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Ly-6\textsuperscript{Ch}\hi monocytes dominate hypercholesterolemia-associated monocytosis and give rise to macrophages in atheromata

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Macrophage accumulation participates decisively in the development and exacerbation of atherosclerosis. Circulating monocytes, the precursors of macrophages, display heterogeneity in mice and humans, but their relative contribution to atherogenesis remains unknown. We report here that the Ly-6\textsuperscript{Ch} monocyte subset increased dramatically in hypercholesterolemic apoE\textsuperscript{−/−} mice consuming a high-fat diet, with the number of Ly-6\textsuperscript{Ch} cells doubling in the blood every month. Ly-6\textsuperscript{Ch} monocytes adhered to activated endothelium, infiltrated lesions, and became lesional macrophages. Hypercholesterolemia-associated monocytosis (HAM) developed from increased survival, continued cell proliferation, and impaired Ly-6\textsuperscript{Ch} to Ly-6\textsuperscript{Cin} conversion and subsided upon statin-induced cholesterol reduction. Conversely, the number of Ly-6\textsuperscript{Cin} cells remained unaffected. Thus, we believe that Ly-6\textsuperscript{Ch} monocytes represent a newly recognized component of the inflammatory response in experimental atherosclerosis.

Introduction
During atherogenesis, an inflammatory process, leukocytes and lipids accumulate in the aortic intima (1, 2). Lipid-rich macrophages, known as foam cells in atheroma, secrete inflammatory mediators that stimulate smooth muscle cell migration and proliferation and participate in plaque development and rupture as well as thrombosis. Serum C-reactive protein and other molecular mediators of inflammation have broadened our understanding of the disease by illustrating that peripheral blood can afford valuable prognostic information (3, 4). Leukocyte counts rise in atherosclerotic patients. Yet, the extent to which circulating leukocyte subsets reflect the inflammatory response during atherogenesis remains less defined (5–9). This study tested the hypothesis that leukocyte heterogeneity in atherosclerosis could provide novel markers of and mechanistic insights into atherogenesis.

Prevailing concepts view monocytes as intermediary cells that continuously develop in the bone marrow, circulate in the bloodstream, and migrate unselected into tissue, where they become macrophages, dendritic cells, or other tissue descendants (10–12). Studies in atherosclerotic mice have shown that bone marrow–derived circulating monocytes populate atherosclerotic lesions (13–17), and many studies support an active role for monocytes/macrophages in atherosclerosis (reviewed in refs. 1, 2). The appreciation of monocyte heterogeneity (18–20) has led to the hypothesis that monocytes commit for specific functions while still in the circulation. Indeed, both human and mouse monocytes fall into at least 2 phenotypically distinct subsets: Ly-6\textsuperscript{Ch} (which are also phenotypically Gr-1 CCR2 CX3CR1\textsuperscript{hi}) and Ly-6\textsuperscript{Cin} (which are phenotypically Gr-1 CCR2 CX3CR1\textsuperscript{lo}) mouse monocytes correspond to human CD14\textsuperscript{lo}CD16\textsuperscript{−} and CD14\textsuperscript{−}CD16\textsuperscript{+} monocytes, respectively (19, 21–24). Ly-6\textsuperscript{Cin} cells selectively populate sites of experimentally induced inflammation, while their Ly-6\textsuperscript{Ch} counterparts can enter lymphoid and nonlymphoid tissues under homeostatic conditions (24).

This study explored Ly-6\textsuperscript{Ch} and Ly-6\textsuperscript{Cin} monocytes in atherosclerotic mice. The results show that hypercholesterolemia induced a surprisingly profound expansion of blood Ly-6\textsuperscript{Ch} but not Ly-6\textsuperscript{Cin} monocytes, a process we termed hypercholesterolemia-associated monocytosis (HAM). Our results also establish a direct link between circulating Ly-6\textsuperscript{Ch} monocytes and lesional macrophages.

Results
Hypercholesterolemic apoE\textsuperscript{−/−} mice undergo gradual and systemic monocytosis of the Ly-6\textsuperscript{Ch} subset. To test the hypothesis that high-fat feeding alters the repertoire of circulating monocytes, we analyzed peripheral blood mononuclear cells from C57BL/6 wild-type (referred to as apoE\textsuperscript{−/−}) and apoE\textsuperscript{−/−} mice that consumed either regular chow or Western diet (high in cholesterol and fat) for 25 weeks. Monocytes were defined as CD11b\textsuperscript{hi}CD90\textsuperscript{−}B220\textsuperscript{−}CD49b\textsuperscript{−}NK1.1\textsuperscript{−}Ly-6\textsuperscript{Gin} mononuclear cells by flow cytometry, as previously reported (16), and further divided into Ly-6\textsuperscript{Ch} and Ly-6\textsuperscript{Cin} fractions (Figure 1A). apoE\textsuperscript{−/−} mice on Western diet had a 4-fold increase of total circulating monocytes when compared with the same mice on chow (Figure 1B). Monocytosis in apoE\textsuperscript{−/−} mice on Western diet resulted from a 4-fold increase of the Ly-6\textsuperscript{Ch} subset (Figure 1C), whereas the Ly-6\textsuperscript{Cin} population remained unchanged (Figure 1D). Consumption of a Western diet increased slightly the number of total circulating leukocytes in apoE\textsuperscript{−/−} mice (mean ± SEM, chow, 3.0 ± 0.5 × 10\textsuperscript{6} cells/ml; Western diet, 3.9 ± 0.4 × 10\textsuperscript{6} cells/ml; Figure 1E). Blood smear counts showed that this increase arose primarily from monocytes (chow, 0.14 ± 0.03 × 10\textsuperscript{6} cells/ml; Western diet, 0.94 ± 0.11 × 10\textsuperscript{6} cells/ml), although granulocytes also increased (chow,
0.27 ± 0.03 × 10^6 cells/ml; Western diet, 0.96 ± 0.11 × 10^6 cells/ml).

