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Protective Effects of C-Type Natriuretic Peptide on Linear Growth and Articular Cartilage Integrity in a Mouse Model of Inflammatory Arthritis

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Objective. The C-type natriuretic peptide (CNP) signaling pathway is a major contributor to postnatal skeletal growth in humans. This study was undertaken to investigate whether CNP signaling could prevent growth delay and cartilage damage in an animal model of inflammatory arthritis.

Methods. We generated transgenic mice that overexpress CNP (B6.SJL-Col2a1-NPPC) in chondrocytes. We introduced the CNP transgene into mice with experimental systemic inflammatory arthritis (K/BxN T cell receptor [TCR]) and determined the effect of CNP overexpression in chondrocytes on the severity of arthritis, cartilage damage, and linear growth. We also examined primary chondrocyte cultures for changes in gene and protein expression resulting from CNP overexpression.

Results. K/BxN TCR mice exhibited linear growth delay ($P < 0.01$) compared to controls, and this growth delay was correlated with the severity of arthritis. Diminished chondrocyte proliferation and matrix production was also seen in K/BxN TCR mice. Compared to non–CNP-transgenic mice, K/BxN TCR mice with overexpressed CNP had milder arthritis, no growth delay, and less cartilage damage. Primary chondrocytes from mice overexpressing CNP were less sensitive to inflammatory cytokines than wild-type mouse chondrocytes.

Conclusion. CNP overexpression in chondrocytes can prevent endochondral growth delay and protect against cartilage damage in a mouse model of inflammatory arthritis. Pharmacologic or biologic modulation of the CNP signaling pathway may prevent growth retardation and protect cartilage in patients with inflammatory joint diseases, such as juvenile idiopathic arthritis.

Linear growth results from the action of multiple signaling pathways. The pathway that includes C-type natriuretic peptide (CNP) contributes to chondrocyte proliferation, differentiation, and matrix synthesis in skeletal growth plates (1–4). Patients with homozygous loss-of-function mutations in natriuretic peptide receptor B (NPR-B), the CNP receptor, have acromesomelic dysplasia (5), and carriers of the heterozygous mutation frequently have short stature (6). Also, patients with chromosome translocations that cause CNP overexpression develop skeletal overgrowth (7). These conditions indicate a dose-dependent effect of CNP signaling on linear growth. Therefore, linear growth may be affected...
in patients with acquired diseases that alter the production of CNP or the expression of its receptor, NPR-B. Children with juvenile inflammatory arthritis (JIA) have impaired linear growth, and 40% develop bone degeneration and deformity adjacent to affected joints (8–11). The mechanism by which chronic systemic inflammation suppresses linear growth in children with JIA is not completely understood (12, 13). Alterations in growth hormone and insulin-like growth factor (IGF) signaling pathways have been associated with growth delay in children with chronic arthritis. Although serum growth hormone levels were unchanged in patients with JIA, patients did have reduced levels of IGF-1 and IGF binding protein 3 (IGFBP-3) (14), suggesting that inflammation causes growth hormone resistance. The CNP signaling pathway has not been studied in children with JIA.

Animal models of inflammatory arthritis have been used to delineate pathways that contribute to impaired linear growth and joint deformity (15). Local elevations of tumor necrosis factor α (TNFα), interleukin-6 (IL-6), and IL-1β levels have been observed in affected growth plates. Furthermore, overexpression of IL-6 (16) or TNFα (17) in transgenic mice causes systemic inflammatory arthritis and growth retardation. By 25 days of age, K/BxN T cell receptor (TCR) mice exhibit pronounced joint inflammation that resembles rheumatoid arthritis (17–19). Herein, we report that these mice also develop linear growth delay and cartilage damage that can be lessened by overexpression of CNP in chondrocytes.

MATERIALS AND METHODS

These experiments were approved by the Institutional Animal Care and Use Committee at Case Western Reserve University.

Generation of CNP-transgenic mice. We cloned full-length human CNP (NPPC) complementary DNA (cDNA) into the pKN185 vector, which drives CNP expression under the control of Col2a1 promoters and enhancers (20). Col2a1-NPPC–transgenic mice were produced following pronuclear injection into zygotes on a mixed C57BL/6J-SJL background. NPPC–transgenic mice were produced following pronuclear injection into zygotes on a mixed C57BL/6J-SJL background. One founder animal that had ~4 copies of the CNP transgene integrated into a single locus, as determined by Southern blotting and Mendelian segregation of the transgene, was then backcrossed for >20 generations onto the C57BL/6J-SJL background. C57BL/6J mice were also used as wild-type (WT) controls.

