Large-Scale Genome-Wide Association Studies and Meta-Analyses of Longitudinal Change in Adult Lung Function

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Abstract

Background: Genome-wide association studies (GWAS) have identified numerous loci influencing cross-sectional lung function, but less is known about genes influencing longitudinal change in lung function.

Methods: We performed GWAS of the rate of change in forced expiratory volume in the first second (FEV_1) in 14 longitudinal, population-based cohort studies comprising 27,249 adults of European ancestry using linear mixed effects model and combined cohort-specific results using fixed effect meta-analysis to identify novel genetic loci associated with longitudinal change in lung function. Gene expression analyses were subsequently performed for identified genetic loci. As a secondary aim, we estimated the mean rate of decline in FEV_1 by smoking pattern, irrespective of genotypes, across these 14 studies using meta-analysis.

Results: The overall meta-analysis produced suggestive evidence for association at the novel *IL16/STARD5/TMC3* locus on chromosome 15 ($P = 5.71 \times 10^{-7}$). In addition, meta-analysis using the five cohorts with $\geq$3 FEV_1 measurements per participant identified the novel *ME3* locus on chromosome 11 ($P = 2.18 \times 10^{-8}$) at genome-wide significance. Neither locus was associated with FEV_1 decline in two additional cohort studies. We confirmed gene expression of *IL16*, *STARD5*, and *ME3* in multiple lung tissues. Publicly available microarray data confirmed differential expression of all three genes in lung samples from COPD patients compared with controls. Irrespective of genotypes, the combined estimate for FEV_1 decline was 26.9, 29.2 and 35.7 mL/year in never, former, and persistent smokers, respectively.

Conclusions: In this large-scale GWAS, we identified two novel genetic loci in association with the rate of change in FEV_1 that harbor candidate genes with biologically plausible functional links to lung function.

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Introduction

Forced expiratory volume in the first second (FEV_1) is a reliable spirometric parameter that reflects the physiological state of the lungs and airways. Reduced FEV_1 relative to forced vital capacity (FVC), is a defining feature of chronic obstructive pulmonary disease (COPD), a leading cause of death globally.[1] FEV_1 is also a predictor of morbidity and mortality in the general population.[2,3] Lung function reaches its peak in early adulthood, followed by a plateau, and then subsequently declines. As first reported by Fletcher and Peto,[4] decline in lung function is accelerated in smokers, leading to increased risks of COPD and premature death. While cigarette smoking is a key risk factor for accelerated loss of lung function, genetic variation is hypothesized to also play an important role.[5,6] Family and twin studies of the longitudinal change in lung function report heritability estimates between 10 and 39%.[7,8]

Recent large-scale genome-wide association studies (GWAS) identified 26 novel loci for cross-sectional lung function,[9–11] demonstrating the power of GWAS with large sample size to identify common genetic variants with modest effect sizes. However, cross-sectional measurements in adults reflect the combination of maximal attained lung growth and subsequent decline. GWAS that specifically study the longitudinal change in lung function are needed to distinguish the genetic contributions to age-related decline. To date, only one population-based GWAS meta-analysis of longitudinal change in lung function has been reported.[12] Separate analyses were conducted in 1,441 asthmatic and 2,667 non-asthmatic participants; association was found at one novel locus in each analysis, though only the locus in non-asthmatics replicated.

In this study, we conducted primary GWAS of the rate of change in FEV_1 in each of 14 population-based cohort studies from the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) and SpiroMeta consortia, comprising 27,249 adult participants of European ancestry and 62,130 FEV_1 measurements. We then performed meta-analysis of the cohort-specific results, followed up our most statistically significant associations in the AGES-Reykjavík cohort study and the Lung Health Study (LHS) for corroborative evidence, and explored the biological basis for identified associations using cell-specific gene expression studies, and expression quantitative trait loci (eQTL) look-up.

Methods

Study populations

All 14 cohort studies are members of the CHARGE or SpiroMeta Consortium (Table 1). The respective local Institutional Review Boards approved all study protocols, and written informed consent for genetic studies was obtained from all participants. Spirometry tests were performed at baseline and at least one follow-up time point by trained technicians and in accordance with the American Thoracic Society or European Respiratory Society recommendations (Methods S1 in File S1 for further details).[13] FEV_1 measurements meeting acceptability criteria were included in the current study.

Studies performed genotyping following standard quality control measures; imputation was conducted based on the HapMap CEU reference panel to generate genotype dosages for $\sim$ 2.5 million autosomal single nucleotide polymorphisms (SNPs) (Table S1 in File S1).

Statistical analysis

For the analysis of repeated measurement data such as longitudinal change in lung function, mixed effects models offer more flexibility and statistical power than alternative approaches; the model allows for the use of unbalanced data and does not exclude individuals with incomplete records. Each cohort study performed the GWAS using a linear mixed effects model. The model included a random intercept and a random slope, and fixed effects for time (a continuous variable quantifying the time distance between each FEV_1 measurement and baseline), SNP and its interaction with time (SNP-by-time), baseline age, gender, standing height, smoking pattern during follow-up and its interaction with time (smoking-by-time), baseline smoking pack-years, study site, and principal components for genetic ancestry (as needed). Cohort-specific results for the SNP-by-time interaction term, which estimates the effect of genotype on the rate of change in FEV_1, were shared, and two meta-analyses, one using all 14 studies and the other using the five studies with $\geq$3 FEV_1

Table 1. Baseline characteristics of cohort studies included in the meta-analysis*.

Cohort:	**ARIC**	**B58C**	**BHS**	**CARDIA**	**CHS**	**FHS**	**Health ABC**
No. of participants	8,242	827	1,009	1,492	3,159	3,230	1,586
No. of FEV_1 measurements	15,582	1,653	3,073	6,140	7,140	11,275	4,426
No. of FEV_1 per person	2	2	7	5	3	5	4
Follow-up duration, yr	5.6	10	29	20.1	7.9	14.7	9.5
Males, %	46.5	48.6	41.6	46.9	39	47	52.7
Baseline age, yr	54.6 (5.7)	35.0 (0.2)	37.5 (12.8)	27.5 (2.3)	72.3 (5.4)	50.9 (10.3)	73.8 (2.8)
Baseline height, cm	168.7 (9.4)	170.1 (9.5)	168.1 (8.9)	171.2 (9.3)	164.6 (9.4)	168.4 (9.3)	166.8 (9.3)
Current smokers, %	20.2	27.1	20.9	24.8	10.8	24.6	6.4
Former smokers, %	32.6	41.5	16.5	17.3	35.7	39.8	49.9
Baseline pack-years†	25.9 (21.7)	7.5 (11.4)	8.2 (17.8)	6.0 (6.5)	33.2 (27.0)	25.4 (21.3)	36.8 (32.2)
Baseline FEV_1, mL	2972 (758)	3631 (744)	3230 (927)	3818 (781)	2123 (652)	2989 (806)	2308 (649)
Baseline FEV_1/FVC, %	74.1 (7.1)	80.6 (5.8)	78.2 (9.2)	81.6 (6.5)	70.5 (10.5)	75.7 (8.0)	74.7 (7.8)
Cohort:	**KORA**	**LBC1921**	**LBC1936**	**PIVUS**	**RS**	**SAPALDIA**	**SHIP**
No. of participants	890	512	1,002	818	1,321	1,401	1,760
No. of FEV_1 measurements	1,597	706	1,790	1,469	2,016	2,692	2,571
No. of FEV_1 per person	2	2	2	2	2	2	2
Follow-up duration, yr	3.2	8.9	4.8	5.8	8.3	10.9	7.9
Males, %	47.2	41.4	50.8	49.9	45.1	48	49.4
Baseline age, yr	53.8 (4.5)	79.1 (0.6)	69.6 (0.8)	70.2 (0.2)	74.4 (5.6)	41.1 (11.2)	52.4 (13.6)
Baseline height, cm	169.3 (9.3)	163.2 (9.4)	166.5 (8.9)	169.0 (9.3)	167.3 (9.1)	169.4 (9.1)	169.5 (9.7)
Current smokers, %	20.5	7.0	12.9	10.2	11.1	26.9	32.8
Former smokers, %	40.9	50.4	42.6	39.6	56.7	25.8	23.8
Baseline pack-years†	11.2 (17.1)	15.3 (22.3)	16.9 (25.8)	14.3 (15.8)	25.7 (21.3)	17.4 (18.0)	11.3 (11.9)
Baseline FEV_1, mL	3280 (792)	1887 (625)	2371 (687)	2452 (682)	2215 (652)	3516 (861)	3238 (876)
Baseline FEV_1/FVC, %	77.5 (6.2)	79.0 (11.8)	78.3 (10.2)	76.0 (10.0)	74.8 (7.9)	78.5 (8.2)	83.1 (6.6)

