



Ancient human genomes suggest three ancestral populations for present-day Europeans

Citation

Lazaridis, I., N. Patterson, A. Mitnik, G. Renaud, S. Mallick, K. Kirsanow, P. H. Sudmant, et al. 2014. "Ancient human genomes suggest three ancestral populations for present-day Europeans." *Nature* 513 (7518): 409-413. doi:10.1038/nature13673. <http://dx.doi.org/10.1038/nature13673>.

Published Version

doi:10.1038/nature13673

Permanent link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:14351136>

Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA>

Share Your Story

The Harvard community has made this article openly available.
Please share how this access benefits you. [Submit a story](#).

[Accessibility](#)

Published in final edited form as:

Nature. 2014 September 18; 513(7518): 409–413. doi:10.1038/nature13673.

Ancient human genomes suggest three ancestral populations for present-day Europeans

A full list of authors and affiliations appears at the end of the article.

Abstract

We sequenced the genomes of a ~7,000 year old farmer from Germany and eight ~8,000 year old hunter-gatherers from Luxembourg and Sweden. We analyzed these and other ancient genomes^{1–4} with 2,345 contemporary humans to show that most present Europeans derive from at least three highly differentiated populations: West European Hunter-Gatherers (WHG), who contributed ancestry to all Europeans but not to Near Easterners; Ancient North Eurasians (ANE) related to Upper Paleolithic Siberians³, who contributed to both Europeans and Near Easterners; and Early European Farmers (EEF), who were mainly of Near Eastern origin but also harbored WHG-related ancestry. We model these populations' deep relationships and show that EEF had ~44% ancestry from a “Basal Eurasian” population that split prior to the diversification of other non-African lineages.

Near Eastern migrants played a major role in the introduction of agriculture to Europe, as ancient DNA indicates that early European farmers were distinct from European hunter-gatherers^{4,5} and close to present-day Near Easterners^{4,6}. However, modelling present-day Europeans as a mixture of these two ancestral populations⁴ does not account for the fact that they are also admixed with a population related to Native Americans^{7,8}. To clarify the prehistory of Europe, we sequenced nine ancient genomes (Fig. 1A; Extended Data Fig. 1): “Stuttgart” (19-fold coverage), a ~7,000 year old skeleton found in Germany in the context of artifacts from the first widespread farming culture of central Europe, the *Linearbandkeramik*; “Loschbour” (22-fold), an ~8,000 year old skeleton from the

†Correspondence and requests for materials should be addressed to David Reich (reich@genetics.med.harvard.edu) or Johannes Krause (johannes.krause@uni-tuebingen.de).

^{5,3}Currently employed by AMGEN; 33 Kazantzaki Str, Ilioupolis 16342, Athens, Greece

^{8,3}Present address: Banaras Hindu University, Varanasi, 221 005, India

Supplementary Information is linked to the online version of the paper at www.nature.com/nature. The fully public version of the Human Origins dataset can be found at http://genetics.med.harvard.edu/reichlab/Reich_Lab/Datasets.html. The full version of the dataset (including additional samples) is available to researchers who send a signed letter to DR indicating that they will abide by specified usage conditions (SI9).

Author contributions

BB, EEE, JBu, MS, SP, JKe, DR and JKr supervised the study. IL, NP, AM, GR, SM, KK, PHS, JGS, SC, ML, QF, HL, CdF, KP, WH, MMeT, MMey and DR analyzed genetic data. FH, EF, DD, MF, J-MG, JW, AC and JKr obtained human remains. AM, CE, RBo, KB, SS, CP, NR and JKr processed ancient DNA. IL, NP, SN, NR, GA, HAB, GBa, EB, OB, RBa, GBe, HB-A, JBe, FBe, CMB, FBr, GBJB, FC, MC, DECC, DCoR, LD, GvD, SD, J-MD, SAF, IGR, MG, MH, BH, TH, UH, ARJ, SK-Y, RKH, EK, RKi, TK, WK, VK, AK, LL, SL, TL, RWM, BM, EM, JMol, JMou, KN, DN, TN, LO, JP, FP, OLP, VR, FR, IR, RR, HS, ASaj, ASal, EBS, ATar, DT, ST, IU, OU, RVa, MVi, MVo, CW, LY, PZ, TZ, CC, MGT, AR-L, SAT, LS, KT, RVi, DCom, RS, MMeT, SP and DR assembled the genotyping dataset. IL, NP, DR and JKr wrote the manuscript with help from all co-authors.

The aligned sequences are available through the European Nucleotide Archive under accession number PRJEB6272.

The authors declare competing financial interests: UH is an employee of Illumina, TL is an employee of AMGEN, and JM is an employee of 23andMe.

Loschbour rock shelter in Luxembourg, discovered in the context of hunter-gatherer artifacts (SI1; SI2); and seven ~8,000 year old samples (0.01–2.4-fold) from a hunter-gatherer burial in Motala, Sweden (the highest coverage individual was “Motala12”).

Sequence reads from all samples revealed >20% C→T and G→A deamination-derived mismatches at the ends of the molecules that are characteristic of ancient DNA^{9,10} (SI3). We estimate nuclear contamination rates to be 0.3% for Stuttgart and 0.4% for Loschbour (SI3), and mitochondrial (mtDNA) contamination rates to be 0.3% for Stuttgart, 0.4% for Loschbour, and 0.01–5% for the Motala individuals (SI3). Stuttgart has mtDNA haplogroup T2, typical of Neolithic Europeans¹¹, and Loschbour and all Motala individuals have the U5 or U2 haplogroups, typical of hunter-gatherers^{5,9} (SI4). Stuttgart is female, while Loschbour and five Motala individuals are male (SI5) and belong to Y-chromosome haplogroup I, suggesting that this was common in pre-agricultural Europeans (SI5).

We carried out large-scale sequencing of libraries prepared with uracil DNA glycosylase (UDG), which removes deaminated cytosines, thus reducing errors arising from ancient DNA damage (SI3). The ancient individuals had indistinguishable levels of Neanderthal ancestry when compared to each other (~2%) and to present-day Eurasians (SI6). The heterozygosity of Stuttgart (0.00074) is at the high end of present-day Europeans, while that of Loschbour (0.00048) is lower than in any present humans (SI2), reflecting a strong bottleneck in Loschbour’s ancestors as the genetic data show that he was not recently inbred (Extended Data Fig. 2). High copy numbers for the salivary amylase gene (*AMY1*) have been associated with a high starch diet¹²; our data are consistent with this finding in that the ancient hunter gatherers La Braña (from Iberia)², Motala12, and Loschbour had 5, 6 and 13 copies respectively, whereas the Stuttgart farmer had 16 (SI7). Both Loschbour and Stuttgart had dark hair (>99% probability); and Loschbour, like La Braña and Motala12, likely had blue or intermediate-colored eyes (>75%) while Stuttgart likely had brown eyes (>99%) (SI8). Neither Loschbour nor La Braña carries the skin-lightening allele in *SLC24A5* that is homozygous in Stuttgart and nearly fixed in Europeans today², but Motala12 carries at least one copy of the derived allele, showing that this allele was present in Europe prior to the advent of agriculture.

We compared the ancient genomes to 2,345 present-day humans from 203 populations genotyped at 594,924 autosomal single nucleotide polymorphisms (SNPs) with the Human Origins array⁸ (SI9) (Extended Data Table 1). We used ADMIXTURE¹³ to identify 59 “West Eurasian” populations that cluster with Europe and the Near East (SI9 and Extended Data Fig. 3). Principal component analysis (PCA)¹⁴ (SI10) (Fig. 1B) indicates a discontinuity between the Near East and Europe, with each showing north-south clines bridged only by a few populations of mainly Mediterranean origin. We projected¹⁵ the newly sequenced and previously published^{1–4} ancient genomes onto the first two principal components (PCs) (Fig. 1B). Upper Paleolithic hunter-gatherers³ from Siberia like the MA1 (Mal’ta) individual project at the northern end of the PCA, suggesting an “Ancient North Eurasian” meta-population (ANE). European hunter-gatherers from Spain², Luxembourg, and Sweden⁴ fall beyond present-day Europeans in the direction of European differentiation from the Near East, and form a “West European Hunter-Gatherer” (WHG) cluster including Loschbour and La Braña², and a “Scandinavian Hunter-Gatherer” (SHG) cluster including

the Motala individuals and ~5,000 year old hunter-gatherers from the Pitted Ware Culture⁴. An “Early European Farmer” (EEF) cluster includes Stuttgart, the ~5,300 year old Tyrolean Iceman¹ and a ~5,000 year old Swedish farmer⁴.

Patterns observed in PCA may be affected by sample composition (SI10) and their interpretation in terms of admixture events is not straightforward, so we rely on formal analysis of f -statistics⁸ to document mixture of at least three source populations in the ancestry of present Europeans. We began by computing all possible statistics of the form $f_3(\text{Test}; \text{Ref}_1, \text{Ref}_2)$ (SI11), which if significantly negative show unambiguously⁸ that *Test* is admixed between populations anciently related to Ref_1 and Ref_2 (we choose Ref_1 and Ref_2 from 5 ancient and 192 present populations). The lowest f_3 -statistics for Europeans are negative (93% are >4 standard errors below 0), with most showing strong support for at least one ancient individual being one of the references (SI11). Europeans almost always have their lowest f_3 with either (EEF, ANE) or (WHG, Near East) (SI11, Table 1, Extended Data Table 1), which would not be expected if there were just two ancient sources of ancestry (in which case the best references for all Europeans would be similar). The lowest f_3 -statistic for Near Easterners always takes Stuttgart as one of the reference populations, consistent with a Near Eastern origin for Stuttgart’s ancestors (Table 1). We also computed the statistic $f_4(\text{Test}, \text{Stuttgart}; \text{MA1}, \text{Chimp})$, which measures whether MA1 shares more alleles with a *Test* population or with Stuttgart. This statistic is significantly positive (Extended Data Fig. 4, Extended Data Table 1) if *Test* is nearly any present-day West Eurasian population, showing that MA1-related ancestry has increased since the time of early farmers like Stuttgart (the analogous statistic using Native Americans instead of MA1 is correlated but smaller in magnitude (Extended Data Fig. 5), indicating that MA1 is a better surrogate than the Native Americans who were first used to document ANE ancestry in Europe^{7,8}). The analogous statistic $f_4(\text{Test}, \text{Stuttgart}; \text{Loschbour}, \text{Chimp})$ is nearly always positive in Europeans and negative in Near Easterners, indicating that Europeans have more ancestry from populations related to Loschbour than do Near Easterners (Extended Data Fig. 4, Extended Data Table 1). Extended Data Table 2 documents the robustness of key f_4 -statistics by recomputing them using transversion polymorphisms not affected by ancient DNA damage, and also using whole-genome sequencing data not affected by SNP ascertainment bias. Extended Data Fig. 6 shows the geographic gradients in the degree of allele sharing of present-day West Eurasians (as measured by f_4 -statistics) with Stuttgart (EEF), Loschbour (WHG) and MA1 (ANE).

