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The Effect of Hepatitis C Virologic Clearance on Cardiovascular Disease Biomarkers in Human Immunodeficiency Virus/Hepatitis C Virus Coinfection

Kara W. Chew,1 Lei Hua,2,a Debika Bhattacharya,1 Adeel A. Butt,3,4 Lorelei Bornfleth,1 Raymond T. Chung,5 Janet W. Andersen,6 and Judith S. Currier1

1Department of Medicine, David Geffen School of Medicine at University of California, Los Angeles; 2Department of Biostatistics, Harvard School of Public Health, Boston, Massachusetts; 3Department of Medicine, University of Pittsburgh School of Medicine, Pennsylvania; 4Sheikh Khalifa Medical City, Abu Dhabi, United Arab Emirates; 5Department of Medicine, Harvard Medical School and Massachusetts General Hospital; and 6Center for Biostatistics in AIDS Research, Harvard School of Public Health, Boston, Massachusetts

Background. Successful hepatitis C virus (HCV) treatment may reduce cardiovascular disease (CVD) risk and improve levels of CVD biomarkers produced outside the liver (nonhepatic biomarkers).

Methods. Stored serum or plasma from before and 24 weeks after end of HCV treatment (EOT) from human immunodeficiency virus (HIV)/HCV-coinfected subjects who received up to 72 weeks of peginterferon/ribavirin, 27 with and 27 without sustained virologic response (SVR) matched by race, ethnicity and sex, were tested for nonhepatic (soluble intercellular adhesion molecule-1 [sICAM-1], soluble P-selectin [sP-selectin], interleukin [IL]-6, D-dimer, and lipoprotein-associated phospholipase A2 [Lp-PLA2]) and hepatic (cholesterol and high-sensitivity C-reactive protein) CVD and macrophage activation markers (soluble CD163 [sCD163] and soluble CD14). Changes in biomarkers and their association with SVR were examined by t tests or Wilcoxon tests and regression models.

Results. Of the 54 subjects, 30 were white, 24 were black, and 44 were male. Pretreatment levels of nonhepatic biomarkers were high: sICAM-1 overall median, 439.2 ng/mL (interquartile range [IQR], 365.6–592.8); sP-selectin, 146.7 ng/mL (IQR, 94.1–209.9), and IL-6, 2.32 pg/mL (IQR, 1.61–3.49). Thirty-seven of 52 (71%) subjects had Lp-PLA2 >235 ng/mL. Sustained virologic response was associated with decrease in sICAM-1 (P = .033) and sCD163 (P = .042); this result was attenuated after controlling for changes in the alanine aminotransferase level. At 24 weeks after EOT, 17 (63%) SVRs had Lp-PLA2 >235 ng/mL vs 25 (93%) non-SVRs (P = .021).

Conclusions. Hepatitis C virus clearance may reduce hepatic and, subsequently, systemic inflammation and CVD risk in HIV/HCV coinfection.

Keywords. cholesterol; HIV/HCV coinfection; macrophage activation; sustained virologic response; vascular adhesion molecules.

Studies in the current era of effective antiretroviral therapy (ART) demonstrate increased overall mortality in human immunodeficiency virus (HIV)/hepatitis C virus (HCV)-coinfected persons compared with HIV-monoinfected control groups, with cardiovascular disease (CVD) being a leading cause of nonliver, nonacquired immune deficiency syndrome (AIDS) deaths [1, 2]. Large observational studies suggest an increased risk of CVD events in HIV-monoinfected and HIV/HCV-coinfected persons compared with uninfected controls [3–6]. Hepatitis C virus may contribute to CVD risk through several mechanisms, including insulin...
resistance, hepatic steatosis, and increased chronic inflammation and immune activation [7, 8].

Hepatitis C viral eradication may provide benefits beyond improvement in liver-specific outcomes. A recent study demonstrated the association of sustained virologic response (SVR) to peginterferon alfa and ribavirin (PEG/RBV) with reduced nonliver morbidity and mortality in HIV-infected persons, including CVD [9]. Other studies have demonstrated that cardiovascular risk factors such as hepatic steatosis and insulin resistance, as well as T-cell activation, improve with HCV clearance [8, 10, 11]. Paradoxically, successful HCV treatment may unmask underlying CVD risk, through rise in liver-produced CVD biomarkers such as cholesterol and pro-atherogenic lipoproteins, reaching indication for lipid-lowering therapy in some patients [12, 13].