Development of the K/BxN TCR mouse model of arthritis. KRN TCR–transgenic mice, as well as K/BxN TCR mice that develop spontaneous arthritis, have been previously described (21–23). KRN TCR–transgenic mice were a gift from CBDM Lab (Joslin Diabetes Center/Harvard Medical School). K/BxN TCR mice were obtained by crossing KRN TCR–transgenic mice with nonobese diabetic mice (NOD/ShiLtJ; The Jackson Laboratory). Only offspring that inherited the TCR transgene developed inflammatory arthritis on this background; mice that did not inherit the TCR transgene were used as nontransgenic controls.

Breeding the CNP transgene into the mice with experimental arthritis. We bred Col2a1-NPPC–transgenic mice with KRN TCR mice. Offspring that carried both the Col2a1-NPPC and the TCR transgenes were then bred with NOD mice. The phenotypes of offspring that were double heterozygous for the TCR and Col2a1-NPPC transgenes (K/BxN TCR, Col2a1-NPPC) were compared to the phenotypes of offspring that only inherited the TCR transgene (K/BxN TCR). Other offspring resulting from this cross included mice heterozygous for the Col2a1-NPPC transgene only and mice that inherited neither transgene (BxN). Offspring were examined at birth and weekly, from 3 weeks of age until 20 weeks of age.

Small-animal radiography. A Faxitron radiographic inspection unit (model 8050-010; Field Emission) was used to obtain radiographic images of the mice postmortem. Legs were exposed for 1.5 minutes at 35 kVp; the entire body was exposed for 1.5 minutes at 30 kVp.

Clinical scoring of arthritis severity. The severity of clinical arthritis was determined using a previously described scoring system (24, 25), with modifications (available online at http://www.case.edu/artsci/biol/skeletal/24114569s.html). Higher scores indicate increased severity of clinical arthritis.

Sample collection. Mice were bled under sedation and then euthanized. Samples were collected from mice that were 1, 2, 3, 4, 6, 8, 12, 16, and 24 months old. To evaluate the growth plates, one hind leg (femur and tibia) from each mouse was dissected, fixed with 4% formaldehyde in phosphate buffered saline (PBS) for 24 hours at 4°C, decalcified with 0.5M EDTA for 1 week, and then embedded in paraffin. Coronal sections measuring 4 μm across the femorotibial joint were stained with Safranin O–fast green or hematoxylin and eosin and analyzed by immunohistochemical staining or in situ hybridization. For accurate measurement of articular cartilage and growth plate cartilage, sections were obtained at the point where the anterior cruciate ligament inserts into the tibia.

Histologic scoring system. Histologic evaluation of inflammation in the knee joint and cartilage integrity was performed using two scoring systems. The histologic scoring system described by Pettit et al (26) was used to assess the severity of inflammatory arthritis. The International Cartilage Repair Society scoring system (27) was used to assess the status of the extracellular matrix repair in joint cartilage (available online at http://www.case.edu/artsci/biol/skeletal/24114569s.html). An investigator who was not otherwise involved with the study evaluated the mouse knee sections under blinded conditions and assigned a score based on these systems.

Cell proliferation analysis by bromodeoxyuridine (BrdU) labeling. BrdU (300 mg/kg dose; Zymed) was injected intraperitoneally. Mice were euthanized 2 hours after injection. Tissue was processed and embedded in paraffin as
described above. BrdU incorporation was detected using a BrdU-staining kit according to the instructions of the manufacturer (Zymed). The percentage of BrdU-positive cells was determined by dividing the number of BrdU-positive chondrocytes by the total number of chondrocytes that were counted in multiple sections of the growth plate.