To determine the minimum number of source populations needed to explain the data for many European populations taken together, we studied the matrix of all possible statistics of the form $f_4(\text{Test}_{base}, \text{Test}_i; \text{O}_{base}, \text{O}_j)$ (SI12). Test_{base} is a reference European population, Test_i is the set of all other European *Test* populations, O_{base} is a reference outgroup, and O_j is the set of other outgroups (ancient DNA samples, Onge, Karitiana, and Mbuti). The rank of the (i, j) matrix reflects the minimum number of sources that contributed to the *Test* populations^{16,17}. For a pool of individuals from 23 *Test* populations representing most present-day European groups, this analysis rejects descent from just two sources ($P < 10^{-12}$ by a Hotelling T-test¹⁷). However, three source populations are consistent with the data after excluding the Spanish who have evidence for African admixture^{18–20} ($P = 0.019$, not

significant after multiple-hypothesis correction), consistent with the results from ADMIXTURE (SI9), PCA (Fig. 1B, SI10) and f_4 -statistics (Extended Data Table 1, Extended Data Fig. 6, SI11, SI12). We caution that the finding of three sources could be consistent with a larger number of mixture events. Moreover, the source populations may themselves have been mixed. Indeed, the positive $f_4(\text{Stuttgart, Test; Loschbour, Chimp})$ statistics obtained when *Test* is Near Eastern (Extended Data Table 1) imply that the EEF had some WHG-related ancestry, which was greater than 0% and as high as 45% (SI13).

We used the ADMIXTUREGRAPH software^{8,15} to fit a model (a tree structure augmented by admixture events) to the data, exploring models relating the three ancient populations (Stuttgart, Loschbour, and MA1) to two eastern non-Africans (Ongce and Karitiana) and sub-Saharan Africans (Mbuti). We found no models that fit the data with 0 or 1 admixture events, but did find a model that fit with 2 admixture events (SI14). The successful model (Fig. 2A) confirms the existence of MA1-related admixture in Native Americans³, but includes the novel inference that Stuttgart is partially ($44 \pm 10\%$) derived from a lineage that split prior to the separation of eastern non-Africans from the common ancestor of WHG and ANE. The existence of such “Basal Eurasian” admixture into Stuttgart provides a simple explanation for our finding that diverse eastern non-African populations share significantly more alleles with ancient European and Upper Paleolithic Siberian hunter-gatherers than with Stuttgart (that is, $f_4(\text{Eastern non-African, Chimp; Hunter-gatherer, Stuttgart})$ is significantly positive), but that hunter-gatherers appear to be equally related to most eastern groups (SI14). We verified the robustness of the model by reanalyzing the data using the unsupervised MixMapper⁷ (SI15) and TreeMix²¹ software (SI16), which both identified the same admixture events. The ANE/WHG split must have occurred >24,000 years ago (as it must predate the age of MA1³), and the WHG/Eastern non-African split must have occurred >40,000 years ago (as it must predate the Tianyuan²² individual from China which clusters with Asians to the exclusion of Europeans). The Basal Eurasian split must be even older, and might be related to early settlement of the Levant²³ or Arabia^{24,25} prior to the diversification of most Eurasians, or more recent gene flow from Africa²⁶. However, the Basal Eurasian population shares much of the genetic drift common to non-African populations after their separation from Africans, and thus does not appear to represent gene flow between sub-Saharan Africans and the ancestors of non-Africans after the out-of-Africa bottleneck (SI14).

Fitting present-day Europeans into the model, we find that few populations can be fit as 2-way mixtures, but nearly all are compatible with 3-way mixtures of ANE/EEF/WHG (SI14). The mixture proportions from the fitted model (Fig. 2B; Extended Data Table 3) are encouragingly consistent with those obtained from a separate method that relates European populations to diverse outgroups using f_4 -statistics, assuming only that MA1 is an unmixed descendent of ANE, Loschbour of WHG, and Stuttgart of EEF (SI17). We infer that EEF ancestry in Europe today ranges from ~30% in the Baltic region to ~90% in the Mediterranean, consistent with patterns of identity-by-descent (IBD) sharing^{27,28} (SI18) and shared haplotype analysis (chromosome painting)²⁹ (SI19) in which Loschbour shares more segments with northern Europeans and Stuttgart with southern Europeans. Southern Europeans inherited their European hunter-gatherer ancestry mostly via EEF ancestors

(Extended Data Fig. 6), while Northern Europeans acquired up to 50% of WHG ancestry above and beyond the WHG-related ancestry which they received through their EEF ancestors. Europeans have a larger proportion of WHG than ANE ancestry in general. By contrast, in the Near East there is no detectable WHG ancestry, but up to ~29% ANE in the Caucasus (SI14). A striking feature of these findings is that ANE ancestry is inferred to be present in nearly all Europeans today (with a maximum of ~20%), but was absent in both farmers and hunter-gatherers from central/western Europe during the Neolithic transition. At the same time, we infer that ANE ancestry was not completely absent from the larger European region at that time: we find that it was present in ~8,000 years old Scandinavian hunter-gatherers, since MA1 shares more alleles with Motala12 (SHG) than with Loschbour, and Motala12 fits as a mixture of 81% WHG and 19% ANE (SI14).

Two sets of European populations are poor fits for the model. Sicilians, Maltese, and Ashkenazi Jews have EEF estimates of >100% consistent with their having more Near Eastern ancestry than can be explained via EEF admixture (SI17). They also cannot be jointly fit with other Europeans (SI14), and they fall in the gap between European and Near Easterners (Fig. 1B). Finns, Mordovians and Russians (from the northwest of Russia) also do not fit (SI14; Extended Data Table 3) due to East Eurasian gene flow into the ancestors of these northeastern European populations. These populations (and Chuvash and Saami) are more related to East Asians than can be explained by ANE admixture (Extended Data Fig. 7), likely reflecting a separate stream of Siberian gene flow into northeastern Europe (SI14).

Several questions will be important to address in future ancient DNA work. Where and when did the Near Eastern farmers admix with European hunter-gatherers to produce the EEF? How did the ancestors of present-day Europeans first acquire their ANE ancestry? Discontinuity in central Europe during the late Neolithic (~4,500 years ago) associated with the appearance of mtDNA types absent in earlier farmers and hunter-gatherers³⁰ raises the possibility that ANE ancestry may have also appeared at this time. Finally, it is important to study ancient genome sequences from the Near East to provide insights into the history of the Basal Eurasians.

Online Methods

Archeological context, sampling and DNA extraction

The Loschbour sample stems from a male skeleton excavated in 1935 at the Loschbour rock shelter in Heffingen, Luxembourg. The skeleton was AMS radiocarbon dated to $7,205 \pm 50$ years before present (OxA-7738; 6,220-5,990 cal BC)³¹. At the Palaeogenetics Laboratory in Mainz, material for DNA extraction was sampled from tooth 16 (an upper right M1 molar) after irradiation with UV-light, surface removal, and pulverization in a mixer mill. DNA extraction took place in the palaeogenetics facilities in the Institute for Archaeological Sciences at the University of Tübingen. Three extracts were made in total, one from 80 mg of powder using an established silica based protocol³² and two additional extracts from 90 mg of powder each with a protocol optimized for the recovery of short DNA molecules³³.

The Stuttgart sample was taken from a female skeleton excavated in 1982 at the site Viesenhäuser Hof, Stuttgart-Mühlhausen, Germany. It was attributed to the

Linearbandkeramik (5,500-4,800 BC) through associated pottery artifacts and the chronology was corroborated by radiocarbon dating of the stratigraphy³⁴. Both sampling and DNA extraction took place in the Institute for Archaeological Sciences at the University of Tübingen. Tooth 47 (a lower right M2 molar) was removed and material from the inner part was sampled with a sterile dentistry drill. An extract was made using 40 mg of bone powder³³.

The Motala individuals were recovered from the site of Kanaljorden in the town of Motala, Östergötland, Sweden, excavated between 2009 and 2013. The human remains at this site are represented by several adult skulls and one infant skeleton. All individuals are part of a ritual deposition at the bottom of a small lake. Direct radiocarbon dates on the remains range between $7,013 \pm 76$ and $6,701 \pm 64$ BP (6,361-5,516 cal BC), corresponding to the late Middle Mesolithic of Scandinavia. Samples were taken from the teeth of the nine best preserved skulls, as well as a femur and tibia. Bone powder was removed from the inner parts of the teeth or bones with a sterile dentistry drill. DNA from 100 mg of bone powder was extracted³⁵ in the ancient DNA laboratory of the Archaeological Research Laboratory, Stockholm.

Library preparation

Illumina sequencing libraries were prepared using either double- or single-stranded library preparation protocols^{36,37} (SI1). For high-coverage shotgun sequencing libraries, a DNA repair step with Uracil-DNA-glycosylase (UDG) and endonuclease VIII (endo VIII) treatment was included in order to remove uracil residues³⁸. Size fractionation on a PAGE gel was also performed in order to remove longer DNA molecules that are more likely to be contaminants³⁷. Positive and blank controls were carried along during every step of library preparation.

Shotgun sequencing and read processing

All non-UDG-treated libraries were sequenced either on an Illumina Genome Analyzer IIx with $2 \times 76 + 7$ cycles for the Loschbour and Motala libraries, or on an Illumina MiSeq with $2 \times 150 + 8 + 8$ cycles for the Stuttgart library. We followed the manufacturer's protocol for multiplex sequencing. Raw overlapping forward and reverse reads were merged and filtered for quality³⁹ and mapped to the human reference genome (hg19/GRCh37/1000Genomes) using the Burrows-Wheeler Aligner (BWA)⁴⁰ (SI2). For deeper sequencing, UDG-treated libraries of Loschbour were sequenced on 3 Illumina HiSeq 2000 lanes with 50-bp single-end reads, 8 Illumina HiSeq 2000 lanes of 100-bp paired-end reads and 8 Illumina HiSeq 2500 lanes of 101-bp paired-end reads. The UDG-treated library for Stuttgart was sequenced on 8 HiSeq 2000 lanes and 101-bp paired-end reads. The UDG-treated libraries for Motala were sequenced on 8 HiSeq 2000 lanes of 100-bp paired-end reads, with 4 lanes each for two pools (one of 3 individuals and one of 4 individuals). We also sequenced an additional 8 HiSeq 2000 lanes for Motala12, the Motala sample with the highest percentage of endogenous human DNA. For the Loschbour and Stuttgart high coverage individuals, diploid genotype calls were obtained using the Genome Analysis Toolkit (GATK)⁴¹.

Enrichment of mitochondrial DNA and sequencing

To test for DNA preservation and mtDNA contamination non-UDG-treated libraries of Loschbour and all Motala samples were enriched for human mitochondrial DNA using a bead-based capture approach with present-day human DNA as bait⁴². UDG-treatment was omitted in order to allow characterization of damage patterns typical for ancient DNA¹⁰. The captured libraries were sequenced on an Illumina Genome Analyzer IIx platform with $2 \times 76 + 7$ cycles and the resulting reads were merged and quality filtered³⁹. The sequences were mapped to the Reconstructed Sapiens Reference Sequence, RSRS⁴³, using a custom iterative mapping assembler, MIA⁴⁴ (SI4).