The optimal method for CVD risk assessment in HIV/HCV-coinfected persons is currently unknown. Total cholesterol, low-density lipoprotein (LDL) cholesterol, and high-sensitivity C-reactive protein (hsCRP), which are routinely used, are reduced in the setting of HCV-associated chronic liver disease [14, 15] and may underestimate CVD risk in coinfected persons. Alternatively, nonhepatically produced markers of endothelial dysfunction and inflammation, such as soluble intercellular adhesion molecule-1 (sICAM-1), soluble P-selectin (sP-selectin), lipoprotein-associated phospholipase A2 (Lp-PLA2), d-dimer, and interleukin-6 (IL-6), may be better CVD risk predictors in this setting. Emerging evidence suggests such markers may improve CVD risk stratification in HIV-infected individuals [16, 17]. Their predictive value in coinfection is unknown.

Additional markers that may be useful in assessing CVD risk in coinfected patients are measures of innate immune activation, which have been linked to all-cause mortality and CVD in the setting of HIV. Soluble CD163 (sCD163), a marker of monocyte/macrophage activation, is associated with the presence or burden of atherosclerotic plaque and arterial wall inflammation in predominantly HIV-monoinfected cohorts [18, 19]. Soluble CD14 (sCD14), another marker of macrophage activation and an endotoxin receptor, predicts mortality [20] and subclinical atherosclerosis progression [21] in patients infected with HIV. Levels of sCD163 and sCD14 are increased in chronic liver disease, including viral hepatitis, in the absence of HIV infection [22, 23]. In the context of chronic HIV infection and associated immune activation, HIV coinfection may further accelerate atherosclerosis through stimulating hepatic inflammation, and specifically macrophage activation, thus promoting systemic inflammation and end organ disease such as CVD.

Our aims were to characterize hepatic and nonhepatic CVD biomarker and macrophage activation levels in a HIV/HCV-coinfected cohort and explore the potential benefit of HCV treatment on cardiovascular outcomes, hypothesizing that HCV clearance would be associated with a favorable reduction in nonhepatic CVD biomarkers and markers of macrophage activation.

METHODS

Study Design

We conducted a retrospective case control study analyzing stored serum and plasma samples and clinical data from HIV/HCV-coinfected subjects who participated in the AIDS Clinical Trials Group study A5178, a study of maintenance PEG for reducing fibrosis progression [24]. All subjects included in this analysis achieved an early virologic response (defined as ≥2 log decrease in or undetectable HCV RNA at 12 weeks of treatment) and were assigned to an extended treatment arm that was offered a 72-week course of PEG/RBV. Serum and plasma samples from before treatment at study entry (baseline) and from 24 weeks after end of treatment (EOT+24 weeks) were tested for nonhepatic (sICAM-1, sP-selectin, IL-6, d-dimer, Lp-PLA2) and hepatic (hsCRP and lipids) CVD risk biomarkers and markers of macrophage activation (sCD163 and sCD14). Included were 54 subjects with available stored serum and plasma, 27 with SVR (defined as HCV RNA <60 IU/mL at 24 weeks after EOT) and 27 nonresponders/relapers (non-SVR), matched simultaneously on race, ethnicity and sex, to exclude their potential confounding [25].

Clinical Data

Baseline and on-treatment data abstracted from the original study dataset included: age; gender; race and ethnicity; presence or absence of hypertension and diabetes; concomitant medications; body mass index; intravenous drug use history; fasting glucose and insulin; aspartate aminotransferase (AST) and alanine aminotransferase (ALT) levels; comorbid disease; HCV treatment history; HCV RNA levels at baseline, EOT, and 24 weeks after EOT; HCV genotype; hepatic fibrosis scores by liver biopsy (scoring systems included Knodell, Ludwig, META-VIR, Modified HAI, and Scheuer); CD4 cell count; and HIV RNA level. The homeostatic model assessment of insulin resistance (HOMA-IR) was calculated as fasting glucose (mg/dL) × fasting insulin (µU/mL)/405.

