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Cytosolic Phospholipase A2 Protein as a Novel Therapeutic Target for Spinal Cord Injury

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Objective: The objective of this study was to investigate whether cytosolic phospholipase A2 (cPLA2), an important isoform of PLA2 that mediates the release of arachidonic acid, plays a role in the pathogenesis of spinal cord injury (SCI).

Methods: A combination of molecular, histological, immunohistochemical, and behavioral assessments were used to test whether blocking cPLA2 activation pharmacologically or genetically reduced cell death, protected spinal cord tissue, and improved behavioral recovery after a contusive SCI performed at the 10th thoracic level in adult mice.

Results: SCI significantly increased cPLA2 expression and activation. Activated cPLA2 was localized mainly in neurons and oligodendrocytes. Notably, the SCI-induced cPLA2 activation was mediated by the extracellular signal-regulated kinase signaling pathway. In vitro, activation of cPLA2 by ceramide-1-phosphate or A23187 induced spinal neuronal death, which was substantially reversed by arachidonyl trifluoromethyl ketone, a cPLA2 inhibitor. Remarkably, blocking cPLA2 pharmacologically at 30 minutes postinjury or genetically deleting cPLA2 in mice ameliorated motor deficits, and reduced cell loss and tissue damage after SCI.

Interpretation: cPLA2 may play a key role in the pathogenesis of SCI, at least in the C57BL/6 mouse, and as such could be an attractive therapeutic target for ameliorating secondary tissue damage and promoting recovery of function after SCI.

T raumatic spinal cord injury (SCI) leads to neurological deficits and motor and sensory dysfunctions. In the United States alone, there were approximately 270,000 people living with SCI in 2012, and an additional 12,000 new SCI cases occur every year, most of them younger than 30 years (https://www.nscisc.uab.edu).1 To date, there is no effective pharmacological treatment for SCI.2 SCI is caused by mechanical damage that triggers cellular events culminating in the secondary injury phase, which provides an important therapeutic window for neuroprotective strategies to improve recovery of function after SCI. Previous studies indicate that multiple injury mechanisms, including inflammation, oxidative stress, and glutamate excitotoxicity,3–6 are involved in the secondary injury process after initial trauma, but exact mechanisms remain to be fully elucidated.

Several lines of evidence suggest that phospholipase A2 (PLA2) may play a key role in mediating multiple injury insults, as mentioned above, after SCI.7–11 PLA2 is a diverse family of enzymes that hydrolyze the acyl bond at the sn-2 position of glycerophospholipids to produce free fatty acids and lysophospholipids.8,12,13 These products are precursors of bioactive eicosanoids and platelet...
activating factor, which are well-known mediators of inflammation and tissue damage implicated in pathological states of several acute and chronic neurological disorders.8,12–14 Our previous study showed that PLA2 activity and expression increased after SCI in rats.15 Injections of exogenous PLA2 or melittin, a potent activator of endogenous PLA2, into the normal spinal cord resulted in inflammation and tissue damage.15 Administration of annexin A1, a nonselective inhibitor of PLA2, inhibited SCI-induced inflammation and reduced tissue damage after SCI.16 These findings suggest that PLA2 may be a potential therapeutic target for SCI.

PLA2 can be broadly classified into 3 major categories: secretory PLA2 (sPLA2), cytosolic PLA2 (cPLA2), and Ca2+-independent PLA2 (iPLA2).8 Among them, cPLA2 is considered to be the most important PLA2 isoform, because it has been implicated as an effector in receptor-mediated release of arachidonic acid (AA) and exhibits strong preference for deacylation of AA over other fatty acids.13,17 However, the role of cPLA2 in the pathogenesis of SCI has not yet been fully understood, and is even controversial.15,18 Here, we report that SCI significantly induced cPLA2 activation and expression. Blocking cPLA2 pharmacologically and genetically ameliorated motor deficits, and reduced cell loss and tissue damage after SCI in mice. Thus, cPLA2 may represent a therapeutic target for treatment after traumatic SCI.

Materials and Methods
All of the chemicals used in this study were from Sigma (St Louis, MO), except for those specifically indicated. Antibodies used in this study were from Cell Signaling Technology (Boston, MA), except for those specifically indicated.

Mice and Rats
Female C57BL/6 mice (12 weeks, 18–24g) were purchased from Jackson Laboratories (Bar Harbor, ME). Breeding pairs of male and female heterozygous (cPLA2+/−) mice were kindly provided by Dr J. Bonventre (Harvard Medical School). The breeding was carried out at Indiana University School of Medicine Laboratory Animal Resource Center. Female cPLA2+/− mice and wild-type (WT) littermates (12 weeks, 18–24g) generated from heterozygous breeding pairs were used in this study. The genotypes of the yielded litters were determined by polymerase chain reaction (PCR). All mice were on a C57/BL6 background.19,20 Female Sprague–Dawley rats (210–230g) were purchased from Harlan (Indianapolis, IN). Female animals are routinely used in SCI studies, because female animals allow for easier manual expression of bladders after SCI, less urinary tract infection, and less mortality.21–24 In addition, there are reports showing that no significant differences were detected in histological and behavioral outcomes between male and female animals after SCI.25,26 The animals were maintained on a 12/12-hour light/dark cycle with food and water freely available. All surgical interventions, treatments, and postoperative animal care were performed in accordance with the Guide for the Care and Use of Laboratory Animals (National Research Council) and the Guidelines of the Institutional Animal Care and Use Committee of the Indiana University School of Medicine.