Contamination estimates

We assessed if the sequences had the characteristics of authentic ancient DNA using four approaches. First we searched for evidence of contamination by determining whether the sequences mapping to the mitochondrial genome were consistent with deriving from more than one individual^{44,45}. Second, for the high-coverage Loschbour and Stuttgart genomes, we used a maximum-likelihood-based estimate of autosomal contamination that uses variation at sites that are fixed in the 1000 Genomes data to estimate error, heterozygosity and contamination⁴⁶ simultaneously. Third, we estimated contamination based on the rate of polymorphic sites on the X chromosome of the male Loschbour individual⁴⁷ (SI3). Fourth, we analyzed non-UDG treated reads mapping to the RSRS to search for aDNA-typical damage patterns resulting in C→T changes at the 5'-end of the molecule¹⁰ (SI3).

Phylogenetic analysis of the mitochondrial genomes

All nine complete mitochondrial genomes that fulfilled the criteria of authenticity were assigned to haplogroups using Haplofind⁴⁸. A Maximum Parsimony tree including present day humans and previously published ancient mtDNA sequences was generated with MEGA⁴⁹. The effect of branch shortening due to a lower number of substitutions in ancient lineages was studied by calculating the nucleotide edit distance to the root for all haplogroup R sequences (SI4).

Sex determination and Y-chromosome analysis

We assessed the sex of all sequenced individuals by using the ratio of (chrY) to (chrY +chrX) aligned reads⁵⁰. We downloaded a list of Y-chromosome SNPs curated by the International Society of Genetic Genealogy (ISOGG, <http://www.isogg.org>) v. 9.22 (accessed Feb. 18, 2014) and determined the state of the ancient individuals at positions where a single allele was observed and MAPQ ≥ 30 . We excluded C/G or A/T SNPs due to uncertainty about the polarity of the mutation in the database. The ancient individuals were assigned haplogroups based on their derived state (SI5). We also used BEAST v1.7.51⁵¹ to assess the phylogenetic position of Loschbour using 623 males from around the world with 2,799 variant sites across 500kb of non-recombining Y-chromosome sequence⁵² (SI5).

Estimation of Neanderthal admixture

We estimate Neanderthal admixture in ancient individuals with the f_4 -ratio or S -statistic^{8,53,54} $\hat{\alpha} = f_4(\text{Altai}, \text{Denisova}; \text{Test}, \text{Yoruba})/f_4(\text{Altai}, \text{Denisova}; \text{Vindija}, \text{Yoruba})$

which uses whole genome data from Altai, a high coverage (52×) Neanderthal genome sequence⁵⁵, Denisova, a high coverage sequence³⁷ from another archaic human population (31×), and Vindija, a low coverage (1.3×) Neanderthal genome from a mixture of three Neanderthal individuals from Vindija Cave in Croatia⁵³.

Inference of demographic history and inbreeding

We used the Pairwise Sequentially Markovian Coalescent (PSMC)⁵⁶ to infer the size of the ancestral population of Stuttgart and Loschbour. This analysis requires high quality diploid genotype calls and cannot be performed in the low-coverage Motala samples. To determine whether the low effective population size inferred for Loschbour is due to recent inbreeding, we plotted the time-to-most-recent common ancestor (TMRCA) along each of chr1-22 to detect runs of low TMRCA.

Analysis of segmental duplications and copy number variants

We built read-depth based copy number maps for the Loschbour, Stuttgart and Motala12 genomes in addition to the Denisova and Altai Neanderthal genome and 25 deeply sequenced modern genomes⁵⁵ (SI7). We built these maps by aligning reads, subdivided into their non-overlapping 36-bp constituents, against the reference genome using the mrsFAST aligner⁵⁷, and renormalizing read-depth for local GC content. We estimated copy numbers in windows of 500 unmasked base pairs slid at 100 bp intervals across the genome. We called copy number variants using a scale space filter algorithm. We genotyped variants of interest and compared the genotypes to those from individuals sequenced as part of the 1000 Genomes Project⁵⁸.

Phenotypic inference

We inferred likely phenotypes (SI8) by analyzing DNA polymorphism data in the VCF format⁵⁹ using VCFtools (<http://vcftools.sourceforge.net/>). For the Loschbour and Stuttgart individuals, we included data from sites not flagged as LowQuality, with genotype quality (GQ) of ≥ 30 , and SNP quality (QUAL) of ≥ 50 . For Motala12, which is of lower coverage, we included sites having at least 2× coverage and that passed visual inspection of the local alignment using samtools tview (<http://samtools.sourceforge.net/>)⁶⁰

Human Origins dataset curation

The Human Origins array consists of 14 panels of SNPs for which the ascertainment is well known^{8,61}. All population genetics analysis were carried out on a set of 594,924 autosomal SNPs, after restricting to sites that had >90% completeness across 7 different batches of sequencing, and that had >97.5% concordance with at least one of two subsets of samples for which whole genome sequencing data was also available. The total dataset consists of 2,722 individuals, which we filtered to 2,345 individuals (203 populations) after removing outlier individuals or relatives based on visual inspection of PCA plots^{14,62} or model-based clustering analysis¹³. Whole genome amplified (WGA) individuals were not used in analysis, except for a Saami individual who we included because of the special interest of this population for Northeastern European population history (Extended Data Fig. 7).

ADMIXTURE analysis

We merged all Human Origins genotype data with whole genome sequencing data from Loschbour, Stuttgart, MA1, Motala12, Motala_merge, and LaBrana. We then thinned the resulting dataset to remove SNPs in linkage-disequilibrium with PLINK 1.07⁶³, using a window size of 200 SNPs advanced by 25 SNPs and an r^2 threshold of 0.4. We ran ADMIXTURE 1.23^{13,64} for 100 replicates with different starting random seeds, default 5-fold cross-validation, and varying the number of ancestral populations K between 2 and 20. We assessed clustering quality using CLUMPP⁶⁵. We used the ADMIXTURE results to identify a set of 59 “West Eurasian” (European/Near Eastern) populations based on values of a “West Eurasian” ancestral population at $K=3$ (SI9). We also identified 15 populations for use as “non-West Eurasian outgroups” based on their having at least 10 individuals and no evidence of European or Near Eastern admixture at $K=11$, the lowest K for which Near Eastern/European-maximized ancestral populations appeared consistently across all 100 replicates.

Principal Components Analysis

We used *smartpca*¹⁴ (version: 10210) from EIGENSOFT^{62,66} 5.0.1 to carry out Principal Components Analysis (PCA) (SI10). We performed PCA on a subset on individuals and then projected others using the *lsqproject: YES* option that gives an unbiased inference of the position of samples even in the presence of missing data (especially important for ancient DNA).

f_3 -statistics

We use the f_3 -statistic⁸ $f_3(\text{Test}; \text{Ref}_1 \text{Ref}_2) = \frac{1}{N} \sum_{i=1}^N (t_i - r_{1,i})(t_i - r_{2,i})$, where t_i , $r_{1,i}$ and $r_{2,i}$ are the allele frequencies for the i^{th} SNP in populations *Test*, *Ref*₁, *Ref*₂, respectively, to determine if there is evidence that the *Test* population is derived from admixture of populations related to *Ref*₁ and *Ref*₂ (SI11). A significantly negative statistic provides unambiguous evidence of mixture in the *Test* population⁸. We allow *Ref*₁ and *Ref*₂ to be any Human Origins population with 4 or more individuals, or Loschbour, Stuttgart, MA1, Motala12, LaBrana. We assess significance of the f_3 -statistics using a block jackknife⁶⁷ and a block size of 5cM. We report significance as the number of standard errors by which the statistic differs from zero (Z-score). We also perform an analysis in which we constrain the reference populations to be (i) EEF (Stuttgart) and WHG (Loschbour or LaBrana), (ii) EEF and a Near Eastern population, (iii) EEF and ANE (MA1), or (iv) any two present-day populations, and compute a Z_{diff} score between the lowest f_3 -statistic observed in the dataset, and the f_3 -statistic observed for the specified pair.

f_4 -statistics

We analyze f_4 -statistics⁸ of the form $f_4(A, B; C, D) = \frac{1}{N} \sum_{i=1}^N (a_i - b_i)(c_i - d_i)$ to assess if populations A, B are consistent with forming a clade in an unrooted tree with respect to C, D. If they form a clade, the allele frequency differences between the two pairs should be uncorrelated and the statistic has an expected value of 0. We set the outgroup *D* to be a sub-Saharan African population or Chimpanzee. We systematically tried all possible

combinations of the ancient samples or 15 “non-West Eurasian outgroups” identified by ADMIXTURE analysis as A, B, C to determine their genetic affinities (SI14). Setting A as a present-day test population and B as either Stuttgart or BedouinB, we documented relatedness to C=(Loschbour or MA1) or C=(MA1 and Karitiana) or C=(MA1 or Han) (Extended Data Figs. 4, 5, 7). Setting C as a test population and (A, B) a pair from (Loschbour, Stuttgart, MA1) we documented differential relatedness to ancient populations (Extended Data Fig. 6). We computed D -statistics⁵³ using transversion polymorphisms in whole genome sequence data⁵⁵ to confirm robustness to ascertainment and ancient DNA damage (Extended Data Table 2).

Minimum number of source populations for Europeans

We used *qpWave*^{16,17} to study the minimum number of source populations for a designated set of Europeans (SI12). We use f_4 -statistics of the form $X(l, r) = f_4(l_0, l; r_0, r)$ where l_0, r_0 are arbitrarily chosen “base” populations, and l, r are other populations from two sets L and R respectively. If $X(l, r)$ has rank r and there were n waves of immigration into R with no back-migration from R to L , then $r+1 \leq n$. We set L to include *Stuttgart, Loschbour, MA1, Onge, Karitiana, Mbuti* and R to include 23 modern European populations who fit the model of SI14 and had admixture proportions within the interval [0,1] for the method with minimal modeling assumptions (SI17).

Admixture proportions for Stuttgart in the absence of a Near Eastern ancient genome

We used Loschbour and BedouinB as surrogates for “Unknown hunter-gatherer” and Near Eastern (NE) farmer populations that contributed to Stuttgart (SI13). Ancient Near Eastern ancestry in Stuttgart is estimated by the f_4 -ratio^{8,15} $f_4(\text{Outgroup}, X; \text{Loschbour}, \text{Stuttgart})/f_4(\text{Outgroup}, X; \text{Loschbour}, \text{NE})$. A complication is that BedouinB is a mixture of NE and African ancestry. We therefore subtracted¹⁷ the effects of African ancestry using estimates of the BedouinB African admixture proportion from ADMIXTURE (SI9) or ALDER⁶⁸.

Admixture graph modeling

We used ADMIXTUREGRAPH⁸ (version 3110) to model population relationships between Loschbour, Stuttgart, Onge, and Karitiana using Mbuti as an African outgroup. We assessed model fit using a block jackknife of differences between estimated and fitted f -statistics for the set of included populations (we expressed the fit as a Z score). We determined that a model failed if $|Z| > 3$ for at least one f -statistic. A basic tree model failed and we manually amended the model to test all possible models with a single admixture event, which also failed. Further manual amendment to include 2 admixture events resulted in 8 successful models, only one of which could be amended to also fit MA1 as an additional constraint. We successfully fit both the Iceman and LaBranca into this model as simple clades and Motala12 as a 2-way mixture. We also fit present-day West Eurasians as clades, 2-way mixtures, or 3-way mixtures in this basic model, achieving a successful fit for a larger number of European populations ($n=26$) as 3-way mixtures. We estimated the individual admixture proportions from the fitted model parameters. To test if fitted parameters for different populations are consistent with each other, we jointly fit all pairs of populations A and B by modifying

ADMIXTUREGRAPH to add a large constant (10,000) to the variance term $f_3(A_0, A, B)$. By doing this, we can safely ignore recent gene flow within Europe that affects statistics that include both A and B .

