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Green Herring Syndrome: Bacterial Infection in Patients With Mucormycosis Cavitary Lung Disease

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Mucormycosis is a life-threatening fungal disease in patients with hematological malignancies. The diagnosis of pulmonary mucormycosis is particularly challenging. We describe 3 mucormycosis cases with an uncommon presentation in patients whose cavitary lung disease was attributed to well documented bacterial infection, although evolution and reassessment established mucormycosis as the underlying disease.

Keywords: bacterial coinfection; cavitary lung disease; mucormycosis; transplantation.

Mucormycosis is a life-threatening, angioinvasive fungal disease. It is caused by ubiquitous saprophytic fungi of the order Mucorales and predominantly occurs in immunocompromised patients [1]. Although uncommon, its frequency appears to have increased in recent years, particularly in transplant recipients and patients with hematological malignancies [2]. In a recent study of the frequency of invasive fungal disease in hematopoietic stem cell transplant (HSCT) recipients, mucormycosis was the third most common fungal infection, after candidiasis and aspergillosis [3], accounting for up to 8% of invasive fungal disease in HSCT recipients [4].

Pulmonary mucormycosis is usually a rapidly progressive infection associated with debilitating morbidity and high mortality. Early recognition of this diagnosis, along with aggressive management, is critical to effective therapy and patient survival [5]. The diagnosis of pulmonary mucormycosis can be particularly challenging. Not only is this condition relatively rare, but it is also challenging to differentiate from other causes of pneumonia and other angioinvasive pulmonary diseases caused by molds [6].

In this report, we present 3 cases that highlight a misleading presentation of mucormycosis in which bacteria were identified as the causal etiology of necrotizing cavitary lung disease on initial diagnostic evaluation, but subsequent evolution and reassessment of the same cavitary disease demonstrated underlying mucormycosis. This bacterial and apparently parsimonious presentation, which we have termed the green herring syndrome, masked and delayed the diagnosis and treatment of the underlying cause of the pulmonary cavitary disease.

CASES

Patient A
A 60-year-old male with non-Hodgkin follicular lymphoma underwent reduced-intensity allogeneic HSCT from a related donor. On day +140 posttransplantation, the patient developed diarrhea and was found to have severe gastrointestinal acute graft-versus-host disease (GVHD) for which he received high-dose prednisone and tacrolimus (Table 1). He was admitted to the hospital with 2 days of shortness of breath and cough on day +177 posttransplantation. Admission chest computed tomography (CT) demonstrated a cavitary lesion surrounded by consolidation in his left lower lobe (Figure 1). He received metronidazole, levofloxacin, and voriconazole empirically.

During admission, the cavitary lung lesion progressed with development of a worsening ipsilateral pleural effusion with new pleural gas. On day +186 posttransplantation, he underwent a bronchoalveolar lavage (BAL) with left lower lobe washings and left video-assisted thoracoscopic surgical exploration. Cloudy, foul-smelling pleural fluid was obtained, suggestive of bacterial empyema. Lung biopsy was not pursued given this finding. Cultures grew fluoroquinolone-resistant Escherichia coli, enterococci, and Peptostreptococcus magnus. Voriconazole was discontinued, and he received cefotaxime, metronidazole, and vancomycin.

Two days later, he developed altered mental status, and a head CT showed an area of heterogeneous attenuation in the left frontal lobe with surrounding mass effect. A repeat chest
CT demonstrated new multifocal ground-glass opacities in the right lung. On day +188 posttransplant, the patient was intubated due to worsening dyspnea, and voriconazole was restarted. The patient died on day +190 posttransplant.

On autopsy, he had angioinvasive broad, ribbon-like hyphal forms with 90-degree branching suggestive of invasive mucormycosis in his lung, brain, and thyroid. Lung cultures grew *E coli*, enterococci, and *Rhizopus* species. The causative fungal species was identified by tissue polymerase chain reaction (PCR) as *Rhizopus microsporus/Rhizopus azygosporus*.

**Patient B**
A 57-year-old male with relapsed B-cell chronic lymphocytic leukemia developed fevers, productive cough, and hemoptysis while receiving posaconazole prophylaxis after recent treatment with alemtuzumab and high-dose methylprednisolone (Table 1).

A chest CT demonstrated (1) a large dominant consolidative opacity in the right upper lobe (RUL) with focal dense consolidation and peripheral ground-glass and (2) another focal opacity superiorly to this dominant opacity with a dense peripheral rind and central ground-glass opacity (reversed halo sign [RHS], Figure 1). He was started on ceftriaxone, levofloxacin, and liposomal amphotericin B (5 mg/kg q24h) empirically. Bronchoalveolar lavage bacterial and fungal cultures were sterile; serum galactomannan and (1→3)-β-D-glucan were also negative. He improved clinically and his nodules were less prominent on a repeat chest CT. He was discharged on 400 mg of posaconazole twice daily.

