Influence of sulfate reduction rates on the Phanerozoic sulfur isotope record

William D. Leavitt¹, I. Itay Halevy², Alexander S. Bradley³, and David T. Johnston⁴,¹

¹Department of Earth and Planetary Sciences, Harvard University, Cambridge, MA 02138; ²Department of Environmental Sciences and Energy Research, Weizmann Institute of Science, Rehovot 76100, Israel; and ³Department of Earth and Planetary Sciences, Washington University in St. Louis, St. Louis, MO 63130

Edited by Mark H. Thiemens, University of California San Diego, La Jolla, CA, and approved May 8, 2013 (received for review October 30, 2012)

Phanerozoic levels of atmospheric oxygen relate to the burial histories of organic carbon and pyrite sulfur. The sulfur cycle remains poorly constrained, however, leading to concomitant uncertainties in O₂ budgets. Here we present experiments linking the magnitude of fractionations of the multiple sulfur isotopes to the rate of microbial sulfate reduction. The data demonstrate that such fractionations are controlled by the availability of electron donor (organic matter), rather than by the concentration of electron acceptor (sulfate), an environmental constraint that varies among sedimentary burial environments. By coupling these results with a sediment biogeochemical model of pyrite burial, we find a strong relationship between observed sulfur isotope fractionations over the last 200 Ma and the areal extent of shallow seafloor environments. We interpret this as a global dependency of the rate of microbial sulfate reduction on the availability of organic-rich sea-floor settings. However, fractionation during the early/mid-Paleozoic fails to correlate with shelf area. We suggest that this decoupling reflects a shallower paleo-redox boundary, primarily confined to the water column in the early Phanerozoic. The transition between these two states begins during the Carboniferous and concludes approximately 1 Ma around the Triassic–Jurassic boundary, indicating a prolonged response to a Carboniferous rise in O₂. Together, these results lay the foundation for decoupling changes in sulfate reduction rates from the global average record of pyrite burial, highlighting how the local nature of sedimentary processes affects global records. This distinction greatly refines our understanding of the S cycle and its relationship to the history of atmospheric oxygen.

Phanerozoic oxygen | sulfate-reducing bacteria

The marine sedimentary sulfur isotope record encodes information on the chemical and biological composition of Earth’s ancient oceans and atmosphere (1, 2). However, our interpretation of the isotopic composition of S-bearing minerals is only as robust as our understanding of the mechanisms that impart a fractionation. Fortunately, decades of research identify microbial sulfate reduction (MSR) as the key catalyst of the marine S cycle, both setting the S cycle in motion and dominating the mass-dependent fractionation preserved within the geological record (1, 3, 4). Despite the large range of S-isotope variability observed in biological studies (4–6), attempts to calibrate the fractionations associated with MSR are less mechanistically definitive (7, 8) than analogous processes influencing the carbon cycle (9, 10). What is required is a means to predict S isotope signatures as a function of the physiological response to environmental conditions (e.g., reduction-oxidation potential).

Microbial sulfate reduction couples the oxidation of organic matter or molecular hydrogen to the production of sulfide, setting in motion a cascade of reactions that come to define the biogeochemical S cycle. In modern marine sediments, sulfide is most commonly shuttled back toward sulfate through oxidation reactions (biotic and abiotic) or scavenged by iron and buried as pyrite (11). It is the balance of oxidation reactions and pyrite burial that influences geological isotope records, which in turn carry historical information on the oxidation state of Earth’s biosphere. Such records generally are thought to indicate that oxidant availability has increased with each passing geologic eon (12). Although playing prominent roles in sedimentary redox cycles (13), oxidation reactions carry only modest S isotopic fractionations (14, 15). In typical modern marine sediments, the oxidative region of aerobic organic carbon remineralization is separated from the zone of sulfate reduction (where MSR takes place) by an intermediate layer in which both sulfide oxidation and sulfur disproportionation occur (16, 17). Despite sulfur recycling across that boundary layer, the sulfur that is eventually buried as pyrite predominately reflects the isotopic fractionation associated with MSR (18).