and lymphocytes decreased slightly (chow, 2.4 ± 0.6 × 10^6 cells/ml; Western diet, 2.0 ± 0.2 × 10^6 cells/ml). As expected (25, 26), apoE<sup>−/−</sup> mice on Western diet had increased serum cholesterol levels (479 ± 20 mg/dl) when compared with apoE<sup>−/−</sup> mice on chow (286 ± 25 mg/dl). Macroscopic and histologic examination of aortas revealed fatty streaks and fibrous plaque lesions in the root and descending aorta of apoE<sup>−/−</sup> mice regardless of diet. Lesions were identified along the entire aorta in older mice (i.e., mice that consumed Western diet for 50 weeks; data not shown). apoE<sup>−/−</sup> mice consuming Western diet had 3–5 times more extended and widespread atherosclerotic lesions than did apoE<sup>−/−</sup> mice on chow.

Figure 1
Hypercholesterolemia induces peripheral blood Ly-6C<sup>hi</sup> monocytosis. (A) Mononuclear cells from blood of apoE<sup>+/+</sup> and apoE<sup>−/−</sup> mice consuming either chow or Western diet were stained with anti-CD11b, -CD90, -B220, -CD49b, -NK1.1, -Ly-6G, and -Ly-6C mAbs. Living cells were gated to determine presence and percentage of CD11b<sup>hi</sup>/CD90<sup>lo</sup>/B220<sup>lo</sup>/CD49b<sup>lo</sup>/NK1.1<sup>lo</sup>/Ly-6G<sup>lo</sup> monocytes (top row) and further divided into Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> subsets (bottom row). Representative dot plots and histograms from individual mice are depicted. Percentages of cells are shown as mean ± SEM. (B) Total blood monocytes in apoE<sup>+/+</sup> and apoE<sup>−/−</sup> mice consuming either Western diet (+) or chow (−). (C) Total blood Ly-6C<sup>hi</sup> monocytes. (D) Total blood Ly-6C<sup>lo</sup> monocytes. (E) Total peripheral blood leukocytes. (F) Representative dot plots depicting expression of CD62L and CD11c among Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocytes. Percentages of cells in each quadrant are shown as mean ± SEM. (G) Representative cyto
disp preparation of purified blood Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocytes in apoE<sup>+/+</sup> mice on chow and apoE<sup>−/−</sup> mice on Western diet. Scale bar: 10 μm. Student's t test was used. Results are representative of 8 independent experiments with 5–14 mice per group.
apoE+/+ mice fed a Western diet had serum cholesterol levels of 227 ± 39 mg/dl, lower than those observed in apoE–/– mice on Western diet but higher than those in apoE+/+ mice on chow (101 ± 10 mg/dl). apoE+/+ mice fed a Western diet did not show a significant increase in the number of circulating monocytes or Ly-6C+ monocytes (Figure 1, B–D) and did not develop atherosclerotic lesions during the 25 weeks of diet consumption (data not shown).

apoE–/– mice also had elevated numbers of CD11b+CD90−B220−CD49b+MK1.1+Ly-6G− cells (chow, 3.8% ± 0.4% cells; Western diet, 3.8% ± 0.4% cells) than apoE+/+ mice (chow, 1.8% ± 0.3% cells; Western diet, 1.9% ± 0.2% cells). These cells were phenotypically distinct from monocytes and were not examined further.

Although the number of circulating monocytes increased dramatically in apoE–/– mice fed Western diet, Ly-6Chi cells consistently expressed CD62 ligand (CD62L; also known as L-selectin) but not CD11c, while Ly-6Clo cells consistently expressed low levels of CD11c but not CD62L (Figure 1F), as previously reported for these monocyte subsets (21). Morphologic analysis of flow-sorted cells also showed that cells of both subsets retained their size as well as their characteristic kidney- or horse-shoe-shaped nuclei (Figure 1G).

Having determined that numbers of circulating monocytes increased in apoE–/– mice on Western diet, we assessed the spatial and temporal course of monocytosis development by quantification of monocytes and their subsets in the bone marrow, peripheral blood, and spleen over 250 days of Western diet consumption (Figure 2A). Analysis included additional compartments because the bone marrow produces monocytes and the spleen may serve as a reservoir for monocytes in the periphery. Monocytosis developed progressively in all 3 compartments, and the blood and spleen showed predominant expansion of the Ly-6Chi subset. Statistical analysis matched the data to an exponential growth curve, permitting determination of doubling time for each tissue. The Ly-6Chi subset showed the lowest doubling times (95% confidence interval, 33 to 38 days in the blood), while, as expected, the Ly-6Clo monocyte subset had the highest doubling times (95% confidence interval, 145 to 256 days in the blood).
Control experiments used blood from apoE^{+/+} and apoE^{−/−} mice during 250 days of chow consumption (Figure 2B). apoE^{+/+} mice on chow did not develop monocytosis, excluding the possibility that age drives the increase. apoE^{−/−} mice on chow showed moderate monocytosis. Statistical analysis matched the data to an exponential growth curve, though doubling times (e.g., 95% confidence interval, 63 to 86 days for Ly-6C^{hi} monocytes in the blood) were 1.9–2.3 times longer than in apoE^{−/−} mice on Western diet. Thus, atherosclerosis and Ly-6C^{hi} monocytosis arise concomitantly in apoE^{−/−} mice and show aggravation by Western diet.