Ancestry estimates from f_4 -ratios

We estimate EEF ancestry using the f_4 -ratio^{8,15} $f_4(\text{Mbuti, Onge; Loschbour, European})/f_4(\text{Mbuti, Onge; Loschbour, Stuttgart})$, which produces consistent results with ADMIXTUREGRAPH (SI14). We use $f_4(\text{Stuttgart, Loschbour; Onge MA1})/f_4(\text{Mbuti, MA1; Onge, Loschbour})$ to estimate Basal Eurasian admixture into Stuttgart. We use $f_4(\text{Stuttgart, Loschbour; Onge Karitiana})/f_4(\text{Stuttgart, Loschbour; Onge MA1})$ to estimate ANE mixture in Karitiana (Fig. 2B). We use $f_4(\text{Test, Stuttgart; Karitiana, Onge})/f_4(\text{MA1, Stuttgart; Karitiana, Onge})$ to lower bound ANE mixture into North Caucasian populations.

MixMapper analysis

We carried out *MixMapper* 2.0⁷ analysis, a semi-supervised admixture graph fitting technique. First, we infer a scaffold tree of populations without strong evidence of mixture relative to each other (Mbuti, Onge, Loschbour and MA1). We do not include European populations in the scaffold as all had significantly negative f_3 -statistics indicating admixture. We then ran *MixMapper* to infer the relatedness of the other ancient and present-day samples, fitting them onto the scaffold as 2- or 3-way mixtures. The uncertainty in all parameter estimates is measured by block bootstrap resampling of the SNP set (100 replicates with 50 blocks).

TreeMix analysis

We applied *TreeMix*²¹ to Loschbour, Stuttgart, Motala12, and MA1³, LaBran² and the Iceman¹, along with the present-day samples of Karitiana, Onge and Mbuti. We restricted the analysis to 265,521 Human Origins array sites after excluding any SNPs where there were no-calls in any of the studied individuals. The tree was rooted with Mbuti and standard errors were estimated using blocks of 500 SNPs. We repeated the analysis on whole-genome sequence data, rooting with Chimp and replacing Onge with Dai since we did not have Onge whole genome sequence data⁵⁵. We varied the number of migration events (m) between 0 and 5.

Inferring admixture proportions with minimal modeling assumptions

We devised a method to infer ancestry proportions from three ancestral populations (EEF, WHG, and ANE) without strong phylogenetic assumptions (SI17). We rely on 15 “non-West Eurasian” outgroups and study $f_4(\text{European, Stuttgart; } O_1, O_2)$ which equals $\alpha\beta f_4(\text{Loschbour, Stuttgart; } O_1, O_2) + \alpha(1-\beta) f_4(\text{MA1, Stuttgart; } O_1, O_2)$ if *European* has $1-\alpha$ ancestry from EEF and β , $1-\beta$ ancestry from WHG and ANE respectively. This defines a

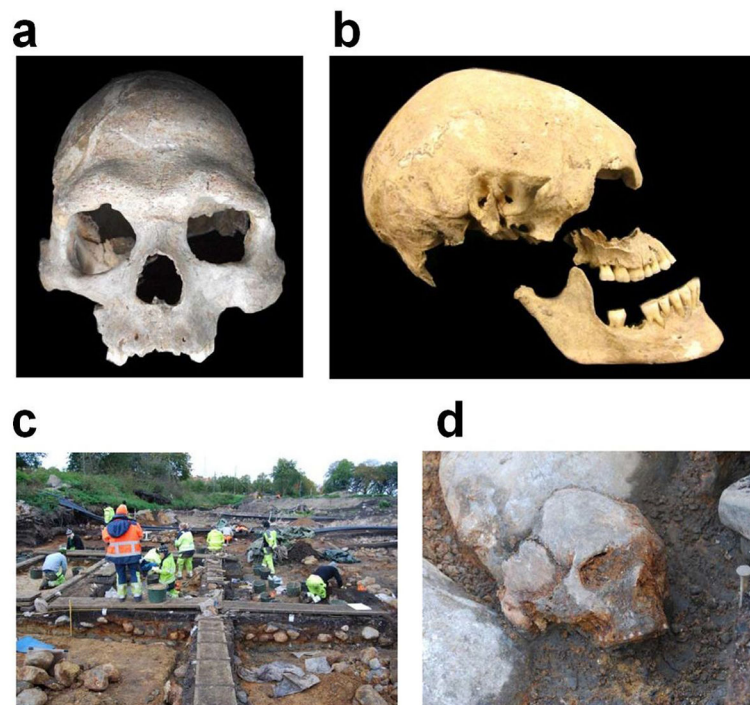
system of $\binom{15}{2} = 105$ equations with unknowns $\alpha\beta$, $\alpha(1-\beta)$, which we solve with least squares implemented in the function *lsfit* in *R* to obtain estimates of α and β . We repeated this computation 22 times dropping one chromosome at a time²⁰ to obtain block jackknife⁶⁷ estimates of the ancestry proportions and standard errors, with block size equal to the

number of SNPs per chromosome. We assessed consistency of the inferred admixture proportions with those derived from the ADMIXTUREGRAPH model based on the number of standard errors between the two (Extended Data Table 1).

Haplotype-based analyses

We used RefinedIBD from BEAGLE 4²⁷ with the settings *ibdtrim*=20 and *ibdwindow*=25 to study IBD sharing between Loschbour and Stuttgart and populations from the POPRES dataset⁶⁹. We kept all IBD tracts spanning at least 0.5 centimorgans (cM) and with a LOD score >3 (SI18). We also used ChromoPainter²⁹ to study haplotype sharing between Loschbour and Stuttgart and present-day West Eurasian populations (SI19). We identified 495,357 SNPs that were complete in all individuals and phased the data using Beagle 4²⁷ with parameters *phase-its*=50 and *impute-its*=10. We did not keep sites with missing data to avoid imputing modern alleles into the ancient individuals. We used both unlinked (-k 1000) and linked modes (estimating -n and -M by sampling 10% of individuals). We combined ChromoPainter output for chromosomes 1-22 using ChromoCombine²⁹. We carried out a PCA of the co-ancestry matrix using fineSTRUCTURE²⁹.

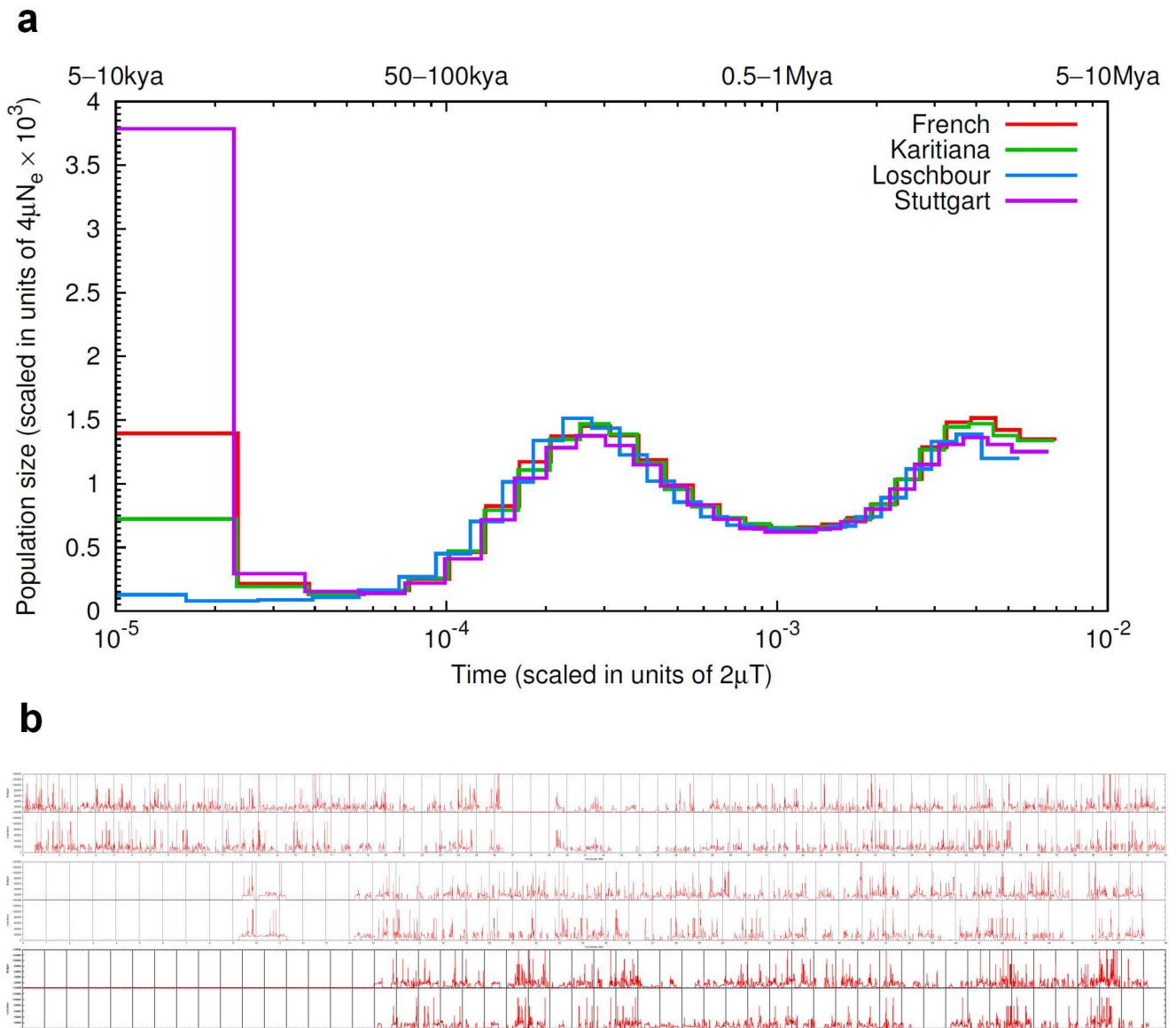
Extended Data



Extended Data Figure 1.

Photographs of analyzed ancient samples.

