Genetic Deletion of SEPT7 Reveals a Cell Type-Specific Role of Septins in Microtubule Destabilization for the Completion of Cytokinesis

Manoj B. Menon1*, Akihiro Sawada2, Anuhar Chaturvedi3, Pooja Mishra4, Karin Schuster-Gossler5, Melanie Galla6, Axel Schambach6,7, Achim Gossler5, Reinhold Förster4, Michael Heuser3, Alexey Kotlyarov1, Makoto Kinoshita2,*, Matthias Gaestel1*

1 Institute of Physiological Chemistry, Hannover Medical School, Hannover, Germany, 2 Department of Molecular Biology, Division of Biological Science, Nagoya University Graduate School of Science, Furo, Chikusa, Nagoya, Japan, 3 Department of Hematology, Hemostasis, Oncology and Stem Cell Transplantation, Hannover Medical School, Hannover, Germany, 4 Institute of Immunology, Hannover Medical School, Hannover, Germany, 5 Institute of Molecular Biology, Hannover Medical School, Hannover, Germany, 6 Institute of Experimental Hematology, Hannover Medical School, Hannover, Germany, 7 Division of Hematology/Oncology, Boston Children’s Hospital, Harvard Medical School, Boston, Massachusetts, United States of America

Abstract

Cytokinesis terminates mitosis, resulting in separation of the two sister cells. Septins, a conserved family of GTP-binding cytoskeletal proteins, are an absolute requirement for cytokinesis in budding yeast. We demonstrate that septin-dependence of mammalian cytokinesis differs greatly between cell types: genetic loss of the pivotal septin subunit SEPT7 in vivo reveals that septins are indispensable for cytokinesis in fibroblasts, but expendable in cells of the hematopoietic system. SEPT7-deficient mouse embryos fail to gastrulate, and septin-deficient fibroblasts exhibit pleiotropic defects in the major cytokinetic machinery, including hyperacetylation/stabilization of microtubules and stalled midbody abscission, leading to constitutive multinucleation. We identified the microtubule depolymerizing protein stathmin as a key molecule aiding in septin-independent cytokinesis, demonstrated that stathmin supplementation is sufficient to override cytokinesis failure in SEPT7-null fibroblasts, and that knockdown of stathmin makes proliferation of a hematopoietic cell line sensitive to the septin inhibitor forchlorfenuron. Identification of septin-independent cytokinesis in the hematopoietic system could serve as a key to identify solid tumor-specific molecular targets for inhibition of cell proliferation.

Introduction

Cytokinesis as final step of cell division is essential for cell proliferation, but there is a considerable degree of diversity in its underlying mechanisms among eukaryotes. Even within one organism, such as the amoeba Dictyostelium discoideum, cytokinesis may proceed by different mechanisms for cells growing in suspension or in an attachment-dependent manner. This has been impressively demonstrated for the myosin II-deletion mutant of D. discoideum, which could not complete cytokinesis in suspension but successfully proliferates when attached to surfaces [1]. Hence, it could be speculated that other cells also confine different molecular requirements for attachment-dependent and -independent cytokinesis, although there is little molecular proof for this idea in mammalian cells. Recent support for this idea comes from the observation that in lymphocytes the hematopoietic lineage-specific Rho-GAP ARHGAP19 is essential for cytoskeleton remodeling resulting in cell division [2] while in most other cells M-phase GAP (MP-GAP) is the major factor restraining RhoA during cell division [3].

Septins, a conserved family of polymerizing GTP-binding proteins regarded as the forth component of the cytoskeleton [4], organize a ring that serves as a submembranous scaffold and diffusion barrier for various molecules, which is an absolute requirement for cytokinesis in budding yeast [5,6]. In metazoans, septins associate with the mitotic spindle, contractile ring, intercellular bridge and midbody at varying degrees [7,8]. For example, anillin-dependent recruitment of septins to the intercellular bridge is required for constriction site formation and ingress in HeLa cells [9], maturation of the midbody ring in Drosophila melanogaster requires septin-dependent removal of anillin via its C-terminal PH-domain [10], and septins are required for the release of midbody and midbody ring into daughter cells during the subsequent cell division in Caenorhabditis elegans [11]. Perturbation or depletion of one of the major septin subunits, such as the pivotal subunit SEPT7 [12,13], affects multiple steps in mitosis [4,14]. In vitro studies with mammalian cell lines have revealed pleiotropic defects in mitotic spindle organization and chromosome alignment [15], cleavage furrow


Editor: Susan K. Dutcher, Washington University School of Medicine, United States of America

Received March 12, 2014; Accepted June 25, 2014; Published August 14, 2014

Copyright: © 2014 Menon et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper and its Supporting information files.