Although we used the same criteria to define monocytes in the bone marrow and spleen as in blood (e.g., the CD11b^{+}CD90^{−}B220^{−}CD49b^{−}I-A^{b}−Ly-6G^{−}−Ly-6C^{lo} phenotype), the preponderance of macrophages and dendritic cells in the spleen necessitated a secondary step to ensure that the cells were indeed monocytes. In the spleen, monocytes were further defined as F4/80^{−}CD11c^{−}I-A^{b}−Ly-6G^{−}−Ly-6C^{lo}−. These cells had the same morphology as blood monocytes, whereas cells positive for F4/80, I-A^{b}, and/or CD11c resembled macrophages or dendritic cells (Figure 2C). Because of the low number of monocytes in the blood, the spleen therefore furnished a rich source of mononuclear cells for further study.

**Monocytosis results from increased survival, continued proliferation, and impaired Ly-6C^{hi} to Ly-6C^{lo} conversion.** Given that the progressive and peripheral monocytosis of the Ly-6C^{hi} subset in apoE^{−/−} mice was most robust during consumption of Western diet, we next sought to determine whether a model modified lipoprotein, such as acetylated low-density lipoprotein (AcLDL), directly influences monocyte survival and/or proliferation. Initially, splenic Ly-6C^{hi} and Ly-6C^{lo} monocytes from apoE^{−/−} mice on Western diet were isolated and cultured for 24 hours in medium supplemented or not with 100 μg/ml AcLDL. The presence of AcLDL allowed Ly-6C^{hi} cells to survive (Figure 3A) while retaining their monocytic Ly-6C^{hi}F4/80^{−}CD11c^{−}I-A^{b}−low phenotype. Conversely, survival of Ly-6C^{lo} cells did not change in the presence of AcLDL (data not shown). Because LDL can induce aortic endothelial cells to synthesize and secrete M-CSF (27), we assessed whether this factor can also influence the fate of monocytes in vitro. Supplementation of medium with M-CSF (50 μg/ml) for 24 hours partially fostered Ly-6C^{lo} monocyte survival (Figure 3A).

These data suggest that LDL and/or its derivatives directly and indirectly promote the survival of Ly-6C^{lo} monocytes. Nevertheless, the in vitro conditions used may not reproduce the in vivo environment in apoE^{−/−} mice on Western diet, since the number of Ly-6C^{lo} monocytes did not increase in vitro. Therefore, we sought to compare directly the proliferation and survival of monocytes in vivo. apoE^{−/−} animals consuming either chow or Western diet received daily i.p. injections of BrdU for 3 days and were sacrificed 1 day later. Mononuclear cells were purified from the bone marrow, blood, and spleen, and monocytes were analyzed for apoptosis (annexin V) and proliferation (anti-BrdU mAb). The Western diet decreased the number of annexin V− cells within Ly-6C^{lo} monocytes in the bone marrow and peripheral blood but not in the spleens of apoE^{−/−} mice (Figure 3, B and C). The Western diet concurrently increased the number of BrdU+ cells within Ly-6C^{lo} monocytes in peripheral blood and the spleen (Figure 3, B and C).
The vast majority of bone marrow monocytes incorporated BrdU regardless of diet, likely reflecting the continuous medullary production of these cells. These observations reflect the increased survival of dividing Ly-6C<sup>hi</sup> monocytes in apoE<sup>+/+</sup> mice consuming Western diet, but may result from accelerated production in the bone marrow and/or from increased extramedullary proliferation. Interestingly, the higher levels of BrdU incorporation by Ly-6C<sup>hi</sup> monocytes in the spleen (mean fluorescence intensity, 528 ± 111) compared with the bone marrow (mean fluorescence intensity, 293 ± 63) indicate either continued proliferation in the periphery or selective emigration from the bone marrow of cells with longer proliferative histories.

Analysis of apoptosis and proliferation in apoE<sup>+/+</sup> mice showed that Western diet increased survival and proliferation of monocytes in the spleen but not in the bone marrow or blood (Figure 3, B and C). Because apoE regulates apoptosis and cell cycle (28, 29), it may participate in the differences observed between apoE<sup>+/+</sup> and apoE<sup>−/−</sup> mice.