Definition of abbreviations: ARIC = Atherosclerosis Risk in Communities; B58C = British 1958 Birth Cohort; BHS = Busselton Health Study; CARDIA = Coronary Artery Risk Development in Young Adults; CHS = Cardiovascular Health Study = FHS, Framingham Heart Study; Health ABC = Health, Aging, and Body Composition; KORA = Cooperative Health Research in the Region of Augsburg; LBC1921 = Lothian Birth Cohort 1921; LBC1936 = Lothian Birth Cohort 1936; PIVUS = Prospective Investigation of the Vasculature in Uppsala Seniors; RS = Rotterdam Study; SAPALDIA = Swiss Study on Air Pollution and Lung Diseases in Adults; SD = standard deviation; SHIP = Study of Health in Pomerania.

*Data are presented as mean (SD) unless otherwise indicated; total no. participants = 27,249, total no. FEV_1 measurements = 62,130.

†Pack-years are calculated among current and former smokers at study baseline.

doi:10.1371/journal.pone.0100776.t001

Table 2. Model estimates for the rate of change in FEV_1 in never smokers and effects of other smoking patterns (compared with never smokers) on the rate of change in FEV_1 (mL/year)*.

Study	Annual FEV_1 change in never smokers		Additional Effect† of smoking patterns on annual FEV_1 change					
	(referent group)		Persistent smokers		Intermittent smokers		Former smokers	
	β	SE	β	SE	β	SE	β	SE
ARIC	−14.0	1.3	−12.4	1.7	−5.5	2.1	−5.3	1.4
B58C	−29.6	1.5	−9.4	2.8	−2.2	3.4	−3.0	3.0
BHS	−23.0	1.0	−20.0	3.0	−8.0	2.0	−9.0	2.0
CARDIA	−26.4	0.5	−6.7	1.3	−0.2	1.0	1.0	1.2
CHS	−35.0	1.1	−2.2	3.3	−4.6	2.2	−2.4	1.7
FHS	−26.0	0.6	−8.1	1.3	−2.9	1.0	−1.1	0.8
Health ABC	−39.7	1.3	−12.9	6.1	−6.8	4.4	−2.6	1.7
KORA	−22.1	3.7	2.2	7.2	−10.4	9.3	2.8	5.2
LBC1921	−10.0	3.6	−11.6	15.7	2.8	14.4	−18.8	4.9
LBC1936	−32.3	3.6	−19.0	9.9	40.1	16.8	4.3	5.3
PIVUS	−21.1	2.5	−15.9	8.2	−21.7	13.4	−3.9	3.9
RS	−27.5	3.7	−1.8	9.0	9.3	8.6	−4.6	4.5
SAPALDIA	−29.7	1.2	−7.4	2.3	−2.0	2.6	−2.8	2.1
SHIP	−31.8	2.8	−0.4	10.9	−0.1	3.9	−15.0	7.3
14-cohort meta-analyzed estimate	−26.9	0.3	−8.8	0.7	−2.6	0.6	−2.3	0.5

Definition of abbreviations: ARIC = Atherosclerosis Risk in Communities; B58C = British 1958 Birth Cohort; BHS = Busselton Health Study; CARDIA = Coronary Artery Risk Development in Young Adults; CHS = Cardiovascular Health Study; FHS = Framingham Heart Study; Health ABC = Health, Aging, and Body Composition; KORA = Cooperative Health Research in the Region of Augsburg; LBC1921 = Lothian Birth Cohort 1921; LBC1936 = Lothian Birth Cohort 1936; PIVUS = Prospective Investigation of the Vasculature in Uppsala Seniors; RS = Rotterdam Study; SAPALDIA = Swiss Study on Air Pollution and Lung Diseases in Adults; SE = standard error; SHIP = Study of Health in Pomerania.

*Data shown are the effect estimates (β and SE) of the time and smoking-by-time interaction terms in the preliminary mixed effects model fully adjusted for all specified variables except the SNP terms. Time represents the rate of change in FEV_1 in never smokers and the smoking-by-time interaction term represents the effects of the other three smoking patterns on the rate of change in FEV_1, compared with never smokers. Smoking categories are defined as persistent (smoke throughout follow-up), intermittent (stop and/or start smoking during follow-up) and former (smoke only prior to start of follow-up).

†Effect estimates in smoking categories are added to estimates in never smokers to compute the actual rate of change in each group (for example, in ARIC, the point estimate of the rate of change in FEV_1 in persistent smokers was −14.0 − 12.4 = −26.4 mL/year).

doi:10.1371/journal.pone.0100776.t002

Table 3. Association of the most statistically significant SNPs with the rate of change in FEV_1 (mL/year) in the meta-analysis of 14 cohort studies (n = 27,249)*.

SNP	Chr	Position	Closest Gene(s)	Coded Allele	Frequency	β	SE	*P* Value
rs12137475	1	44059735	*ST3GAL3*	T	0.11	−3.5	0.8	3.90×10^{-6}
rs766488	1	61583103	*NFIA*	A	0.31	1.4	0.3	6.60×10^{-6}
rs17698444	1	215483178	*ESRRG/GPATCH2*	C	0.89	−2.2	0.5	2.62×10^{-6}
rs12692550	2	159958017	*BAZ2B*	T	0.17	−1.7	0.4	5.16×10^{-6}
rs2260722	13	113236292	*TMCO3*	A	0.72	−1.5	0.3	1.83×10^{-6}
rs4077833	15	79419738	*IL16/STARD5/TMC3*	C	0.10	2.3	0.5	5.71×10^{-7}
rs8027498	15	89595638	*SV2B*	A	0.25	1.4	0.3	9.41×10^{-6}
rs8051319	16	15794449	*MYH11*	T	0.60	1.7	0.3	5.12×10^{-6}
rs740557	17	62451139	*CACNG4*	C	0.85	−2.3	0.5	3.59×10^{-6}

Definition of abbreviations: Chr = chromosome; SE = standard error; SNP = single-nucleotide polymorphism.