Funding: This work was funded by the Deutsche Forschungsgemeinschaft Grant KO2091/1-3 to AK and MGae, Sonderforschungsbereich 738-B5 to RF and the Cluster of Excellence “From Regenerative Biology to Reconstructive Therapy” to AG. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: There are no competing interests.

* Email: menon.manoj@mh-hannover.de (MBM); kinoshita.makoto@c.mbox.nagoya-u.ac.jp (MK); gaestel.matthias@mh-hannover.de (MG)
Cytokinesis is the finalizing step of the complex scenario of mitosis, leading to separation of two sister cells. The cellular mechanism of cytokinesis in eukaryotes differs at least between yeasts, plants and animals. So far, it is also not clear whether all mammalian cells follow the same mechanistic rules of cytokinesis. Here, we demonstrate that, depending on the mammalian cell type, two different pathways could result in completion of cytokinesis, a septin-dependent pathway and a distinct mechanism, which does not require septins prevalent in the hematopoietic system. Using multiple conditional knockouts, we demonstrate this cell type specificity in vitro and in vivo, and present evidence for the involvement of cell-type specific alteration of the microtubule cytoskeleton. Our data, together with the previously available septin knockdown data in cancer cell lines, suggest septins as plausible antitumor targets with high therapeutic index due to lack of off-target effects on hematopoiesis.

**Results**

**Deletion of Sept7 causes embryonic lethality**

We floxed Sept7 (exon 4, encoding the GTP-binding P-loop) in the mouse genome using the Cre-loxP system (Figure 1A). The Sept7lox allele was converted to Sept7fllox (null) allele by using oocyte-specific expression of Cre-recombinase (ZP3-Cre). Sept7fllox (KO) embryos were found in utero up to embryonic day 6.5 (E6.5-7.0, but not after E10.5, indicating early embryonic lethality (Figure 1B). As the genetic loss of Sept3 or Sept11 causes embryonic death by E10 [21] and E13 [22] respectively, Sept7 appears no less vital than these major subunits. These data obviously indicate that septins are dispensable for the majority of cells to execute mitosis in early mouse embryo.

**SEPT7-deficient fibroblasts display incomplete cytokinesis and constitutive multinucleation**

To probe the impact of the genetic loss of SEPT7 on mitosis in vitro, we prepared primary fibroblasts (MEFs) and SV40-large T-immortalized tail fibroblasts (TFs) from Sept7fllox/fllox mice. Cre-transduction via adenov- or retroviral vectors caused significant reduction of SEPT7 and collateral depletion of SEPT2, SEPT6, transduction via adeno- or retroviral vectors caused significant reduction of SEPT7 and collateral depletion of SEPT2, SEPT6, transduction via adeno- or retroviral vectors caused significant recombination (Figure 2G) caused a

Impaired cytokinesis and stalled midbody abscission in SEPT7-deficient fibroblasts

Time-lapse observation of the same population identified two subsets; one completed cytokinesis normally within 70–130 min (about 70% of cells), while another could not complete cytokinesis within 130 min, displaying stalled cytokinesis yielding binucleated cells after unsuccessful severing of the intercellular bridge (about 30% of cells) (Figure 3A, Figure S4 and video S1). Immunofluorescence analysis of the intercellular bridges and midbodies did not show obvious disorganization of α-tubulin and F-actin in the absence of SEPT7 (Figure 3B and Figure S3A, S3B, S3D). Improper segregation of chromosomes can lead to the formation of chromatin-bridges associated with a delay in abscission and multinucleation [24]. Analysis of the arrested midbody structures in the Sept7fllox/−− revealed absence of persistent chromatin bridges as shown by LAP2 staining (Figure S5). However, Sept7fllox/−− cells were often accompanied by unresolved α-tubulin aggregates (arrowheads in Figure 3C) and about two-fold hyperacetylation of α-tubulin (Figure 3D and Figure S3C, S3E). These data indicate hyperstabilization of microtubules in Sept7fllox/−− cells, as has been observed in interphase HeLa cells [25] and postmitotic primary neurons [26]. Anillin, a contractile ring organizer which interacts with actomyosin and septins, was reduced in interphase nuclei of Sept7fllox/−− cells (Figure 3E, cf. Figure 2C and Figure S1). However, SEPT7 was dispensable for the targeting of anillin to the cleavage furrow (Figure 3F). Thus, genetic loss of SEPT7 in fibroblasts appeared to affect mitotic spindle and midbody rather than the contractile ring.