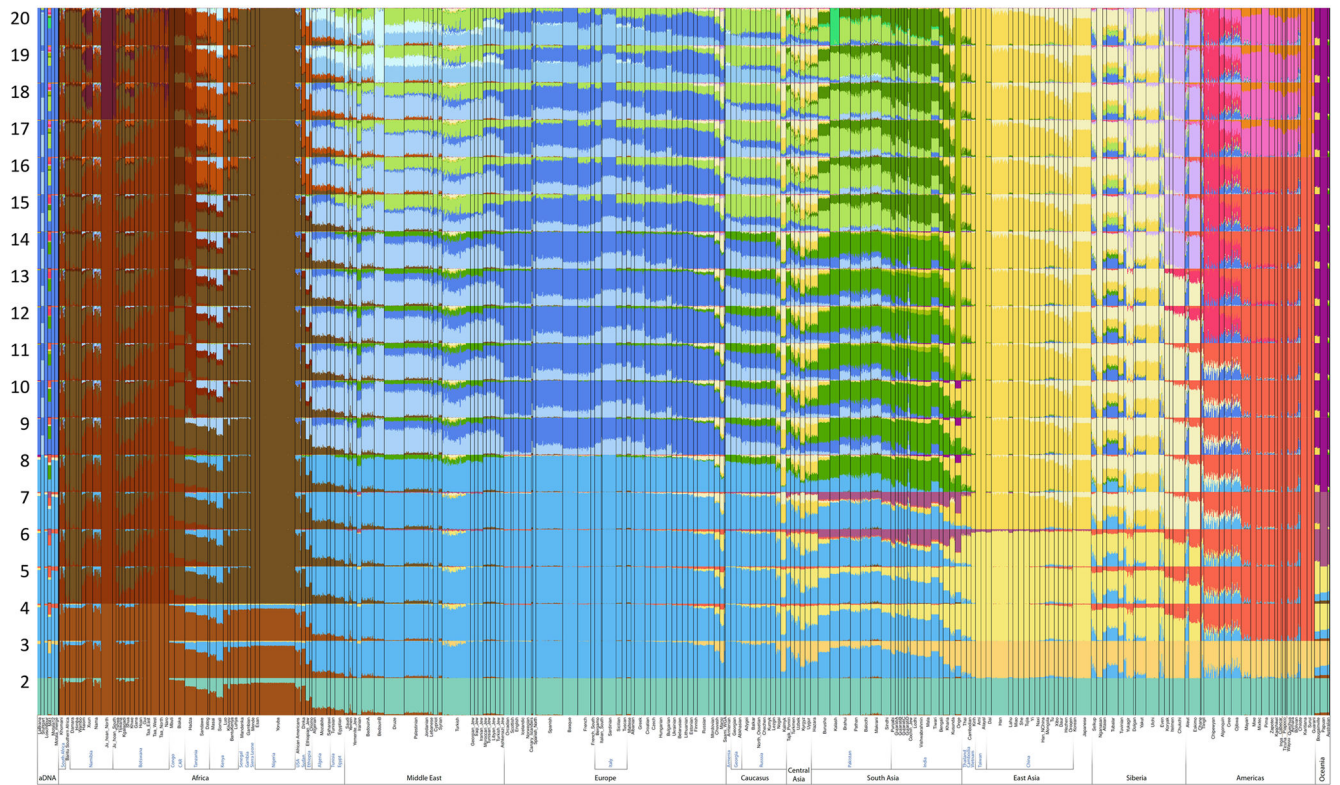
(A) Loschbour skull; (B) Stuttgart skull, missing the lower right M2 we sampled; (C) excavation at Kanaljorden in Motala, Sweden; (D) Motala 1 in situ.



Extended Data Figure 2.

Pairwise Sequential Markovian Coalescent (PSMC) analysis.

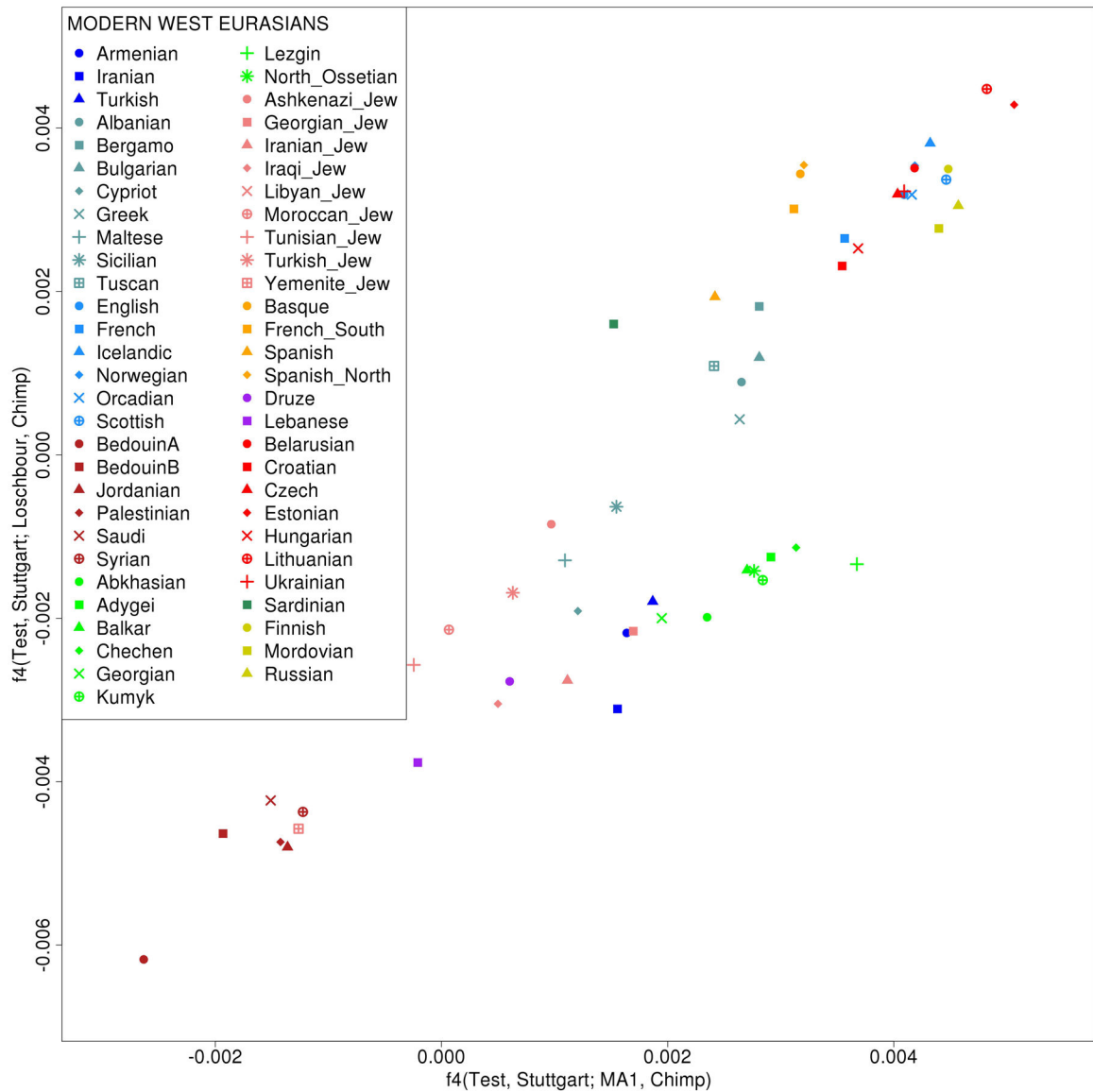
(A) Inference of population size as a function of time, showing a very small recent population size over the most recent period in the ancestry of Loschbour (at least the last 5–10 thousand years). (B) Inferred time since the most recent common ancestor from the PSMC for chromosomes 20, 21, 22 (top to bottom); Stuttgart is plotted on top and Loschbour at bottom.



Extended Data Figure 3.

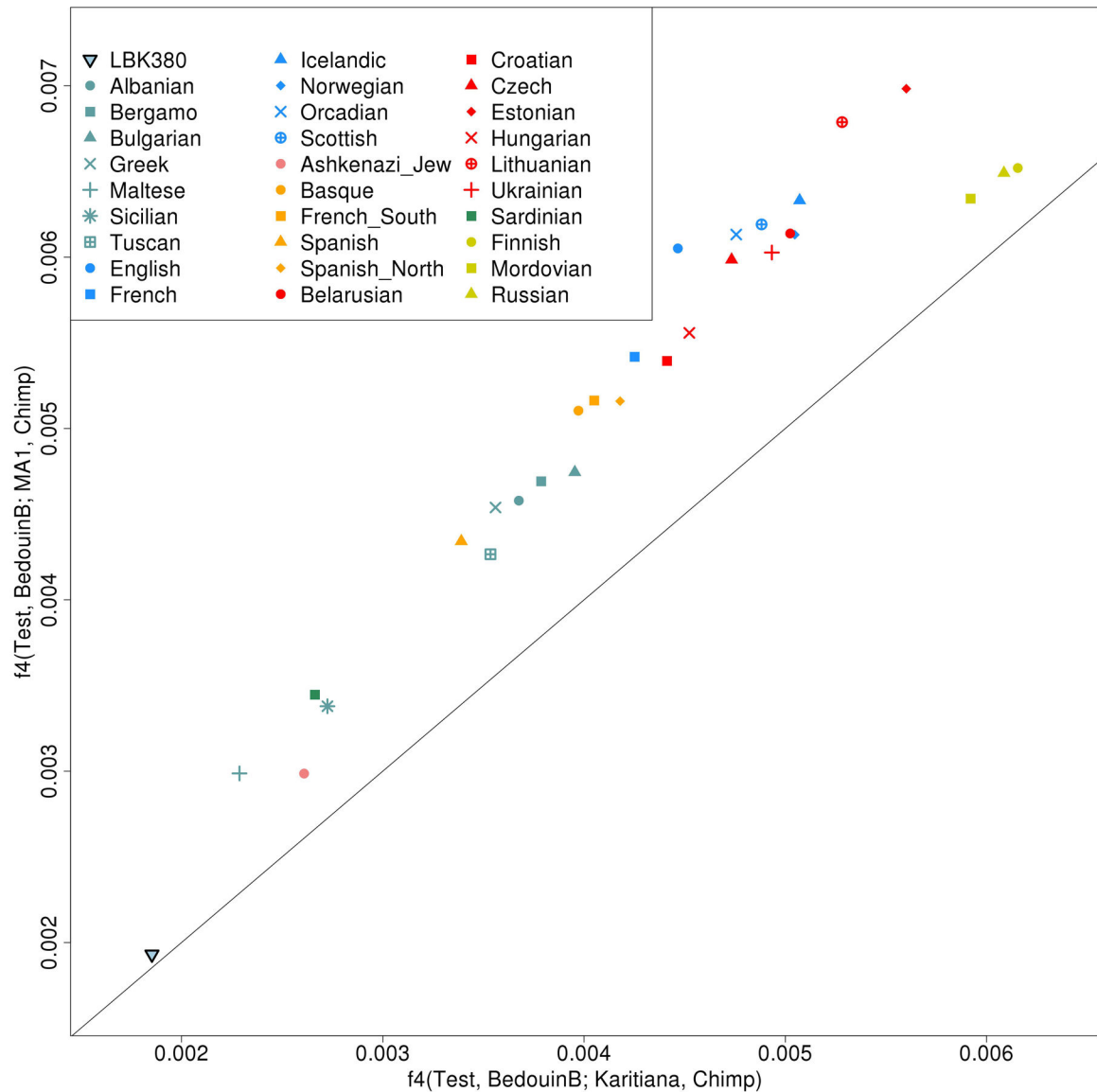
ADMIXTURE analysis (K=2 to K=20).

Ancient samples (Loschbour, Stuttgart, Motala_merge, Motala12, MA1, and LaBrana) are at left.