*Data reported are the meta-analysis results of the SNP-by-time interaction term from the GWAS mixed effects model. A positive β-coefficient indicates an attenuation of FEV_1 decline and a negative β-coefficient an acceleration of FEV_1 decline.

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measurements per participant, were performed using METAL software with inverse variance weighting to combine effect estimates after applying genomic control correction.[14]

We sought corroborative evidence for SNPs with $P < 1 \times 10^{-5}$ in the AGES-Reykjavík cohort study (n = 1,494), and in LHS (n = 4,048), a clinical cohort study of smokers with mild COPD, in which a longitudinal GWAS was recently reported.[15]

Gene expression analyses

Expression profiles of genes at the novel loci were evaluated in human lung tissues and primary cell samples using RT-PCR (Table S7 in File S1). Using publicly available data from the Lung Genomics Research Consortium (LGRC), expression profiles of these genes were compared in lung specimens of 219 COPD patients and 137 controls, and sentinel (most associated) SNPs at the novel loci were also searched against an eQTL database of lymphoblastoid cell lines.[16]

This manuscript follows the PRISMA statement and a checklist is available online (Checklist S1).

Results

Population characteristics

The majority of the 14 cohort studies had FEV_1 at two times, but five studies (BHS, CARDIA, CHS, FHS, Health ABC) had ≥ 3 FEV_1 measurements per participant. The maximum length of follow-up ranged from 4 to 29 years. Studies with older participants generally had fewer current smokers and more former smokers, and had lower mean baseline FEV_1.

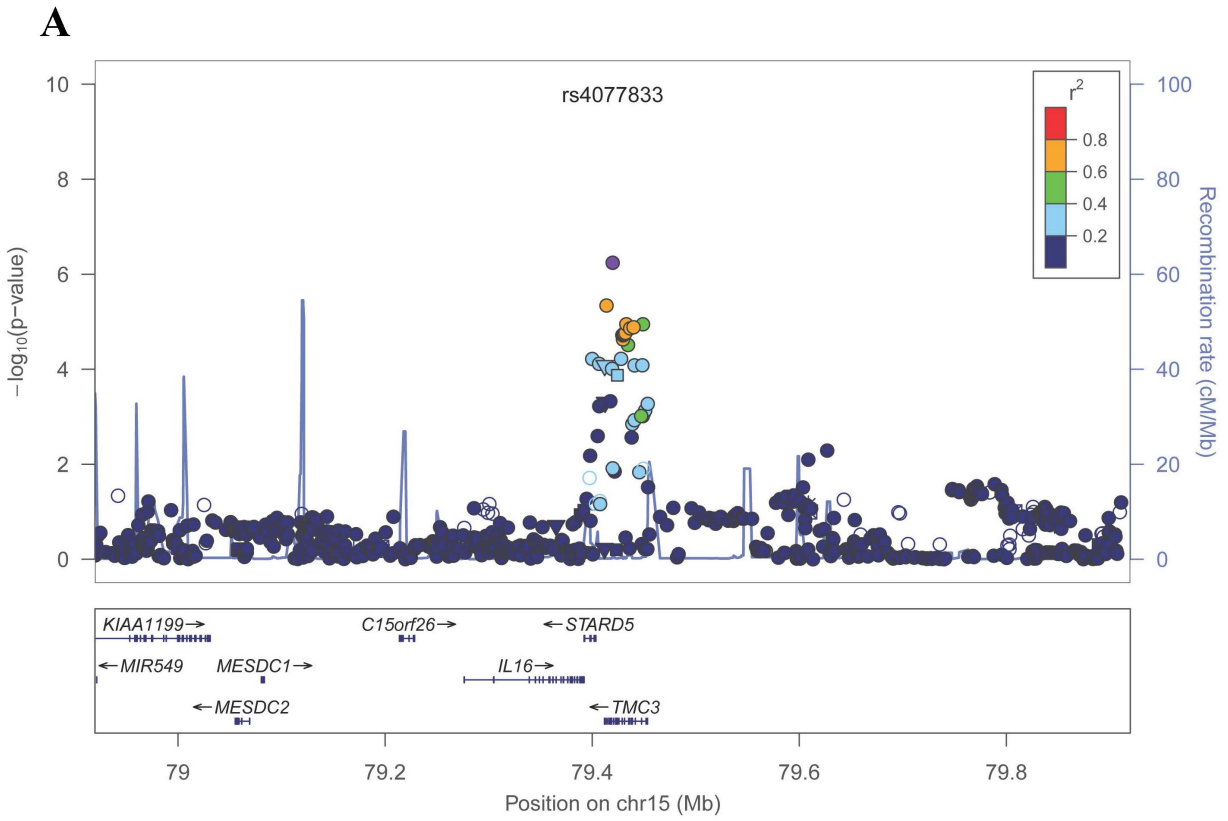
Smoking patterns and rate of decline in FEV_1

All 14 studies implemented a preliminary mixed model adjusted for all specified variables except the SNP terms and reported the estimated rate of change in FEV_1 by smoking pattern (Table 2). The rate of decline in FEV_1 in never smokers ranged from 10.0 to 39.7 mL/year, and was generally steeper in studies with older participants, as expected.[4] Across all 14 studies, the meta-analyzed rate of change in FEV_1 was a decline of 26.9±0.3 mL/year in never smokers, and was 8.8±0.7, 2.6±0.6, and 2.3±0.5 mL/year *steeper* in persistent, intermittent, and former smokers, respectively (Table 2). We repeated the meta-analyses in the five cohort studies with ≥3 FEV_1 measurements per participant, and found similar, although less statistically significant results.

Discovery meta-analyses

Study-specific genomic inflation factors (λ_{gc}) were calculated for the SNP-by-time interaction term and used for study-level genomic control prior to the meta-analyses. Study-specific λ_{gc} values ranged from 0.96 to 1.11 (Table S1 in File S1) and the meta-analysis λ_{gc} was 1.01 for both the 14-study and five-study meta-analyses. Figures S1 and S2 in File S1 present the Manhattan and quantile-quantile (QQ) plots.