**SEPT7 is dispensable for the cytokinesis of myeloid and lymphoid cells**

Next, we examined the aforementioned presumable dispensability of SEPT7 in non-adherent cell lineages. We introduced a bidirectional γ-retroviral mCherry-Cre construct [27] (Figure S6A, 6b) into Sept7fllox/−− bone marrow cells, which successfully induced recombination (Figure 4A). An interleukin (IL)-3/IL-6/SCF-dependent myeloid colony formation assay (Figure S6C) revealed that almost every subpopulation of the Sept7fllox/−− leukocytes exhibited subnormal but sufficient proliferative activity in vitro (Figure 4B). Given that most of these Sept7fllox/−− cells (Figure 4C) had undergone more than 10 replication cycles, SEPT7 protein carried over from the original Sept7fllox/−− cell had been eliminated. These data indicate that the resistance to the loss of SEPT7 in mitosis is a common trait of the myeloid lineage.

To corroborate the dispensability of SEPT7 in myeloid cell mitosis in vivo, we generated lymphocyte-specific Sept7fllox/−− mice, by intercrossing Sept7fllox/−− and CD2-Cre lines [28]. We detected efficient recombination in the bone marrow (Figure S7A), spleen, thymus, and lymph nodes (Figure 4D) with recognizable volume loss in the spleen and thymus (Figure 4E). Flow cytometric analysis demonstrated complete loss of SEPT7 in cells collected from thymus, while those from spleen contained a minor population that fully expressed SEPT7 (Figure 4F). Viability of lymphocytes from spleen, peripheral lymph nodes (Figure 4G), thymus (Figure 4H), bone marrow (Figure S7B) and a
Figure 1. Generation of Sept7 floxed mice and characterization of embryonic lethality of the Sept7 knockout. A, Strategy for conditional targeting of Sept7. A neomycin cassette with flanking FRT and lox-P sites was incorporated by homologous recombination in ES cells. The resulting mice were crossed with Flippase expressing mice to remove the neomycin cassette retaining the lox-P flanked (floxed) exon 4. Cre expression leads to excision of exon4 and a downstream frame shift. B, Analysis of embryonic lethality in Sept7 knockout mice. Analysis of progeny by

<table>
<thead>
<tr>
<th></th>
<th>Sept7/ Sept7</th>
<th>Sept7/Δ</th>
<th>Δ/Δ</th>
<th>χ² (p&lt;0.05, test 5.99)</th>
</tr>
</thead>
<tbody>
<tr>
<td>E 6.5-7.0</td>
<td>7</td>
<td>9</td>
<td>4</td>
<td><strong>1.100</strong></td>
</tr>
<tr>
<td>E 10.5</td>
<td>8</td>
<td>16</td>
<td>-</td>
<td><strong>8.000</strong></td>
</tr>
<tr>
<td>P1</td>
<td>9</td>
<td>28</td>
<td>-</td>
<td><strong>13.82</strong></td>
</tr>
</tbody>
</table>
number of peripheral blood cells (Figure S8) showed no differences with or without Sept7. Although SEPT7/6/2/9 had been depleted from Sept7+/−/− thymocytes (Figure S9), flow cytometric DNA content analysis did not detect any multineucleated population (Figure 4I). Intriguingly, as opposed to fibroblasts, HeLa cells [25] and neurons [26], thymocytes did not exhibit microtubule hyperacetylation after septin depletion (Figure S10). Sept7+/−/− splenocytes proliferated normally in vitro in response to concanavalin A and IL-2 (Figure 4J, 4K), without forming multinucleated cells (Figure 4L). Taken together, we conclude that Sept7 is dispensable in the proliferation and maturation of B- and T-lymphocytes in vivo, and in the proliferation of splenocytes and myeloid progenitors in vitro.

Elevated levels of stathmin enable SEPT7-deficient cells to complete cytokinesis

In our search for the factor enabling diverse hematopoietic cell lineages to go through the cell cycle without SEPT7, we compared the proteome between the fibroblasts and myeloid cells. From a number of candidate proteins we focused our studies on stathmin (STMN1) because of its specific abundance in the blood cell lineages (Figure 3A) and biochemical activity. The stathmin family is known to facilitate microtubule depolymerization by sequestering α/β-tubulin heterodimers [29,30]. We hypothesized that the scarcity of stathmin in fibroblasts contributes to the stability of the microtubule network, while the abundance of stathmin in hematopoietic cells facilitates the disassembly of spindle microtubules and the disposal of midbodies. To test the latter possibility, we generated Sept7flx/flx MEFs that express stathmin via a doxycycline-regulatable promoter (Figure 5B). Indeed, stathmin overexpression (to the level of thymocytes) was sufficient to rescue the mitotic failure of Sept7+/−/− MEFs (Figure 5C, 5D) without changing other complex cellular properties as represented by cell mobility and adhesion measured in a scratch assay (Figure S11). We then asked whether stathmin overexpression also rescues multinucleation of the Sept7+/−/− MEFs. For this reason we cotransduced MEFs with pR Bid-Cre and the doxycyclin-inducible stathmin construct and DAPI-stained and counted mCherry-positive mono- and multinucleated cells after 5 days of cultivation in the presence or absence of doxycycline (Figure 5E and Figure S12). While the majority of control cells are multinucleated, overexpression of stathmin clearly shifted the MEFs to the mononucleated phenotype.