Contusive Spinal Cord Injury and Treatment
A contusive SCI in mice was performed at the T10 level using an Infinite Horizon Impactor (Infinite Horizons, Lexington, KY) at an impact force of 60 kdyne as described.27 The rats underwent a T10 contusive spinal cord injury using a New York University Impactor (10g, 12.5mm) as was described previously.16,28 After injury, the muscles and skin were closed in layers, and animals were placed in a temperature- and humidity-controlled chamber overnight. Manual bladder expression was carried out at least 3× daily until reflex bladder emptying was established. Control animals received laminectomy only. At 30 minutes after contusion injury, mice were treated with arachidonyl trifluoromethyl ketone (AACOCF3; Cayman Chemicals, Ann Arbor, MI), delivered intraperitoneally (50μl of 4mM), followed by intraperitoneal injections of the compound (200μl of 4mM) every other day up to 2 weeks postinjury. The dose and treatment regimen were selected based on our pilot study and a previous published report.29 Another group of SCI animals received vehicle injections.

Western Blotting
Western blot analysis was performed as described previously with minor modification.16,28 For cPLA2 expression, primary antibodies included mouse monoclonal anti-cPLA2 antibody (1:100; Santa Cruz Biotechnology, Santa Cruz, CA), polyclonal rabbit anti–phospho(p)-cPLA2 antibody (1:500), and mouse anti–β-tubulin antibody (1:1,000; Sigma). For active caspase-3 and poly(adenosine diphosphate ribose) polymerase (PARP) expression, primary antibodies included rabbit anti–caspase-3 antibody (cleaved, 1:1,000), rabbit anti–PARP-1 (cleaved, 1:500), and mouse anti–β-tubulin antibody (1:1,000, Sigma). For extracellular signal-regulated kinase (ERK) expression, primary antibodies included rabbit anti–ERK1/2 antibody (1:1,000) and monoclonal mouse anti–p-ERK1/2 antibody (1:2,000). Secondary Alexa Fluor 680 goat antimouse (1:10,000; Invitrogen, Grand Island, NY) and IRDye 800 goat antimouse (1:400; Vector Laboratories, Burlingame, CA) antibodies were used. The Western blot was imaged and quantified using a Li-Cor Odyssey Infrared Imaging system (LI-COR Biosciences, Lincoln, NE) according to the manufacturer’s instruction.

Immunohistochemistry
Immunohistochemistry followed procedures described previously.15 One set of the sections was incubated with primary polyclonal rabbit anti–p-cPLA2 antibody (1:100) overnight at 4°C. On the second day, the sections were incubated with secondary biotinylated goat antirabbit immunoglobulin G antibody (1:400; Vector Laboratories, Burlingame, CA) for 1 hour at room temperature. Primary antibody omission controls were
used to further confirm the specificity of the immunohistochemical labeling.

**Immunofluorescence Double Labeling**

This method has been described in our previous publication. Briefly, a mixture of rabbit polyclonal anti–phospho(p)-cPLA₂ (1:100) and mouse anti–NeuN (1:100; Chemicon, Temecula, CA), anti–SMI-31 (1:2,000; Sigma), and anti–CC1 (APC-7, 1:100; Calbiochem, San Diego, CA) antibodies were used to examine colocalization of p-cPLA₂ in neurons, axons, and oligodendrocytes, respectively. For colocalization of p-cPLA₂ and p-ERK1/2, polyclonal anti–p-cPLA₂ antibody (1:100) and monoclonal mouse anti–p-ERK1/2 antibody (1:400) were used. On the following day, the sections were incubated with fluorescein-conjugated goat antirabbit (1:100; ICN Biochemicals, Aurora, OH) and rhodamine-conjugated goat antimouse (1:100; ICN Biochemicals) antibodies. Primary antibody omission controls and cPLA₂ knockout (KO) spinal cord sections were used to further confirm the specificity of the immunofluorescence double labeling, and secondary antibody omission controls were used to determine the degree of autofluorescence. Single-labeling controls were also used to assess any bleed-through. Images were acquired using an FluoView 500 Confocal Laser Scanning Microscope (Olympus America, Melville, NY) with a sequential scanning mode to minimize crosstalk among channels in multicolor images.