In the meta-analysis including all 14 cohort studies, 15 SNPs at nine independent loci were associated with the rate of change in FEV_1 at $P < 1 \times 10^{-5}$, and none reached the genome-wide significance threshold of $P < 5 \times 10^{-8}$. The association results for the sentinel SNPs at these nine loci are presented in Table 3, and more detailed results for all 15 SNPs are included in Table S2 in File S1. The most statistically significant association, and the only one that reached $P < 1 \times 10^{-6}$, was for rs4077833, an intronic SNP located in the novel *IL16/STARD5/TMC3* gene region on chromosome 15 ($P = 5.71 \times 10^{-7}$; Figures 1A and 1B). The C allele of rs4077833, with a frequency of 10%, was associated with


A
rs4077833
r2
0.8
0.6
0.4
0.2
−log10(p−value)
Recombination rate (cM/Mb)
KIAA1199
C15orf26
STARD5
MIR549
MESDC1
IL16
MESDC2
TMC3
Position on chr15 (Mb)

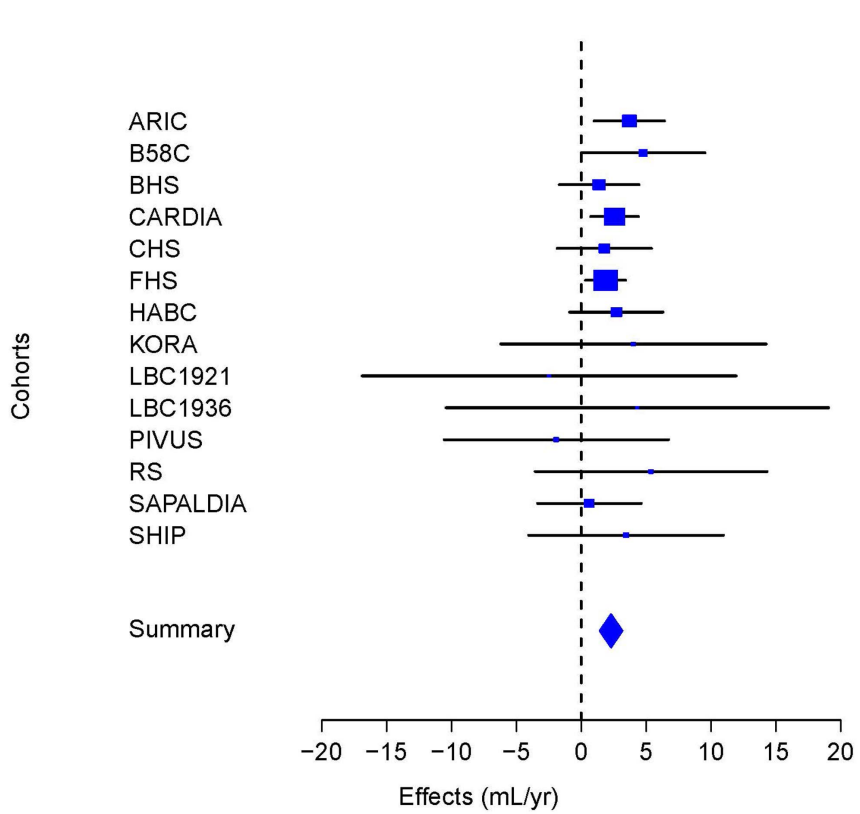

B
Cohorts
ARIC
B58C
BHS
CARDIA
CHS
FHS
HABC
KORA
LBC1921
LBC1936
PIVUS
RS
SAPALDIA
SHIP
Summary
Effects (mL/yr)

Figure 1. Association of the chromosome 15 locus with the rate of change in FEV_1 in the meta-analysis of 14 cohort studies. A) Regional association plot, where the X-axis is Megabase (Mb) position and Y-axes are the negative log of the *P* value on the left and recombination rate on the right. The sentinel SNP is colored in purple and linkage disequilibrium to the sentinel SNP is depicted by degree of color according to the legend. **B**) Forest plot for rs4077833, where the size of the square for each study represents its contributing weight to the meta-analysis.
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an attenuation of the rate of decline in FEV_1 by 2.3 mL/year in comparison to the G allele.

For estimation of longitudinal trajectory in lung function, having more than two measurements over time provides greater precision.[4] We performed a further meta-analysis with the five cohort studies (BHS, CHS, CARDIA, FHS, Health ABC) having $\geq$3 FEV_1 measurements per participant, with a combined sample size of 10,476 participants and 32,054 FEV_1 measurements (Methods S1 in File S1 for further details). A novel region on chromosome 11 had a genome-wide significant association ($P < 5 \times 10^{-8}$) with the rate of change in FEV_1 (Table 4). The most statistically significant finding at this locus was for rs507211, an intronic SNP located in *ME3* (Figures 2A and 2B). Six other SNPs, which are in linkage disequilibrium (LD) with rs507211 and are located in *ME3*, were identified at $P < 1 \times 10^{-6}$ (Table S3 in File S1). The rs507211 A allele, with a frequency of 25%, was associated with an attenuation of the rate of decline in FEV_1 by 2.09 mL/year in comparison to the G allele ($P = 2.18 \times 10^{-8}$). Besides the *ME3* locus, 17 SNPs from four other chromosomal regions had *P* values between 5×10^{-8} and 1×10^{-5} for associations with the rate of change in FEV_1 (Tables 4 and Table S3 in File S1).

Additional analyses

Corroborative evidence was sought for the sentinel SNP at each of the 14 loci associated at $P < 1 \times 10^{-5}$ (from both the 14-study and five-study meta-analyses) in 1,494 adults from the AGES-Reykjavík population-based cohort study (Table S4 in File S1). A *P* value of 0.004, representing the Bonferroni correction for 14 tests at the $\alpha = 0.05$ level, was selected *a priori* as the threshold for statistical significance. No SNPs achieved this threshold. The lowest *P* value was for rs740577 in *CACNG4* ($P = 0.08$), which showed consistent effect direction and magnitude with the original meta-analysis.

These same 14 SNPs were further examined in LHS, a clinical cohort study of 4,048 smokers with mild COPD for evidence of consistent association between healthy and diseased individuals.[17] None of the 14 SNPs were associated with the rate of change in FEV_1 in LHS at $P < 0.004$ (Table S4 in File S1).

Previous meta-analyses in the CHARGE and SpiroMeta consortia identified 26 novel loci associated with cross-sectional FEV_1 and/or FEV_1/FVC at genome-wide significance.[9-11] We examined the sentinel SNPs from these loci in the meta-analysis of the 14 cohort studies for association with the rate of change in FEV_1 (Table S5 in File S1). Given the *a priori* association with cross-sectional lung function, a *P* value threshold of 0.05 was used. Sentinel SNPs in *PID1*, *HHIP*, *GPR126*, and *CFDP1* showed association with the rate of change in FEV_1 ($0.005 \leq P \leq 0.048$).

Gene expression analyses

Three genes (*IL16*, *STARD5*, and *TMC3*) at the novel chromosome 15 locus and *ME3* at the novel chromosome 11 locus were selected for follow-up mRNA expression profiling in human lung tissue, and primary cultures of human bronchial epithelial and airway smooth muscle cells, together with control tissues (peripheral blood mononuclear cells and brain). Transcripts of *STARD5* and *ME3* were found in all lung-derived tissues, transcripts of *IL16* were found in lung tissue and smooth muscle cells, but not in epithelial cells, and *TMC3* was not expressed in any of the lung-derived tissues (Table S6 in File S1).

Using the public LGRC data repository, we found that the expression profiles of *IL16*, *STARD5*, and *ME3* in human lung samples showed statistically significant differences ($P < 0.05$) between COPD patients and controls (Figure S3 in File S1). Lower levels of *IL16* ($P = 0.004$) were observed in COPD patients compared with controls, whereas higher levels of *STARD5* ($P = 3.22 \times 10^{-9}$) and *ME3* ($P = 0.044$) were observed in COPD patients compared with controls. Data on *TMC3* expression were not available.