Numerous studies show a correlation between microbial sulfate reduction rate (mSRR; SI Text) and the expressed magnitudes of the MSR S isotopic fractionations (19–22). The mSRR depends on a suite of physiological controls (i.e., metabolic enzymes) (23) having variable efficiencies in response to environmental conditions (e.g., nutrient availability, redox potential, etc.; refs. 8 and 24). Such reactions can be presumed to be first-order with respect to a limiting reactant (SI Text and refs. 25 and 26). In the case of MSR, either the electron acceptor (sulfate; ref. 7) or the electron donor (generally, organic carbon (OC); refs. 19, 20, 27, and 28) plays this role. It has been hypothesized that since the Archean–Proterozoic boundary sulfur (oxidant) limitation has not occurred in the water column, but rather has been restricted to significantly below the sediment–water interface. This is consistent with estimates through the Proterozoic and Phanerozoic Eons (29) that suggest seawater sulfate concentrations in excess of the physiologically inferred minimum threshold for MSR (30). It follows that the quantity and quality of OC delivery to the zone of sulfate reduction most often dictates sulfide reduction rates where sulfate is abundant, and consequently play the larger role in determining the sulfur isotope record.

The most direct experimental means of studying the metabolic rate of a bacterial population is to maintain the culture in a chemostat (chemical environment in static). In a chemostat, the input

Author contributions: W.D.L. and D.T.J. designed research; W.D.L. and I.H. performed research; W.D.L., I.H., A.S.B., and D.T.J. analyzed data; and W.D.L., I.H., A.S.B., and D.T.J. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

See Commentary on page 11217.

¹To whom correspondence may be addressed. E-mail: wleavitt@fas.harvard.edu or johnston@eps.harvard.edu.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1218874110/-/DCSupplemental.

²Variability in \( \Delta S \) of a measured pool, \( y \), is tracked through \( \delta^{34}S_y = [(\Delta^{32}S/\Delta^{34}S)_{sample} - 1] \times 1,000 \), where \( x = 3, 4, 6 \), and \( y \) is a distinct S-bearing species or operationally defined pool. The difference between two pools \( (y = A \ or \ B, \ e.g., \ sulfate \ and \ sulfide \ or \ sedimentary \ sulfate \ and \ pyrite) \) is calculated rigorously (i.e., not a simple arithmetic difference), as must be done in studies tracking both major and minor isotope variability: \( \delta^{34}S_{A,B} = \delta^{34}S_{A} - \delta^{34}S_{B} \approx 1 \times 10^{0} \), where \( \delta^{34}S_{A,B} = [(\Delta^{32}S/\Delta^{34}S)_{A} - (\Delta^{32}S/\Delta^{34}S)_{B}] \). Variability in \( \delta^{32}S \) is tracked through \( \delta^{32}S_{\lambda} = \ln \left[ \ln \left( 1 + \delta^{32}S_{\lambda} \right) / \ln \left( 1 + \delta^{34}S_{\lambda} \right) \right] / \left[ \ln \left( 1 + \delta^{34}S_{\lambda} \right) \right] \). Qualitatively, \( \lambda \) describes the slope of a line connecting two points on a \( \delta^{32}S \) vs. \( \delta^{34}S \) plot.
concentration of the limiting substrate (e.g., micronutrient, electron donor, or electron acceptor) dictates the biomass yield (i.e., new biomass per mass substrate used), whereas turnover time of the reactor [dilution rate (D), time$^{-1}$] dictates growth rate (SI Text and ref. 25). It follows that the mSRR (specifically, the rate of reduction per unit biomass) scales with the availability of the limiting nutrient, and thus with D (19, 25). Limited previous experimental work with open and semiopen experimental systems hints that mSRR inversely correlates with fractionation of the major S isotopes ($^{34}$S/$^{32}$S) (19, 31, 32)—a prediction that is reinforced by measurement of the same relationship in modern marine sediments (17, 22). However, the limited range of mSRRs previously explored does not adequately capture the vast range of rates inferred from marine sediments (33–35). In this work we present an empirical calibration of a ~50-fold change in mSRR, nearly doubling the previous experimental ranges. We also target the minor isotope, $^{33}$S, in addition to targeting $^{34}$S fractionations; this supplies another dimension for interpreting the geologic record. We then apply this calibration to Phanerozoic isotope records to reveal how secular changes in S isotopic fractionation reflect a temporal history of paleoredox conditions. This enables us to reassess the burial of pyrite and associated changes in environmental conditions across the Phanerozoic Eon.

Results and Discussion

We conducted a suite of continuous-culture experiments with the model sulfate reducing bacterial strain Desulfovibrio vulgaris Hildenborough (DvH). Strain DvH is among the most well-studied sulfate reducers and is genetically tractable (36, 37). Although DvH is nominally a nonmarine strain, the concentration of sulfate in the chemostat was always near modern marine levels (28 mM) and well above known sulfate affinity constants (38). The sole electron donor (lactate) was always the limiting nutrient in these experiments and was provided at a stoichiometric 1:2 or 1:20 ratio with sulfate (Materials and Methods and SI Text).

