Distinct Malignant Behaviors of Mouse Myogenic Tumors Induced by Different Oncogenetic Lesions

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Citation

Published Version
doi:10.3389/fonc.2015.00050

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INTRODUCTION

Rhabdomyosarcomas (RMS) are heterogeneous cancers with myogenic differentiation features. The cytogenetic and mutational aberrations in RMS are diverse. This study examined differences in the malignant behavior of two genetically distinct and disease-relevant mouse myogenic tumor models. *Kras*; *p1619null* myogenic tumors, initiated by expression of oncogenic *Kras* in *p1619full* mouse satellite cells, were metastatic to the lungs of the majority of tumor-bearing animals and repopulated tumors in seven of nine secondary recipients. In contrast, *SmoM2* tumors, initiated by ubiquitous expression of a mutant Smoothened allele, did not metastasize and repopulated tumors in 2 of 18 recipients only. In summary, genetically distinct myogenic tumors in mice exhibit marked differences in malignant behavior.

**Keywords:** rhabdomyosarcoma, myogenic differentiation, metastasis, transplantation

MATERIALS AND METHODS

MICE

R26-SmoM2 (mixed genetic background including 129/Sv and Swiss Webster as main components) (*11*), CAGGS-CreER (*11*), and NOD.CB17-Prkdc<sup>scid</sup>/J (NOD.SCID) mice were purchased from The Jackson Laboratory. *p1619null* mice (B6.129 background) were obtained from the NIH/Mouse Models of Human Cancer Consortium. Mice were bred and maintained at the Joslin Diabetes Center Animal Facility. All animal experiments were approved by the Joslin Diabetes Center Institutional Animal Care and Use Committee.

SARCOMA INDUCTION

*Kras*; *p1619null* myogenic tumors were initiated by fluorescence-activated cell sorting of *p1619null* satellite cells, followed by lentiviral transduction to introduce oncogenic *Kras*(*G12v*) and implantation in the gastrocnemius muscles (*10*). R26-SmoM2;CAGGS-CreER were injected with Tamoxifen (1 mg/40 g) on postnatal day 10 to activate expression of CRE recombinase and SMOM2. R26-SmoM2;CAGGS-CreER spontaneously developed multifocal skeletal muscle tumors (*SmoM2* tumors) as previously described (*11, 12*).

HISTOPATHOLOGY

Tumor tissue was dissected, fixed in 4% paraformaldehyde for 2 h, and embedded in paraffin. Standard H&E stained sections were prepared. Staining for Actin (Dako, M0635, 1:200), Desmin (Dako, M0760, 1:50), and Ki67 (Vector Labs, VP-K451, 1:250) was performed.

LUNG METASTASES

Tumor-bearing mice were monitored at least twice weekly for health problems, and were sacrificed once tumors reached a volume of 1 cm<sup>3</sup> or were ill. Lungs were dissected, fixed in 4% paraformaldehyde for 2 h, and embedded in paraffin. Standard
As previously reported (12), the Ki67 index of the Kras; p16p19 tumor-repopulating activity in mice (11, 12). The phenotypes of Kras; p16p19 mouse myogenic tumors were previously described (10–12). To assess the metastatic potential of Kras; p16p19 tumors, viable tumor cells were transplanted into the cardiotoxin-pre-injured gastrocnemius muscles of NOD.SCID mice using a transdermally inserted dental needle attached to a Hamilton syringe via polyethylene tubing. Recipient muscles were preinjured 24 h before cell implantation by injection of 25 ml of a 0.03 mg/ml solution of cardiotoxin (from Naja mossambica, Sigma) in order to enhance cell engraftment. Mice were screened once weekly for the development of tumors at the injection sites.

STATISTICS

Differences between Kras; p16p19null and SmoM2 mouse myogenic tumors were evaluated by T-test (Ki67 indices), Fisher’s Exact test (prevalence of lung metastases), and Kaplan–Meier analysis (tumor-repopulating activity).