**Intraspinal Injection of U0126**

The bilateral microinjections (2 injections/side, 1 μl/injection, total = 4 μl) of vehicle (20% dimethylsulfoxide) or U0126 (0.5 μg/μl) from Phoenix Pharmaceuticals (Burlingame, CA) were made into the spinal cord at 0.6 mm from the midline (0.5 total g/l) from Phoenix Pharmaceuticals (Burlingame, CA) were used to examine colocalization of p-cPLA₂ in neurons, axons, and oligodendrocytes, respectively. For colocalization of p-cPLA₂ and p-ERK1/2, polyclonal anti–p-cPLA₂ antibody (1:100) and monoclonal mouse anti–p-ERK1/2 antibody (1:400) were used. On the following day, the sections were incubated with fluorescein-conjugated goat antirabbit (1:100; ICN Biochemicals, Aurora, OH) and rhodamine-conjugated goat antimouse (1:100; ICN Biochemicals) antibodies. Primary antibody omission controls and cPLA₂ knockout (KO) spinal cord sections were used to further confirm the specificity of the immunofluorescence double labeling, and secondary antibody omission controls were used to determine the degree of autofluorescence. Single-labeling controls were also used to assess any bleed-through. Images were acquired using an FluoView 500 Confocal Laser Scanning Microscope (Olympus America, Melville, NY) with a sequential scanning mode to minimize crosstalk among channels in multicolor images.

**Reverse Transcription PCR**

Reverse transcription PCR (RT-PCR) was performed with the Access RT-PCR system (Promega, Madison, WI) according to the manufacturer’s instructions. Sense primer 5’-AAG GCC AAG TGA CAC CAG CC-3’ and antisense primer 5’-GAA ACA GAG CAA CGA GAT GGG-3’ were used to yield a 452–base pair cPLA₂ product. Primers for cyclophilin were used for control.

**Spinal Cord Neuronal Culture, Cell Treatment, and Viability Assessment**

Cells were obtained from embryonic day 14 rat spinal cords by gentle trituration according to our previously described protocol. Under this culture condition, a purity of >85% spinal cord neuronal population was obtained at the seventh day in vitro. Cultures were then treated with the designated concentration of ceramide-1-phosphate (C-1-P), A23187, and/or AAMCCOF3 for the designated time. The cultures were maintained for an additional 24 hours, and the culture medium of each well was removed for lactate dehydrogenase release assay using a CytoTox 96 Non-Radioactive Cytotoxicity Assay kit (Promega). In a subset of cultures, spinal cord neurons were treated using terminal deoxynucleotide transferase–mediated deoxyuridine triphosphate nick-end labeling (TUNEL) and immunofluorescent double labeling as well as Western blot.

**TUNEL Assay**

Apoptotic spinal cord neurons were detected by TUNEL and the immunostaining of the neuronal marker neurofilament protein (NFP) using an in situ cell death detection kit (TMR red; Roche Applied Science, Mannheim, Germany), according to the manufacturer’s instructions. Primary rabbit polyclonal NFP antibody (1:400; Sigma) and secondary fluorescein-conjugated goat antirabbit antibody (1:100; ICN Biochemicals) were used.

**Behavioral Assessments**

All behavioral tests were blindly performed. The Basso Mouse Scale (BMS) locomotor test was performed weekly up to 6 weeks post-SCI by 2 observers lacking knowledge of the experimental groups according to a method published previously. Briefly, mice were placed in an open field (diameter = 42 in) and observed for 4 minutes by 2 trained observers. The scores were on a scale of 0 to 9 (9 = normal locomotion; 0 = complete hind limb paralysis), which is based on hind limb movements made in an open field including hind limb joint movement, weight support, plantar stepping, coordination, paw position, and trunk and tail control.

Footprint analysis was used to examine the stepping patterns of the mice. The animals’ hind paws were inked with blue dye, and the animals were required to traverse a narrow runway (100 × 4 × 4 cm) lined with white paper. Only mice with frequently or consistently plantar stepping were tested (BMS score ≥ 5 for both hind limbs). Three separate traverses of the track (trials) were recorded per testing session. A minimum of 5 consecutive footprints were assessed to determine values for the trial, and the 3 trials were averaged to obtain the values for each parameter assessed per session. Six parameters including toe spread, paw length, paw rotation, stride length, stride width, and intermediary toes were analyzed.

Beam walking was evaluated in mice that showed frequent or consistent plantar stepping using a graded series of rough metal beams (24 cm long) of various widths: 0.4, 0.8, 1.2, 1.6, and 2.0 cm. The narrowest beam each mouse could traverse was recorded, along with the number of errors across 4 trials. Hind paw and whole body falls were both counted as errors. If an animal could not maintain placement of its hind paws on the beam, or if the animal was dragging its hindquarters across the beam, this was considered failing the task, and no score was recorded for the animal. The scoring was based on beam size and the number of errors. Data for this test were obtained by taking the average of 4 trials per beam per animal.