**Extended Data Figure 4.**

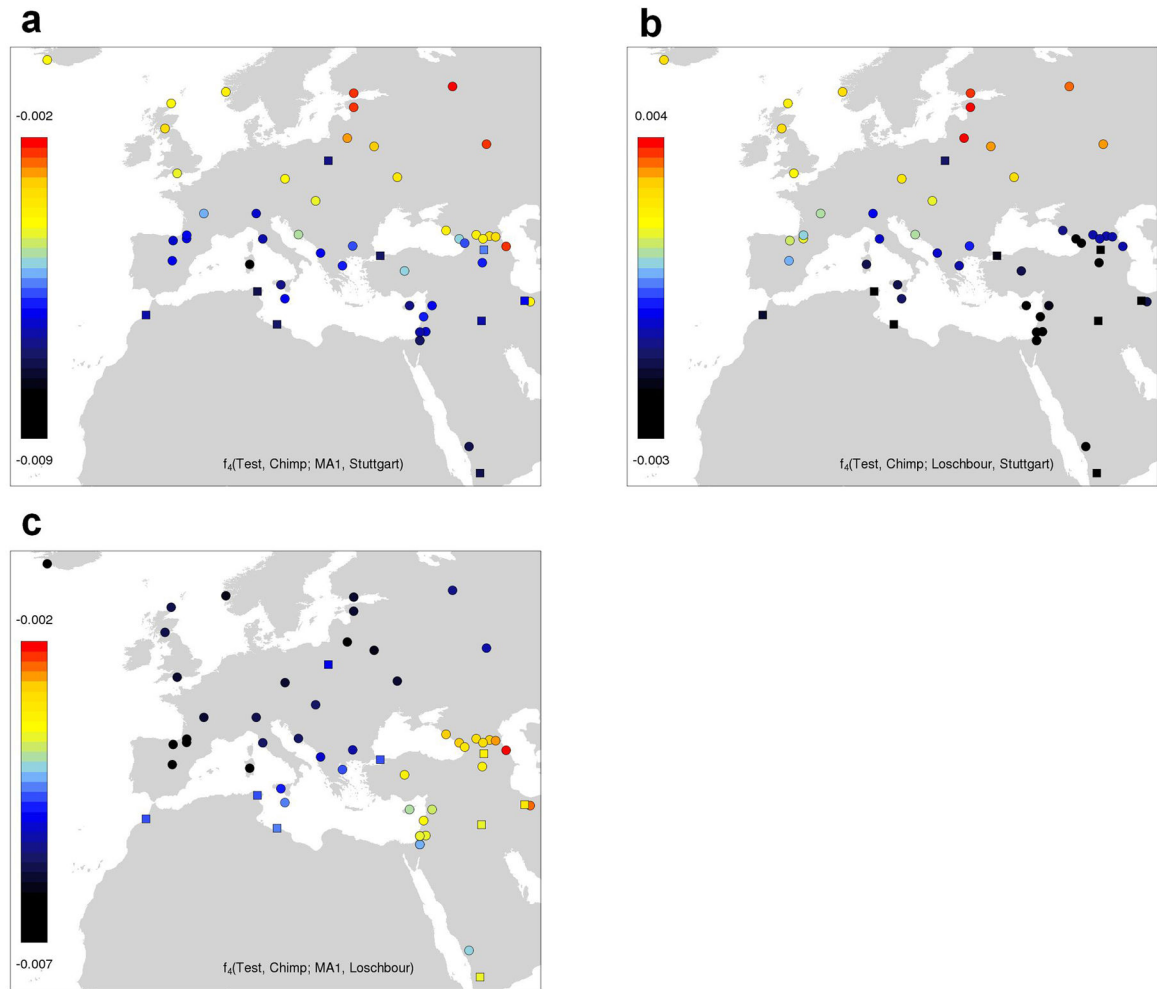
ANE ancestry is present in both Europe and the Near East but WHG ancestry is restricted to Europe, which cannot be due to a single admixture event.

(x-axis) We computed the statistic $f_4(\text{Test}, \text{Stuttgart}; \text{MA1}, \text{Chimp})$, which measures where MA1 shares more alleles with a test population than with Stuttgart. It is positive for most European and Near Eastern populations, consistent with ANE (MA1-related) gene flow into both regions. (y-axis) We computed the statistic $f_4(\text{Test}, \text{Stuttgart}; \text{Loschbour}, \text{Chimp})$, which measures whether Loschbour shares more alleles with a test sample than with Stuttgart. Only European populations show positive values of this statistic, providing evidence of WHG (Loschbour-related) admixture only in Europeans.

**Extended Data Figure 5.**

MA1 is the best surrogate for ANE for which we have data.

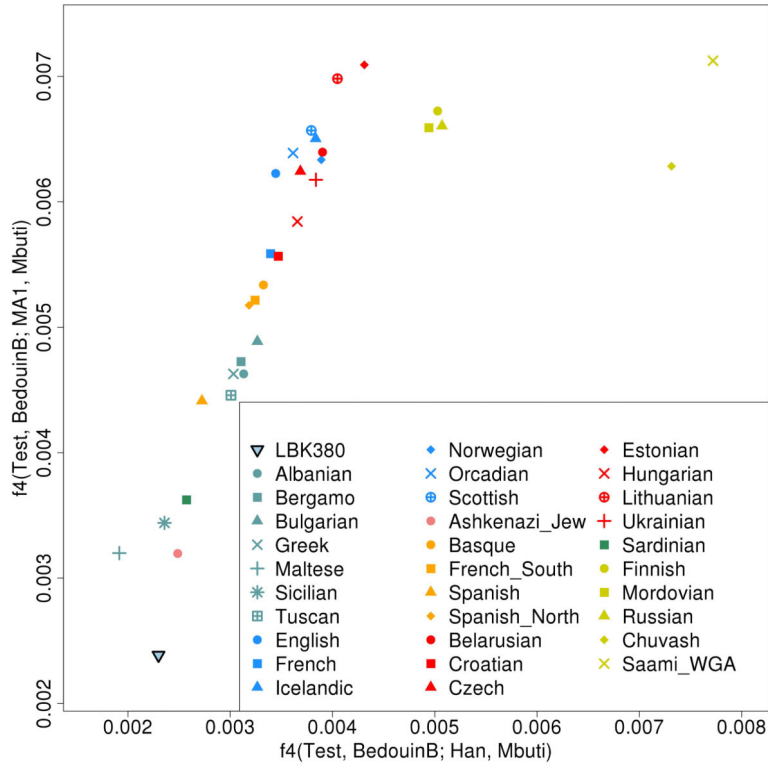
Europeans share more alleles with MA1 than with Karitiana, as we see from the fact that in a plot of $f_4(\text{Test}, \text{BedouinB}; \text{MA1}, \text{Chimp})$ and $f_4(\text{Test}, \text{BedouinB}; \text{Karitiana}, \text{Chimp})$, the European cline deviates in the direction of MA1, rather than Karitiana (the slope is >1 and European populations are above the line indicating equality of these two statistics).



Extended Data Figure 6.

The differential relatedness of West Eurasians to Stuttgart (EEF), Loschbour (WHG), and MA1 (ANE) cannot be explained by two-way mixture.

We plot on a West Eurasian map the statistic $f_4(\text{Test}, \text{Chimp}; A_1, A_2)$, where A_1 and A_2 are a pair of the three ancient samples representing the three ancestral populations of Europe. (A) In both Europe and the Near East/Caucasus, populations from the south have more relatedness to Stuttgart than those from the north where ANE influence is also important. (B) Northern European populations share more alleles with Loschbour than with Stuttgart, as they have additional WHG ancestry beyond what was already present in EEF. (C) We observe a striking contrast between Europe west of the Caucasus and the Near East in degree of relatedness to WHG. In Europe, there is a much higher degree of allele sharing with Loschbour than with MA1, which we ascribe to the 60–80% WHG/(WHG+ANE) ratio in most Europeans that we report in SI14. In contrast, the Near East has no appreciable WHG ancestry but some ANE ancestry, especially in the northern Caucasus. (Jewish populations are marked with a square in this figure to assist in interpretation as their ancestry is often anomalous for their geographic regions.)



Extended Data Figure 7.

Evidence for Siberian gene flow into far northeastern Europe.

Some northeastern European populations (Chuvash, Finnish, Russian, Mordovian, Saami) share more alleles with Han Chinese than with other Europeans who are arrayed in a cline from Stuttgart to Lithuanians/Estonians in a plot of $f_4(\text{Test}, \text{BedouinB}; \text{Han}, \text{Mbuti})$ against $f_4(\text{Test}, \text{BedouinB}; \text{MA1}, \text{Mbuti})$.

Extended Data Table 1

West Eurasians genotyped on the Human Origins array and key f -statistics.