These experiments show that MSR isotope fractionation is strongly inversely dependent on electron donor concentration (Fig. 1). In detail, mSRR follows a first-order nonlinear relationship to D (Fig. 1): as we decrease D, lactate availability and mSRR decline, whereas $^{34}$eMSR increases. The values of $^{34}$eMSR and $^{33}$MSR refer to the isotopic differences between the sulfate and sulfide from chemostat experiments (SI Text), whereas $^{34}$Egeo and $^{33}$GEO refer to those differences calculated from sulfate and pyrite sedimentary records as time-binned averages (Dataset S1, Table S1, and Eqs. S4–S7). Further definitions are presented in SI Text. These open-system experiments (Fig. S1 and SI Text) allow for the direct calculation of fractionation factors from the isotopic composition of output sulfate and sulfide (39), without having to apply Rayleigh distillation models addressing closed system (batch) dynamics (40). We demonstrated conservation of elemental and isotopic mass balance throughout the entire experiment (Dataset S1) via the direct measurement of SO$_2$, H$_2$S/HS$, S_2O$_3$, and S$_2$O$_3$, the latter two of which were always below detection (2.5 μM) but have been previously detected in semicontinuous system experiments (31).

Our data complement and significantly extend previous open-system experiments (8, 19, 27, 41, 42). Over a ~50-fold change in mSRR (Fig. 1 and Figs. S2 and S3), $^{34}$eMSR ranged from 10.9 to 54.9‰ (Fig. 1A), whereas $^{33}$MSR varied from 0.5079 to 0.5144 (Fig. 1B). In addition to aiding in our understanding of environmental/geologic records (discussed below), these data allow for an empirically derived fractionation limit for DvH under conditions of electron donor limitation. We apply a nonlinear regression model (43) based on a pseudo-first-order rate expression, appropriate for a reaction in which rate depends on the concentration of a single reactant (Eqs. S8 and S9) (26, 44). The results of the model illustrate the capacity of MSR to exceed the classic 47‰ limit for $^{34}$eMSR (Fig. 2 and ref. 45), here suggesting an upper limit of 56.5 ± 2.6‰. The same nonlinear regression model (Eq. S9) predicts a minor isotope limit (in $^{33}$λ) at 0.5143 ± 0.0004. Given that our experiments were performed with an axenic population of sulfate reducers, we can definitively rule out contributions from intermediate S-oxidizing or disproportionating organisms (46). However, although the magnitudes of these fractionations exceed the canonical MSR limits ($^{34}$e = 47‰), they do not reach the low-temperature equilibrium predictions between sulfate and sulfide in $^{34}$e (at 20 °C, $^{34}$e = 71.3‰; ref. 40).

Decoupling the Isotopic Effects of Sulfate Reduction Rates from Pyrite Burial Records. In modern marine sediments, OC availability is strongly tied to sedimentation rate (47–49). Faster sedimentation (such as in river deltas or on continental shelves) generally leads to more efficient delivery of OC below the depth of oxygen penetration, into the zone of sulfate reduction. (Extreme sedimentation rates deviate from this prediction and can lead to OC dilution.) The absolute flux of OC also scales directly with primary production. The balance of these processes dictates where sulfate reduction occurs with respect to the sediment–water interface, as well as how much sulfate ultimately is reduced (50). Because

Fig. 1. Multiple sulfur isotope fractionation as a function of dilution rate in chemostat experiments. In these experiments, growth and sulfate reduction rates scale inversely with organic carbon (lactate) delivery rate, expressed here as dilution rate (D, hours$^{-1}$). This rate dictates the magnitude of major (A) and minor (B) isotope fractionation between sulfate and sulfide. Error bars in A are smaller than the symbols (2σ = 0.3%), and vertical error bars in B are 2σ SDs. Other studies cited refer to Chambers et al. (19) and Sim et al. (27).