Two weeks later, he underwent a reduced-intensity allogeneic HSCT from a matched unrelated donor. Posaconazole and levofloxacin were continued posttransplant. Posaconazole levels were not obtained. Serial chest CT scans performed to follow-up the RUL consolidation showed progression, and on day +6 posttransplant 2 new areas of focal consolidation were noted. He underwent a fine-needle aspiration under CT guidance of this RUL lesion, and bacterial cultures grew fluoroquinolone-resistant *E coli*. No fungus was cultured or seen on cytopathology. He was treated with ceftriaxone. On day +34 posttransplant, a repeat chest CT demonstrated increasing dense RUL consolidation with increased cavitation and liposomal amphotericin B was started.

### Table 1. Clinical Characteristics of Patients With Mucormycosis Presenting With Well Documented Bacterial Infection

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Patient A</th>
<th>Patient B</th>
<th>Patient C</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age at diagnosis (years)</td>
<td>60</td>
<td>57</td>
<td>49</td>
</tr>
<tr>
<td>Sex</td>
<td>Male</td>
<td>Male</td>
<td>Male</td>
</tr>
<tr>
<td>Underlying disease</td>
<td>Non-Hodgkin’s lymphoma</td>
<td>Chronic lymphocytic leukemia</td>
<td>T-cell prolymphocytic leukemia</td>
</tr>
<tr>
<td>Oncologic status</td>
<td>Remission</td>
<td>Relapsed</td>
<td>Remission</td>
</tr>
<tr>
<td>GVHD</td>
<td>Yes</td>
<td>No</td>
<td>Yes</td>
</tr>
<tr>
<td>GVHD treatment</td>
<td>Prednisone, tacrolimus</td>
<td>No</td>
<td>Prednisone, interleukin-2, extracorporeal photopheresis</td>
</tr>
<tr>
<td>Initial bacterial infection</td>
<td>Pneumonia</td>
<td>Pneumonia</td>
<td>Pneumonia and bacteremia with <em>Pseudomonas aeruginosa</em></td>
</tr>
<tr>
<td>Fungal disease diagnosis</td>
<td>Autopsy</td>
<td>Transbronchial biopsy</td>
<td>Brain mass biopsy and bronchoalveolar lavage</td>
</tr>
<tr>
<td>Days posttransplant</td>
<td>D+186</td>
<td>Before the transplant</td>
<td>D+650</td>
</tr>
<tr>
<td>Fungal species</td>
<td><em>Rhizopus microsporus/azygosporus</em></td>
<td><em>Rhizopus species</em></td>
<td><em>Rhizomucor pusillus</em></td>
</tr>
<tr>
<td>Clinical presentation</td>
<td>Shortness of breath and cough</td>
<td>Cough</td>
<td>Shortness of breath and pleuritic pain</td>
</tr>
<tr>
<td>Hemoptysis</td>
<td>No</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Neutropenia at diagnosis</td>
<td>No</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td>Steroids before diagnosis</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Antifungals before diagnosis</td>
<td>Fluconazole, voriconazole</td>
<td>Posaconazole, liposomal amphotericin B</td>
<td>Voriconazole</td>
</tr>
<tr>
<td>Site of infection</td>
<td>Lung, brain, thyroid</td>
<td>Lung</td>
<td>Lung and brain</td>
</tr>
<tr>
<td>Therapy</td>
<td>None</td>
<td>Liposomal amphotericin B, isavuconazole</td>
<td>Liposomal amphotericin B, isavuconazole</td>
</tr>
<tr>
<td>Days between mucormycosis diagnosis and death</td>
<td>0</td>
<td>229</td>
<td>29</td>
</tr>
</tbody>
</table>

Abbreviation: GVHD, graft-versus-host disease.
Three days later, the patient developed massive hemoptysis and acute hypoxemic respiratory failure requiring intubation and endovascular right bronchial artery embolization. A transbronchial lung biopsy demonstrated hyphal forms consistent with mucormycosis; cultures were negative. The mold was identified as *Rhizopus* species by PCR/sequencing [7]. The patient continued treatment with liposomal amphotericin B for several months and developed acute renal failure requiring hemodialysis. He developed pancytopenia due to graft loss, and a donor-lymphocyte infusion was administered 7 months post-transplant. Isavuconazole was used for further treatment given ongoing renal dysfunction. One month later, the patient developed severe gastrointestinal and cutaneous GVHD and died from septic shock due to a carbapenem-resistant *Enterobacter cloacae* bacteremia on day +276 posttransplantation. There was no radiological progression of his lung cavity during this period.