**Histological Assessments**

Spinal cord segments containing the epicenter were isolated from each animal, embedded, and cut into 25 μm-thick serial
sections (250μm apart and spanning the entire rostrocaudal extent of the lesion). One set of the sections was stained for myelin with Luxol fast blue, and the other was counterstained with cresyl violet–eosin. The lesion and spared white matter area of the injured cord were visualized, outlined, and quantified using an Olympus BX60 microscope equipped with a Neulucida system (MicroBrightField, Colchester, VT). An unbiased estimation of the percentage of spared tissue and lesion volume were calculated using the Cavalieri method.\(^{16}\)

**Measurement of Na\(^+\), K\(^+\)-Adenosine Triphosphatase Activity**

Membrane-bound Na\(^+\), K\(^+\)-adenosine triphosphatase (ATPase) was isolated as described previously with minor modification.\(^{32,33}\) Spinal cord tissue was homogenized in ice-cold isolation buffer (30mM histidine, 0.32M sucrose, and 1mM ethylenediaminetetraacetic acid [EDTA], pH 7.4). The homogenates were centrifuged at 1,000 \(\times\) g for 20 minutes at 4°C. The pellet was discarded, and the supernatant was centrifuged at 14,000 \(\times\) g for 60 minutes at 4°C. The pellet was resuspended in isolation buffer and stored at −80°C. The enzyme activity was assayed by the method previously described.\(^{34}\)

Briefly, the Na\(^+\), K\(^+\)-ATPase activity was assayed in an incubation medium consisting of 30mM histidine, 130mM NaCl, 20mM KCl, 5mM MgCl\(_2\), and 6.5μg membrane protein with or without 1mM ouabain. Inorganic phosphate was measured by the method of Fiske and Subbarow.\(^{34}\)

**Tissue of Prostaglandin E\(_2\) Determination**

Tissue levels of prostaglandin E\(_2\) (PGE\(_2\)) in the spinal cord were assayed using enzyme immunoassay (EIA; Prostaglandin E\(_2\), EIA kit; Cayman Chemical). Spinal cord segments containing the epicenter were removed and were homogenized in ice-cold lysis buffer (0.1M phosphate, pH 7.4, 1mM EDTA, 10μM indomethacin; Cayman Chemical) using a tube pestle. Acetone was added (2 \(\times\) sample volume), and samples were centrifuged at 1,500 \(\times\) g for 10 minutes. The supernatants were then stored at −80°C, and assay followed the manufacturer’s instructions.

**Myeloperoxidase Activity Assay**

Myeloperoxidase (MPO) activity, an indicator of polymorphonuclear leukocyte accumulation, was performed as previously described.\(^{16}\) Briefly, the injured spinal cord segment (10mm) was removed and homogenized. The supernant, after centrifugation at 14,000 \(\times\) g for 25 minutes, was assayed for MPO activity.

**PLA\(_2\) Activity Assay**

A 10mm spinal cord segment containing the injury epicenter was dissected after intracardial perfusion of the mice with 10ml of saline under anesthesia. The cord segment was homogenized in 0.4ml of 50mM hydroxyethylpiperazine ethanesulfonic acid (HEPES), pH 7.4, containing 1mM EDTA and centrifuged at 10,000 \(\times\) g for 15 minutes at 4°C. Supernatant was removed, and PLA\(_2\) activity was measured in the presence and absence of calcium according to the protocol in the PLA\(_2\) Assay Kits with minor modification (Cayman Chemical Company). Briefly, Total PLA\(_2\) activity was measured by incubating the samples with a substrate, arachidonoyl thio-PC, for 1 hour at room temperature in the assay buffer. The reactions were stopped by dithiobis nitrobenzoic acid (DTNB)/ethyleneglycoltetraacetic acid (EGTA) for 5 minutes, and the absorbances were determined at 405nm using a VICTOR\(_3\) V 1420 Multilabel Counter (PerkinElmer Wallac Oy, Turku, Finland). To detect the activity of iPLA\(_2\), the assay buffer was modified to Ca\(^{2+}\)-free buffer (4mM EGTA, 160mM HEPES, pH 7.4, 300mM NaCl, 8mM Triton X-100, 60% glycerol, 2mg/ml of bovine serum albumin) as described previously.\(^{35}\) The iPLA\(_2\) activity was assayed by incubating the samples with the substrate, arachidonyl thio-PC, for 1 hour at room temperature in the modified Ca\(^{2+}\)-free buffer. The reaction was stopped by addition of DTNB/EGTA for 5 minutes, and the absorbance was determined at 405nm using the PerkinElmer VICTOR\(_3\) V 1420 Multilabel Counter. Activity of Ca\(^{2+}\)-dependent PLA\(_2\) = total PLA\(_2\) activity – iPLA\(_2\) activity. Because C57BL/6 mice have a naturally occurring null mutation of the major form of sPLA\(_2\),\(^{36}\) Ca\(^{2+}\)-dependent PLA\(_2\) activity reflects cPLA\(_2\) activity.

**Statistical Analysis**

All statistical analyses were performed using Prism software (version 6.00; GraphPad, La Jolla, CA) except for number of neurons. All data are presented as mean ± standard error of the mean values, and were analyzed by Student t tests or analysis of variance (ANOVA; 1-way, 2-way, or repeated measures as appropriate) followed by post hoc Dunnett or Tukey multiple comparison test. For the number of neurons, nonparametric repeated measures ANOVA was performed using SAS software (SAS Institute, Cary, NC), where the ranks of the outcome variable were used as the dependent variable, with group (WT and cPLA\(_2\) KO) and location as independent variables, including interaction. Potential correlation was adjusted for measurements obtained from the same animals. Bonferroni adjustment was used in post hoc analysis comparing WT to cPLA\(_2\) KO within each specific location. A \(p\) value of \(<0.05\) was considered statistically significant.