**In situ hybridization.** Slides were deparaffinized and fixed in 4% formaldehyde. Sections were digested with protease K (1 μg/ml) for 20 minutes at 37°C and acetylated in 0.25% acetic anhydride in 0.1M triethanolamine HCl. After fixation in 4% formaldehyde, sections were hybridized with 32P-labeled riboprobes in hybridization buffer (50% deionized formamide, 300 mM NaCl, 20 mM Tris HCl, pH 8.0, 5 mM EDTA, 0.5 mg/ml yeast transfer RNA, 10% dextran sulfate, and 1× Denhardt’s solution) in a humidified chamber at 55°C overnight. After hybridization, slides were washed with 5× saline–sodium citrate (SSC) at 50°C, 50% formamide, 2× SSC, and 1× EDTA, 0.5 mg/ml yeast transfer RNA, 10% dextran sulfate, and 1× Denhardt’s solution in a humidified chamber at 65°C, and 1× washing with Tris HCl, pH 8.0, 5 mM EDTA, 0.5 mg/ml yeast transfer RNA, 10% dextran sulfate, and 1× Denhardt’s solution in a humidified chamber at 37°C overnight. After hybridization, slides were washed with 5× saline–sodium citrate (SSC) at 50°C, 50% formamide, 2× SSC at 65°C, and 1× NTE (0.5 M NaCl, 10 mM Tris HCl, pH 8.0, 5 mM EDTA) at 37°C, and then treated with RNase A (20 μg/ml) and RNase T1 (1 unit/μl) in 1× NTE at 37°C for 20 minutes. Slides were further washed in 1× NTE at 37°C, 50% formamide, 2× SSC at 65°C, 2× SSC, 0.1× SSC, and then dehydrated with graded concentrations of ammonium acetate and ethanol. Slides were dipped in NTB emulsion (Kodak) and exposed to the emulsion for 6 weeks, after which they were developed and counterstained with Hoechst 33258 (Sigma). The NPPC probe was made from the pKN185-hCNP vector construct that was originally used to create the transgenic mice. A 5’ 280-bp fragment of the human CNP cDNA sequence plus 118 bp of upstream vector sequence was used for sense and antisense probes (398 bp).

**Primary chondrocyte isolation and cytokine treatment.** Primary chondrocytes from rib cages of ~5-day-old mice were isolated by enzymatic digestion using standard methods (28). Briefly, rib cages were collected in sterile PBS. The ribs were rinsed several times before digestion with 10 ml of 3 mg/ml collagenase B (Worthington) in Dulbecco’s modified Eagle’s medium (DMEM) for ~1 hour at 37°C. The ribs were then washed several times with PBS and digested overnight at 37°C with collagenase solution (2 ml of 3 mg/ml collagenase B/DMEM plus 10 ml DMEM with 10% fetal bovine serum). The cell suspension was strained and the cells were pelleted at 1,000 revolutions per minute for 10 minutes and then resuspended in DMEM containing 10% fetal bovine serum, penicillin (100 units/ml), streptomycin (100 μg/ml), and amphotericin B (0.25 μg/ml). Viable cell counts were determined by trypsin blue staining, and cells were seeded at a density of 1 × 106 cells per 35-mm culture plate in 2.5 ml DMEM (~1.2 × 105 cells/cm²). Cells were grown in culture medium until ~90% confluent, with a change of medium every 3 days. Once cells became ~90% confluent, chondrocyte cultures were serum-starved and treated with cytokines (10 ng/ml TNFα or 10 ng/ml IL-1β; both from R&D Systems) overnight (18 hours) and messenger RNA (mRNA) and proteins were extracted as previously described (29).

**Western blotting.** Cells were lysed in 500 μl of buffer (50 mM Tris HCl, pH 7.4, 150 mM NaCl, 1% Triton X-100, 0.1% sodium dodecyl sulfate, 0.5% sodium deoxycholate, 1 mM EDTA, 1 mM EGTA, and Complete protease inhibitor cocktail), and total lysate protein (60 μg/lane) was resolved by sodium dodecyl sulfate–polyacrylamide gel electrophoresis, transferred to nitrocellulose membranes (Bio-Rad), blocked with nonfat dry milk, resuspended in Tris buffered saline, and probed with primary and secondary antibodies as previously described (29,30). Primary antibodies included anti-phosphorylated and anti-nonphosphorylated p38 MAPK and ERK-1/2 (p38 MAPK phosphorylated #4631, p38 MAPK nonphosphorylated #9212, phospho–ERK-1/2–phospho-MAPK [Thr202/Tyr204] [Cell Signaling Technologies]) and anti–NPR-B (sc-25486; Santa Cruz Biotechnology). Immunoreactive proteins were visualized by using 1:1,000 diluted horseradish peroxidase (HRP)–linked secondary antibodies and enhanced chemiluminescence (GE Healthcare) as described (31). HRP-conjugated goat anti-rabbit IgG at a dilution of 1:2,000 (sc-2004; Santa Cruz Biotechnology) was used as the secondary antibody for NPR-B.