Biomarkers

Biomarker testing of stored serum and EDTA plasma specimens was performed at a central laboratory. Serum sICAM-1 and plasma sP-selectin were measured by enzyme-linked immunosorbent assay (ELISA) (SearchLight Custom Human ICAM-1 and P-Selectin Assays, Pierce Biotechnology, Inc.); plasma Lp-PLA2 mass was measured by ELISA (PLAC Test, diaDexus, Inc.); plasma d-dimer was measured by immuno-turbidimetric assay (STA-Liatest D-DI, Diagnostica Stago); serum IL-6 was measured by ELISA (Quantikine HS ELISA, R&D Systems, Inc.); plasma sCD163 and sCD14 were measured by ELISA.
(Macro163, Trillium Diagnostics, LLC and Quantikine Human sCD14 ELISA, R&D Systems, Inc.); serum hsCRP was measured by immunonephelometry (CardioPhase hsCRP, Siemens Healthcare Diagnostics, Inc.); and serum lipid panel was measured by automated enzymatic and automated spectrophotometry (Beckman Coulter analyzer). Forty-one of the 54 available subjects had fasting lipids, insulin, and glucose levels available from the parent ACTG study for analysis. A lipid panel was repeated on all subjects, including nonfasting lipids on the 13 subjects without fasting samples.

### Table 1. Baseline Characteristics of the Study Cohort

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Overall (n = 54) Median (Q1, Q3) or n (%)</th>
<th>SVR (n = 27) Median (Q1, Q3) or n (%)</th>
<th>Non-SVR (n = 27) Median (Q1, Q3) or n (%)</th>
<th>P Value (SVR vs Non-SVR)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>48 (44, 52)</td>
<td>47 (42, 52)</td>
<td>48 (45, 52)</td>
<td>.069</td>
</tr>
<tr>
<td>Sex: Male</td>
<td>44 (81)</td>
<td>22 (81)</td>
<td>22 (81)</td>
<td>N/A (matched)</td>
</tr>
<tr>
<td>Race</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>White</td>
<td>30 (56)</td>
<td>15 (56)</td>
<td>15 (56)</td>
<td>N/A (matched)</td>
</tr>
<tr>
<td>Black</td>
<td>24 (44)</td>
<td>12 (44)</td>
<td>12 (44)</td>
<td></td>
</tr>
<tr>
<td>Body mass index</td>
<td>25.9 (23.8, 29.5)</td>
<td>26.1 (23.2, 30.0)</td>
<td>25.5 (24.1, 28.5)</td>
<td>.592</td>
</tr>
<tr>
<td>HIV-1 RNA Undetectable (&lt;50 copies/mL)</td>
<td>42 (78)</td>
<td>20 (74)</td>
<td>22 (81)</td>
<td>.745</td>
</tr>
<tr>
<td>CD4 cell count</td>
<td>544 (370, 734)</td>
<td>571 (378, 747)</td>
<td>536 (357, 734)</td>
<td>.616</td>
</tr>
<tr>
<td>HCV genotype</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>41 (76)</td>
<td>18 (87)</td>
<td>23 (85)</td>
<td>.145</td>
</tr>
<tr>
<td>2</td>
<td>9 (17)</td>
<td>7 (26)</td>
<td>2 (7)</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>3 (6)</td>
<td>2 (7)</td>
<td>1 (4)</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>1 (2)</td>
<td>0 (0)</td>
<td>1 (4)</td>
<td></td>
</tr>
<tr>
<td>Cirrhosis</td>
<td>2 (4)</td>
<td>0 (0)</td>
<td>2 (7)</td>
<td>.491</td>
</tr>
<tr>
<td>Fibrosis stage ≥3</td>
<td>18 (33)</td>
<td>5 (19)</td>
<td>13 (48)</td>
<td>.042</td>
</tr>
<tr>
<td>HCV RNA (log₁₀ copies/mL)</td>
<td>6.60 (6.10, 6.98)</td>
<td>6.41 (5.73, 6.92)</td>
<td>6.70 (6.31, 7.04)</td>
<td>.084</td>
</tr>
<tr>
<td>History of prior HCV treatment</td>
<td>15 (28)</td>
<td>4 (15)</td>
<td>11 (41)</td>
<td>.066</td>
</tr>
<tr>
<td>History of CVD</td>
<td>7 (13)</td>
<td>3 (11)</td>
<td>4 (15)</td>
<td>1.000</td>
</tr>
<tr>
<td>Hypertension</td>
<td>14 (26)</td>
<td>5 (19)</td>
<td>9 (33)</td>
<td>.352</td>
</tr>
<tr>
<td>Diabetes</td>
<td>7 (13)</td>
<td>2 (7)</td>
<td>5 (19)</td>
<td>.420</td>
</tr>
<tr>
<td>History of injection drug use</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Current</td>
<td>1 (2)</td>
<td>1 (4)</td>
<td>0 (0)</td>
<td>.785</td>
</tr>
<tr>
<td>Former</td>
<td>30 (56)</td>
<td>14 (52)</td>
<td>16 (59)</td>
<td></td>
</tr>
<tr>
<td>Never</td>
<td>23 (43)</td>
<td>12 (44)</td>
<td>11 (41)</td>
<td></td>
</tr>
<tr>
<td>HOMA-IR (SVR, n = 22; non-SVR, n = 16)</td>
<td>4.00 (1.65, 6.27)</td>
<td>2.97 (1.43, 4.49)</td>
<td>6.11 (3.73, 8.77)</td>
<td>.007</td>
</tr>
<tr>
<td>AST (U/L)</td>
<td>50 (33, 72)</td>
<td>41 (30, 60)</td>
<td>63 (42, 85)</td>
<td>.007</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>64 (43, 79)</td>
<td>58 (37, 73)</td>
<td>68 (51, 91)</td>
<td>.045</td>
</tr>
<tr>
<td>Antiretroviral therapy (ART)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Unboosted protease inhibitor</td>
<td>15 (28)</td>
<td>9 (33)</td>
<td>6 (22)</td>
<td>.498</td>
</tr>
<tr>
<td>Ritonavir-boosted protease inhibitor</td>
<td>13 (24)</td>
<td>5 (19)</td>
<td>8 (30)</td>
<td></td>
</tr>
<tr>
<td>No protease inhibitor</td>
<td>19 (35)</td>
<td>11 (41)</td>
<td>8 (30)</td>
<td></td>
</tr>
<tr>
<td>No ART</td>
<td>1 (2)</td>
<td>0 (0)</td>
<td>1 (3)</td>
<td></td>
</tr>
<tr>
<td>Unknown</td>
<td>6 (11)</td>
<td>2 (7)</td>
<td>4 (15)</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase; CVD, cardiovascular disease; HCV, hepatitis C virus; HIV, human immunodeficiency virus; HOMA-IR, homeostatic model assessment of insulin resistance; N/A, not applicable; SVR, sustained virologic response.

