for genes and gene products are given for the publically available database Wormbase (http://www.wormbase.org). The accession numbers for the principal genes mentioned in this paper are: afi-7 (C07G2.2), C32H11.1, cyp-33A1 (C036G14.14), cyp-35B2 (K07G3.3), cyp-35C1 (C06G3.3), F35E12.5, F08G5.6, F01D5.5, fsh-1 (C50H2.1), ig-1 (C07G5.2), ig-2 (C49G7.5), mdt-15 (R12B2.5), nyc-1 (F59A5.1), pmk-1 (B0218.3), sek-1 (R03G5.2), skn-1 (T19E7.2), tir-1(F13B10.1), and zip-2 (K02F3.4). Other accession numbers are given in Figure 2, Figure 6, Table S1 and Table S2.

Supporting Information

Figure S1 P. aeruginosa strains with greater virulence towards C. elegans cause more robust induction of the immune response gene F08G5.6. (A) Wild-type N2 C. elegans were exposed for eight hours to eight P. aeruginosa strains, each with a different virulence potential toward C. elegans [13]. Data are the average of two biological replicates, each normalized to a control gene with error bars representing SEM and are presented as the fold induction of F08G5.6 compared to its expression in C. elegans exposed to E. coli OP50. *p<0.05 for the difference from the F08G5.6 induction in the most virulent P. aeruginosa strain (CF18). (B) Worms carrying the F08G5.6::GFP reporter were exposed to P. aeruginosa PA14 and PAK for 18 hours, and photographed under the same conditions.

Figure S2 The Mediator subunit MDT-15 regulates immune and detoxification genes in response to RPW-24. (A) The expression of putative C. elegans immune effector genes was analyzed by qRT-PCR in vector control (L4440) and mdt-15(RNAi) animals exposed to either 70 μM RPW-24 or the solvent control DMSO, as indicated. The difference in expression of each of the genes in both mdt-15(RNAi) conditions was significantly different from vector control (L4440) pt<0.001). (B) The RPW-24-induced expression levels for the indicated genes were compared in mdt-15(tm2182) mutant animals and wild-type controls. Data are presented as the average expression value of the indicated gene following RPW-24 exposure in mdt-15(tm2182) animals versus wild-type animals. (C) The expression levels of three components of the p38 MAP kinase PMK-1 pathway were compared in animals exposed to 70 μM RPW-24 or DMSO. (D) RPW-24-mediated induction levels of cyp-35B1, cyp-35B2 and cyp-35C1 were assessed in pmk-1(tm25) and wild-type N2 animals as described above. There was no significant difference in the induction of these genes (p>0.05). All data in this figure are the average of two or three replicates, each normalized to a control gene with error bars representing SEM. For the analyses in A and C, data are presented as the value relative to the average expression from all replicates of the indicated gene in the baseline condition [L4440 (A) or N2 (C) animals exposed to DMSO].

Figure S3 The Mediator subunit MDT-15 is required for defense against P. aeruginosa infection. C. elegans wild-type N2 (A) and fer-15(h26)严重影响 (hcl17) sterile (C) animals were exposed to vector control (L4440) or mdt-15(RNAi) and infected with P. aeruginosa in the presence of the solvent control DMSO or 70 μM RPW-24. There is a significant difference in susceptibility to P. aeruginosa infection between the L4440/DMSO and mdt-15(RNAi)/DMSO conditions in A and C (p<0.001). In addition, the magnitude of lifespan prolongation conferred by RPW-24 during P. aeruginosa infection was significantly reduced in mdt-15(RNAi) animals compared to control animals for both assays (p<0.001). (B) C. elegans wild-type N2 or mdt-15(tm2182) animals were infected with P. aeruginosa in the presence of the solvent control DMSO or 70 μM RPW-24. There is a significant difference between the N2/DMSO and mdt-15(tm2182)/DMSO conditions, which was also significant in a biological replicate of this experiment (p<0.001). The degree of lifespan extension conferred by RPW-24 exposure during P. aeruginosa infection was reduced in mdt-15(tm2182) animals compared to wild-type animals, but the difference did not reach statistical significance (p=0.09). In A, B and C, data at each time point are the average of three plates per strain, each with approximately 30–40 animals per plate (see Table S3 for sample sizes). Each assay presented in this figure is representative of two independent experiments.

Figure S4 Protection from the toxic effects of the xenobiotic RPW-24 requires MDT-15, but not PMK-1. The percentage of the population of animals that were at the indicated development stage after 70 hours of incubation at 20°C is presented. Adult animals all had a fully formed vulva and were divided into three categories based on the number of eggs carried in the animal. Larval stages were determined based on microscopic examination of the gonad. See Table S3 for the sample sizes. (EPS)

Figure S5 Neither MDT-15 nor PMK-1 are required for the behavioral avoidance response to RPW-24. (A) The percentage of wild-type N2, pmk-1(tm25) and mdt-15(tm2182) animals that were off a lawn of E. coli OP50 supplemented with either DMSO or 20 μg RPW-24 (average of three replicates) 16 hours after exposure is presented with error bars representing standard deviation. There is no significant difference in the percentage of animals off a lawn containing RPW-24 among the conditions in this experiment (p>0.05). See Table S3 for the sample sizes. (B) Representative photographs of each condition are shown. (EPS)

Figure S6 Resistance to P. aeruginosa phenazine toxins requires MDT-15. The controls for the experiment in Figure 7 are presented. (A) C. elegans wild-type N2 and mdt-15(tm2182) animals at the L4 stage are sensitive to killing by P. aeruginosa in the “fast killing” toxic phenazine assay. (B) Likewise, L4 staged nematodes are rapidly killed by toxic phenazines added to assay
media in the absence of pathogen. These experiments were conducted in parallel with and as described for the data presented in Figure 7.

EPS

Table S1 Genes identified in an RNAi screen for regulators of the p38 MAP kinase PMK-1 pathway. (A) Presented are the 29 genes that were identified in a screen of 1,420 RNAi clones, the complete list of which is given (B), because they reduce or abrogate the induction of the p38 MAP kinase PMK-1-dependent immune reporters F08G5.6::GFP and F35E12.5::GFP. Knockdown of these genes does not affect the induction of mg-1::GFP by an E. coli strain engineered to express ToxA. qRT-PCR of three replicates was used to confirm that the RNAi-mediated knockdown of the indicated gene in wild-type animals reduced the induction of F08G5.6 and F35E12.5 by RPW-24. We previously showed that aff-7 and pmk-1 are required for the RPW-24-mediated induction of F08G5.6 and F35E12.5 [9]. These 29 genes were identified in an RNAi screen of 1,420 clones, which are presented in (B). Of note, the 29 genes presented in (A) were sequenced confirmed.

(XLSX)

Table S2 Relative expression of the 118 genes in the nCounter Gene Expression Analysis. Nanosting nCounter Gene Expression Analysis data for (A) N2 wild-type and pmk- (km25), and (B) Vector control (L4440) and mdt-15(RNAi), each exposed to DMSO or RPW-24, are presented. Expression values are the average of two independent replicates each of which was normalized to three control genes and are presented as the value relative to the average expression of the indicated gene in the baseline condition [N2 animals (A) or vector control (L4440) animals (B) exposed to DMSO]. p values for the comparison of the indicated conditions are presented.

(XLSX)

Table S3 Samples sizes for the assays.

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Author Contributions

Conceived and designed the experiments: RPW RLF FMA. Performed the experiments: RPW RLF DLM ALC. Analyzed the data: RPW RLF. Contributed reagents/materials/analysis tools: RPW RLF DLM ALC. Wrote the paper: RPW RLF FMA.

References


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