Stathmin knockdown renders proliferation of Jurkat cells septin-dependent

Finally, we ask whether hematopoietic cells proliferating septin-independently require stathmin and whether stathmin-knockdown renders these cells sensitive to septin inactivation. To answer these questions we used the Jurkat human lymphocyte cell line, because manipulation of primary mouse hematopoietic cells in culture was not feasible. To inactivate septins in Jurkat cells, we applied the septin inhibitor forchlorfenuron (FCF) [31], which dampens septin dynamics and induces the assembly of abnormally large septin structures [32]. Stathmin knockdown by siRNA was performed and cells were further cultivated for 48 hours in the presence or absence of different concentrations of FCF (Figure 5F, 5G). siSTMN1 treatment efficiently reduced stathmin levels while the control siRNA did not (Figure 5F). Remarkably, while 50 μM FCF did not inhibit proliferation of Jurkat cells transfected with the control siRNA, siSTMN1-treated Jurkat cells displayed a clear proliferation defect at this concentration of FCF. At higher concentrations of FCF (100 μM) slight cytotoxic effects also reduced proliferation of the control, but the stronger reduction in the siSTMN1-treated cells remained. Taken together, we demonstrated that stathmin can rescue the proliferation block in SEPT7-deficient MEFs and that stathmin is necessary for proliferation of hematopoietic cells in the absence of functional septins. Thus, stathmin is a critical permissive factor whose abundance enables cells to proliferate without septins.

Discussion

This study has revealed two distinct types of mammalian cytokinesis which vary by the requirement for SEPT7/septins. Consistent with previous studies [5,16,18] our findings indicate that cell division requires septins in two spatiotemporally distinct processes, first for the organization of the contractile ring and later for midbody abscission. The former became known early on due to the high prominence and its evolutionarily conservation from budding yeast to humans, while the latter had remained unknown due to its cell-type-dependence. Fibroblasts, typical adherent cells, divide in contact with other cells and/or connective tissue in vivo and extracellular matrices and artificial substrate in vitro. In contrast, amoeboid hematopoietic cells grow planktonically in vivo and divide individually in suspension. Our study confirm the role of septins in the recruitment of the microtubule cleaving machinery (multi-protein membrane associated abscission machinery probably including spastin for local microtubule destabilization) [7,8] to the midbody for final microtubules scission. This system seems to be inactive in the absence of SEPT17 in fibroblasts, leading to midbody stabilization. In the hematopoietic system the abundance of stathmin leads to a passive rescue due to general microtubule destabilization and thus cytokinesis proceeds in a septin independent manner. The supplementation of stathmin is sufficient for fibroblasts to override the loss of SEPT17 and to complete cytokinesis. The abundant expression of stathmin in early embryo [33,34] may account for the dispensability of septins up to midgestation. These data indicate that the synergy between septins and stathmin, among other microtubule-regulating proteins, is critical for completion of cytokinesis and midbody abscission. The entire process should depend not only on the quantitative balance of tubulin/stathmin/septin but also on the phosphorylation level of stathmin [29,30]. Of note, β1-integrin-blocking antibodies can inhibit cytokinesis of adherent cells, but not their cytokinesis in suspension [35]. Given these and our findings, it is conceivable that non-adherent cells develop less cytoskeletal network than adherent cells, which should reduce the burden for midbody abscission. Conversely, myosin II-deficient Dictostelium cells can complete cytokinesis on a substrate but not in suspension [36], indicating that microtubule is not a critical determinant in this case. A recent study with Drosophila revealed that the SEPT7 ortholog peanut (Pnut) and other septins are required for planar cell cytokinesis but dispensable for orthogonal cell division in the single-layered neuroepithelium of the dorsal thorax [37]. This finding supports our notion that SEPT7/septins play a context-dependent role in mammalian cytokinesis. Accordingly, SEPT7 is a promising target for the development of solid tumor-selective anti-proliferative therapy without damaging genotyping at postnatal day 1 (P1) and embryos at E6.5–7 and E10.5, shows embryonic lethality between E7.5 and E10.5. Significance of the χ² test is given where a difference to Mendelian distribution is indicated by values greater than 5.99 (p = 0.05). doi:10.1371/journal.pgen.1004558.g001
Figure 2. Cre-induced deletion of SEPT7 in fibroblasts leads to proliferation block and obligate multinucleation. A, Adenoviral Cre-expression in Sept7floxflox primary mouse embryonic fibroblasts (MEFs) [26] leads to deletion of SEPT7 and, as judged by immunofluorescence, to a strong reduction of expression of the other core-septins SEPT6/2. B, Retroviral Cre-transduction of Sept7floxflox immortalized mouse tail fibroblasts causes complete deletion of the Sept7 gene in two different experiments (#1, #2, independently immortalized lines) (control: non-integrating control virus). Genotyping by PCR as indicated in Supporting Figure S1. C, Western blot detection of multiplicity of infection (MOI)-dependent reduced SEPT7 levels in Sept7floxflox immortalized mouse tail fibroblasts. Expression of SEPT6/2/9 and anillin is significantly reduced. D, E, Adenoviral Cre-expression in primary Sept7floxflox MEFs decreases immunofluorescence detection of the proliferation marker Ki67 (D) and increases histone H3 phosphorylation (E). The percentage of positive cells is indicated. F, SEPT7-depleted Sept7floxflox immortalized mouse tail fibroblasts fail to proliferate (3 days post transduction). G, Retroviral Cre-transduced SEPT7-depleted cells are overgrown by non-depleted cells, as indicated by the relative increase in the floxed allele after 12 days in culture. H, Immunofluorescence analysis showing obligatory multi-nucleation of Sept7-KO tail fibroblasts after 11 days post transduction. DAPI is used for nuclear staining and the WGA as counter stain. Arrowheads indicate multi-nucleated, SEPT7-negative cells.