* Categorical variables compared by Fisher exact test, continuous variables by Wilcoxon test.

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**Data Analysis**

The primary outcome was the change in sICAM-1 levels from baseline to 24 weeks after EOT. Secondary outcomes were changes in sP-selectin, IL-6, d-dimer, Lp-PLA₂, sCD163, sCD14, total cholesterol, high-density lipoprotein cholesterol, LDL, and triglyceride levels between the same timepoints. Biomarkers below the limit of detection were assigned the lowest level of detection. Analyses of lipid levels were done separately for fasting lipids collected during A5178 and combining retesting of fasting samples and nonfasting samples. Lp-PLA₂ levels
were analyzed as a continuous variable and categorized into high (>235 ng/mL), intermediate (200–235 ng/mL), or low predicted CVD risk (<200 ng/mL). Liver fibrosis scores were dichotomized as <3 or ≥3, regardless of scoring method. Baseline characteristics and biomarker levels between the SVR and non-SVR groups were compared by Fisher’s exact test for categorical variables and Wilcoxon rank-sum test for continuous variables. Log10 transformations were utilized for biomarkers with skewed distribution. Associations between biomarkers and baseline variables were examined by simple linear regressions. Within each group, change in each biomarker from baseline to 24 weeks after EOT was compared to zero by paired sample test or Wilcoxon signed-rank test and between groups by two-sample t test or Wilcoxon test. Linear regressions for the association between SVR and change in each biomarker were performed, adjusting for (1) sex and race or ethnicity to account for the matching in study design, (2) baseline variables that were significantly different by SVR status at a 0.10 significance level with a backward variable selection procedure, (3) change in ALT from baseline to 24 weeks after EOT, (4) duration of PEG/ RBV treatment, (5) occurrence of serious infections on study, and (6) concomitant medication use (grouped a priori as immunomodulators, antiplatelet/aspirin, antihypertensives, nonsteroidal anti-inflammatory drugs, and lipid-lowering agents), to examine the potential confounding effect of these covariates. Sensitivity analyses excluding subjects with use of these medications were also conducted to evaluate their effect on the results of analyses. Given concern that HIV viremia might confound biomarker levels, sensitivity analyses were also conducted limiting analyses to those with documented HIV virologic suppression (<50 copies/mL) for the entire duration of the study. Given the expected reduction in power with sensitivity analysis, parameter estimates in these subsets were examined for large shifts.