Fig. 2. Demonstration of new fractionation limits under electron-donor limitation. A nonlinear regression model (Eqs. S8 and S9) was used to calculate the empirical fractionation limit in (A) $^{34}$eMSR ($r^2 = 0.9515$) and (B) $^{33}$MSR ($r^2 = 0.8895$), as a function of mSRR (see also Fig. S2). Bold lines indicate the best-fit estimates with 95% confidence intervals as the thin dashed lines ($^{34}$eMSR: 51–62‰, $^{33}$MSR: 0.5135–0.5151). Inset values are the calculated fitting parameters (along with $k_{MSR}$, $0.054 ± 0.0012$, and $k_{MSR}$, $0.0395 ± 0.0158$) with 95% confidence interval.
It is critical to determine how the $m_{\text{SRR}}$-dependent parameter $\lambda_{\text{s}}$, and $\lambda_{\text{fl}}$, tend to decrease and increase (47, 52). To provide a context for evaluating the potential variability in $s_{\text{SRR}}$ through time. Records of $^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$ from Phanerozoic sedimentary basins provide a time-series approximation of the mean isotopic difference between coeval sulfates and sulfides (Fig. 3 and Table S1) (13, 54). In parallel, we use recent estimates for the areal extent of continental shelf and abyssal ocean across Phanerozoic time (Table S1) (55, 56). Assuming that the physical processes dictating OC delivery to sediments today hold throughout the Phanerozoic, shelf environments are generally expected to be OC-rich and support higher sulfate reduction rates than deep-water settings (17, 22). This is reinforced by modern observations, where in shallow water sediments (<1,000 m deep), the area-weighted average $s_{\text{SRR}}$ is 96 $\mu$m SO$_4$~$^{2-}$ cm$^{-2}$ yr$^{-1}$, whereas in deep-water sediments (>1,000 m deep) it is $\sim$1 $\mu$m SO$_4$~$^{2-}$ cm$^{-2}$ yr$^{-1}$.

Some fraction of the resulting sulfide will be buried as pyrite—either its concomitant release of oxidant to the ocean-atmosphere system—it is critical to determine how the $m_{\text{SRR}}$-dependent parameter $\lambda_{\text{s}}$, and $\lambda_{\text{fl}}$, influences the $S$-isotopic records of pyrite relative to coeval sulfate ($^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$).

Analogous to the influence of lactate on $m_{\text{SRR}}$ in our experimental system, the availability of greater quantities of more highly labile OC to the sulfate reduction zone in modern sediments translates to an increase in sulfate reduction rates, with a corresponding decrease in the magnitude of $^{34}_{\text{MSRR}}$ (24, 35, 51). Conversely, decreases in sulfate reduction rates down a sediment column owing to the modeled loss of OC quality and quantity are consistent with observations that $^{34}_{\text{MSRR}}$ increases (47, 52).

Together these ideas describe the concept of net sedimentary sulfate reduction rate ($s_{\text{SRR}}$; moles of S per square meter per year). The $s_{\text{SRR}}$, because it integrates the depth-dependent rate through the entire zone of sulfate reduction, must encompass a depth-weighted average of the variable $^{34}_{\text{MSRR}}$ as well as be tightly linked to the initial OC delivery to the sediment–water interface. Data from the modern sea floor (18, 22, 53) confirm the expected inverse relation between bulk in situ $s_{\text{SRR}}$ and the magnitude of the net sedimentary $S$ isotope fractionation ($^{34}_{\text{GEO}}e$).

The concept of $s_{\text{SRR}}$ also contains another important distinction from $m_{\text{SRR}}$. In most environments, measures of microbial population density are lacking, as are measures of the metabolically available OC compounds. Our experimental system quantifies these parameters and enables direct calculation of $m_{\text{SRR}}$. However, in the environment, the useful metrics are the bulk $s_{\text{SRR}}$ and the global sulfate reduction rate ($g_{\text{SRR}}$, moles of S per year). The $s_{\text{SRR}}$ is converted to $g_{\text{SRR}}$ through an areal normalization and a reoxidation coefficient (1, 17); further details are presented in Table S2. It is $g_{\text{SRR}}$ that is necessary for determining long-term oxidant budgets and global pyrite burial.

We propose that it is possible to decouple the isotopic effects caused by variable $s_{\text{SRR}}$ from the ultimate sedimentary record of $S$ isotopes. For example, under iron-replete conditions, net pyrite burial may increase as a result of more sulfide production and/or an increased sulfide scavenging efficiency by iron (i.e., less oxidation), but at a constant $s_{\text{SRR}}$. Alternatively, pyrite burial could increase in response to a higher $s_{\text{SRR}}$, if it were driven by a higher flux of metabolizable OC to the sedimentary zone of sulfate reduction. This comparison can also be extended to include the roles for weathering and changes in the abundance of shallow-water environments (shelf area). We posit that it is possible to differentiate between these scenarios and determine the ultimate control (tectons vs. OC) on pyrite burial at the global scale: The first case would maintain a constant $^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$, whereas varying $s_{\text{SRR}}$ would be recorded by a change in the fractionation patterns.