Sample Location	N	Lat.	Long.	Locus 3 (X, E227, W2)				Locus 3 (X, E227, W2) (G and A2601 frequency)				Locus 3 (X, E227, A262)				Locus 4 (X, E227, C262)		Locus 4 (X, E227, C262)								
				R ₁	R ₂	statistic	Z	R ₁	R ₂	statistic	Z	Z _{diff}	R ₁	R ₂	statistic	Z	Z _{diff}	statistic	Z							
Abkhazian	9	41	41.02	Str	MA1	-0.0013	-1.9			Georgian	Lab	-0.004	-6.5	2.6	Str	MA1	-0.0021	-2.9	0.0	0.020	4.2	-0.021	-4.7			
Ache	17	44	39	Popco	Str	-0.0073	-5.9								Str	MA1	-0.0067	-4.1	0.1	0.013	2.6	-0.029	-6.0			
Adhama	4	41.33	19.83	Str	MA1	-0.0121	-7.0			Irqi_Juv	Lon	-0.0049	-4.1	1.7	Str	MA1	-0.0121	-7.0	0.0	-0.009	-1.8	-0.027	-5.4			
Armenian	10	40.19	44.55	CapracC	Str	-0.0079	-4.2								Str	MA1	-0.0066	-4.1	0.1	0.022	4.5	-0.016	-3.3			
Ashkenazi_Jew	7	52.33	21.62	Str	MA1	-0.0087	-5.4			Irqi_Juv	Lon	-0.0042	-4.7	1.6	Str	MA1	-0.0087	-5.4	0.0	0.008	1.7	-0.010	-2.0			
Bakur	10	43.48	45.62	Popco	Str	-0.0113	-6.9								Str	MA1	-0.0082	-5.5	1.1	0.014	2.9	-0.027	-5.6			
Baqra	20	43.04	-6.65	Irqi_Juv	Lon	-0.0083	-5.3	Str	Lon	-0.0061	-3.8	1.3	Irqi_Juv	Lon	-0.0083	-5.3	0.0	Str	MA1	-0.0041	-2.8	2.2	-0.014	-7.2	-0.012	-4.7
BulgariaA	25	31	35	Str	Str	-0.0162	-10.2													0.042	11.0	0.026	5.4			
BulgariaB	10	31	35	Str	Str	0.0089	7.8													0.046	9.3	0.019	3.9			
Belarusian	10	53.02	28.05	Georgian	Lon	-0.0133	-17.6			Georgian	Lon	-0.0133	-17.6	0.0	Str	MA1	-0.0102	-6.1	1.9	-0.015	-6.9	-0.042	-8.6			
Berzop	12	46	10	Str	MA1	-0.0106	-6.2	Str	Lon	-0.0068	-4.2	1.7	Irqi_Juv	Lon	-0.0100	-11.9	0.1	Str	MA1	-0.0106	-6.2	0.0	-0.018	-3.9	-0.028	-5.8
Bulgaria	10	42.36	24.74	Str	MA1	-0.0130	-8.2	Str	Lab	-0.0074	-4.9	2.8	Irqi_Juv	Lon	-0.0106	-12.4	1.5	Str	MA1	-0.0130	-8.2	0.0	-0.012	-2.5	-0.028	-5.9
Chelms	9	43.33	45.65	Str	MA1	-0.0096	-5.2			Georgian	Lon	-0.0082	-6.3	2.8	Str	MA1	-0.0096	-5.2	0.0	0.011	2.3	-0.013	-4.2			
Croatian	10	45.51	16.45	Str	MA1	-0.0114	-6.7	Str	Lon	-0.0067	-3.8	2.1	Irqi_Juv	Lon	-0.0112	-12.0	0.2	Str	MA1	-0.0114	-6.7	0.0	-0.023	-4.7	-0.015	-3.8
Cypriot	8	35.13	33.43	Str	MA1	-0.0087	-5.2			Yamnaya_Iuv	Lon	-0.0013	-1.5	2.5	Str	MA1	-0.0087	-5.2	0.0	0.019	3.9	-0.012	-2.5			
Czech	10	50.1	14.4	Georgian	Lon	-0.0137	-17.9	Str	Lon	-0.0068	-5.3	3.0	Georgian	Lon	-0.0137	-17.9	0.0	Str	MA1	-0.0121	-7.2	0.9	-0.012	-4.6	-0.040	-8.2
Dinar	30	32	35	Str	MA1	-0.0024	-1.5								Str	MA1	-0.0024	-1.5	0.0	0.028	5.9	-0.006	-1.3			
English	10	50.75	-1.09	Irqi_Juv	Lon	-0.0129	-14.9	Str	Lon	-0.0090	-5.5	2.2	Irqi_Juv	Lon	-0.0129	-14.8	0.0	Str	MA1	-0.0125	-7.8	0.1	-0.012	-4.5	-0.041	-8.5
Estonian	10	58.54	24.89	Abkhazian	Lon	-0.0124	-15.1			Abkhazian	Lon	-0.0124	-15.1	0.0	Str	MA1	-0.0094	-6.6	1.9	-0.043	-8.3	-0.015	-3.3			

Interpretation	D(A, B; C, D) on Human Origins genotype data								D(A, B; C, D) on whole genome sequence data transversions							
	A	B	C	D	594,924 SNPs		110,817 transversions		A	B	C	D	statistic	Z		
					statistic	Z	statistic	Z								
Eastern non-Africans closer to WHG/ANE/S HG than to EEF	Loschbour	Stuttgart	Onge	Chimp	0.0196	3.5	0.0202	2.5								
	Loschbour	Stuttgart	Papuan	Chimp	0.0142	2.6	0.0127	1.5	Loschbour	Stuttgart	Papuan2	Chimp	0.017	2.7		
	Loschbour	Stuttgart	Dai	Chimp	0.0164	3.2	0.021	2.8	Loschbour	Stuttgart	Dai2	Chimp	0.018	2.9		
	MA1	Stuttgart	Papuan	Chimp	0.0139	2.2	0.0103	1.0	MA1	Stuttgart	Papuan2	Chimp	0.018	2.8		
	MA1	Stuttgart	Dai	Chimp	0.0174	3.0	0.016	1.7	MA1	Stuttgart	Dai2	Chimp	0.028	4.3		
	Motala12	Stuttgart	Papuan	Chimp	0.0182	3.2	0.011	1.1	Motala12	Stuttgart	Papuan2	Chimp	0.023	3.7		
	Motala12	Stuttgart	Dai	Chimp	0.0156	2.8	0.0149	1.6	Motala12	Stuttgart	Dai2	Chimp	0.02	3.2		
	LaBrana	Stuttgart	Papuan	Chimp	0.0123	2.3	0.0101	1.1	LaBrana	Stuttgart	Papuan2	Chimp	0.02	3.2		
Native Americans closer to ANE than to WHG	LaBrana	Stuttgart	Dai	Chimp	0.0149	2.9	0.0228	2.5	LaBrana	Stuttgart	Dai2	Chimp	0.024	3.7		
	Karitiana	Chimp	MA1	Loschbour	0.0467	7.1	0.0467	4.4	Karitiana2	Chimp	MA1	Loschbour	0.052	7.1		
West Eurasians closer to Native Americans than to other Eastern non-Africans	Stuttgart	Chimp	Karitiana	Papuan	0.0559	10.9	0.0474	6.6	Stuttgart	Chimp	Karitiana2	Papuan2	0.052	7.6		
	Stuttgart	Chimp	Karitiana	Onge	0.0237	5.1	0.0179	2.6								
Ancient Eurasian hunter-gatherers equally related to Eastern non-Africans other than Native Americans	Loschbour	MA1	Dai	Chimp	-0.0015	-0.2	0.0016	0.2	Loschbour	MA1	Dai2	Chimp	-0.013	-1.9		
	Loschbour	MA1	Papuan	Chimp	0.0002	0.0	0.0012	0.1	Loschbour	MA1	Papuan2	Chimp	-0.003	-0.4		
	Loschbour	Motala12	Dai	Chimp	0.0024	0.4	0.009	0.9	Loschbour	Motala12	Dai2	Chimp	-0.002	-0.3		
	Loschbour	Motala12	Papuan	Chimp	-0.0028	-0.4	0.0046	0.5	Loschbour	Motala12	Papuan2	Chimp	-0.004	-0.6		
	MA1	Motala12	Dai	Chimp	0.0026	0.4	0.0047	0.4	MA1	Motala12	Dai2	Chimp	0.01	1.5		
LaBrana and Loschbour are a clade	MA1	Motala12	Papuan	Chimp	-0.0047	-0.7	-0.001	-0.1	MA1	Motala12	Papuan2	Chimp	-0.004	-0.5		
	LaBrana	Loschbour	Dai	Chimp	-0.0028	-0.5	0.0024	0.3	LaBrana	Loschbour	Dai2	Chimp	0.007	1.1		
	LaBrana	Loschbour	Papuan	Chimp	-0.0031	-0.5	-0.0012	-0.1	LaBrana	Loschbour	Papuan2	Chimp	0.002	0.3		
SHG closer to ANE than to WHG	LaBrana	Loschbour	MA1	Chimp	-0.006	-0.8	0.0101	0.7	LaBrana	Loschbour	MA1	Chimp	0.005	0.7		
	Motala12	Loschbour	MA1	Chimp	0.0425	5.3	0.0353	2.6	Motala12	Loschbour	MA1	Chimp	0.042	5.9		
LaBrana and Loschbour equally related to Stuttgart	Motala12	LaBrana	MA1	Chimp	0.0465	5.8	0.0347	2.4	Motala12	LaBrana	MA1	Chimp	0.038	5.4		
	LaBrana	Loschbour	Stuttgart	Chimp	-0.0176	-2.6	-0.0106	-1.0	LaBrana	Loschbour	Stuttgart	Chimp	-0.012	-1.8		

Extended Data Table 3

Admixture proportions for European populations. The estimates from the model with minimal assumptions are from SI17. The estimates from the full modeling are from SI14 either by single population analysis or co-fitting population pairs and averaging over fits (these averages are the results plotted in Fig. 2B). Populations that do not fit the models are not reported.

	Full modeling of population relationships (individual fits)			Full modeling of population relationships (averaged fits)				Modeling of population relationships with minimal assumptions			Model-based (averaged) - Model with minimal assumptions (Z-score)				
	EEF	WHG	ANE	EEF		WHG		ANE		EEF	WHG	ANE	EEF	WHG	ANE
				Mean	Range	Mean	Range	Mean	Range						
Albanian	0.781	0.092	0.127	0.781	0.772-0.819	0.082	0.032-0.098	0.137	0.129-0.158	0.595 ± 0.112	0.353 ± 0.150	0.052 ± 0.049	1.658	-1.807	1.741
Ashkenazi_Jew	0.931	0	0.069							0.938 ± 0.146	-0.021 ± 0.185	0.083 ± 0.049			
Basque	0.593	0.293	0.114	0.569	0.527-0.616	0.335	0.255-0.392	0.096	0.076-0.129	0.569 ± 0.091	0.315 ± 0.124	0.115 ± 0.041	-0.001	0.165	-0.472
Belarusian	0.418	0.431	0.151	0.426	0.397-0.464	0.408	0.338-0.443	0.167	0.150-0.199	0.272 ± 0.094	0.554 ± 0.131	0.174 ± 0.047	1.637	-1.118	-0.158
Bergamo	0.715	0.177	0.108	0.721	0.704-0.793	0.163	0.061-0.189	0.117	0.104-0.147	0.644 ± 0.125	0.248 ± 0.170	0.108 ± 0.053	0.615	-0.503	0.162