To determine whether the Western diet also affects conversion of Ly-6C<sup>hi</sup> to Ly-6C<sup>lo</sup> monocytes, apoE<sup>−/−</sup> mice on chow and Western diet received clodronate liposomes to deplete endogenous circulating monocytes (24). Mice administered clodronate liposomes had a dramatically reduced number of Ly-6C<sup>lo</sup>F4/80<sup>−/−</sup>CD11c<sup>−/−</sup>I-A<sup>b</sup>-low monocytes in blood (chow, 0.6 ± 0.2 x 10<sup>4</sup> versus 8.7 ± 4.6 x 10<sup>4</sup> cells/ml; Western diet, 1.1 ± 0.6 x 10<sup>4</sup> versus 9.6 ± 1.3 x 10<sup>4</sup> cells/ml) and spleen (chow, 2.4 ± 1.1 x 10<sup>5</sup> versus 32.5 ± 7.3 x 10<sup>4</sup> cells/ml; Western diet, 3.8 ± 0.9 x 10<sup>5</sup> versus 381.0 ± 84.2 x 10<sup>4</sup> cells/ml) 1 day after injection. Clodronate also strongly reduced the number of Ly-6C<sup>hi</sup>F4/80<sup>−/−</sup>CD11c<sup>−/−</sup>I-A<sup>b</sup>-high macrophages/dendritic cells and moderately reduced the number of Ly-6C<sup>lo</sup>F4/80<sup>−/−</sup>CD11c<sup>−/−</sup>I-A<sup>b</sup>-low monocytes (Figure 3D and data not shown). The near absence of Ly-6C<sup>lo</sup> monocytes on day 1 allowed us to study their reemergence from the Ly-6C<sup>hi</sup> repertoire (Figure 3, D and E). Five days after clodronate injection, Ly-6C<sup>lo</sup> monocytes had repopulated both blood and spleen in animals on chow but not in animals consuming Western diet (Figure 3, D and E, and data not shown). Impaired Ly-6C<sup>lo</sup> to Ly-6C<sup>hi</sup> conversion in apoE<sup>−/−</sup> mice consuming Western diet fostered Ly-6C<sup>hi</sup> monocytosis.

Ly-6C<sup>lo</sup> monocytes accumulate selectively in atherosclerotic lesions. Adoptively transferred EGFP<sup>+</sup> monocytes accumulate in atherosclerotic lesions (16), but low numbers of EGFP<sup>+</sup> signals detected by immunohistochemistry prevent quantification of cell accumulation. Here we employed a recently established flow cytometry method (30) to phenotype single-cell suspensions of enzyme-digested aortas and determine the in vivo relevance of Ly-6C<sup>lo</sup> monocytosis to atherosclerosis. Aortas from apoE<sup>−/−</sup> mice on chow and Western diet were digested with a protease cocktail. Cells were dispersed and stained with anti-CD11b, -CD90, -B220, -CD49b, -NK1.1, -Ly-6G, -F4/80, -I-A<sup>b</sup>, -CD11c, and -Ly-6C mAb. Percent (mean ± SEM) are shown for each quadrant. (B) Number of retrieved Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocytes per aorta in apoE<sup>+/+</sup> and apoE<sup>−/−</sup> mice on chow and Western diet. Results are pooled from 5 independent experiments with 2–5 mice per group. Mean and SEM are shown. *P < 0.001 versus all other groups (1-way ANOVA with Tukey’s multiple comparison test). (C) Immunohistochemistry depicts the intima at the aortic root of a representative apoE<sup>−/−</sup> mouse on Western diet and an apoE<sup>+/+</sup> mouse on chow. Sections stained with anti-CD31, -Ly-6C, and -Mac-3 mAbs are shown. Original magnification, ×400.
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Figure 5
Ly-6C<sup>+</sup> monocytes adhere preferentially to TNF-α-activated endothelium, accumulate in lesions, and differentiate to macrophages in vivo. (A) Purified Ly-6C<sup>+</sup> monocyte phenotype after 24 hours in culture. (B) Adherence of blood monocytes on TNF-α-treated MHECs under laminar flow conditions. Monocyte subsets were isolated from the blood of apoE<sup>−/−</sup> and apoE<sup>+/+</sup> mice on chow and Western diet. (C) Relative proportion of blood monocytes expected to adhere to activated endothelium, based on the capacity of each subset to adhere and their average abundance in peripheral blood of apoE<sup>−/−</sup> mice on Western diet for 25 weeks. (D) Ly-6C, F4/80, and I-A<sup>+</sup> expression of CD45.2<sup>+</sup> Ly-6C<sup>+</sup> monocytes retrieved from aortas and spleens 24 hours after transfer into CD45.1<sup>+</sup> mice (both donor and recipient apoE<sup>−/−</sup> mice consuming a Western diet). Monocytes cultured in vitro were also analyzed. (E) F4/80 and Ly-6C coexpression on CD45.2<sup>+</sup> donor cells retrieved from aortas. (F) In vivo aortic accumulation of [111<sup>II</sup>]oxine-labeled Ly-6C<sup>+</sup> and Ly-6C<sup>+</sup> monocytes 24 hours after adoptive transfer in apoE<sup>−/−</sup> mice on Western diet. (G) Phosphorimager plates depicting relative distribution of signal in aortas of apoE<sup>−/−</sup> mice that received equal numbers of Ly-6C<sup>+</sup> apoE<sup>−/−</sup> or Ly-6C<sup>+</sup> apoE<sup>−/−</sup> monocytes. (H) Relative proportion of Ly-6C<sup>+</sup> and Ly-6C<sup>+</sup> monocytes expected to accumulate in atherosclerotic aortas, based on the capacity of each subset for aortic accumulation and their average abundance in peripheral blood of apoE<sup>−/−</sup> mice on Western diet for 25 weeks. Shown are 1 of 2–3 independent experiments. Student’s t test was used.

and/or dendritic cells in apoE<sup>+</sup> mice on Western diet (gate ii; 4.9 ± 1.7 × 10<sup>4</sup> cells) than on chow (1.1 ± 0.3 × 10<sup>4</sup> cells).