### Sample Size Determination

The study was powered to detect a difference in change in sICAM-1 between the SVR and non-SVR groups. We anticipated a decrease of 45 ng/mL in the change of sICAM-1 from baseline to 24 weeks after EOT in SVRs compared with no change for non-SVRs, the difference associated with improved cardiovascular outcome was reported by Hwang et al [25]. Assuming a standard deviation (SD) of 40.4 for both SVR and non-SVR groups, 20% correlation between baseline and 24 weeks after EOT, and 20% reduction in the SD by matching on race or ethnicity and sex, 54 subjects provided 95% power to detect the projected difference in change in sICAM-1.

### Institutional Review Board Approval

Informed consent was obtained from participants for the parent A5178 study, including use of stored samples for research testing. The current analysis was reviewed and approved by the University of California, Los Angeles Institutional Review Board.

### RESULTS

Baseline characteristics of the cohort are summarized in Table 1. Median age overall was 48 years (interquartile range [IQR], 44–52). Forty-four of the 54 subjects were male, 30 were white, 24 were black, and all were non-Hispanic. Median CD4 cell count was similar between the groups, and the majority (78%) had HIV-1 RNA <50 copies/mL at baseline. Antiretroviral therapy did not differ between the groups (P = .498) (Table 1). The

### Table 2. Baseline Biomarker Levels by SVR Status

<table>
<thead>
<tr>
<th>Biomarker</th>
<th>Overall (n = 54)</th>
<th>SVR (n = 27)</th>
<th>Non-SVR (n = 27)</th>
<th>P Value (SVR vs Non-SVR)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>sICAM-1 (ng/mL)</td>
<td>439.2 (365.6, 592.8)</td>
<td>428.6 (355.1, 478.8)</td>
<td>476.6 (366.9, 744.5)</td>
<td>.156</td>
</tr>
<tr>
<td>sP-selectin (ng/mL)</td>
<td>146.7 (94.1, 209.9)</td>
<td>153.6 (110.8, 225.9)</td>
<td>146.4 (89.4, 170.6)</td>
<td>.276</td>
</tr>
<tr>
<td>IL-6 (pg/mL)</td>
<td>2.32 (1.61, 3.49)</td>
<td>2.43 (1.56, 3.60)</td>
<td>2.32 (1.61, 3.49)</td>
<td>.943</td>
</tr>
<tr>
<td>Lp-PLA2 (ng/mL)</td>
<td>312.5 (234.5, 382.5)</td>
<td>315.5 (231.0, 394.0)</td>
<td>297.5 (235.0, 371.0)</td>
<td>.934</td>
</tr>
<tr>
<td>sCD14 (ng/mL)</td>
<td>2464 (1522, 3480)</td>
<td>2056 (1172, 3006)</td>
<td>2933 (2094, 4129)</td>
<td>.013</td>
</tr>
<tr>
<td>Total cholesterol (mg/dL)</td>
<td>168 (139, 192)</td>
<td>188 (169, 222)</td>
<td>153 (135, 167)</td>
<td>.002</td>
</tr>
<tr>
<td>LDL (mg/dL)</td>
<td>96 (76, 124)</td>
<td>112 (85, 125)</td>
<td>81 (68, 108)</td>
<td>.015</td>
</tr>
<tr>
<td>Triglycerides (mg/dL)</td>
<td>143 (102, 225)</td>
<td>144 (107, 243)</td>
<td>141 (97, 215)</td>
<td>.665</td>
</tr>
<tr>
<td>HDL (mg/dL)</td>
<td>36 (31, 48)</td>
<td>38 (31, 50)</td>
<td>36 (31, 46)</td>
<td>.545</td>
</tr>
<tr>
<td>hsCRP (mg/L)</td>
<td>1.10 (0.70, 2.60)</td>
<td>1.15 (0.70, 3.20)</td>
<td>1.00 (0.60, 2.60)</td>
<td>.476</td>
</tr>
</tbody>
</table>

Abbreviations: HDL, high-density lipoprotein cholesterol; hsCRP, high-sensitivity C-reactive protein; IL-6, interleukin-6; LDL, low-density lipoprotein cholesterol; Lp-PLA2, lipoprotein-associated phospholipase A2; sCD14, soluble CD14; sCD163, soluble CD163; sICAM-1, soluble ICAM-1; sP-selectin, soluble P-selectin; SVR, sustained virologic response.

* Wilcoxon rank-sum test.