We performed additional follow-up analysis of the sentinel SNPs at the two novel loci using an eQTL database of lymphoblastoid cell lines (Table S8 in File S1). Trans-eQTL associations were observed between rs4077833 at the *IL16/STARD5/TMC3* locus and a nuclear receptor, *NR1I2* (chromosome 3; $P = 6.84 \times 10^{-4}$) and between rs507211 at the *ME3* locus and *KIAA1109* (chromosome 4; $P = 5.20 \times 10^{-4}$), which is part of a gene cluster (*KIAA1109-TENR-IL2-IL21*) that encodes two interleukins (IL2 and IL21).[18]

Discussion

Although the genetic contribution to cross-sectional lung function phenotypes has been addressed by large-scale GWAS, much less information is available for longitudinal lung function phenotypes. To identify novel loci that specifically affect lung function change over time, we performed a large-scale GWAS of the rate of change in FEV_1 in 27,249 participants from 14 population-based cohort studies. We identified a novel locus (*IL16/STARD5/TMC3*) on chromosome 15 with suggestive evidence for association with the rate of change in FEV_1. Given the greater precision to estimate longitudinal trends with more measurements, a meta-analysis of the five cohort studies with $\geq$3 FEV_1 measurements per participant was performed, and it identified a second novel locus (*ME3*) on chromosome 11 at genome-wide statistical significance. For both loci, the minor allele was protective, and the magnitude of the association with the rate of change in FEV_1 was similar to that of being an intermittent or former smoker versus a never-smoker.

The sentinel SNP at the novel chromosome 15 locus is located in *TMC3*, although two neighboring genes, *IL16* and *STARD5* both harbor SNPs that are in modest LD with the sentinel SNP (Figure 1A). TMC3, a member of the transmembrane channel-like gene family, likely functions as an ion channel, transporter, or modifier,[19] and has been associated with deafness and skin cancer.[20,21] IL16 is a pleiotropic immunomodulatory cytokine that acts as a chemoattractant for $CD4^+$ cells and contributes to their recruitment and activation in response to inflammation.[22] Notably, asthma was the first disease where increased *IL16* expression was observed.[23] Subsequent studies confirmed that in the non-diseased state *IL16* is almost exclusively expressed by T lymphocytes in lymphatic tissue, whereas in asthmatic patients IL16 is also synthesized by airway epithelial cells to inhibit airway inflammation.[24-26] A promoter polymorphism (T-295C) in *IL16* was associated with asthma in a Caucasian population in England,[27] although this finding was not confirmed in an Australian study.[28] STARD5 belongs to the steroidogenic acute

Table 4. Association of the most statistically significant SNPs with the rate of change in FEV_1 (mL/year) in the meta-analysis of the five cohort studies with ≥3 FEV_1 measurements per participant (n = 10,476).

SNP	Chr	Position	Closest Gene(s)	Coded Allele	Frequency	β*	SE	*P* Value
rs10209501	2	28536881	*FOSL2/PLB1*	A	0.33	1.6	0.4	7.09×10^{-6}
rs12692550	2	159958017	*BAZ2B*	T	0.18	−2.0	0.4	2.02×10^{-6}
rs1729588	3	110790025	*FLJ25363/MIR4445*	A	0.30	1.6	0.4	8.38×10^{-6}
rs10764053	10	19863644	*C10orf112*	T	0.47	1.5	0.3	4.15×10^{-6}
rs507211	11	86054387	*ME3*	A	0.25	2.1	0.4	2.18×10^{-8}

Definition of abbreviations: Chr = chromosome; SE = standard error; SNP = single-nucleotide polymorphism.

*Data reported are the meta-analysis results of the SNP-by-time interaction term from the GWAS mixed effects model. A positive β-coefficient indicates an attenuation of FEV_1 decline and a negative β-coefficient an acceleration of FEV_1 decline.

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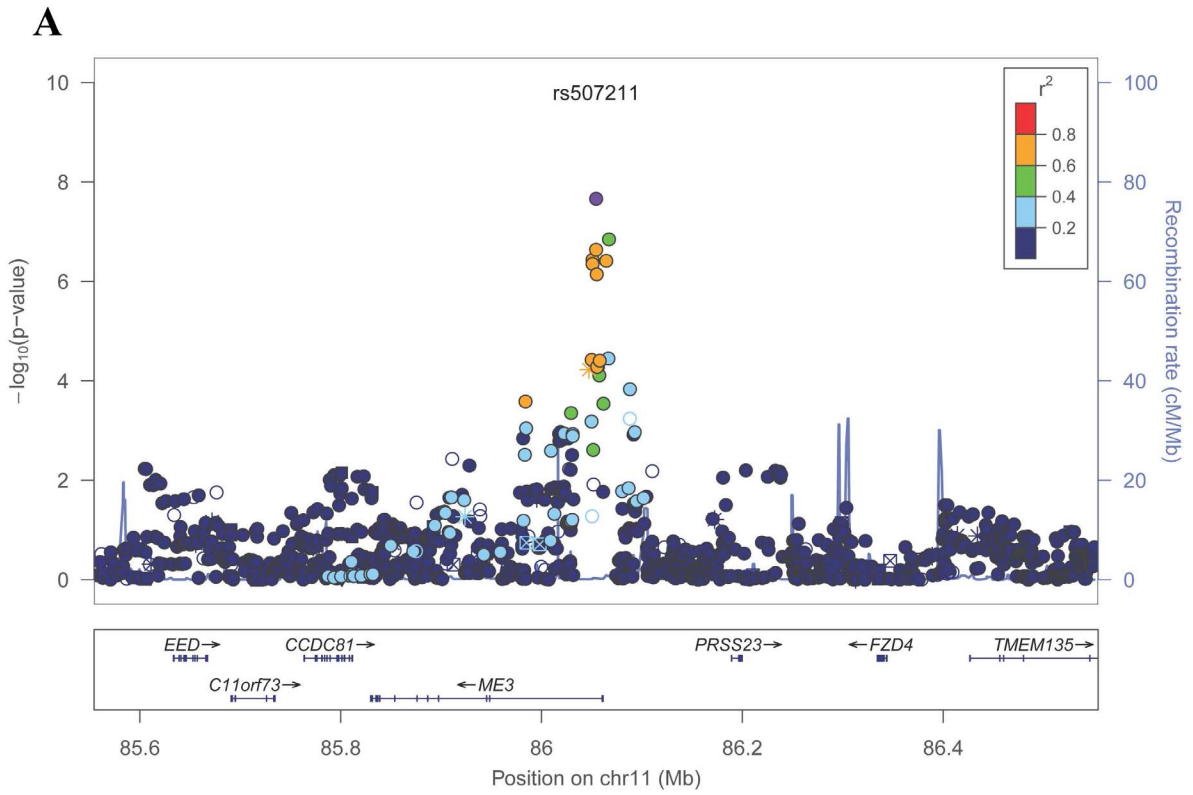
regulatory lipid transfer domain protein superfamily, and is involved in the trafficking of cholesterol and other lipids between intracellular membranes.[29] Recent *in vitro* studies showed increased *STARD5* expression and protein redistribution as a protective mechanism in response to induced endoplasmic reticulum (ER) stress and consequent over-accumulation of intracellular free cholesterol.[30] We confirmed the expression of *STARD5* in all human lung tissues examined and of *IL16* in human lung smooth muscle cells, but not epithelial cells, in line with previous observations. In contrast, no expression of *TMC3* was detected in any of the tested human lung tissues. We also found significantly lower levels of *IL16* in whole lung samples from COPD patients compared with controls, in contrast to its increased expression in asthma, and significantly higher levels of *STARD5* in COPD patients compared with controls. Taken together, these results suggest *IL16* as the most likely candidate accounting for the observed association, but further investigation is needed to elucidate underlying mechanisms.