Figure 3. Defective cytokinesis and unresolved midbody in SEPT7-deficient fibroblasts. A, Time lapse differential interference contrast (DIC) microscopy of Cre-transduced floxed Sept7 tail fibroblasts. The green arrow indicates a SEPT7-positive, normal dividing cell (about 70% of cells). The process of cell division is completed in less than 120 min and two daughter cells appear. The red arrow indicates a SEPT7-negative cell which could not complete cytokinesis after nuclear division (about 30% of cells). Even after 400 min the daughter cells do not separate and the cell becomes multi-nucleated. B, Localization of microtubules (α-tubulin) and microfilaments (phalloidin) by immunofluorescence in the midbody zone of dividing fibroblasts in the absence (SEPT7−/−) or presence of SEPT7 (WT). Two different cells each were analyzed. Arrowheads indicate the midbodies. C, Detection of unresolved midbody tubulin bundles (indicated by arrowheads) in multinucleated SEPT7-deficient cells. D, E, Immunofluorescence of tail fibroblasts transduced with pRbid-Cre-mCherry. D, About two-fold increased acetyl-tubulin detection and E, about 2–3-fold decreased nuclear anillin staining in mCherry-Cre-positive, SEPT7-negative cells. F, Intact recruitment of the remaining anillin to the midbody zone in mCherry-Cre-positive, SEPT7-negative cells.

doi:10.1371/journal.pgen.1004558.g002

doi:10.1371/journal.pgen.1004558.g003
hematopoietic cells. Reciprocally, stathmin could be selectively targeted in hematopoietic malignancies and p53-compromised cancer [38,39].

Materials and Methods

Generation of SEPT7 conditional knockout mice

Two independently developed Sep7 floxed mice strains were used in this study, both targeting exon4 of mouse Sep7 gene using similar targeting strategies. Sep7floxflox/mx mice (Sep7floxflox/Mgx) were generated as indicated in Figure 1A. Briefly, the targeting vector containing lox sites and FRT sites flanked neomycin cassette was linearized and electroporated in 129Ola ES-cells. Two positive clones (42A3 and 44A1) obtained by PCR screen were injected into blastocysts for the generation of chimeric mice. Agouti germ line pups were derived from the mating of chimeric male mice, obtained following the blastocyst injection of Sep7 targeted ES-cell clone 44A1, with C57Bl/6 Flp females. The resulting sept7loxNeo mice were crossed with C57BL/6-(C3-Tg(Pgk1-FLPo)10Syrk/J Flipase- expressing mice [40] to delete the neomycin cassette retaining the lox-P-flanked (floxed) exon 4 leading to Sep7lox mice. Subsequent Cre-recombinase expression will then catalyze exon 4 excision resulting in an additional frame-shift mutation downstream in the exon. For generation of Oocyte specific knockout animals, Sep7 homozygous floxed mice were crossed with B6-Zp3CrehCD2-tcre mice to breed sepa7lox mice. Lymphocyte-specific Sep7 knockout mice were generated by mating floxed animals with B6-hCD2-tcre mice [28]. In animal experiments age and sex matched, Cre-expressing Sep7 wt and floxed mice were used.

DNA isolation and genotyping

Tail biopsies, cells and colonies were overnight digested at 53 °C in lysis buffer (50 mM Tris-Cl (pH 8.0), 100 mM EDTA, 100 mM NaCl and 1% SDS) containing proteinase-K (0.5 mg/ml). For tissue samples proteins were salted out with extra NaCl. DNA was precipitated with isopropanol, washed with 70% ethanol and dissolved in water. Genotyping PCR were performed with Hotstar Taq (Qiagen) with extra Mg2+ under standard conditions (5° – TTA CGT CCA TCG TGG AGA GC -3°), Myo-rev (5°- TGG GCT GGG TGT TAG CCT TA -3°). Myogenin and IL2 gene fragments were amplified as controls for Cre and iCre genotyping respectively. PCR reactions were separated on 2% agarose gels and images acquired using INTAS Gel documentation system.