† One subject in each group missing Lp-PLA2, one subject in SVR group missing IL-6, one subject in SVR group missing sCD163 and sCD14.
HOMA-IR, AST, and ALT levels were significantly higher in non-SVR subjects at baseline (see Table 1). Few subjects (3 in the SVR group, 4 in the non-SVR group) had known CVD at baseline. Only 2 of the 54 subjects, both in the non-SVR group, had cirrhosis; 5 (19%) of the SVR group and 13 (48%) of the non-SVR group had significant fibrosis, with a fibrosis
alanine aminotransferase (ALT) level. (B) Regression of change in log10 or experienced), baseline aspartate aminotransferase (AST), and baseline hepatitis C virus (HCV) RNA level, HCV treatment history (naive included in backward variable selection: baseline hepatic molecule-1 (sICAM-1) after backward variable selection. Variables in separation of sICAM-1 with backward variable selection and adjusting for change in Figure 2. (A) Regression of change in log10 soluble intercellular adhesion molecule-1 (sICAM-1) after backward variable selection. Variables included in backward variable selection: baseline hepatic fibrosis stage, baseline hepatitis C virus (HCV) RNA level, HCV treatment history (naive or experienced), baseline aspartate aminotransferase (AST), and baseline alanine aminotransferase (ALT) level. (B) Regression of change in log10 sICAM-1 with backward variable selection and adjusting for change in ALT level from baseline to 24 weeks after EOT. 

score of at least 3. At baseline, 19 (35%) subjects were taking antihypertensive agents, 5 (9%) were taking antiplatelet medications including aspirin, 24 (44%) were taking nonsteroidal anti-inflammatory drugs (NSAIDs), 4 (7%) were taking 3-hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors (statins), and 4 (7%) were taking other lipid-lowering agents. Twenty-four subjects (44%) received immunomodulators on study, primarily granulocyte colony-stimulating factor. Median duration of PEG/RBV treatment for the SVR and non-SVR groups was 71.9 (IQR, 71.0–72.1) and 71.4 (IQR, 47.3–72.3) weeks. Of 48 subjects with baseline ART information available, 38 (79%; 21 of 25 with SVR and 17 of 25 non-SVRs) did not change their ART while on study. No subjects initiated ART while on study. No subjects reported a CVD event during the course of the study. Two subjects, both in the non-SVR group, developed diabetes during the study. Eight subjects (15%) experienced a serious infection (3 SVR, 5 non-SVR), including bacterial pneumonia, sepsis, pulmonary histoplasmosis, and cutaneous varicella zoster.

Baseline (pretreatment) levels of the biomarkers by SVR status are shown in Table 2. Overall, levels of the nonhepatic biomarkers (sICAM-1, sP-selectin, IL-6, and Lp-PLA2) and macrophage activation markers were high. Thirty-seven of 52 (71%) subjects with available Lp-PLA2 measures had Lp-PLA2 level >235 ng/mL. Of note, 24 (44%) subjects had undetectable D-dimer (<0.22 mcg/mL) at baseline (13 SVR, 11 non-SVR). Biomarker levels were similar between the SVR and non-SVR groups for all biomarkers except sCD163, total cholesterol, and LDL. Baseline sCD163 levels were significantly lower in the SVR group (median, 2056 ng/mL [IQR, 1172–3006] vs 2933 ng/mL [IQR, 2094–4129]; P = .013) and were positively associated with fibrosis stage (P < .001) and baseline ALT (P < .001) in univariate analysis. Adjusting for fibrosis, there was no significant difference in baseline sCD163 by SVR status (P = .094). Subjects with a fibrosis score of 3 or greater also had a significantly lower total cholesterol level (median, 157 mg/dL [IQR, 135–157]) than those with less fibrosis (177 mg/dL [IQR, 146–207]; P = .034). Likewise, LDL level was lower among subjects with higher fibrosis score (86 mg/dL [IQR, 51–109] vs 103 mg/dL [IQR, 78–124]), but the difference did not reach statistical significance (P = .130).