**Interpreting Phanerozoic Records.** Phanerozoic compilations provide a context for evaluating the potential variability in $s_{\text{SRR}}$ through time. Records of $^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$ from Phanerozoic sedimentary basins provide a time-series approximation of the mean isotopic difference between coeval sulfates and sulfides (Fig. 3 and Table S1) (13, 54). In parallel, we use recent estimates for the areal extent of continental shelf and abyssal ocean across Phanerozoic time (Table S1) (55, 56). Assuming that the physical processes dictating OC delivery to sediments today hold throughout the Phanerozoic, shelf environments are generally expected to be OC-rich and support higher sulfate reduction rates than deep-water settings (17, 22). This is reinforced by modern observations, where in shallow water sediments (<1,000 m deep), the area-weighted average $s_{\text{SRR}}$ is 96 $\mu$m SO$_4$~$^{2-}$ cm$^{-2}$ yr$^{-1}$, whereas in deep-water sediments (>1,000 m deep) it is $\sim$1 $\mu$m SO$_4$~$^{2-}$ cm$^{-2}$ yr$^{-1}$.

It has long been appreciated that the $^{34}_{\text{GEO}}e$ record through the Phanerozoic carries a definitive structure (13, 57, 58)—the earliest Paleozoic mean $s_{\text{GEO}}$ has mean fractionations of $\sim$30‰ and transitions through the Permain and Triassic to a Meso-Cenozoic average near $\sim$45‰ (Fig. 3). This temporal distinction also exists for estimates of $^{33}_{\text{AGEO}}$ (Fig. 3) (54). Interestingly, by comparing both isotopic metrics ($^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$) with estimates of shelf area, we are able to resolve temporal patterns (Fig. 4 and Fig. S4). From our analysis of the compiled datasets (SI Text), we find that as shelf area increases, both $^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$ tend to decrease (Fig. 4 and Fig. S4). Closer examination of these data illustrates, however, that the statistical significance of these correlations rests largely on the tightly coupled behavior in the Meso-Cenozoic. In contrast, the Paleozoic has a less coherent relationship (Fig. 4 and Fig. S4). We thus interpret the multiple $S$ isotope record as being divisible into two states offset by a transition. The first state is the early Paleozoic (540–300 Ma), where $^{34}_{\text{GEO}}e$ is lower, shelf areas are larger, and $^{33}_{\text{AGEO}}$ is highly variable; the second state is the Meso-Cenozoic (200–0 Ma), where $^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$ are larger and associated with less shelf area. The transition spans at least the Permian and the Triassic (ca. 300–200 Ma).

Although there is relatively less sulfur isotopic variability in the Meso-Cenozoic compared with the Paleozoic, the interval from the Cretaceous to the present (ca. 100–0 Ma) shows increases in $^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$ of $\sim$40‰ and $\sim$0.0002, respectively (Fig. 3). [The opposite trends, and by analogy the converse arguments, apply to the Mesozoic interval from 200 to 100 Ma (Fig. 3).]

In our conceptual framework, this may represent a constant $s_{\text{SRR}}$ through time. Records of pyrite relative to coeval sulfate ($^{34}_{\text{GEO}}e$ and $^{33}_{\text{AGEO}}$) gives compilation, binning strategy, and data handling.

![Fig. 3. The Phanerozoic S isotope record. SI Text gives compilation, binning strategy, and data handling.](image-url)
with a decreasing shelf area (thereby decreasing both gsrr and pyrite burial), or it may represent a decrease in sedSRR with concomitant decline in shelf area. We first consider the case of constant sedSRR: Here, a ~10% decrease in pyrite burial is required to accommodate the 7%/e increase in 34/Sgeo (SI Text). Such a fluctuation in pyrite burial is reasonable. However, changes in pyrite burial cannot affect 33/Sgeo and, as such, the observed variance in 33/Sgeo means that a change in shelf area alone (as it relates to gsrr) cannot explain the Meso-Cenozoic S isotope record (i.e., sedSRR also must have changed).