The sentinel SNP at the novel chromosome 11 locus is located in *ME3*, whose protein product is a mitochondrial NADP(+)-dependent malic enzyme that catalyzes the oxidative decarboxylation of malate to pyruvate using NADP+ as a cofactor.[31] Mitochondrial malic enzymes play a role in the energy metabolism in tumors, and are considered potential therapeutic targets in cancer.[32,33] We performed independent expression profiling of *ME3* and confirmed its expression in all human lung tissues examined, and found significantly higher levels of *ME3* in lung samples from COPD patients compared with controls. In addition, we looked up the sentinel SNP in *ME3* in a recent GWAS of airway obstruction and found a *P* value of 0.049.[34] Taken together, these results support *ME3* as a biologically plausible candidate in the regulation of lung function and pathogenesis of COPD.

The identification of trans-eQTL associations for the sentinel SNPs at both the *IL16/STARD5/TMC3* and *ME3* loci is interesting, and while the interpretation of trans-eQTL associations is ambiguous,[35] the regions these SNPs regulate merit further study.

Besides the GWAS meta-analyses, the assembly of 14 longitudinal cohort studies allowed us to meta-analyze the association of cumulative smoking patterns with the rate of change in FEV_1 in the general population. The meta-analyzed estimate for the rate of decline in FEV_1 in never smokers was 26.9 mL/year, and the annual decline was steeper in persistent, intermittent, and former smokers by 8.8, 2.6, and 2.3 mL/year, respectively. These findings provide a reference point for the effect of cigarette smoking on longitudinal lung function change in the general population.

There is phenotypic variation among the 14 cohort studies in aspects such as baseline age and cigarette smoking, and in factors that are of special importance to this longitudinal GWAS, such as the number of FEV_1 measurements per participant and follow-up duration. Phenotypic heterogeneity represents a general challenge in genetic epidemiology, particularly in the investigation of longitudinal phenotypes. Thus, we performed a meta-analysis using the subset of cohort studies with ≥3 FEV_1 measurements per participant, given that longitudinal trajectories are best estimated over longer time periods and with more measurements. There was little overlap between the top loci identified in the two meta-analyses at $P < 1 \times 10^{-5}$, suggesting that phenotypic heterogeneity affected the association results. Future meta-studies of lung function decline should aim to increase sample size while maintaining high phenotypic comparability among participating studies. In addition, the trajectory of lung function change, especially over a long period of time, is known to be nonlinear,


A
rs507211
r2
0.8
0.6
0.4
0.2
−log10(p−value)
Recombination rate (cM/Mb)
EED→
CCDC81→
PRSS23→
←FZD4
TMEM135→
C11orf73→
←ME3
85.6
85.8
86
86.2
86.4
Position on chr11 (Mb)

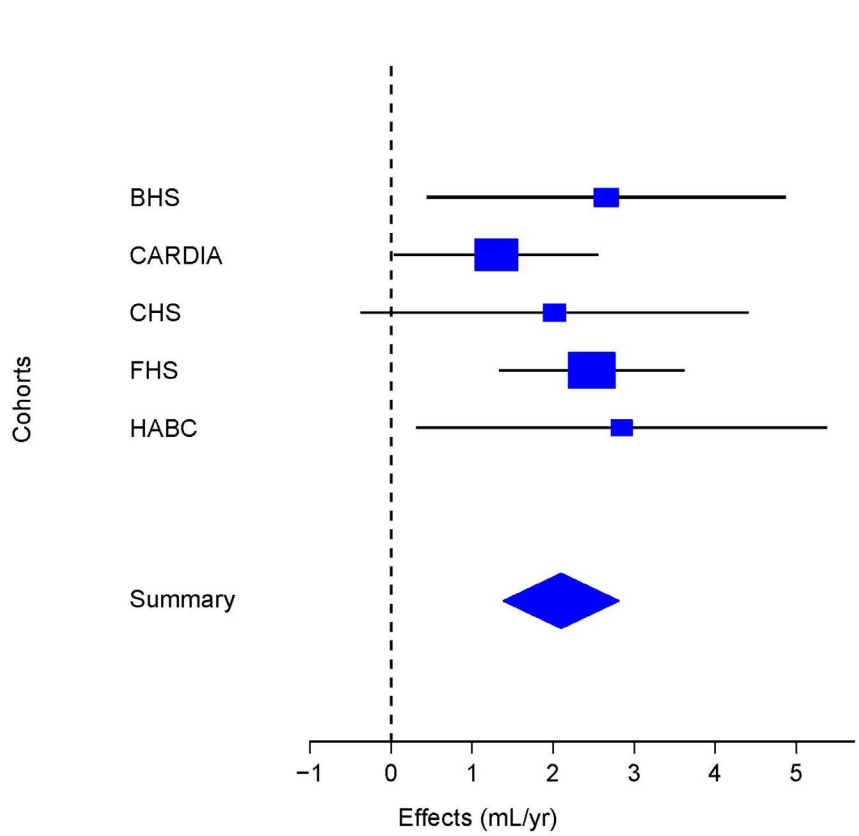

B
BHS
CARDIA
CHS
FHS
HABC
Cohorts
Summary
−1
0
1
2
3
4
5
Effects (mL/yr)

Figure 2. Association of the chromosome 11 locus with the rate of change in FEV_1 in the meta-analysis of the five cohort studies with ≥3 FEV_1 measurements per participant. A) Regional association plot, where the X-axis is Megabase (Mb) position, and the Y-axes are the negative log of the *P* value on the left and recombination rate on the right. The sentinel SNP is colored in purple and linkage disequilibrium to the sentinel SNP is depicted by degree of color according to the legend. **B**) Forest plot for rs507211, where the size of the square for each study represents its contributing weight to the meta-analysis.
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which may require the use of nonlinear time effects in the statistical model. In this study, given that over half of the included cohort studies have FEV_1 measurements at only two time points, our consideration was limited to a linear time effect. Further, the outcome studied, the rate of change in lung function, represents one of many ways to describe lung function change. Additional studies of other aspects of lung function change, such as reduced growth and premature decline, would be of interest.

We sought corroborative evidence in a single cohort study of 1,494 participants. This sample size is much smaller and arguably insufficient compared with replications applied to previous studies of cross-sectional lung function phenotypes. Thus, despite the lack of corroboration for the two novel loci identified in the meta-analyses, results from the complementary gene expression analyses provide compelling evidence for biologically plausible roles of the implicated genes in the longitudinal change in lung function.