**Results**

**cPLA\(_2\) Activation in the Injured Spinal Cord following SCI**

Mouse SCI models are being increasingly used because transgenic and KO mice are available for the study of cellular mechanisms. We found that cPLA\(_2\) expression in mice significantly increased after SCI, peaked at 7 days post-SCI, and remained highly expressed at 14 days (Fig 1). This profile of cPLA\(_2\) expression is consistent with our previous observation in rats.\(^{15}\) Because cPLA\(_2\) activation requires phosphorylation of cPLA\(_2\) by MAPK,\(^{17}\) we also examined p-cPLA\(_2\) as an indicator of cPLA\(_2\) activation. The p-cPLA\(_2\) expression was also significantly increased after SCI, with a similar expression profile to that of cPLA\(_2\). The expression of specific p-cPLA\(_2\) (ie, ratio of p-cPLA\(_2\)/cPLA\(_2\)) was also significantly increased...
at as early as 1 day and reached the highest at 14 days after SCI. Immunohistochemistry further revealed that the expression of p-cPLA2 increased as early as 8 hours post-SCI. Extensive p-cPLA2 expression was found in axons, particularly in those that underwent degeneration (see Fig 1E, right column, white matter) and in neurons and glial cells (see Fig 1E, left column, gray matter) between 1 and 7 days post-SCI. The p-cPLA2 expression was found not only in regions close to the injury (1.5mm) but also in areas distant from it (5mm). Immunofluorescence double labeling further confirmed that p-cPLA2 was expressed in neurons, swollen axons, and oligodendrocytes (Fig 2). The c-PLA2 expression and activation in mice was also confirmed in rats by immunohistochemistry (data not shown), protein,15 and mRNA (Fig 3) analyses.

**ERK1/2 Signaling Pathway Mediates SCI-Induced cPLA2 Phosphorylation In Vivo**

To assess the mechanism of cPLA2 activation, we asked whether ERK1/2 signaling pathway plays a role in mediating cPLA2 phosphorylation. Our results showed colocalization of p-cPLA2 and p-ERK1/2 in neurons, degenerated axons, and glial cells at 24 hours after SCI (Fig 4). In sham-operated controls, p-ERK1/2 immunoreactivity (IR) at a very low level was observed in morphologically characteristic neurons; however, no p-cPLA2 IR was detected in these neurons or any other cells. Two-way ANOVA analysis showed that there were statistically significant effects of treatment group ($F_{2,18} = 137.6, \ p < 0.0001$), ERK1/2 ($F_{1,18} = 83.38, \ p < 0.0001$), and the interaction of treatment group and ERK1/2 ($F_{2,18} = 11.86, \ p = 0.0005$). Western blot analysis revealed that expressions of p-p44 (p-ERK1) and p-p42 (p-ERK2) were increased by 581.11% and 514.92%, respectively ($p < 0.01$) at 24 hours after SCI. Importantly, the increased expression of p-p44 and p-p42 were reversed by 59.82% ($p < 0.01$) and 41.61% ($p < 0.05$), respectively, by U0126, an ERK1/2 inhibitor. In the same animal model, SCI increased p-cPLA2 expression by 298.85% ($p < 0.01$). Interestingly, administration of the ERK1/2 inhibitor U0126 inhibited p-cPLA2 expression by 42.84% ($p < 0.01$).

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**FIGURE 1: Cytosolic phospholipase A2 (cPLA2) activation following spinal cord injury.** (A) Representative time courses of phosphorylated cPLA2 (p-cPLA2), cPLA2, and β-tubulin expression. (B) Compiled results in a bar graph for the ratio of p-cPLA2/β-tubulin expression. (C) Compiled results in a bar graph for the ratio of cPLA2/β-tubulin expression. (D) Compiled results in a bar graph for the ratio of p-cPLA2/cPLA2 expression. **p < 0.01 versus sham (1-way analysis of variance, Dunnett post hoc test, n = 6 mice/group). Error bars represent mean ± standard error of the mean. (E) Immunohistochemistry of mouse spinal cord sections at 1.5 and 5mm caudal to the injury epicenter shows increased p-cPLA2 expression in neurons (arrows), axons (open arrows), and glial cells (arrowheads). GM = gray matter; KO = knockout; WM = white matter. Bars = 40μm.
Activation of cPLA2 Induces Spinal Cord Neuronal Death In Vitro

Because we observed that cPLA2 activation was induced following SCI, the next question would be: could cPLA2 activation induce spinal cord neuronal death? To address this issue, we first examined the effects of C-1-P and A23187 on spinal cord neuronal death in vitro. C-1-P is a direct activator of cPLA2 through interaction with the CaLB/C2 domain.37 The calcium ionophore A23187 is an indirect activator of cPLA2 through elevations of intracellular free calcium.38 Our results showed that both C-1-P and A23187 induced cultured spinal neuronal death in a dose-dependent manner (Fig 5). Importantly, such C-1-P- or A23187-induced neuronal death could be significantly reversed by AACOCF3, a cPLA2 inhibitor. TUNEL staining revealed that C-1-P–induced spinal cord neuronal death took the form of apoptosis. Western blot analysis further confirmed that C-1-P–induced death of spinal cord neurons expressed apoptotic markers active caspase-3 ($p < 0.05$) and active PARP-1 ($p < 0.01$).