We counted relatively low numbers of Ly-6C<sup>+</sup> and Ly-6C<sup>+</sup> monocytes and macrophages/dendritic cells in the aortas of apoE<sup>−/−</sup> mice regardless of diet (Western diet, 260 ± 10 Ly-6C<sup>+</sup> monocytes, 310 ± 160 Ly-6C<sup>+</sup> monocytes, 1.3 ± 0.5 × 10<sup>4</sup> macrophages/dendritic cells; chow, 220 ± 170 Ly-6C<sup>+</sup> monocytes, 230 ± 120 Ly-6C<sup>+</sup> monocytes, 1.2 ± 0.3 × 10<sup>4</sup> macrophages/dendritic cells; Figure 4B).

The above cell numbers likely underestimate actual values, since the enzymatic digestion of aortic tissue required to obtain single-cell suspensions caused the death of many cells (90% ± 1% of events and Ly-6C<sup>+</sup> monocyte subsets in apoE<sup>−/−</sup> mice on Western diet suggests that greater than 95% of cells capable of binding to activated endothelium would belong to the Ly-6C<sup>+</sup> subset (Figure 5C).

Further exploration used adoptive transfer of Ly-6C<sup>+</sup> monocytes from CD45.2 apoE<sup>−/−</sup> mice into congenic CD45.1 apoE<sup>−/−</sup> recipients (both donor and recipient mice on Western diet for 25 weeks). Because peripheral blood contains few monocytes, splenic monocytes served as surrogates for circulating monocytes. After 24 hours, aortas from recipient mice were digested enzymatically, and single-cell suspensions were analyzed by flow cytometry. We counted 135 ± 4 donor cells in recipient aortas, of which approxi-
mately 25% were Ly-6C<sup>hi</sup> and approximately 75% were Ly-6C<sup>lo</sup> (Figure 5D). Many donor cells showed enhanced expression of F4/80 and I-A<sub>b</sub> (Figure 5D). Remarkably, the combined analysis of Ly-6C and F4/80 expression by donor cells revealed the existence of at least 3 distinct populations, Ly-6C<sup>hi</sup>F4/80–, Ly-6C<sup>hi</sup>F4/80+, and Ly-6C<sup>lo</sup>F4/80+, resembling monocytes, monocytes in the process of differentiating into macrophages, and mature macrophages, respectively (Figure 5E). These phenotypic relationships are comparable to those observed among endogenous populations (Figure 4A). In contrast, donor cells retrieved from the spleens of the same mice remained phenotypically unchanged (Figure 5D).

These results demonstrate recruitment of Ly-6C<sup>hi</sup> monocytes to atherosclerotic aortas, followed by local and rapid (<24 hours) differentiation into macrophages. Combined with the restricted localization of Ly-6C<sup>hi</sup> monocytes to the luminal face of the endothelium, these data suggest that differentiation into macrophages accompanies transmigration and also support the notion that aortic monocyte detection did not result from contamination of circulating cells.

To determine the relative capacity of Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocytes to migrate to atherosclerotic aortas and to determine whether accumulation of Ly-6C<sup>hi</sup> cells mapped to lesions, we labeled equal numbers of splenic Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocytes from apoE<sup>−/−</sup> mice on Western diet with [111<sup>In</sup>]oxine and injected them separately into apoE<sup>−/−</sup> mice on Western diet. After 24 hours, we excised the aortas and calculated the percent injected dose per gram of tissue, which revealed that Ly-6C<sup>hi</sup> cells preferentially accumulated in aortas (Figure 5F). Autoradiography showed discrete regions of activity only in recipients of Ly-6C<sup>hi</sup> cells (Figure 5G). The lack of such regions in animals receiving Ly-6C<sup>lo</sup> cells suggests that the signals detected in these mice corresponded to background activity. The radioactive signal observed for Ly-6C<sup>hi</sup> cells always mapped directly to areas containing lesions as determined microscopically, but not all lesions showed focal areas of radioactivity (data not shown). The relative proportion of circulating Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> monocyte subsets in apoE<sup>−/−</sup> mice on Western diet suggests that greater than 90% of cells accumulating in atherosclerotic lesions originate from the Ly-6C<sup>hi</sup> subset (Figure 5H). Taken together, these results suggest that circulating Ly-6C<sup>hi</sup> monocytes are direct precursors of lesional macrophages.

Expression of MCP-1 by a subset of cells in atherosclerotic lesions suggests active recruitment of monocytes to developing lesions in vivo (31, 32), and mice lacking MCP-1 or C-C motif chemokine receptor 2 (CCR2) show reduced atherosclerosis (33–35). Thus monocyte recruitment into lesions may require CCR2 expression, a feature of the Ly-6C<sup>hi</sup> subtype (19). Testing this hypothesis involved the adoptive transfer of Ly-6C<sup>hi</sup> monocytes from bone marrow of CCR2<sup>−/−</sup> mice into peripheral blood of atherosclerotic (CCR2<sup>−/−</sup>) apoE<sup>−/−</sup> mice. We counted only 14 ± 2 donor cells in recipient aortas, suggesting that Ly-6C<sup>hi</sup> monocyte accumulation in lesions does indeed depend on CCR2.