**RNA extraction and cDNA synthesis.** Total cytoplasmic RNA was prepared from primary chondrocytes using an RNeasy kit (Qiagen). RNA (2 μg) was reverse transcribed using a SuperScript II reverse-transcriptase kit (Invitrogen), and the cDNA mixture was diluted 5-fold in nuclease-free water; 5 μl was used for real-time quantitative polymerase chain reaction (qPCR).

**Real-time qPCR.** Using a StepOnePlus thermocycler (Applied Biosystems), reactions were performed in a 20-μl volume with 0.5 μM of each primer and SYBR Green Master Mix according to the instructions of the manufacturer (Qiagen). The expression level of the housekeeping gene Actb was measured (31). HRP-conjugated goat anti-rabbit IgG was used to normalize mRNA expression. Information on the primers used to amplify specific cDNA sequences is available online at http://www.case.edu/artsci/biol/skeletal/24114569s.html. All reactions were performed in duplicate.

**Statistical analysis.** Disease severity and histopathologic scores were compared between groups, using chi-square tests for categorical variables (clinical and histologic scores) and t-tests for continuous variables. Results are expressed as the mean ± SD. To test differences between the groups of mice, data on the osteo–tail tip length of mice were examined by analysis of variance (ANOVA). ANOVA F test results were reported. P values less than 0.05 were considered significant. We also used Pearson correlation matrices to test for correlations between the clinical and histopathologic data. The data were analyzed using SAS 9.0 and Stata 11.0.

**RESULTS**

**Col2a1-NPPC–transgenic mice exhibit linear bone overgrowth.** Mice carrying the CNP transgene driven by the Col2a1 promoter/enhancer (Col2a1-NPPC) demonstrated skeletal overgrowth at 4 weeks of age (available online at http://www.case.edu/artsci/biol/skeletal/24114569s.html). Overgrowth affected the long bones and the vertebrae (Figures 1A and B) and was associated with increases in growth plate width, the size of hypertrophic chondrocytes (P < 0.01 by F test) (Figures 1C and D), nose-to–tail tip length (Figure 1E),
and the number of proliferating chondrocytes ($P < 0.01$ by F test) (Figure 1F). The mean ± SD width of the growth plate was 148 ± 14 μm in Col2a1-NPPC–transgenic mice ($n = 12$) and 99 ± 5.3 μm in nontransgenic control mice ($n = 5$) at 4 weeks of age ($P < 0.05$ by Student’s t-test). CNP transgene expression in growth plate and articular chondrocytes was detected by in situ hybridization (Figures 1G and H). Col2a1-NPPC–transgenic mice developed thoracic kyphosis and joint dislocations over time, but no abnormalities outside of the skeletal system were detected.

**Arthritic K/BxN TCR mice exhibit linear growth delay and articular cartilage damage.** K/BxN TCR mice developed arthritis by age 3 weeks and exhibited visible growth delay by age 12 weeks when compared to littermates that did not inherit the TCR transgene (BxN) (Figure 2A). Significant differences in linear growth (as determined by nose-to–tail tip length) between K/BxN TCR mice and BxN littermates were detected by 4 weeks of age ($P < 0.05$ by F test) (Figure 2B). To prevent length discrepancies, we did not obtain tissue for DNA isolation by clipping tails in order to genotype the mice. In the
K/BxN TCR mice, the severity of arthritis was inversely correlated with longitudinal growth; this correlation was strongest at age 14 weeks (available online at http://www.case.edu/artsci/biol/skeletal/24114569s.html). The growth plates in the K/BxN TCR mice were narrower and had fewer cells than growth plates in the BxN littermates (Figure 2C). The articular cartilage in K/BxN TCR mice also became less cellular, had less cartilage matrix, and had a more irregular surface than the articular cartilage in BxN mice (Figures 2D and E).