Embryonic lethality analysis

Sep7floxed mice were mated and plug checked for embryo analysis. Pregnant mice were sacrificed between embryonic day 6–7.0 or 10.5 days. The embryos were dissected out in cold PBS and cleaned up from extra-embryonic tissues. Whole embryos were overnight digested for DNA isolation and genotyping. Deviations from Mendelian ratios were calculated by Chi-squared test.

Cell culture methods

Sep7 floxed mouse embryonic fibroblasts were generated from E13 day embryos and maintained under standard conditions. Sep7 floxed adult tail fibroblasts (TFs) were isolated from 6–8 weeks old mice tail tips. Minced tip were sequentially digested with collagenase and trypsin at 37°C and plated on collagen coated dishes in DMEM supplemented with 20% serum, non-essential amino acids and antibiotics. The cells were splitted 1:4 and maintained in the same growth medium without coated dishes. To immortalize primary TFs, cells were co-transfected with pSV40Tag encoding simian virus 40 large T antigen and pREP8 plasmid (Invitrogen) in a 10:1 mixture; colonies were selected with 2 mM histidinol (Sigma). Jurkat cells were maintained in RPMI-1640 medium supplemented with 15% serum, 1 mM pyruvate and antibiotics. Post electroporation cells were additionally supported by 2 ng/ml IL2.

Antibodies and reagents

Antibody against SEPT7 was from IBL international (JP18991), Rabbit anti-anillin antibodies were reported earlier (Watanabe et al., 2010). Antibodies used for western blot analysis were SEPT2 (#11397-1-AP, Acris), SEPT9 (#10769-1-AP, Acris), SEPT6 (sc-20180, Santa Cruz Biotech), SEPT8 (sc-48937, Santa Cruz Biotech), EF2 (sc-13004-R, Santa Cruz Biotech), GAPDH (#MAB374, Millipore), GFP (sc-9996, Santa Cruz Biotech) and Stathmin (#3352, Cell Signaling Technology). Antibodies used for Immunofluorescence staining were rabbit anti SEPT7, SEPT2, SEPT6 [23], K67, phospho-Histone-H3, cleaved caspase-3 (Cell Signaling Technology), tubulin-α (T6199, Sigma and sc-31779, Santa Cruz Biotech), LAP2 (#611000, BD Transduction lab) and acetyl tubulin (T6793, Sigma). All alexa-dye labeled secondary antibodies, tetramethyl rhodamine-conjugated WGA (#W849) and Alexa fluor-647-conjugated phalloidin (#A2287) were from Invitrogen. DAPI for DNA staining was from Carl Roth.
Polybrene (H9268), doxycycline (D9891), RNAse A (R4875) and propidium iodide (P4170) were from Sigma. Forchlorfenuron (FCF) was obtained from Santa Cruz Biotech. IL2 was from ImmunoTools. IL3, IL6 and SCF were from Peprotech.

Virus transduction

Primary MEFs were transduced with commercially available adenoviral Cre particles (AxCANCre2, TaKaRa, Japan). Gammaretroviral particles (SF91-nlsCre and pRBid-Cre) were packaged as described previously [27]. Doxycycline inducible retroviral

Figure 5. Stathmin expression levels correlate with SEPT7 dependence on cytokinesis. A, Lysates prepared from mouse embryonic fibroblasts (MEF), tail fibroblasts (TF), thymocytes (Thy) and bone marrow cells (BM) were probed with indicated antibodies. B, SEPT7flox/flox MEFs transduced with inducible stathmin expression construct were treated with 1 μg/ml doxycycline, to induce stathmin expression similar to thymocytes (Thy). C, Cre-induced SEPT7 depletion in SEPT7flox/flox cells in the presence and absence of doxycycline induced stathmin expression (5 days). D, WST-1 assay showing the rescue of Cre-transduction-induced proliferation defect in fibroblasts by doxycycline-induced stathmin expression. E, Effect of doxycycline induced stathmin expression on Rbid-Cre induced multinucleation analyzed and quantified from fluorescent images as indicated in Figure S12. F, G, Knockdown of stathmin in Jurkat cells. F, Stathmin expression was reduced to about 28% by knockdown. G, Stathmin knockdown renders Jurkat cell proliferation sensitive to the septin inhibitor forchlorfenuron (FCF) applied at two different concentrations (50 and 100 μM). doi:10.1371/journal.pgen.1004558.g005
expression vector used for generating Stathmin-IRES-EGFP cell line was packaged as described previously [42]. Immortalized tail fibroblasts were seeded in 24 well plates (2.5 x 10^5 cells/well) day before transduction. Plates with viral particles in the presence of polybrene (8 μg/mL) were spun at 1200 x g for 1 h at 32°C. After overnight virus treatment, cells were washed, medium changed and processed as indicated. For Sept7 deletion in primary lineage negative bone marrow progenitors, cells were trasduced by spinoculation with pRBid-Cre as described for tail fibroblasts. The transduction was repeated to achieve better transduction efficiency.