None of the 14 sentinel SNPs were associated with the rate of change in FEV_1 in the COPD patient-based LHS cohort. Similarly, a previous population-based GWAS of lung function decline noted a high degree of heterogeneity in findings when analyses were stratified by presence/absence of asthma.[12] The observed discrepancy of association results suggests that the genetic determination of lung function decline may be different in healthy individuals compared with COPD patients, may contribute differentially in a pre-diseased vs. post-diseased state in which medications may influence the rates of decline, or that LHS was underpowered for confirming our findings.

In this study, statistical models included a comprehensive list of confounders that are commonly adjusted for when modeling lung function phenotypes. Given the study's meta-analysis design and the objective to carry out the same statistical model in all cohort studies, additional covariates that were not available in all cohort studies could not be included. In addition, the adjustment of certain confounders, such as smoking, is challenging in a longitudinal study, and although we accounted for the two most important aspects of smoking, cumulative pattern and dosage, residual confounding due to smoking cannot be excluded.

In summary, we performed GWAS of the longitudinal change in lung function and subsequent meta-analyses, using harmonized data from more than 27,000 participants of European ancestry to identify genetic loci influencing the rate of change in FEV_1. We identified the novel *ME3* locus on chromosome 11 at genome-wide significance and found suggestive evidence for association at the novel *IL16/STARD5/TMC3* locus on chromosome 15. Additional expression analyses confirmed the expression of *ME3*, *IL16*, and *STARD5* in multiple lung tissues, and found differential expression profiles of these three genes in the lungs of COPD patients compared to non-COPD controls. These results support the involvement of these implicated genes in the longitudinal change in lung function in adults of European ancestry. Additional studies with larger sample size and in populations of other races/ethnicities are warranted.

Supporting Information

File S1 This is a single file that contains all supporting information for the paper. Briefly, File S1 contains the following items: Methods S1, which describes further details of the cohort studies and the statistical methodology; Table S1, Details of SNP genotyping, quality control (QC), imputation, and statistical analysis across the 14 cohort studies; Table S2, Regression results for single nucleotide polymorphisms associated with the rate of change in FEV_1 (mL/year) at $P < 1 \times 10^{-5}$ in the meta-analysis of 14 cohort studies (N = 27,249); Table S3, Regression results for single nucleotide polymorphisms associated with the rate of change in FEV_1 (mL/year) at $P < 1 \times 10^{-5}$ in the meta-analysis of the five cohort studies with three or more FEV_1 measurements per participant (N = 10,476); Table S4, Association of the 14 sentinel SNPs from the meta-analyses in the AGES-Reykjavík study (AGES) and the Lung Health Study (LHS) for the rate of change in FEV_1 (mL/year); Table S5, Association of previously reported loci in GWAS of cross-sectional lung function with the rate of change in FEV_1 (mL/year) in the meta-analysis of 14 cohort studies (N = 27,249); Table S6, mRNA expression profiling of the implicated genes at the two novel loci in human lung and control tissues; Table S7, Primers for mRNA expression profiling; Table S8, Summary of eQTL look-up for the most significant SNPs at the novel chromosome 11 and 15 loci; Figure S1, Manhattan and QQ plots for the meta-analysis of the rate of change in FEV_1 in 14 cohort studies; Figure S2, Manhattan and QQ plots for the meta-analysis of the rate of change in FEV_1 in the five cohort studies with three or more FEV_1 measurements per participant; Figure S3, mRNA expression profiling in human lung samples from 219 COPD patients and 137 controls for A) *IL16*, B) *STARD5*, and C) *ME3*, using publicly available microarray data from the Lung Genomics Research Consortium site (http://www.lung-genomics.org/). The y-axes reflect the probe intensities of each gene transcript in the binary logarithm form, with the red dots indicating the average probe intensities and the red bars indicating standard deviation. The *P* values were calculated using the two-sample t-test.
(DOCX)

Checklist S1 PRISMA Checklist.
(DOCX)

Author Contributions

Wrote the paper: All authors. Drafted the manuscript: WT PAC. AGES Study concept, design: TBH VG LJL. ARIC Study concept, design: DJC NF DBH BRJ SJL ACM KEN. B58C Study concept and design: DPS. BHS Study concept, design: AJ B. Musk. CARDIA Study concept, design: M. Fornage AS. CHS Study concept, design: SRH SAG BMP. FHS Study concept, design: JD GTO JBW. Health ABC Study concept, design: PAC SBK WT. KORA Study concept, design: JH HS. LBC Study concept, design: IJD JMS. LHS Study concept, design: KCB NNH RAM. RS Study concept, design: GGB AH FR BHS AGU. SAPALDIA Study concept, design: MI NMP-H. SHIP Study concept, design: BK SG HV. SpiroMeta Study concept, design: IPH MDT. AGES Genotype data/QC: AVS. ARIC Phenotype data/QC: DJC. Genotype data/QC: ACM KEN. B58C Phenotype data/QC: DPS. Genotype data/QC: WLM. BHS Phenotype data/QC: AJ B. Musk. Genotype data/QC: AJ B. Musk LJP. CARDIA Phenotype data/QC: LJS AS ODW. Genotype data/QC: M. Fornage M. Foy XG. CHS Phenotype data/QC: BMP. Genotype data/QC: TL BMP JIR. FHS Phenotype data/QC: GTO. Genotype data/QC: GTO. Health ABC Phenotype data/QC: PAC SBK B. Mwibohm WT. Genotype data/QC: SBK YL KL. KORA Phenotype data/QC: JK SK HS. LBC Phenotype data/QC: IJD JMS. Genotype data/QC: GD. LHS Genotype

data/QC: KCB RAM IR. RS Phenotype data/QC: GGB L. Lahousse DWL BHS. Genotype data/QC:FR AGU. SAPALDIA Phenotype data/QC: IC MI NMP-H. Genotype data/QC: IC AK MI NMP-H. SHIP Phenotype data/QC: BK SG HV. Genotype data/QC: BK SG AT HV. SpiroMeta Genotype data/QC: IPH MDT. PIVUS Phenotype data/QC: EI L. Lind. Genotype data/QC: EI L. Lind APM. AGES Data analysis: AVS. ARIC Data analysis: BRJ SJL. B58C Data analysis: DPS LVW. BHS Data analysis: MK LP. CARDIA Data analysis: M. Fornage M. Foy XG. CHS Data analysis: SAG SRH GL TL AV. FHS Data analysis: JD WG JBW. Health ABC Data analysis: PAC YL KL WT MTW. KORA Data analysis: EA. LBC Data analysis: MA GD. LHS Data analysis: KCB NNH RAM IR. RS Data analysis: L. Lahousse DWL. SAPALDIA Data analysis: MI. SHIP Data analysis: AT. SpiroMeta Data analysis: MSA IPH MDT. PIVUS Data analysis: TF.