Box plots depicting the change in biomarker levels by SVR status are provided in Figure 1 and Supplementary Figure 1. Supplementary Figure 2 provides spaghetti plots of the within-subject change in biomarker levels. Levels of sICAM-1 decreased significantly from week 0 to 24 weeks after EOT in the SVR group (mean change in log10 sICAM-1 = −0.09 [SD = 0.13]; P < .001), but they remained unchanged in the non-SVR group (mean change in log10 sICAM-1 = −0.01 [SD = 0.14]; P = .632). The change in sICAM-1 comparing SVR and non-SVR groups was statistically significant (P = .047). In regression analysis controlling for sex and race, SVR status was significantly associated with decrease in log10 sICAM-1 (P = .042). After a backward variable selection procedure including liver fibrosis stage, HCV RNA level, HCV treatment history, and baseline AST and ALT level, which differed between the SVR and non-SVR groups at baseline, only baseline AST and ALT were retained, and SVR status remained statistically significantly associated with decrease in log10 sICAM-1 (P = .033) (Figure 2A). After further adjusting for change in ALT from baseline to 24 weeks after EOT, SVR status was no longer associated with the change in log10 sICAM-1 (P = .105) (Figure 2B). Change in ALT was significantly associated with change in log10 sICAM-1 (P = .021). There was no association between duration of PEG/RBV treatment and change in sICAM-1.

On-study occurrence of serious infections was not significantly associated with SVR status (P = .365). Adjusting for occurrence of serious infection, change in log10 sICAM-1 remained significantly associated with SVR (P = .025). Because more than 20% of subjects received medications that may have affected biomarker levels (24 with immunomodulators, 26 with antihypertensives, and 32 with NSAIDS), sensitivity analyses were conducted excluding these subjects. Statistically significant between-group differences in log10 sICAM-1 persisted with exclusion of subjects with antihypertensive use. No statistically
significant between-group differences were seen after excluding subjects with immunomodulator use (n = 25) or with NSAID use (n = 20). Fourteen SVR subjects and 8 non-SVR subjects had documented HIV virologic suppression for the entire PEG/RBV treatment period. Restricting analyses to these subjects, the mean change in log_{10} sICAM-1 among SVRs was −0.09 (SD = 0.12) compared with 0.02 (SD = 0.17) among non-SVRs (within group \( P = .015 \) and \( P = .750 \); between-group \( P = .129 \)).

Soluble CD163 levels decreased significantly from week 0 to 24 weeks after EOT in SVRs (mean change in log_{10} sCD163 = −0.11 [SD = 0.19]; \( P = .009 \)) but not in non-SVRs (mean change in log_{10} sCD163 = −0.04 [SD = 0.15]; \( P = .205 \)). After backward variable selection, only baseline AST, in addition to race and sex, were retained. Sustained virologic response remained significantly associated with change in log_{10} sCD163 (\( P = .042 \)) (Figure 3A). Adjusting further for change in ALT resulted in loss of the association between SVR status and change in log_{10} sCD163 (\( P = .969 \)), as was seen with sICAM-1 models; in this model, change in ALT was significantly associated with change in sCD163 (\( P = .002 \)) (Figure 3B). With further adjustment for duration of PEG/RBV, SVR and duration of PEG/RBV were associated with change in sCD163, but statistical significance was not reached (\( P = .071 \) and \( P = .052 \), respectively) (Figure 3C).

At baseline, there was no difference in CVD risk class by Lp-PLA2 level, whereas after PEG/RBV treatment, Lp-PLA2 levels decreased, although not statistically significantly, among SVRs (median change = −17.5 ng/mL [IQR, −67.0 to 47.0]) but not non-SVRs (median change = 9.50 ng/mL [IQR, −49.0 to 67.0]). There was a statistically significant difference in CVD risk class...
distribution by Lp-PLA2 level at 24 weeks after EOT (Figure 4), with 25 of 27 (93%) of the non-SVR group in the high-risk category compared with 17 of 27 (63%) of the SVR group ($P = .021$). There were no statistically significant differences in the other CVD biomarkers comparing SVR and non-SVR groups.

**DISCUSSION**

Prior to HCV treatment, subjects had high levels of the nonhepatic CVD biomarkers sICAM-1, sP-selectin, IL-6, and Lp-PLA2, exceeding levels in HIV-uninfected persons at high risk for CVD or with known baseline CVD [25–29], suggesting higher CVD risk in HIV/HCV co-infection. Levels of sICAM-1, sP-selectin, and IL-6 in our analysis were consistent with higher CVD risk in HIV/HCV coinfection. Levels of sICAM-1, and it is a new observation for sCD163 [36, 37]. The association of sCD163 with sICAM-1 and sCD163 no longer exist- ed after adjusting for change in ALT, suggesting that the effect of SVR on sICAM-1 and sCD163 levels may be mediated by reduction in hepatic inflammation. This result is a distinct and new finding from other studies that have explored the effect of HCV treatment and eradication on biomarkers such as sICAM-1, and it is a new observation for sCD163 [36, 37].