CNP overexpression in chondrocytes improves linear growth and reduces articular cartilage damage in K/BxN TCR mice. Since CNP overexpression enhanced linear growth in Col2a1-NPPC–transgenic mice, we sought to determine whether overexpression could protect against growth impairment in mice with inflammatory arthritis. We first bred Col2a1-NPPC–transgenic mice onto the KRN TCR background and then onto the K/BxN background (Figure 3A).

In K/BxN TCR mice that overexpressed CNP...
growth retardation was seen (P < 0.05 by F test) (Figure 3B), growth plates were increased in width (Figure 3C and online at http://www.case.edu/artsci/biol/skeletal/24114569s.html), and long bones were increased in length (Figure 3D) compared to K/BxN TCR arthritic mice. The increase in growth plate width in mice with the Col2a1-NPPC transgene was associated with an increase in BrdU incorporation (http://www.case.edu/artsci/biol/skeletal/24114569s.html). CNP overexpression reduced the clinical arthritis score in the K/BxN TCR mice, with a mean ± SD arthritis score of 4.37 ± 1.38 in CNP-overexpressing mice (n = 8) compared to 8.66 ± 3.26 in the K/BxN TCR mice (n = 14) (P < 0.05 by t-test).

CNP overexpression also reduced the severity of articular cartilage inflammation and damage in the K/BxN TCR mice (Figure 3E). Thirteen-week-old male mice with CNP overexpression had better articular cartilage chondrocyte distribution and organization and cartilage matrix content than the controls. Although CNP overexpression appeared to protect articular cartilage against damage caused by inflammation (Figures 4A–C), overexpression did not reduce inflammation-
related changes that occurred in the synovium (Figure 4D).

**CNP overexpression enhances signal transduction via NPR-B and reduces the sensitivity of this signaling pathway to the proinflammatory cytokines TNFα and IL-1β.** We performed cGMP enzyme-linked immunosorbent assays, real time qPCR, and Western blotting using cultures of rib cartilage primary chondrocytes harvested from Col2a1-NPPC–transgenic and WT control mice. The mean ± SD intracellular cGMP level in the Col2a1-NPPC–transgenic mouse chondrocytes was ~8 ± 0.7-fold higher than in the WT mouse chondrocytes, confirming enhanced CNP signaling. In Col2a1-NPPC–transgenic mouse chondrocytes, levels of protein and mRNA for NPR-B (Npr2) were increased (Figures 5, 6A, and online at http://www.case.
transgenic mice and control mice with the proinflamma-
treated primary chondrocytes from Col2a1-NPPC–
in vivo against the damaging effect of inflammation, we
transgenic mice appeared to protect articular cartilage
mouse chondrocytes (Figure 6C).

MMP-13 was mildly increased in Col2a1-NPPC–
chondrocytes compared to WT
mice (available online at http://www.case.edu/artsci/biol/skeletal/24114569s.html). In addition, as has
been previously reported (32), we confirmed that p38
phosphorylation in WT mouse chondro-
ment with TNFα and treatment with IL-1β increased
p38 MAPK phosphorylation in WT mouse chondro-
cytes. The expression of mRNA for the MMPs was
either the same or less elevated in Col2a1-NPPC–
transgenic mouse chondrocytes compared to WT mouse chondrocytes following exposure to these cytokines (P >
0.05) (Figure 6C).

It has been reported that IGF-1, IGF receptor I,
and IGFBP-3 levels are diminished in serum of chil-
dren with JIA, accounting for the delay in longitudinal
growth. To understand whether CNP overexpression
affects Igf1, Igf1r, and Igfbp3 mRNA expression, we
performed real-time qPCR using primary chondrocyte
cultures of rib cartilage harvested from Col2a1-NPPC–
transgenic and WT mice. Col2a1-NPPC–transgenic
mouse primary chondrocytes did not have increased
levels of mRNA for Igf1 or Igf1r, but did have ~6–7-fold
increased levels of mRNA for Igfbp3 compared to WT
mouse chondrocytes. When exposed to TNFα (10 ng/ml)
and IL-1β (10 ng/ml) overnight, Igf1 and Igf1r expression
levels in chondrocytes were suppressed significantly in
both Col2a1-NPPC–transgenic and WT mouse chondro-
cytes. Interestingly, Igfbp3 expression levels did not
differ between Col2a1-NPPC–transgenic and WT mouse
chondrocytes exposed to proinflammatory cytokines; in
addition, Igfbp3 expression remained up-regulated in the
Col2a1-NPPC–transgenic mouse chondrocytes (available online at http://www.case.edu/artsci/biol/skeletal/
24114569s.html).