Figure 6

Statin treatment limits monocytosis. apoE<sup>−/−</sup> mice consumed Western diet supplemented or not with atorvastatin for 25 weeks. A control group of apoE<sup>−/−</sup> mice received regular chow. (A) Serum cholesterol after 25 weeks of diet. (B) Number of leukocytes, monocytes, and Ly-6C<sup>hi</sup> and Ly-6C<sup>lo</sup> subtypes in bone marrow, blood, and spleen. (C) Association between serum cholesterol and number of circulating Ly-6C<sup>hi</sup> or Ly-6C<sup>lo</sup> monocytes after 25 weeks of diet. Mean ± SEM are shown for apoE<sup>−/−</sup> mice on chow (filled circles), Western diet (open circles), and Western diet supplemented with atorvastatin (gray circles). Results are pooled from 9 independent experiments (n = 3–14 per group). Student’s t test was used.
reduction of Ly-6C\textsuperscript{hi} monocytosis attenuates disease. Repeated administration of anti-Ly-6C mAb could theoretically control Ly-6C\textsuperscript{hi} monocytosis. This approach is impractical, given the chronic nature of atherogenesis, and may not keep the size of the Ly-6C\textsuperscript{hi} monocyte population at homeostatic levels, but rather trigger transient cell depletion upon each mAb injection. In contrast, since inhibitors of hydroxymethylglutaryl coenzyme A reductase (statins) decrease cholesterol levels, exert antiinflammatory effects, and attenuate atherosclerosis (36, 37), we sought to determine whether concurrent treatment of apoE \textsuperscript{−/−} mice on Western diet with statin also modulates the extent of monocytosis. Mice analyzed after 25 weeks of atorvastatin treatment had significantly attenuated serum cholesterol levels when compared with age-matched littermates on Western diet (Figure 6A). The statin treatment also reduced monocytosis (Figure 6B). Specifically, the numbers of Ly-6C\textsuperscript{hi} monocytes declined significantly in the spleen and peripheral blood (although they remained higher than in apoE \textsuperscript{−/−} mice on chow), whereas the numbers of Ly-6C\textsuperscript{lo} monocytes in the bone marrow fell to levels found in apoE \textsuperscript{−/−} mice on chow (Figure 6B). The statin treatment also reduced the numbers of the Ly-6C\textsuperscript{lo} monocytes in the spleen. Antiinflammatory effects of statins beyond LDL lowering could also participate in the attenuation of Ly-6C\textsuperscript{lo} cell numbers.

Blood Ly-6C\textsuperscript{hi} monocyte counts positively associated with serum cholesterol levels in apoE \textsuperscript{−/−} (data not shown) and apoE \textsuperscript{−/−} mice (Figure 6C). Furthermore, correlation between Ly-6C\textsuperscript{hi} numbers from statin-treated apoE \textsuperscript{−/−} mice conformed to a linear axis between chow- and Western diet–fed animals. Such correlutive analysis did not apply for blood Ly-6C\textsuperscript{lo} monocytes, since Ly-6C\textsuperscript{lo} cell counts were similar for all serum cholesterol concentrations.

Discussion

This study shows that hypercholesterolemia in apoE \textsuperscript{−/−} mice induced the progressive and selective expansion of Ly-6C\textsuperscript{hi} monocytes, a phenomenon we have termed HAM. Ly-6C\textsuperscript{hi} monocytes participate in atherosclerosis because they preferentially adhere to activated endothelium, accumulated in lesions, and locally differentiated into macrophages, the predominant leukocytes during plaque inflammation and development. Although we did not describe a role for Ly-6C\textsuperscript{hi} cells in atherosclerosis, future studies may determine whether this subset, as a possible precursor of dendritic cells (38), influences the adaptive arm of the immune system, particularly with respect to antigen presentation and lymphocyte activation, processes known to occur in atherosclerosis (1).

HAM results from continued bone marrow production of Ly-6C\textsuperscript{hi} monocytes, increased survival of these cells in the periphery, and impaired conversion to the Ly-6C\textsuperscript{lo} subset. Monocytosis was characterized by a doubling of the Ly-6C\textsuperscript{hi} population every month in peripheral blood, generating up to $1.4 \times 10^9$ cells/ml after 250 days of diet; although the rate of Ly-6C\textsuperscript{hi} monocytosis was slightly slower in the spleen (doubling time of around 2 months), this organ served as a much larger reservoir of Ly-6C\textsuperscript{hi} monocytes, reaching approximately $2 \times 10^7$ cells after 250 days of diet. Moreover, since the number of Ly-6C\textsuperscript{lo} monocytes continuously increased during the 250 days of the study, monocytosis expansion would likely have persisted beyond this point.