Western immunoblotting

Cells were lysed directly in SDS gel loading dye and western blotting was performed as previously described using gradient SDS-PAGE gels [43].

Immunofluorescence staining

Cells were grown on glass coverslips and fixed with 4% paraformaldehyde (PFA) in PBS. Fixation was performed for 2–5 min at room temperature (RT) followed by 20 min at 4°C. Cells were permeabilized with 0.25% Triton X-100–PBS for 30 min at RT. Blocking was done using 4% bovine serum albumin (BSA) for 1 h at 4°C. Primary antibodies were used at a 1:50 to 1:200 dilution in 1% BSA–PBS. Fixation was performed for 2–647-conjugated phalloidin/tetramethyl rhodamine conjugated WGA was used at a 1:500 dilution in 1% BSA–PBS for 1–2 h. Secondary antibodies or Alexa Fluor were used at a 1:100 in 1% BSA–PBS at RT for 30 min. After 1 h at 4°C, samples were stained and seeded in 96 well plates in the presence or absence of 2 μg/ml mitomycin A and 10 ng/ml murine IL2. Cell proliferation was assayed using WST1 reagent (Roche Applied Sciences) as per manufacturer protocol. For measuring fibroblast proliferation, 500 cells/100 μl/well were seeded in 96 well plates. Viable cells were quantified daily using WST1 reagent as per manufacturer protocol.

Stathmin knockdown analysis in Jurkat cells

Jurkat cells were microporated with control siRNA (Allstars negative control siRNA-Qiagen) or siRNA against human Stathmin (Hs_STMN1_1: 5'-GCUGAGGUCUUGAAGCAGCTT-3'–Qia-gen) using a Microporator MP-100 system. 200 picomoles of siRNA were used per one million cells microporated at 1400 V/20 msec/ single pulse following the standard manufacturer’s protocol. 24 h post transfection cells were counted and re-seeded at 3 x 10^5/mL density in the presence or absence of FCF in v-bottom 96well plates (triplicate wells). After 48 h of treatment cells were collected and viable cell numbers quantified by flow cytometry in the presence of 2 μg/ml propidium iodide and 2 mM EDTA.

Cell migration assay

Quantitative microplate scratch assays were performed with mitomycin-C treated fibroblasts as described previously [46]. Stathmin-IRES-EGFP transduced and sorted Sept7 knockdown cells were stained and seeded in 96 well plates in the presence or absence of 2 μg/ml doxycyclin and scratches were made 24 h later after mitomycin pre-treatment. Infrared fluorescent images were acquired using a Li-COR odyssey scanner at 0 h, 6 h and 18 h. Migration indices were calculated and plotted.

Ethics statement

All mice experiments were conducted according to German and international guidelines and were approved by the ethics committee of Hannover Medical School (MHf).

Supporting Information

Figure S1 Down-regulation of anillin in SEPT7-depleted fibroblasts. Band intensities for SEPT7 and anillin blots presented in figure 1C were quantified and normalized to GAPDH. The data for two different floxed lines are presented as percentage of non-transduced control. (PDF)
Figure S2  Sept7 deletion does not induce enhanced cell death. Adenoviral Cre-transduced or control treated Sept7flox/flox primary MEFs were analyzed for apoptotic cells by cleaved caspase-3 staining. Cells with typical apoptotic morphology and cleaved caspase-3 staining (shown in set) were counted and plotted. (PDF)

Figure S3  Microfilament and microtubule architecture in the Sept7-deficient fibroblasts. A, Immortalized Sept7flox/flox fibroblasts transduced with retroviral Cre showing unaltered F-actin staining in the absence of Sept7. B, Sept7 knockout primary MEFs showing unaltered F-actin staining and C, enhanced microtubule acetylation. D, General microtubule architecture is unaffected in Sept7-deficient immortalized fibroblasts as shown by α-tubulin staining. E, Intensity of acetyl tubulin (green)/Sept7 (red) staining were quantified from individual cells using ‘Color histogram’ plugin of Image J program (n = 10). Representative images used for analysis are shown in the right panel with the quantified intensity values. (PDF)