**Patient C**

A 49-year-old male with T-cell prolymphocytic leukemia, status post-allogeneic HSCT complicated by chronic GVHD requiring prednisone, presented with fever, bronchopneumonia, and *Pseudomonas aeruginosa* bacteremia 1 year after transplant. Treatment with ciprofloxacin and ceftazidime was started. His serum (1→3)-β-D-glucan was positive at 126 pg/mL and serum galactomannan was negative. Chest CT showed several small cavitory nodules and peribronchial nodular opacities in both lungs. He received treatment with voriconazole, with resolution of his respiratory symptoms and cavitory lesions.
He presented to the hospital on day +650 posttransplantation with dyspnea, fever, and pleuritic chest pain (Table 1). A chest radiograph revealed a nodular right lung opacity with cavitation. Blood cultures grew *P aeruginosa*. He had copious purulent secretions on BAL, so transbronchial biopsy was not performed; bacterial cultures of BAL fluid also grew *P aeruginosa*. The patient was treated with cefepime and levofloxacin (Figure 1).

Ten days later, he developed altered mental status, and a head CT demonstrated a new mass in the right caudate lobe with mass effect. A brain biopsy was performed and showed fungal forms suggestive of mucormycosis; cultures grew *Rhizomucor pusillus*. Repeat BAL cultures eventually grew the same organism. He received a dose of liposomal amphotericin B (5 mg/kg) followed by isavuconazole due to renal dysfunction.

The patient’s neurologic function improved, and his brain and chest mucormycosis lesions decreased in size on isavuconazole therapy. Unfortunately, 1 month later, the patient died of multidrug-resistant *P aeruginosa* pneumonia and septic shock.

**DISCUSSION**

The 3 patients in this series had cavitary pulmonary lesions that were attributed to bacterial infection based on an initial diagnostic evaluation, and they improved transiently with antibiotic therapy directed against the identified bacterial organisms. However, subsequent reassessments given an inadequate clinical response to antibacterial therapy led to the diagnosis of the underlying mucormycosis. The initial diagnosis of bacterial infection was plausible, parsimonious, and based on direct pulmonary cultures but ultimately misleading. This initial diagnosis resulted in delays and interruptions in appropriate antifungal treatment and increased morbidity, a scenario that we have termed the green herring syndrome. Despite a high degree of clinical suspicion and the use of various invasive and non-invasive diagnostic tools, the fungal diagnosis was difficult to establish and delayed.

Previous studies of pulmonary mucormycosis have demonstrated that clinical features of mucormycosis are nonspecific, leading to diagnostic difficulties and late diagnoses, with mortality rates as high as 65% among patients with hematological malignancies and 90% among bone marrow transplant recipients [5,8].

Chest CT imaging can be a useful tool for timely diagnosis of pulmonary mucormycosis, because cavitation is described as the common finding [9]. Moreover, the RHS or atoll sign, defined as a central ground-glass opacity with a surrounding rim of consolidation, is present early in the disease course and is more common among patients with mucormycosis than among those with other forms of invasive fungal pneumonia [10], especially when associated with negative serum galactomannan and (1→3)-β-D-glucan [5]. However, neither of these radiographic findings is unique to pulmonary mucormycosis. The diagnosis often involves a multidisciplinary approach, aided by a combination of imaging features, clinical picture, biochemical and microbiological evaluation, and pathological confirmation. The RHS is not pathognomonic and can be seen in patients with aspiration pneumonia and pulmonary infarction secondary to thromboembolic disease. Of interest, imaging in these patients demonstrated complex cavitary lesions with thick, irregular cavity walls and internal septations secondary to ischemic necrosis, which in our experience is not typically seen in bacterial lung abscesses. These lesions were preceded by the RHS in 2 of the 3 cases.

Due to the complex presentation and lack of reliable biochemical markers, methods to improve diagnosis and identification of mucormycosis and characterization of culture-negative invasive mold infection are essential. Some examples of emerging strategies that could be used to improve the diagnosis of mucormycosis are PCR detection of circulating DNA in serum [11] or detection of fungal species-specific breath volatile organic compound profiles [12].

These cases demonstrate that bacterial coinfection of pulmonary mucormycosis can lead to obscurcation and delay in diagnosis of underlying mucormycosis. This diagnosis should be promptly considered in immunocompromised patients who present with complex cavitary lesions on CT with or without presence of RHS, even if receiving antimold prophylaxis, or when their lung disease progresses despite appropriate therapy against an identified bacterial pathogen.

**Notes**

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