We suggest that Ly-6C\textsuperscript{hi} monocytosis requires elevated concentrations of cholesterol or lipid derivatives, since (a) monocytosis occurred in mice fed Western diet, (b) the presence of a cholesterol-lowering drug of the statin class at least partially controlled monocytosis in vivo, and (c) the addition of LDL for 24 hours to purified Ly-6C\textsuperscript{hi} monocytes in vitro allowed for their increased survival without promoting differentiation into macrophages. These findings demonstrate that augmented serum cholesterol levels, in addition to promoting lipid deposition in lesions (39) and inducing M-CSF production for local monocyte maturation into macrophages (40, 41), also trigger the expansion of circulating monocytes. These data suggest that a cholesterol-rich diet promotes murine atherogenesis, at least in part through the development of monocytosis. Molecules such as M-CSF, GM-CSF, PI3K, and NF-κB are induced by constituents of modified LDL and mediate the expansion and maturation of macrophages (42, 43). It will be of interest to determine whether similar pathways also elicit Ly-6C\textsuperscript{hi} monocytosis and why circulating Ly-6C\textsuperscript{lo} monocytes appear less sensitive to the same stimuli.

Following the fate of adoptively transferred, purified monocye subsets revealed that lesional macrophages derived predominantly from circulating Ly-6C\textsuperscript{hi} monocytes, pointing to an active role in this process for the membrane-bound, glycosylphosphatidylinositol-linked protein Ly-6C (44–46). Future studies will show whether Ly-6C serves only as a marker for or also contributes functionally to inflammatory monocytes, whether Ly-6C characterizes inflammatory cells capable of tissue transmigration, and whether therapeutic targeting of Ly-6C, or a human equivalent, contains monocytosis and consequently reduces atherosclerosis.

The observation that Ly-6C\textsuperscript{hi} cells trafficked differentially to lesions supports the concept of heterogeneity of atheroma (47, 48) and suggests that these cells migrate preferentially to lesions of high inflammatory activity. Interestingly, fatty streaks harbored few Ly-6C\textsuperscript{hi} monocytes, indicating low monocyte accumulation at this stage. Expression of CD62L also suggests a role for Ly-6C\textsuperscript{hi} monocytes in antigen presentation in secondary lymphoid organs and participation in adaptive immunity.

Activated macrophages, operationally categorized as M1 cells (classically activated macrophages), are induced by IFN-γ and generate inflammation, while M2 cells (alternatively activated macrophages) generated in response to IL-4 or IL-13 can exhibit immunoregulatory function (49, 50). Ly-6C\textsuperscript{hi} monocytes selectively populate sites of experimentally induced inflammation and can locally differentiate into M1 cells. However, whether the cytokine milieu recruits and/or activates specific monocyte subsets, or indiscriminationally polarizes already accumulated subsets for particular macrophage function will require further study.

Clinical data have shown a positive correlation between white blood cell count and acute myocardial infarction (5–7). More recently, a study showed that monocytosis is an independent risk factor for coronary artery disease (51). Here we propose that monocytosis is an etiologic constituent of experimental atherogenesis. Future research will determine whether humans with elevated serum cholesterol and/or C-reactive protein exhibit CD14\textsuperscript{hi}CD16\textsuperscript{−} monocytosis, and whether this variable relates to atheroma burden or complications. Overall, our present results indicate a striking role of innate immunity and atherosclerosis that will require future testing experimentally and in humans.
Methods

**Animals.** apoE<sup>+/-</sup> (C57BL/6) and apoE<sup>-/-</sup> (B6.129P2-Apoem<sup>1Unc</sup>) mice were purchased from The Jackson Laboratory. The apoE<sup>-/-</sup> animals had been backcrossed to the C57BL/6 background for at least 10 generations. apoE<sup>-/-</sup> CD45.1<sup>+/+</sup> mice were generated after backcrossing apoE<sup>-/-</sup> mice to C57BL/6 CD45.1<sup>+/+</sup> mice (The Jackson Laboratory). C57BL/6 CCR2<sup>/-/-</sup> mice were a gift from B. Rollins (Dana-Farber Cancer Institute, Boston, Massachusetts, USA), A. Luster (Massachusetts General Hospital and Harvard Medical School), and I. Charo (UCSF, San Francisco, California, USA). At 10 weeks of age, groups of animals were placed on a Western diet (21.2% fat/weight, 0.2% cholesterol; Harlan Teklad), which was defined as day 0 in this study. The remaining animals consumed a regular chow diet. A group of animals received supplemental atorvastatin in their diet (52) (0.01% w/w) starting on day 0. All studies were conducted with age-matched animals after 20–25 weeks of diet unless otherwise stated. All protocols were approved by the Subcommittee on Animal Research Care (SARC) at Massachusetts General Hospital.

**Cells.** Cells were harvested from bone marrow, peripheral blood, spleens, and aortas at the time points indicated in Results. Bone marrow from both tibias was harvested, and the cells were collected by inserting a needle into the bone and washing with HBSS supplemented with 0.2% (w/v) BSA and 1% (w/v) FCS. Peripheral blood was drawn via cardiac puncture with citrate solution (100 mM Na-citrate, 130 mM glucose, pH 6.5) as anticoagulant, and mononuclear cells were purified by density centrifugation (16). Total leukocyte numbers were determined using acetic acid lysis solution (3% HEMA 3 solution II, 94% ddH<sub>2</sub>O, 3% glacial acetic acid). Blood smears were prepared to corroborate monocyte numbers and subtract percent granulocytes from total leukocyte counts. Spleens were removed, triturated in HBSS (cellgro; Mediatech Inc.) at 4°C with the end of a 3-ml syringe, and filtered through nylon mesh (BD Biosciences). Aortas were excised and placed into a cocktail of collagenase I, collagenase XI, DNase I, and hyaluronidase (Sigma-Aldrich) at 37°C for 1 hour, as described previously (30). Cells were then triturated through nylon mesh. The cell suspensions were centrifuged (15 minutes, 500 g, 4°C), red blood cells were lysed with ACK lysis buffer, and the resulting single-cell suspensions were washed with HBSS supplemented with 0.2% (w/v) BSA and 1% (w/v) FCS.