As an alternative to a change only in shelf area, our experimental data provide a means to test the second hypothesis: variable sedSRR. The sediment data for 33/Sgeo vs. 34/Sgeo through the Meso-Cenozoic are statistically within the relationship extracted from our chemostat data (33/SMSR vs. 34/DMSR; Fig. S5). This suggests that the dominant control on fractionation is sulfate reduction, and that the geologic data can be interpreted in the context of where they fall on the slope-rate relationship of MSR (Fig. S5). The isotopic record of the Meso-Cenozoic corresponds to an estimated mean 83%/e variation in sedSRR if based on our experimentally determined 33/SMSR-rate relation, or a mean 43%/e variation in sedSRR if based on the 33/SMSR-rate relation [Fig. 5, Fig. S4, and SI Text]. 34/DMSR or 33/SMSR-derived mSRR is converted to sedSRR by using the Plio-Pleistocene as a reference and assuming that the scaling relationship is constant over time (SI Text)].

We postulate that synergistic changes in sedSRR and shelf area work together to explain the Meso-Cenozoic S isotope record. The pattern of change in sedSRR between 200 Ma and 0 Ma is also significantly correlated to the variability in shelf area over that same interval (Fig. 5 and Fig. S4). This implies that sedSRR and shelf area are at least partially inseparable variables, although the underlying coupling between them is not immediately clear. Importantly, an increase in sedSRR is not simply an increase in the molar flux of sulfate reduction, proportional to increased shelf area. An increase in sedSRR requires a change in the local mSRR integrated in a local sediment column. This must signal a response to availability of the limiting reactant—in this case, OC—either via its quantity or quality delivered to the sedimentary zone of sulfate reduction. We hypothesize that changes in the absolute flux of nutrients (59) to shelf environments is influencing the location and intensity of primary productivity, changing the delivery of OC to the sediments, and influencing the sedSRR. In this way, sedSRR joins the many other sedimentary biogeochemical processes known to respond to varying nutrient regimes (60). Further refining the structure of the Meso-Cenozoic record may provide critical insight into what processes are playing a prominent role in setting sedSRR, and we consider the consequences of this prediction below. However, we first consider the different patterns apparent in the Paleozoic records.

Our mechanistic understanding of the isotopic zonation of the S isotope record of the last 200 Ma does not explain the 33/Sgeo~34/Sgeo relationship of the Paleozoic. Compared with the mid-Cenozoic, the early-mid Paleozoic (540–300 Ma) records a similar variance in 34/Sgeo values (~6%/e around an era mean of ~30%/e), but three times the variability in 33/Sgeo (a continuous decrease of ~0.0008; Fig. 3). This results in a significant departure of the Paleozoic from the 34/Sgeo vs. 33/Sgeo slope-rate relationship that defines the last 200 Ma (Fig. 4 and Fig. S5). The lower overall 34/Sgeo during the Paleozoic (~30%/e), if paired with the lowest Paleozoic 33/Sgeo from that same interval (Pennsylvania; Fig. S4), may indeed relate to an elevated sedSRR and greater areal extent of shallower sea floor (Fig. 4). However, the failure of the Paleozoic data to plot on the 34/SMSR vs. 33/SMSR line (Fig. S5) not only undermines any confidence in applying our experimental calibration to this earlier interval (SI Text), it also is consistent with the observed weak relationship between the 34/Sgeo and 33/Sgeo data to shelf area (Fig. 4). This suggests that the general decoupling of 34/Sgeo from 33/Sgeo within the Paleozoic is not readily attributable only to changes in sedSRR. The breakdown of the observed relationships that applied in the Meso-Cenozoic suggests that, for the Paleozoic, a conceptual model that ascribes changes in 34/Sgeo and 33/Sgeo to a codependent relationship between sedSRR and shelf area is insufficient. Because fluctuations in 33/Sgeo for the Paleozoic are largely independent of changes in 34/Sgeo, there must be additional forcings.