**Inhibition of cPLA2 Reduces SCI-Induced Tissue Damage and Improves Behavioral Recovery**

To further assess whether activation of cPLA2 is sufficient to mediate the secondary SCI, we tested whether blocking cPLA2 activation with the cPLA2 inhibitor AACOCF3 would reduce injury-induced increases in eicosanoids (downstream metabolites of cPLA2), inflammation, and tissue damage and in turn enhance recovery after a contusive SCI in mice. AACOCF3 is a potent and selective inhibitor of cPLA2. This inhibitor shows slow tight binding to cPLA2 in the presence of Ca$^{2+}$ and forms a covalent bond with a serine residue in the active site of the enzyme.39,40 This inhibitor is about 500-fold more potent at inhibiting cPLA2 than sPLA2,39 and may also be a weak inhibitor of iPLA2.41–43 AACOCF3 was delivered intravenously (50 µl of 4mM) at 30 minutes in C57BL/6 mice after the injury followed by intraperitoneal injections of the compound (200 µl of 4mM) every other day up to 2 weeks postinjury. Certain mouse strains, such as C57BL/6, 129/Sv, and B10.rIII, have a naturally occurring null mutation of the major form of sPLA2.36 Therefore, C57BL/6 mice in this study were deficient in sPLA2. To confirm cPLA2 inhibition in AACOCF3-treated mice, we measured PLA2 activity and its metabolite PGE2 at 24 hours after SCI. Our results showed that AACOCF3 treatment significantly reduced cPLA2 activity and PGE2 production by 42.9% ($p < 0.01$) and 35.1% ($p < 0.01$) after SCI (Fig 6). However, SCI-induced iPLA2 activation was not affected significantly by AACOCF3 ($p > 0.05$). AACOCF3

**FIGURE 2:** Cellular localization of phosphorylated cytosolic phospholipase A2 (p-cPLA2) expression in the mouse spinal cord after spinal cord injury (SCI). (A–C) p-cPLA2 immunoreactivity (IR) was localized in neurons indicated by NeuN IR (arrows). (D–F) p-cPLA2 IR was localized in a subpopulation of axons that show morphologically degenerating changes such as swelling (arrows). Axons were SMI-31 immunoreactive. (G–I) p-cPLA2 IR was localized in oligodendrocytes indicated by CC1 IR (arrows). (J–L) In spinal cord sections of cPLA2 knockout (KO) mice after SCI, p-cPLA2 IR was not observed, confirming the specificity of p-cPLA2 antibody (J). Bars = 20 µm.

**FIGURE 3:** Expression of cytosolic phospholipase A2 (cPLA2) mRNA in the rat injured spinal cord at 7 days after spinal cord injury (SCI). Reverse transcription polymerase chain reaction was performed using specific primers designed for rat cPLA2 and cyclophilin. Bar graph indicates the mean ± standard error of the mean; $n = 6$ mice/group, **$p < 0.01$ versus sham-operated group (Student t test).
administration also restored Na⁺, K⁺-ATPase activity (a marker for membrane integrity or damage) by 38.1% \((p < 0.05)\), and reduced SCI-induced MPO activity (a marker for neutrophil infiltration) by 67.1% \((p < 0.01)\).

To determine whether inhibition of cPLA₂ promotes functional recovery, an array of behavior tests were performed on consecutive days following SCI to evaluate motor and sensorimotor functions. There were statistically significant effects of treatment group \((F_{2,21} = 40.17, p < 0.0001)\), test day \((F_{7,147} = 55.32, p < 0.0001)\), and the interaction of treatment group and test day \((F_{14,147} = 15.93, p < 0.0001); \text{ repeated measures ANOVA}\) for BMS for open field locomotion. AACOCF₃ treatments significantly improved BMS scores for up to 6 weeks (Fig 7; \(p < 0.05–0.01\)). Repeated measures ANOVA also revealed that there were statistically significant effects of AACOCF₃ treatment \((F_{2,21} = 30.19, p < 0.0001)\) for beam walking scores at 4 and 6 weeks post-SCI. Footprint analysis showed that administration of AACOCF₃ significantly improved the stride length \((p < 0.05)\), paw rotation angle \((p < 0.01)\), and intermediary toes \((p < 0.01)\) at 5 weeks post-SCI.