Earlier reports indicate that TNFα and IL-1β
inhibit chondrocyte differentiation and DNA synthesis
in growth plate and costal chondrocytes by suppressing
Sox9 expression (33). It is known that CNP up-regulates
Sox9 expression in chondrocytes (34,35). Thus, we
sought to determine whether CNP overexpression in
primary chondrocytes from Col2a1-NPPC–transgenic
mice also up-regulates Sox9 mRNA expression. Results
showed that in primary chondrocytes from Col2a1-
NPPC–transgenic mice, Sox9 expression is up-regulated

duarcticular cartilage in vivo against the damaging effect of inflammation, we
treated primary chondrocytes from Col2a1-NPPC–
transgenic mice and control mice with the proinflamma-
tory cytokines TNFα and IL-1β. TNFα and IL-1β each
significantly reduced Npr2 mRNA expression in both
Col2a1-NPPC–transgenic and WT mouse chondrocytes.
However, Npr2 mRNA expression in Col2a1-NPPC–
transgenic mouse chondrocytes after TNFα or IL-1β
treatment was comparable to Npr2 expression in un-
treated WT mouse chondrocytes (Figure 6A).

In Col2a1-NPPC–transgenic mouse chondro-
cytes, TNFα exposure had no effect on p38 MAPK
phosphorylation, whereas IL-1β reduced p38 MAPK
phosphorylation (Figure 6B). In contrast, both treat-
ment with TNFα and treatment with IL-1β increased
p38 MAPK phosphorylation in WT mouse chondro-
cytes. The expression of mRNA for the MMPs was
either the same or less elevated in Col2a1-NPPC–
transgenic mouse chondrocytes compared to WT mouse chondrocytes following exposure to these cytokines (P >
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showed that in primary chondrocytes from Col2a1-
NPPC–transgenic mice, Sox9 expression is up-regulated

Figure 5. C-type natriuretic peptide overexpression in chondrocytes
increases the expression of genes associated with cartilage anabolism.
Results of quantitative real-time polymerase chain reaction analysis of
primary chondrocyte cultures recovered from Col2a1-NPPC–
transgenic mice and wild-type (WT) mice for several genes associated
with cartilage growth are shown. Values are the mean ± SD. * = P <
0.05.
5-fold as compared to WT mouse chondrocytes. As expected, when chondrocytes were exposed to TNFα overnight, Sox9 expression was reduced in both WT and Col2a1-NPPC–transgenic mouse chondrocytes, but in Col2a1-NPPC–transgenic mouse chondrocytes that reduction brought Sox9 expression to the level seen in untreated WT mice (Figure 6D). We suggest that CNP overexpression in Col2a1-NPPC–transgenic mice protected against down-regulation of Sox9 expression levels during TNFα exposure.

**DISCUSSION**

It has been posited that in children with JIA the inhibitory effects of proinflammatory cytokines (IL-6, TNFα, and IL-1 β) on growth plate chondrocytes contribute to growth suppression (36,37). Elevation of proinflammatory cytokines (e.g., TNFα and IL-1β) in synovial fluid has also been associated with destruction of growth plates adjacent to an arthritic joint. Proinflammatory cytokines decrease the width of the proliferating zone in growth plate cartilage, which leads to a decrease in endochondral bone growth. IL-1β has been shown to reduce expression of cartilage collagens and proteoglycans (13,38).

In this study, we tested whether the growth-promoting effects of the CNP/NPR-B signaling pathway would lessen the severity of growth failure and cartilage degeneration in a mouse model of inflammatory arthri-
chondrocytes when cultures were exposed to TNFα and IL-1β overnight, while Igfbp3 expression remained up-regulated. However, in the absence of a significant change in Igf1 and Igf1r expression in Col2a1-NPPC–transgenic mouse chondrocytes, we do not believe the protective effects of CNP seen in cartilage are due to its regulatory effects on Igf1 signaling.