Modeling studies illustrate how such an isotopic decoupling can be produced, either through an increase in reoxidative fluxes and associated processes (61) or through non-steady-state behavior (62). The former may seem unlikely given lower estimated oxygen partial pressure (pO2) during the first two-thirds of the Paleozoic (63). However, lower pO2 also implies a sulfate reduction zone closer to the sediment–water interface, or perhaps within the water column. Under these conditions, the delivery of O2 to the sulfate reduction zone is no longer diffusion-limited, and as a result reoxidation reactions may be fractionally more important despite lower overall pO2. This includes both classic sulfide oxidation [at the expense of oxygen, nitrate, manganese (IV) oxides, and iron (III) oxide-hydroxides] as well as disproportionation reactions. Both have been tentatively shown to produce a negative slope of the 33/Sgeo~34/Sgeo isolate relationship (15, 61). The shallow slope of the Paleozoic 33/Sgeo vs. 34/Sgeo data suggests that this opposite isotopic directionality partially is expressed and preserved (64). A change in the location and flux through reoxidation reactions is also dependent on the proximity to iron (water column vs. sediment), given that pyrite formation is a terminal sink (i.e., reoxidation inhibitor) for aqueous sulfide (65). Non-steady-state behavior of the Paleozoic S cycle is also possible and necessarily relates to seawater sulfate concentrations; however, current estimates suggest Paleozoic sulfate was abundant enough to circumvent such direct, first-order microbial control on fractionation (29). We therefore favor the explanation that a fundamentally different zonation of the paleo-redox boundary in the Paleozoic decoupled the controls on S isotopes from sedimentary processes (sedSRR) and shelf area effects. Although speculative, this idea is testable using modern environments with strong redox gradients.

Equally as intriguing as the difference between the early Paleozoic and the last 200 Ma is the isotopic transition captured during the Permian and Triassic (Fig. 3 and Fig. S4). The generally weak correlations between 34/Sgeo and 33/Sgeo within the

![Fig. 5](http://example.com/figure5.png)

Fig. 5. Calculated changes in sedimentary sulfate reduction rates over the Meso-Cenozoic. There is a statistically significant relationship between the calculated relative change in sulfate reduction rate and shelf area over the last 200 Ma, with a Cretaceous maximum nearly 40–80% higher than the Plio-Pleistocene average. Before 200 Ma, rate predictions from 33/Sgeo and 33/Ageo are not always convergent or statistically robust (Fig. S4). The sedSRR calculated using the 33/Ageo as input is presented in Fig. S4 but should be interpreted with care given the modestly sized dataset underpinning Phanerozoic predictions (S4).
Paleozoic are followed by a 15% increase in average $^{34}$S_{GEO} and a 0.0005 increase in $^{34}$S_{GEO}. This implies a major state change to the global biogeochemical S cycle across 100 Ma spanning the Permo-Triassic boundary. Complementary geochemical metrics ($\delta^{13}$C, $\delta^{18}$O, and $^{87}$Sr/$^{86}$Sr) also record major changes as Earth transitioned from the later Paleozoic to late Triassic (66). A recent statistical treatment elegantly points to the assembly and breakup of Pangaea as the explanation for this state change (56). Pangaea formed during the end of the Paleozoic and began to rift during the Triassic (67). This clearly relates to shelf area (55), but also affects primary production (OC) through the effect of continental weathering on nutrient budgets (68). Although this transition in the S cycle encapsulates the Permo-Triassic boundary, it began tens of millions of years before the Permian–Triassic extinction. Such a protracted time scale for the isotopic transition (from 300 to 200 Ma) may implicate the possible cause as a change in the ratio of S to Fe from rivers (65) associated with a Carboniferous increase in pO$_2$ (63).

Conclusions

We propose that changes in sulfate reduction rates in marine sediments, rather than global pyrite burial, dictated S isotope records over the last 200 Ma. The post-Triassic geological record suggests that sedSRR increased to a mid-Cretaceous apex and then slowed again over the last 100 Ma (Fig. 5). What, then, controlled sedSRR? During this interval our estimates of sedSRR correlate strongly with estimates for shelf area (Fig. S4B). To the degree that global shelves capture weathering fluxes and modulate nutrient delivery to primary producers, they may not only dictate the quantity and quality of organic matter reaching the zone of sulfate reduction, but they also help dictate the position of this zone with respect to the sediment–water interface. Thus, if the combination of OC delivery and sedimentation rate controls sedSRR, marine sedimentary S isotope compositions may reflect a combination of Meso-Cenozoic productivity and shelf area. For this relationship to hold (i.e., to maintain the direct connection to shelf area), it requires that pO$_2$ be elevated in the Meso-Cenozoic relative to the early/mid-Paleozoic (63), because it must reconcile the locus of sulfate reduction to be (on average) below the sediment–water interface. Encouragingly, this provides a falsifiable testable hypothesis, replicable to high-resolution datasets from Phanerozoic sections where multiple biogeochemical isotope proxies are measurable and interpretable (69).