Because we showed that administration of AACOCF₃ significantly improved behavioral recovery after SCI, we next examined whether such a treatment also would result in tissue protection in vivo. To ensure that the entire rostrocaudal expansion of the lesion was examined, a 1.2cm-long cord segment was serially sectioned. Measurements of percentage total lesion volume, lesion area, and white matter sparing area were made from cresyl violet-stained and eosin-stained transverse sections spanning the entire lesion. Comparison of the lesion area at the injury epicenter demonstrated that AACOCF₃ treatments resulted in a significant reduction of lesion area by 28.1% (see Fig 7; \(p < 0.05\)). Such reduction in lesion area was accompanied by a corresponding increase in the area of white matter sparing by 47.8% \((p < 0.05)\) at 6 weeks post-SCI. In addition, Luxol fast blue staining showed that the AACOCF₃ treatment resulted in a corresponding increase in myelin sparing by 35.5% \((p < 0.01)\). Finally, stereological assessments of the lesion volume showed that AACOCF₃...
treatment resulted in a significant reduction in the percentage total lesion volume by 34.3% ($p < 0.01$).

**Genetic Ablation of cPLA$_2$ Reduces Cell Loss and Tissue Damage, and Improves Behavioral Recovery after SCI**

To definitively determine the role of cPLA$_2$ following SCI, we used cPLA$_2^{-/-}$ mice and compared them with WT littermates (cPLA$_2^{+/+}$). All mice were on a C57/BL6 background and deficient in sPLA$_2$. cPLA$_2^{-/-}$ mice develop normally, gain weight at a rate equal to that of WT animals, and have a lifespan of >1 year. We found that at 24 hours after SCI, there was a marked loss of ventral horn neurons at and near the site of injury in the WT littermates (Fig 8). In contrast, in cPLA$_2$ KO mice, the SCI-induced neuronal loss was significantly reduced at 24 hours postinjury. To test further whether cPLA$_2$ ablation resulted in neuroprotection against cell death, we performed experiments to evaluate the effects of cPLA$_2$...
Outcomes in cPLA2 KO mice and their WT littermates improved in the cPLA2 after SCI. The BMS locomotor scores were significantly increased in the cPLA2 KO mice compared to their WT littermates (see Fig 9). Such reduction in lesion area was accompanied by a corresponding increase in the area of white matter sparing by 56.2% (p < 0.01) in the cPLA2 KO mice. Luxol fast blue staining also showed that there was a corresponding increase in myelin sparing by 33.2% (p < 0.01) in the cPLA2 KO mice. Stereological assessments showed that there was a significant reduction in the percentage total lesion volume by 31.1% (p < 0.01) in the cPLA2 KO mice. Thus, genetic ablation of cPLA2 not only confirmed the previous observation of pharmacological inhibition of cPLA2 by AACOCF3, but also clearly indicated that cPLA2 could be an attractive target for intervention following SCI.

**Discussion**

The goal of this study was to determine whether targeting cPLA2 could be an effective strategy for functional repair after SCI. Our study showed that SCI induced an elevation of cPLA2 expression and activation. The elevated cPLA2 was localized mainly in neurons and oligodendrocytes. We also showed that the SCI-induced cPLA2 activation is mediated, at least in part, by ERK signal, revealing a molecular mechanism of cPLA2 activation. In vitro studies demonstrated that cPLA2 activation induced cultured spinal cord neuronal death. Most importantly, both pharmacological blockade and genetic deletion of cPLA2 significantly reduced inflammation, cell death, and tissue damage, as well as improved behavioral recovery after SCI. These findings collectively suggest that modulation of cPLA2 could represent a new therapeutic strategy for treatment of SCI.}

SCI significantly induced cPLA2 activation, which was observed as early as 8 hours postinjury and peaked at 7 days postinjury. The activated cPLA2 was mainly localized in neurons and oligodendrocytes. The expression of cPLA2 mRNA was increased in the injured cord, which correlated well with increased expression of cPLA2 protein. Several earlier investigators found that AA and eicosanoids, metabolites of cPLA2, increased within 30 minutes after SCI.45,46 Others reported that increased eicosanoids were persistent at least for 3 days (the longest time point studied) after SCI.47 Furthermore, Demediuk et al reported that induced concentrations of free fatty acid quickly increased after SCI, peaked at 3 days, and remained significantly high at 7 days after SCI.48 The induction profiles of these PLA2 metabolites are similar to that of cPLA2 after SCI in the present study. These results indicate that a prolonged effect of cPLA2 exists after SCI, which suggests that there may be a unique role.
therapeutic window for intervention. The finding that cPLA₂ was mainly localized in neurons and oligodendrocytes is particularly interesting, because these 2 cell types not only play important roles in normal central nervous system (CNS) function but also are the most vulnerable cell types to injuries as compared to other CNS cell types such as astrocytes and microglia.⁴⁹

Although cPLA₂ activation and expression were significantly increased after SCI, the mechanism(s) by which they were increased remains to be determined. Our in
vivo experiments revealed that ERK1/2 signaling pathway mediated SCI-induced cPLA2 activation. Our previous in vitro experiments\(^7\) also showed that ERK1/2 signaling pathway mediated cPLA2 phosphorylation, induced by glutamate and \(\text{H}_2\text{O}_2\), two important injury mediators of secondary SCI\(^5\). We and others have reported that cPLA2 is induced by several toxic factors that are generated in the injured cord, including inflammatory cytokines,\(^8,17\) free radicals,\(^7-9,11\) and excitatory amino acids.\(^7,8,10\) Therefore, cPLA2 may serve as a central or convergence molecule that mediates multiple key mechanisms of secondary injury, making it an attractive therapeutic target to improve tissue protection and function recovery.