Earlier reports have indicated that TNFα and IL-1β inhibit chondrocyte differentiation by suppressing DNA synthesis in the growth plate and costal chondrocytes (33). TNFα suppresses DNA synthesis, chondrocyte differentiation, and matrix synthesis by unknown mechanisms (41). It has been suggested that TNFα suppresses expression of Sox9, a key chondrocyte transcription factor, in inflamed cartilage. Our data suggest that CNP might improve chondrocyte proliferation and differentiation during inflammation by its regulatory effect on Sox9 expression. Sox9 mRNA expression was up-regulated 5-fold in chondrocytes of Col2a1-NPPC–transgenic mice. SOX9 and CNP are both known to increase matrix synthesis (42). Although Sox9 expression was significantly reduced following TNFα exposure in Col2a1-NPPC–transgenic mouse chondrocytes, expression was comparable to the level found in untreated WT mouse chondrocytes. Thus, it can be speculated that CNP expression overcomes the effect of proinflammatory cytokines on cartilage at the level of transcription factor Sox9 expression. Sox9 regulates chondrocyte differentiation, which is needed not only for longitudinal growth, but also for the matrix production that maintains the cartilage integrity.

We also cannot rule out the possibility that CNP overexpression directly modulates the inflammatory response. It has been reported that CNP inhibits cytokine-induced leukocyte rolling and has an antiplatelet/antithrombotic effect (43). Reports suggest that CNP secretion by vascular endothelial cells is increased in response to inflammatory stimuli such as IL-1β, TNFα, and lipopolysaccharide (44,45). Additionally, CNP suppresses lipopolysaccharide-activated murine macrophage secretion of prostaglandin E2 (46) and inhibits vascular inflammation and intimal hyperplasia in experimental vein grafts (47). Therefore, it is possible that CNP overexpression protects growth and prevents joint damage by diminishing chronic inflammation. Consequently, studies investigating the antiinflammatory effects of CNP in models of arthritis are warranted. While our data demonstrate that linear growth retardation in K/BxN TCR mice can be prevented by CNP overexpression, it remains to be determined whether this protective
effect will be observed in other animal models of inflammation and whether pharmacologic manipulation of the CNP signaling pathway will have similar effects.

Reduction in linear growth is one complication of joint inflammation. Another common complication is widening of the metaphysis, which has been attributed to neovascularization and VEGF overexpression in the growth plate. CNP may play a role in this neovascularization, since chondrocytes that overexpress CNP were shown to produce increased Vegf mRNA levels (Figure 5), and VEGF immunostaining revealed an area that encompasses hypertrophic and proliferating chondrocytes, as well as the trabecular bone where the blood vessels reside (available online at http://www.case.edu/artsci/biol/skeletal/24114569s.html). Consequently, Col2a1–NPPC–transgenic mice developed widening at the ends of their long bones. We did not observe worsening periarticular overgrowth in the K/BxN TCR, Col2a1–NPPC–transgenic mice, perhaps because of the protective effect of CNP overexpression on the chondrocyte response to inflammatory cytokines. However, further studies are needed. An analog of CNP has been shown to improve skeletal growth in mice harboring a knockin Fgfr3 allele that causes thanatophoric dysplasia in humans (48). It will be interesting to determine whether this analog will be able to improve the cartilage phenotype in K/BxN TCR mice.

CNP overexpression by mouse chondrocytes prevented endochondral growth delay and reduced articular cartilage damage in a mouse model of systemic inflammatory arthritis. The likely mechanism for this effect is a cell-autonomous increase in chondrocyte differentiation, proliferation, hypertrophy, and matrix production, and a cell-autonomous resistance to the growth-suppressive effects of proinflammatory cytokines. These data suggest that the CNP/NPR-B pathway may represent a novel therapeutic target to preserve growth plate and joint cartilage integrity during systemic inflammatory diseases. Although our findings are of particular relevance to JIA, their impact may extend to other forms of inflammatory arthritis and other inflammatory diseases of childhood that stunt longitudinal growth, such as inflammatory bowel disease.

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AUTHOR CONTRIBUTIONS

All authors were involved in drafting the article or revising it critically for important intellectual content, and all authors approved the final version to be published. Dr. Bükülmez had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Study conception and design. Bükülmez, Khan, Haqqi, Warman.

Acquisition of data. Bükülmez, Khan, Bartels, Ortiz-Lopez.


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