The Paleozoic $^{34}$S_{GEO} vs. $^{34}$S_{GEO} relationship requires significant contribution from processes other than changing OC delivery rates or total shelf area. As such, building a quantitative understanding of the Paleozoic sulfur cycle remains difficult. The disagreement between Paleozoic sedimentary data and our experimental calibration leaves room for other S metabolisms to contribute significantly to the observed S fractions (biotic and abiotic sulfide oxidation, as well as disproportionation). Indeed, if a major difference between the Paleozoic and Meso-Cenozoic is the locality and magnitude of sulfide reoxidation—the water column in the Paleozoic vs. the sediments in the Meso-Cenozoic—then the delivery of reactive iron to the zone of sulfate reduction (55, 70) will determine whether most iron sulfides are syngenetic or diagenetic. This in turn influences the net pyrite burial flux and the balance of global oxidants. Further measurements and modeling are required to define these fluxes over both eras. Still, we can state with some confidence that recent multiple S isotope records do not require changes in pyrite burial over the last 200 Ma. Recent sediments carry a detailed and rich repository of small-scale microbial activities, and the magnitude of overall S fractionations likely reflects global oxidant budgets working in tandem with major tectonic changes.

Ultimately it is the reduction potential trapped in pyrite and organic matter—rather than the rate of sedimentary sulfate reduction—that influences Earth’s surface oxidant budget on billion-year time scales. Our hypotheses suggest that pyrite burial flux (and by extension its contribution to pO$_2$) has not changed dramatically in recent times. Conversely, changes in pyrite burial flux remain a potentially critical control on the oxidant budget during the first half of the Phanerozoic. Continued measurements of geological materials (marine pyrites, sulfates, and terrestrial deposits), coupled with additional microbial experimentation and biogeochemical modeling, promise to yield further insight into the behavior of the S cycle over Earth’s history.

Materials and Methods

An experiment with Dvh in a custom-built chemostat was run with lactate as the sole electron donor and rate-limiting nutrient for sulfate reduction. The dilution rate varied over 43-fold (D = mSRR), with measurements of the following biological, geochemical, and isotopic parameters on all samples (Dataset S1): temperature, pH, dilution rate, cell density, sulfide production rate (the sole detectable sulfate reduction product), acetate production rate, $^{34}$S$_{NaH}$ and $^{34}$S$_{NaP}$ of sulfate and sulfide, and cell density. By measuring the difference between the S-isotope compositions of sulfate and sulfide, and because S-mass balance is closed with these two pools alone (i.e., no thionates detected), we may directly calculate the $\delta^{34}$S$_{NaH}$ and $\delta^{34}$S$_{NaP}$. Specific growth rate is calculated by measuring the dilution rate and the rate of change in optical density between time points (Dataset S1). The non-linear regression models (based on pseudo-first-order kinetics) are applied to the experimental data ($^{34}$S$_{MSR}$ or $^{34}$S$_{MSSR}$ vs. mSRR, allowing us to calculate theoretical $^{34}$S$_{MSR_{max}}$ and $^{34}$S$_{MSSR_{max}}$ along with corresponding fitting parameters, SEs, and 95% confidence interval (Fig. 2). We compare between models by examining the goodness of fit from each model (Fig. S3). The resultant expressions relate a measured isotopic fractionation (Eqs. 58 and 59) to mSRR and the fitted kinetic constant (k) to extract fractionation limits (e.g., $^{34}$S$_{MSR_{max}}$ and $^{34}$S$_{MSSR_{max}}$). Relative sedSRRs are calculated with Eqs. S12–S14. Phanerozoic data compilations and isotope mass-balance models are detailed in SI Text.

Acknowledgments. We thank A. Pearson for detailed comments that greatly improved this manuscript, C. Cavanaugh, C. Hansel, T. Laakso, and J. C. Creveling for comments that helped align earlier versions of this manuscript, and Tim Lyons and an anonymous reviewer for thoughtful commentary. A. Masterson, E. Beirne, and M. Schmidt provided expert analytical assistance. The National Science Foundation (NSF) Graduate Research Fellowship Program (W.I.L.), Microbial Sciences Initiative at Harvard (W.I.L. and D.T.J.), National Aeronautics and Space Administration (NASA)–NASA Astrobiology Institute, NSF Earth Sciences/Instrumentation and Facilities, NSF Faculty Early Career Development Program, and Harvard University (D.T.J.) all supported this work.