Our results clearly demonstrated that cPLA2 activation induced spinal cord neuronal death. This was in agreement with our previous finding that cPLA2 activation mediated cultured spinal cord neuronal death induced by glutamate and \(\text{H}_2\text{O}_2\).\(^7\) Apoptosis has been considered a key mechanism of cell death following SCI.\(^49,50\) Caspase-3 plays a central role in the execution phase of apoptosis and is responsible for the cleavage of proteins such as the nuclear enzyme PARP. Our results showed that cPLA2 activation induced the expression of active caspase-3 and PARP-1. TUNEL staining further confirmed that cPLA2 activation induced neuronal apoptosis, which was supported by cPLA2-mediated neural apoptosis induced by \(\text{A}\beta\).\(^51\) These results suggest that cPLA2 activation induced neuronal death through apoptosis, at least in part.

A significant finding of this study is that pharmacological blockade of cPLA2 with AACOCF3 inhibited inflammation and membrane injury, reduced tissue damage, and improved behavioral recovery in C57BL/6 mice after SCI. Notably, the cPLA2 inhibitor was administered after trauma. Our results showed a long beneficial effect of targeting cPLA2 on anatomical and functional recoveries. In agreement with our observation, several studies have reported a detrimental effect of cPLA2 in other...
CNS diseases such as ischemia, experimental autoimmune encephalomyelitis, and Alzheimer disease. In contrast, there is a recent report showing that activation of cPLA2 is beneficial. In that study, both BALB/c mice treated with AX059, a cPLA2 inhibitor, and cPLA2-null BALB/c mice displayed greater neuronal and myelin loss after SCI. The contrary results between this mouse strain and others remain unclear, and they may be related to different mouse backgrounds (C57BL/6 vs BALB/c) and inhibitors (AACOCF3 vs AX059) that were used. It has been reported that different strains of mice display distinctly different responses to trauma injury, including inflammation, histology, and behavioral recovery. For example, post-traumatic inflammation was markedly reduced in BALB/c mice compared with C57BL/6 mice. After SCI, a densely packed cellular matrix fills necrotic cavities. The magnitude of this response was greatest for C57BL/6 mice and least for BALB/c mice. Kipnis and colleagues also showed that BALB/c mice exhibited a T-cell–dependent neuroprotective response, whereas trauma- or glutamate-mediated neuronal cell loss was enhanced in C57BL/6 mice. These results suggest that genetic differences may confer different responses to traumatic injury, which may modify the secondary injury processes after SCI. Thus, the contrary results from C57BL/6 and BALB/c mice may be related to genetic differences including sPLA2.

We previously demonstrated increased sPLA2 expression following SCI. Injection of sPLA2 into the normal spinal cord resulted in tissue damage, demyelination, and behavioral impairment in vivo. Importantly, administration of a sPLA2 inhibitor GK511 in BALB/c mice reduced tissue damage and improved behavioral recovery after SCI. In the current study, sPLA2 action...
was excluded in both sham and SCI groups, because C57BL/6 mice have a naturally occurring null mutation of sPLA2. iPLA2 is generally considered to be a housekeeping enzyme for the maintenance of membrane phospholipids. Lopez-Vales et al reported that iPLA2 was upregulated after SCI and was expressed mainly in oligodendrocytes. Using FKGK11, a potent and highly selective iPLA2 inhibitor in BALB/c mice with SCI, they further showed that iPLA2 appeared to play a minor detrimental role in SCI. Although AACOCF3 has been reported to be a weak inhibitor of iPLA2, our results showed that there was no significant difference of iPLA2 activity between the SCI and AACOCF3-treated groups. These results suggest that AACOCF3 exerts neuroprotection mainly via inhibition of cPLA2.

A definitive finding of the present study is that genetic deletion of cPLA2 resulted in neuroprotection and behavioral recovery following SCI. Genetic deletion of cPLA2 also inhibited the expression of active caspase-3 after SCI, suggesting that cPLA2 activation mediates neural apoptosis. Our observation is supported by several other reports that cPLA2−/− mice show protection in ischemic brain damage, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity, and neurodegeneration. cPLA2−/− mice also show significant reductions in AA release and eicosanoid production in response to a variety of stimuli. cPLA2 may contribute to injury by a direct effect on cell membranes and/or indirectly through generation of its metabolites. Our results suggest that cPLA2 contributes to the pathogenesis of SCI and that targeting cPLA2 could be an effective therapeutic strategy for intervention after SCI.

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Authorship
N.-K.L. designed experiments, performed experiments, analyzed data, and wrote the paper. L.-X.D. and J.-G.H. performed in vitro experiments. Y.P.Z., Q.-B.L., and X.-F.W. performed in vivo experiments. E.O. performed some behavioral assessments. J.V.B. provided cPLA2 knockout mice and edited the manuscript. C.B.S. designed and edited the manuscript. X.-M.X. designed experiments, reviewed the data, and wrote the manuscript.

Potential Conflicts of Interest
Nothing to report.

References