References

1. Rabe KF, Hurd S, Anzueto A, Barnes PJ, Buist SA, et al. (2007) Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. Am J Respir Crit Care Med 176: 532–555.
2. Schunemann HJ, Dorn J, Grant BJ, Winkelstein W Jr, Trevisan M (2000) Pulmonary function is a long-term predictor of mortality in the general population: 29-year follow-up of the Buffalo Health Study. Chest 118: 656–664.
3. Young RP, Hopkins R, Eaton TE (2007) Forced expiratory volume in one second: not just a lung function test but a marker of premature death from all causes. Eur Respir J 30: 616-622.
4. Fletcher C, Peto R (1977) The natural history of chronic airflow obstruction. Br Med J 1: 1645–1648.
5. Halbert RJ, Natoli JL, Gano A, Badamgarav E, Buist AS, et al. (2006) Global burden of COPD: systematic review and meta-analysis. Eur Respir J 28: 523–532.
6. Eisner MD, Anthonisen N, Coultas D, Kuenzli N, Perez-Padilla R, et al. (2010) An official American Thoracic Society public policy statement: Novel risk factors and the global burden of chronic obstructive pulmonary disease. Am J Respir Crit Care Med 182: 693–718.
7. Gottlieb DJ, Wilk JB, Harmon M, Evans JC, Joost O, et al. (2001) Heritability of longitudinal change in lung function. The Framingham study. Am J Respir Crit Care Med 164: 1655–1659.
8. Finkel D, Pedersen NL, Reynolds CA, Berg S, de Faire U, et al. (2003) Genetic and environmental influences on decline in biobehavioral markers of aging. Behav Genet 33: 107–123.
9. Hancock DB, Eijgelsheim M, Wilk JB, Gharib SA, Loehr LR, et al. (2009) Meta-analyses of genome-wide association studies identify multiple loci associated with pulmonary function. Nat Genet 42: 45–52.
10. Repapi E, Sayers I, Wain LV, Burton PR, Johnson T, et al. (2009) Genome-wide association study identifies five loci associated with lung function. Nat Genet 42: 36–44.
11. Soler Artigas M, Loth DW, Wain LV, Gharib SA, Obeidat M, et al. (2011) Genome-wide association and large-scale follow up identifies 16 new loci influencing lung function. Nat Genet 43: 1082–1090.
12. Imboden M, Bouzigon E, Curjuric I, Ramasamy A, Kumar A, et al. (2012) Genome-wide association study of lung function decline in adults with and without asthma. The Journal of allergy and clinical immunology 129: 1218–1228.
13. Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, et al. (2005) Standardisation of spirometry. Eur Respir J 26: 319–338.
14. Willer CJ, Li Y, Abecasis GR (2010) METAL: fast and efficient meta-analysis of genomewide association scans. Bioinformatics 26: 2190–2191.
15. Hansel NN, Ruczinski I, Rafaels N, Sin DD, Daley D, et al. (2013) Genome-wide study identifies two loci associated with lung function decline in mild to moderate COPD. Hum Genet 132: 79–90.
16. Dixon AL, Liang L, Moffatt MF, Chen W, Heath S, et al. (2007) A genome-wide association study of global gene expression. Nat Genet 39: 1202–1207.
17. Anthonisen NR, Connett JE, Kiley JP, Altose MD, Bailey WC, et al. (1994) Effects of smoking intervention and the use of an inhaled anticholinergic bronchodilator on the rate of decline of FEV1. The Lung Health Study. JAMA 272: 1497–1505.
18. van Heel DA, Franke L, Hunt KA, Gwilliam R, Zhernakova A, et al. (2007) A genome-wide association study for celiac disease identifies risk variants in the region harboring IL2 and IL21. Nat Genet 39: 827–829.
19. Kurima K, Yang Y, Sorber K, Griffith AJ (2003) Characterization of the transmembrane channel-like (TMC) gene family: functional clues from hearing loss and epidermodysplasia verruciformis. Genomics 82: 300–308.
20. Ramoz N, Rueda LA, Bouadjar B, Montoya LS, Orth G, et al. (2002) Mutations in two adjacent novel genes are associated with epidermodysplasia verruciformis. Nat Genet 32: 579–581.
21. Vreugde S, Erven A, Kros CJ, Marcotti W, Fuchs H, et al. (2002) Beethoven, a mouse model for dominant, progressive hearing loss DFNA36. Nat Genet 30: 257–258.
22. Cruikshank WW, Kornfeld H, Center DM (1998) Signaling and functional properties of interleukin-16. International reviews of immunology 16: 523–540.
23. Bellini A, Yoshimura H, Vittori E, Marini M, Mattoli S (1993) Bronchial epithelial cells of patients with asthma release chemoattractant factors for T lymphocytes. The Journal of allergy and clinical immunology 92: 412–424.
24. Cruikshank WW, Long A, Tarpy RE, Kornfeld H, Carroll MP, et al. (1995) Early identification of interleukin-16 (lymphocyte chemoattractant factor) and macrophage inflammatory protein 1 alpha (MIP1 alpha) in bronchoalveolar lavage fluid of antigen-challenged asthmatics. Am J Respir Cell Mol Biol 13: 738–747.
25. Krug N, Cruikshank WW, Tschernig T, Erpenbeck VJ, Balke K, et al. (2000) Interleukin 16 and T-cell chemoattractant activity in bronchoalveolar lavage 24 hours after allergen challenge in asthma. Am J Respir Crit Care Med 162: 105–111.
26. Laberge S, Ernst P, Ghaffar O, Cruikshank WW, Kornfeld H, et al. (1997) Increased expression of interleukin-16 in bronchial mucosa of subjects with atopic asthma. Am J Respir Cell Mol Biol 17: 193–202.
27. Burkart KM, Barton SJ, Holloway JW, Yang IA, Cakebread JA, et al. (2006) Association of asthma with a functional promoter polymorphism in the IL16 gene. The Journal of allergy and clinical immunology 117: 86–91.
28. Akesson LS, Duffy DL, Phelps SC, Thompson PJ, Kedda MA (2005) A polymorphism in the promoter region of the human interleukin-16 gene is not associated with asthma or atopy in an Australian population. Clinical and experimental allergy: journal of the British Society for Allergy and Clinical Immunology 35: 327–331.
29. Rodriguez-Agudo D, Ren S, Hylemon PB, Redford K, Natarajan R, et al. (2005) Human StarD5, a cytosolic StAR-related lipid binding protein. Journal of lipid research 46: 1615–1623.
30. Rodriguez-Agudo D, Calderon-Dominguez M, Medina MA, Ren S, Gil G, et al. (2012) ER stress increases StarD5 expression by stabilizing its mRNA and leads to relocalization of its protein from the nucleus to the membranes. Journal of lipid research 53: 2708–2715.
31. Chang GG, Tong L (2003) Structure and function of malic enzymes, a new class of oxidative decarboxylases. Biochemistry 42: 12721–12733.
32. Moreadith RW, Lehninger AL (1984) The pathways of glutamate and glutamine oxidation by tumor cell mitochondria. Role of mitochondrial NAD(P)+− dependent malic enzyme. J Biol Chem 259: 6215–6221.
33. Teller JK, Fahien LA, Davis JW (1992) Kinetics and regulation of hepatoma mitochondrial NAD(P) malic enzyme. J Biol Chem 267: 10423–10432.
34. Wilk JB, Shrine NR, Loehr LR, Zhao JH, Manichaikul A, et al. (2012) Genome Wide Association Studies Identify CHRNA5/3 and HTR4 in the Development of Airflow Obstruction. Am J Respir Crit Care Med.
35. Montgomery SB, Dermitzakis ET (2011) From expression QTLs to personalized transcriptomics. Nat Rev Genet 12: 277–282.