RESULTS

Kras; p16p19null AND SmoM2 MOUSE TUMORS EXHIBIT A MYOGENIC TUMOR PHENOTYPE

Kras; p16p19null mouse myogenic tumors were induced by intramuscular implantation of Kras(G12v)-expressing p16p19null muscle satellite cells (10). In contrast, SmoM2 mouse myogenic tumors were initiated by ubiquitous activation of a mutant, constitutively active smoothened (SmoM2) allele in R26-SmoM2;CAGGS-CreER mice (11, 12). The phenotypes of Kras; p16p19null and SmoM2 myogenic tumors were previously described (10–12). In brief, Kras; p16p19null tumors contained bundles of cells with large, atypical nuclei, frequent mitotic figures, and occasional multinucleated giant cells. Subsets of cells (<50% of all tumor cells) expressed terminal muscle differentiation markers such as desmin and actin (Figure 1A), and the proliferative index as evidenced by the percentage of Ki67-expressing nuclei was 41.6 ± 12.5% (range 30.5–59.3%; four tumors evaluated) (Table 1). SmoM2 tumors contained many multinucleated, elongated cells with abundant cytoplasm interspersed with small round cells. SmoM2 tumors lacked cellular atypia and diffusely expressed desmin and actin in many tumor cells (more than 75% of all tumor cells; Figure 1B). As previously reported (12), the Ki67 index of SmoM2 tumors was 19.1 ± 15.9% (range 3.4–41.8%; six tumors evaluated) and lower than that observed in Kras; p16p19null tumors (p = 0.05; Table 1).

Kras; p16p19null AND SmoM2 MOUSE MYOGENIC TUMORS HAVE DIFFERENT METASTATIC POTENTIAL

The lung is the primary organ affected by distant sarcoma metastases in humans. To assess the metastatic potential of Kras; p16p19null and SmoM2 tumors, random lung sections obtained from tumor-bearing animals were screened for the presence of metastases. Six of seven mice with Kras; p16p19null myogenic tumors were found to have lung metastases at the time of death (mice were sacrificed 17–28 days after detection of palpable tumors) (Figure 2). In contrast, 0 of 8 mice with SmoM2 myogenic tumors had lung metastases at the time of death (mice were sacrificed at 38–55 days of age and 5–21 days after detection of palpable tumors). The prevalence of lung metastases in Kras; p16p19null and SmoM2 myogenic tumor-bearing mice was significantly different (p = 0.001).

Kras; p16p19null AND SmoM2 MOUSE MYOGENIC TUMORS DIFFER IN TUMOR-REPOPULATING ACTIVITY

Most malignant tumors contain cells that have the capacity to repopulate secondary tumors when transplanted into a susceptible secondary environment, and this assay has been used as a test of the malignancy of distinct tumors and tumor cell subsets (13). To evaluate the tumor-repopulating activity of Kras; p16p19null and SmoM2 mouse myogenic tumors, viable tumor cells were transplanted into the cardiotoxin-pre-injured gastrocnemius muscles of NOD.SCID mice. The Kras; p16p19null tumor cell pool contains approximately 70% GFP+ cells and 30% GFP− cells (10). Because tumor-repopulating activity in Kras; p16p19null tumors resides within the Kras-expressing, GFP+ subset of tumor cells descended from virally infected satellite cells (Figure S1 in Supplementary Material).
Table 1 | Differences in the malignant behavior of Kras; p16p19null and SmoM2 mouse tumors.

<table>
<thead>
<tr>
<th>Kras; p16p19null tumors</th>
<th>SmoM2 tumors</th>
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<tbody>
<tr>
<td>Terminal muscle differentiation</td>
<td>Actin/desmin expression in &lt;50% of tumor cells</td>
</tr>
<tr>
<td>Ki67 index (p = 0.05)</td>
<td>41.6 ± 12.5%</td>
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<tr>
<td>Metastases (p = 0.001)</td>
<td>7 of 9 mice with lung metastases</td>
</tr>
<tr>
<td>Transplantation (p &lt; 0.001)</td>
<td>7 of 9 transplanted mice developed tumors (50 cells injected)</td>
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</tbody>
</table>

Kras; p16p19null and SmoM2 mouse myogenic tumors exhibit profound differences in tumor-repopulating activity and metastatic behavior.