Figure S4  Dynamics of cell division in Sept7-deficient fibroblasts. Time-lapse images were acquired for Cre-transduced Sept7flox/flox tail fibroblasts as described in methods, Figure 3A and supporting video S1. Total time taken for individual cells to complete cytokinesis was calculated. A, Sample time-lapse analysis showing a cell (indicated by red arrow) undergoing the complete process from cell detachment to complete abscission in 80 min. B, Similar analysis of all successful divisions in 39 distinct cells followed by time lapse. C, Classification of mitotic cells followed by time-lapse including cells completing division (compiled from b) and cells failing to complete cell division. (PDF)

Figure S5  Staining for LAP2 in unresolved midbody structures in Sept7 KO fibroblasts. Upper and lower panel show two representative Sept7 floxed tail fibroblast cells transduced with mCherry-Cre and stained with indicated antibodies. LAP2/Tubulin/DNA triple staining revealed the presence of unresolved midbody structures lacking chromosome bridges. (PDF)

Figure S6  Bidirectional retroviral vector for expression of Cre and mCherry. A, Expression cassette and important features in the bidirectional Cre-mCherry expression vector used in the study. B, mCherry positive cells show efficient Sept7-deletion as shown by Sept7 co staining in Cre-transduced cells. C, Representative images of mCherry positive hematopoietic cell colonies genotyped and enumerated in Figure 4. (PDF)

Figure S7  Analysis of bone marrow in the Lymphocyte specific Sept7 KO. A, Sept7 genotyping showing partial deletion in CD2iCre mouse bone marrow. Tail biopsy DNA is shown as a control tissue. B, Lymphocytes: T cells (CD3+) and B cells (B220+) in the bone marrow from CD2iCre mice (n = 2) were analyzed by surface-staining and flow cytometry. (PDF)

Figure S8  Analysis of peripheral blood from lymphocyte specific Sept7 KO mice. Peripheral blood samples from Sept7flox/flox::CD2-iCre mice (2x) and Sept7flox/flox::CD2-iCre mice (3x) were analyzed using an ABC Vet Automated Blood counter (Sci1 animal care company GmbH, Viernheim, Germany). (PDF)

Figure S9  Co-depletion of other septins in Sept7 KO thymocytes. Similar to fibroblasts, Sept7 deletion (CD2iCre) in thymocytes lead to depletion of SEPT2/6/9. (PDF)

Figure S10  Analysis of tubulin acetylation in Sept7 KO thymocytes. A, Thymocytes from Sept7flox/flox::CD2-iCre mice (KO1 and KO2) and Sept7flox/flox::CD2-iCre mice (WT1) were analyzed by indirect fluorescence staining and flow cytometry analysis for acetylated tubulin. Labeled Secondary antibody only staining is shown as control. B, As positive control for flow-cytometric detection of acetyl tubulin, similar staining was performed with control and taxol (2 μM for 2 h) treated Jurkat cells. (PDF)

Figure S11  Effect of stathmin expression on fibroblast migration. Sept7 floxed MEFs inducibly expressing stathmin were subjected to scratch wound healing assay. A, Representative fluorescent scans of wells showing scratch wound healing. B, Calculated migration index for 6 and 18 h wound healing (n = 19). (PDF)

Figure S12  Analysis of multinucleation in stathmin expressing Sept7 KO fibroblasts. Sept7 floxed MEFs inducibly expressing stathmin were transduced with Rhoi-Cre, maintained in the presence or absence of 2 μg/ml doxycycline and were fixed and stained with DAPI. The extent of multinucleation in mCherry-positive cells in the presence or absence of doxycycline-induced stathmin expression was quantified and is presented in Figure 5E. (PDF)

Video S1  Stalled abscission and cytokinetic failure in Sept7 KO fibroblasts. Sequence from the time-lapse differential interference contrast (DIC) microscopy of Cre-transduced Sept7 floxed tail fibroblasts corresponding to Figure 3A. The cell attempting division (middle right) cannot resolve the intercellular bridge and does not complete cytokinesis after nuclear division. Even after 300 min the daughter cells do not separate and the cell becomes multi-nucleated. (AVI)

Acknowledgments
We thank Dr. S. Selbert (Polygene Transgenics) for help with the generation of the floxed Sept7 allele, Dr. B. Weinhold (MHF) for the ZP3-Cre, Dr. A. Kruger (MHF) for the hCD2-iCre strains, Dr. R. Bauerefeld (MHF) for support in confocal imaging and T. Yakovleva (MHF) for genotyping and breeding the mice. We thank H. Nakashima and D. Kamiya for help in the early part of MEF experiments.

Author Contributions
Conceived and designed the experiments: MBM. Performed the experiments: MBM AAs AK AC PM KSG. Analyzed the data: MBM AK AC PM MH AG RF. Contributed reagents/materials/analysis tools: ASc MGal. Contributed to the writing of the manuscript: MGae MK AK AC PM MK MH AG RF. Contributed conceptual insight: MGae AK MK.

References