The magnitude of the change in sICAM-1 observed with HCV clearance was greater than the difference in sICAM-1 observed in studies comparing subjects who went on to have a coronary disease event and those who did not [25, 35]. The association of SVR with sICAM-1 and sCD163 no longer exist- ed after adjusting for change in ALT, suggesting that the effect of SVR on sICAM-1 and sCD163 levels may be mediated by reduction in hepatic inflammation. This result is a distinct and new finding from other studies that have explored the effect of HCV treatment and eradication on biomarkers such as sICAM-1, and it is a new observation for sCD163 [36, 37].

Also suggestive is that SVR was associated with stable Lp-PLA2 levels over 96 weeks, whereas non-SVRs had a shift to higher CVD risk class by Lp-PLA2 level. It is possible that HCV vireologic clearance attenuates atherosclerotic disease progression in persons infected with HIV. We observed that hepatic markers such as cholesterol in chronic HCV infection, there may be a distinct role for these biomarkers in HIV/HCV-coinfected or HCV-monoinfected persons.

Levels of sCD163 in our study were higher than those seen in cohorts of predominantly HIV-infected persons on ART with significant subclinical atherosclerotic disease [18, 19] and levels of sCD14 similar to those seen in studies of HIV-monoinfected persons, where sCD14 levels predicted subclinical atherosclerotic disease progression [21]. We found sCD163 levels, but not sCD14 levels, were associated with degree of hepatic fibrosis and baseline ALT (greater fibrosis and higher ALT with higher sCD163 levels). Different results observed between sCD163 and sCD14 suggest differential sensitivity of these 2 macrophage activation markers to hepatic inflammation and extent of liver disease. Interpretation of sCD163 levels in HIV-infected persons must take into account degree of liver disease, which may be confounding. Our analyses also support a complex relationship among chronic viral coinfection, liver disease, and immune dysregulation with systemic immune activation.

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Liver-related deaths alone do not explain excess mortality in HIV/HCV-coinfected patients on effective ART. Chronic and high levels of viral replication and possibly increased gut permeability and microbial translocation/endotoxemia are associated
with systemic immune activation in HIV, and they are thought to be major drivers of HIV-1 disease progression. Hepatitis C virus coinfection and ongoing HCV viremia may augment such immune activation, complicating the interaction between HIV and host innate immune response. Impaired lipopolysaccharide tolerance in the setting of chronic HCV infection may lead to chronic intrahepatic monocyte and macrophage activation, hepatic inflammation, and further systemic immune activation and inflammation [38]. As such, it may be that HCV acts additively, or even synergistically, to drive immune activation and extrahepatic complications. Treatment and clearance of HCV may reduce hepatic and, subsequently, systemic inflammation and its associated complications. This finding is supported by several studies to date describing decreased all-cause and nonhepatic mortality with SVR in HCV-monoinfected and coinfected patients [9, 39, 40].

Limitations of the study include its retrospective nature with use of frozen samples for which the stability of all the assays has not been confirmed; lack of smoking history, which is relevant to the outcomes of interest, although we would not expect there to be between-group differences in smoking status that would affect biomarker levels or changes; inclusion of a select group of interferon-responsive individuals, such that generalizability may be limited; and possible confounding by HIV viremia or concomitant medication use, which we explored in sensitivity analyses (finding similar point estimates for change in biomarker levels, with substantially reduced sample size likely limiting our power to find statistically significant differences between the groups). The study was powered to detect a significant change in sICAM-1 levels, and it may have been underpowered for evaluation of the other biomarkers. Interpretation of the favorable changes in sICAM-1 and Lp-PLA₂ is limited given the small size of our cohort, lack of validation of these biomarkers for CVD risk prognostication in HIV/HCV coinfection, and inability to correlate biomarker levels with hard outcomes such as CVD events or validated surrogate measures of cardiovascular risk.

Despite these limitations, our data remain suggestive and further investigation into characterizing the predictive utility of both nonhepatic and hepatic CVD biomarkers and quantifying the potential benefit of HCV treatment on nonliver outcomes such as CVD is warranted. Today, aging HIV-infected patients on effective ART are at increasing risk for non-AIDS complications including CVD, which may be further accelerated by HCV coinfection. Successful HCV treatment may be a viable method for CVD risk reduction and an additional indication for earlier HCV treatment in HIV/HCV-coinfected persons.

**Supplementary Material**

Supplementary material is available at Open Forum Infectious Diseases (http://OpenForumInfectiousDiseases.oxfordjournals.org/).

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