DISCUSSION

Our findings highlight differences in the malignant phenotype and behavior of mouse myogenic tumors driven by activation of distinct RMS-relevant oncogenic pathways. Kras; p16p19null myogenic tumors were metastatic to the lungs of the majority of tumor-bearing animals and contained high tumor-repopulating activity. In contrast, SmoM2 tumors did not metastasize and were substantially less effective in repopulating tumors in secondary recipients. These experiments indicate marked differences in tumor-repopulating activity of Kras; p16p19null and SmoM2 tumors (p < 0.001, Figure 3), in terms of both the frequency of tumor-repopulating cells and the latency of secondary tumor formation.
The two model systems described in this study were induced by different experimental methods. SmoM2 tumors originated from Cre-mediated activation of a conditionally expressed transgene. Kras; p16p19null mouse tumors, on the other hand, were initiated by viral transduction and intramuscular implantation of target satellite cells. We note that Kras; Tp53−/− mouse myogenic tumors (14, 15), induced by Cre-mediated activation of oncogenic hits instead of viral transduction, exhibit a phenotype that closely resembles the Kras; p16p19null mouse tumors described here. For example, Kras; p16p19null share their propensity to metastasize to the lungs of tumor-bearing animals with Kras; Tp53−/− mouse tumors (14). Nevertheless, it is possible that differences in the tumor induction strategy (such as off-target effects of viral transduction) could contribute to the observed differences in malignant behavior between SmoM2 and Kras; p16p19null mouse myogenic tumors.

Similar to mouse myogenic tumors, human fusion-negative RMS comprises a group of tumors with clear differences in histology, myogenic differentiation state, oncogenic pathway activation, and genetic background. In recent years, subsets of human RMS tumors that exhibit a combination of specific genetic and phenotypic characteristics were distinguished. For example, a subset of human fusion-negative RMS with spindle cell/sclerosing histology was recently found to exhibit diffuse MyoD expression, carry frequent somatic MyoD mutations, and portend a poor prognosis (16, 17). Also, children with TP53 germline mutations are predisposed to develop anaplastic RMS at a young age (18), and germline mutations in Dicer1 were linked to a genetic susceptibility to develop RMS of the genitourinary tract (19). Future extended (epi-)genotype/phenotype correlations might pinpoint clinically/biologically distinct subgroups of human fusion-negative RMS and identify biomarkers to facilitate prognostication and/or stratification of therapy.

AUTHOR CONTRIBUTIONS
SH, RB, and AW conceived experiments, analyzed data, wrote, and approved of the manuscript.

ACKNOWLEDGMENTS
We thank C. L. Unitt and T. Bowman at the DF/HCC Histopathology Core for help with immunohistochemistry, D. Tchesalova for excellent animal care, and Joyce LaVecchio, Girijesh Burizula, and Atsuya Wakayabashe in the Joslin Diabetes Center Flow Cytometry Core (supported by the Harvard Stem Cell Institute and NIH P30DK036836) for flow cytometry support. This work was funded in part by a Stand Up To Cancer-American Association for Cancer Research Innovative Research Grant (SU2C-AACR-IRG1111; to AW); by grants from the Burroughs-Wellcome Fund and the Harvard Stem Cell Institute (to AW), and by P.A.L.S. Bermuda/St. Baldrick’s, ALSF, and Bear Necessities (to SH). Content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH or other funding agencies.

SUPPLEMENTARY MATERIAL
The Supplementary Material for this article can be found online at http://www.frontiersin.org/Journal/10.3389/fonc.2015.00050/abstract
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Received: 21 December 2014; accepted: 11 February 2015; published online: 24 February 2015.

This article was submitted to Pediatric Oncology, a section of the journal Frontiers in Oncology.
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