**mAbs and flow cytometry.** For visualization of monocytes, cells were incubated with a cocktail of mAbs against T cells (CD90-PE, 53-2.1), B cells (CD19-FITC, B220-PE, CD23-PE, CD69-PE, CD40-PE, CD132-PE), and macrophages (CD11b-PE, CD11c-FITC (HL3) and CD62L-FITC (MEL-14). CD45.2-FITC and CD45.1-PE (BD Biosciences Inc.). Monocytes were identified as cells 10–30 μm in diameter with a horseshoe- or kidney-shaped nucleus, a variable nucleus-to-cytoplasm ratio, and fine granules and vacuoli.

Mouse heart EC isolation. Mouse heart ECs (MHECs) were isolated and cultured from 7-day-old mice (54). Briefly, harvested tissues were digested with collagenase (Worthington Biochemical Corp.), and ECs were isolated with 2 rounds of magnetic bead purification (Invitrogen), the first with anti–PECAM-1 and the second with anti–ICAM-2 mAbs (BD Biosciences).

Monocyte adhesion under defined laminar flow conditions. Monocytes from blood of apoE<sup>-/-</sup> and apoE<sup>+</sup/>mice were isolated. Interactions with MHECs were examined under conditions of fluid shear stress in a parallel plate flow chamber (55). Briefly, confluent MHECs were grown on glass coverslips (25-mm diameter; Carolina Biological Supply Co.) coated with 5 μg/ml fibronectin (Sigma-Aldrich), stimulated with 120 ng/ml TNF-α for 4 hours, and inserted into the flow chamber. Monocytes (10<sup>6</sup> cells) were suspended in flow buffer (Dulbecco PBS, 0.75 mM Ca<sup>2+</sup> and Mg<sup>2+</sup>, containing 0.1% human serum albumin) and drawn through the flow chamber as a bolus at an estimated shear stress of 0.76 dynes/cm<sup>2</sup>. Monocyte accumulation was measured after 3 minutes of flow by counting stably adherent monocytes in 12–16 different fields. Monocyte–MHEC interactions were visualized using videomicroscopy (x20 phase-contrast objective) and digitally recorded to a PC using VideoLab software (version 2.2.1; Ed Marcus Laboratories).

**Immunohistochemistry.** Aortas were excised, frozen in O.C.T. compound (Sakura), sectioned in 5-μm slices, and stained by the avidin-biotin-peroxidase method as previously described (56). Cell types were identified with primary antibodies: monoclonal anti-CD31 (MEC.13.3) for endothelial cells, rat anti-mouse Mac-3 (M3/84) for macrophages, and anti-Ly-6C (AL-21) antibody to mouse macrophage precursor cells (BD Biosciences). Images were captured with a digital camera (Nikon DXM1200F) using imaging software ACT-1 (version 2.63; Nikon).

**In vivo accumulation.** Purified Ly-6C<sup>+</sup> or Ly-6C<sup>+</sup> monocytes from the spleen of apoE<sup>-/-</sup> or apoE<sup>+</sup>/CCR2<sup>-/-</sup> mice were injected into apoE<sup>-/-</sup> CD45.1<sup>+/+</sup> mice or labeled with <sup>111</sup>Inoxine according to the manufacturer’s protocol (Amersham Biosciences). For imaging and autoradiography experiments, cells were washed with HBSS, spun, and resuspended in <sup>111</sup>Inoxine for 15 minutes at 37°C, pH 6.5–7.5. The cells were then washed twice with HBSS. This labeling protocol keeps the cells viable and in a resting state (16). Approximately 0.8 × 10<sup>6</sup> cells incorporating 50 μCi were injected i.v. into apoE<sup>-/-</sup> mice fed Western diet for 50 weeks. The total amount of activity that was injected into each animal was measured with a radioisotope calibrator (Capintec Inc.). For flow cytometric studies, approximately 4 × 10<sup>6</sup> Ly-6C<sup>+</sup>CD45.2<sup>+</sup> monocytes were injected into CD45.1<sup>-/-</sup> apoE<sup>-/-</sup> mice fed Western diet for 50 weeks. After 24 hours, all animals were euthanized with CO<sub>2</sub>. Aortas were perfused in situ with 10 ml HBSS via the left ventricle and excised from the root to the bifurcation. All heart and fat tissue was excised to eliminate confounding signal. Aortas were then subjected to digestion (flow cytometric studies, see above). For radioactive studies, cpm for each aorta as well as blood were measured using a Wallac 1480 WIZARD gamma counter (PerkinElmer). Percent injected dose per gram tissue was calculated after correcting for