	Full modeling of population relationships (individual fits)			Full modeling of population relationships (averaged fits)				Modeling of population relationships with minimal assumptions			Model-based (averaged) - Model with minimal assumptions (Z-score)				
	EEF	WHG	ANE	EEF		WHG		ANE	EEF	WHG	ANE	EEF	WHG	ANE	
				Mean	Range	Mean	Range	Mean	Range						
Bulgarian	0.712	0.147	0.141	0.718	0.707-0.778	0.132	0.047-0.151	0.151	0.138-0.175	0.556 ± 0.110	0.328 ± 0.143	0.116 ± 0.043	1.469	-1.372	0.804
Croatian	0.561	0.293	0.145	0.564	0.548-0.586	0.285	0.242-0.310	0.151	0.137-0.172	0.453 ± 0.122	0.407 ± 0.159	0.140 ± 0.046	0.911	-0.768	0.238
Czech	0.495	0.338	0.167	0.489	0.460-0.531	0.348	0.273-0.382	0.163	0.145-0.196	0.402 ± 0.117	0.400 ± 0.162	0.198 ± 0.050	0.744	-0.322	-0.698
English	0.495	0.364	0.141	0.503	0.476-0.536	0.353	0.296-0.382	0.144	0.130-0.169	0.475 ± 0.091	0.357 ± 0.125	0.168 ± 0.043	0.304	-0.028	-0.561
Estonian	0.322	0.495	0.183	0.323	0.293-0.345	0.49	0.451-0.520	0.187	0.172-0.205	0.072 ± 0.121	0.778 ± 0.176	0.150 ± 0.064	2.070	-1.636	0.584
French	0.554	0.311	0.135	0.563	0.537-0.601	0.297	0.230-0.328	0.14	0.126-0.169	0.498 ± 0.097	0.359 ± 0.127	0.142 ± 0.039	0.672	-0.487	-0.060
French_South	0.675	0.195	0.13	0.636	0.589-0.738	0.256	0.111-0.323	0.108	0.088-0.151	0.636 ± 0.116	0.225 ± 0.165	0.140 ± 0.057	-0.003	0.189	-0.558
Greek	0.792	0.058	0.151	0.791	0.780-0.816	0.048	0.019-0.060	0.161	0.150-0.171	0.658 ± 0.098	0.255 ± 0.127	0.086 ± 0.039	1.357	-1.627	1.915
Hungarian	0.558	0.264	0.179	0.548	0.520-0.590	0.279	0.199-0.313	0.174	0.156-0.210	0.391 ± 0.109	0.454 ± 0.153	0.155 ± 0.050	1.437	-1.145	0.371
Icelandic	0.394	0.456	0.15	0.409	0.386-0.424	0.448	0.409-0.473	0.143	0.126-0.170	0.342 ± 0.102	0.476 ± 0.137	0.182 ± 0.045	0.654	-0.204	-0.861
Lithuanian	0.364	0.464	0.172	0.352	0.327-0.384	0.488	0.433-0.527	0.16	0.135-0.184	0.248 ± 0.117	0.548 ± 0.163	0.205 ± 0.052	0.886	-0.367	-0.864
Maltese	0.932	0	0.068							1.298 ± 0.185	-0.509 ± 0.248	0.211 ± 0.079			
Norwegian	0.411	0.428	0.161	0.417	0.388-0.438	0.423	0.383-0.450	0.16	0.140-0.181	0.273 ± 0.115	0.557 ± 0.161	0.170 ± 0.055	1.252	-0.831	-0.185
Orcadian	0.457	0.385	0.158	0.465	0.439-0.493	0.378	0.329-0.403	0.157	0.140-0.179	0.395 ± 0.088	0.437 ± 0.122	0.168 ± 0.041	0.798	-0.487	-0.264
Sardinian	0.817	0.175	0.008	0.818	0.791-0.874	0.141	0.058-0.182	0.041	0.026-0.068	0.883 ± 0.128	0.075 ± 0.166	0.042 ± 0.048	-0.510	0.400	-0.024
Scottish	0.39	0.428	0.182	0.408	0.387-0.424	0.421	0.384-0.448	0.171	0.149-0.201	0.286 ± 0.112	0.532 ± 0.156	0.182 ± 0.053	1.091	-0.712	-0.210
Sicilian	0.903	0	0.097							1.012 ± 0.149	-0.131 ± 0.199	0.119 ± 0.060			
Spanish	0.809	0.068	0.123	0.759	0.736-0.804	0.126	0.066-0.170	0.115	0.091-0.151	0.856 ± 0.126	-0.015 ± 0.165	0.160 ± 0.049	-0.769	0.855	-0.922
Spanish_North	0.713	0.125	0.163	0.612	0.561-0.660	0.292	0.214-0.365	0.096	0.072-0.126	0.581 ± 0.120	0.298 ± 0.158	0.121 ± 0.046	0.254	-0.038	-0.533
Tuscan	0.746	0.136	0.118	0.751	0.737-0.806	0.123	0.047-0.145	0.126	0.114-0.150	0.734 ± 0.118	0.153 ± 0.160	0.113 ± 0.054	0.141	-0.188	0.249
Ukrainian	0.462	0.387	0.151	0.463	0.445-0.491	0.376	0.322-0.399	0.16	0.148-0.187	0.259 ± 0.123	0.596 ± 0.173	0.145 ± 0.057	1.661	-1.269	0.269
Finnish										-0.299 ± 0.204	1.194 ± 0.296	0.105 ± 0.105			
Mordovian										-0.255 ± 0.173	1.151 ± 0.246	0.104 ± 0.090			
Russian										-0.303 ± 0.211	1.230 ± 0.301	0.072 ± 0.106			

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Authors

Iosif Lazaridis^{1,2}, Nick Patterson², Alissa Mittnik³, Gabriel Renaud⁴, Swapan Mallick^{1,2}, Karola Kirsanow⁵, Peter H. Sudmant⁶, Joshua G. Schraiber⁷, Sergi Castellano⁴, Mark Lipson⁸, Bonnie Berger^{2,8}, Christos Economou⁹, Ruth Bollongino⁵, Qiaomei Fu^{1,4,10}, Kirsten I. Bos³, Susanne Nordenfelt^{1,2}, Heng Li^{1,2}, Cesare de Filippo⁴, Kay Prüfer⁴, Susanna Sawyer⁴, Cosimo Posth³, Wolfgang Haak¹¹, Fredrik Hallgren¹², Elin Fornander¹², Nadin Rohland^{1,2}, Dominique Delsate^{13,14}, Michael Francken¹⁵, Jean-Michel Guinet¹³, Joachim Wahl¹⁶, George Ayodo¹⁷, Hamza A. Babiker^{18,19}, Graciela Bailliet²⁰, Elena Balanovska²¹, Oleg Balanovsky^{21,22}, Ramiro Barrantes²³, Gabriel Bedoya²⁴, Haim Ben-Ami²⁵, Judit Bene²⁶, Fouad Berrada²⁷, Claudio M. Bravi²⁰, Francesca Brisighelli²⁸, George B. J. Busby^{29,30}, Francesco Cali³¹, Mikhail Churnosov³², David E. C. Cole³³, Daniel Corach³⁴, Larissa Damba³⁵, George van Driem³⁶, Stanislav Dryomov³⁷, Jean-Michel Dugoujon³⁸, Sardana A. Fedorova³⁹, Irene Gallego Romero⁴⁰, Marina Gubina³⁵, Michael Hammer⁴¹, Brenna M. Henn⁴², Tor Hervig⁴³, Ugur Hodoglugil⁴⁴, Aashish R. Jha⁴⁰, Sena Karachanak-Yankova⁴⁵, Rita Khusainova^{46,47}, Elza Khusnutdinova^{46,47}, Rick Kittles⁴⁸, Toomas Kivisild⁴⁹, William Klitz⁷, Vaidutis Ku inksas⁵⁰, Alena Kushniarevich⁵¹, Leila Laredj⁵², Sergey Litvinov^{46,47,51},

Theologos Loukidis⁵³, Robert W. Mahley⁵⁴, Béla Melegh²⁶, Ene Metspalu⁵⁵, Julio Molina⁵⁶, Joanna Mountain⁵⁷, Klemetti Näkkäläjärvi⁵⁸, Desislava Nesheva⁴⁵, Thomas Nyambo⁵⁹, Ludmila Osipova³⁵, Jüri Parik⁵⁵, Fedor Platonov⁶⁰, Olga Posukh³⁵, Valentino Romano⁶¹, Francisco Rothhammer^{62,63,64}, Igor Rudan⁶⁵, Ruslan Ruizbakiev⁶⁶, Hovhannes Sahakyan^{51,67}, Antti Sajantila^{68,69}, Antonio Salas⁷⁰, Elena B. Starikovskaya³⁷, Ayele Tarekegn⁷¹, Draga Toncheva⁴⁵, Shahlo Turdikulova⁷², Ingrida Uktveryte⁵⁰, Olga Utevska⁷³, René Vasquez⁷⁴, Mercedes Villena⁷⁴, Mikhail Voevoda^{35,75}, Cheryl Winkler⁷⁶, Levon Yepiskoposyan⁶⁷, Pierre Zalloua^{77,78}, Tatijana Zemunik⁷⁹, Alan Cooper¹¹, Cristian Capelli²⁹, Mark G. Thomas⁸⁰, Andres Ruiz-Linares⁸⁰, Sarah A. Tishkoff⁸¹, Lalji Singh^{82,83}, Kumarasamy Thangaraj⁸², Richard Villems^{51,55,84}, David Comas⁸⁵, Rem Sukernik³⁷, Mait Metspalu⁵¹, Matthias Meyer⁴, Evan E. Eichler^{6,86}, Joachim Burger⁵, Montgomery Slatkin⁷, Svante Pääbo⁴, Janet Kelso⁴, David Reich^{1,2,87,†}, and Johannes Krause^{3,88,90,†}

Affiliations

¹Department of Genetics, Harvard Medical School, Boston, MA, 02115, USA ²Broad Institute of Harvard and MIT, Cambridge, MA, 02142, USA ³Institute for Archaeological Sciences, University of Tübingen, Tübingen, 72074, Germany ⁴Max Planck Institute for Evolutionary Anthropology, Leipzig, 04103, Germany ⁵Johannes Gutenberg University Mainz, Institute of Anthropology, Mainz, D-55128, Germany ⁶Department of Genome Sciences, University of Washington, Seattle, WA, 98195, USA ⁷Department of Integrative Biology, University of California, Berkeley, CA, 94720-3140, USA ⁸Department of Mathematics and Computer Science and Artificial Intelligence Laboratory, Massachusetts Institute of Technology, Cambridge, MA 02139, USA ⁹Archaeological Research Laboratory, Stockholm University, 114 18, Sweden ¹⁰Key Laboratory of Vertebrate Evolution and Human Origins of Chinese Academy of Sciences, IVPP, CAS, Beijing, 100049, China ¹¹Australian Centre for Ancient DNA, School of Earth and Environmental Sciences, University of Adelaide, Adelaide, South Australia, SA 5005, Australia ¹²The Cultural Heritage Foundation, Västerås, 722 12, Sweden ¹³National Museum of Natural History, L-2160, Luxembourg ¹⁴National Center of Archaeological Research, National Museum of History and Art, L-2345, Luxembourg ¹⁵Department of Paleoanthropology, Senckenberg Center for Human Evolution and Paleoenvironment, University of Tübingen, Tübingen, D-72070, Germany ¹⁶State Office for Cultural Heritage Management Baden-Württemberg, Osteology, Konstanz, D-78467, Germany ¹⁷Center for Global Health and Child Development, Kisumu, 40100, Kenya ¹⁸Institutes of Evolution, Immunology and Infection Research, School of Biological Sciences, University of Edinburgh, Edinburgh, EH9 3JT, UK ¹⁹Biochemistry Department, Faculty of Medicine, Sultan Qaboos University, Alkhod, Muscat, 123, Oman ²⁰Laboratorio de Genética Molecular Poblacional, Instituto Multidisciplinario de Biología Celular (IMBICE), CCT-CONICET & CICPBA, La Plata, B1906APO, Argentina ²¹Research Centre for Medical Genetics, Moscow, 115478, Russia ²²Vavilov Institute for General Genetics, Moscow, 119991, Russia ²³Escuela de Biología, Universidad de Costa Rica, San José, 2060, Costa Rica ²⁴Institute of

Biology, Research group GENMOL, Universidad de Antioquia, Medellín, Colombia
²⁵Rambam Health Care Campus, Haifa, 31096, Israel ²⁶Department of Medical Genetics and Szentagothai Research Center, University of Pécs, Pécs, H-7624 Hungary
²⁷Al Akhawayn University in Ifrane (AUI), School of Science and Engineering, Ifrane, 53000, Morocco ²⁸Forensic Genetics Laboratory, Institute of Legal Medicine, Università Cattolica del Sacro Cuore, Rome, 00168, Italy
²⁹Department of Zoology, University of Oxford, Oxford, OX1 3PS, UK ³⁰Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, OX3 7BN, UK
³¹Laboratorio di Genetica Molecolare, IRCCS Associazione Oasi Maria SS, Troina, 94018, Italy ³²Belgorod State University, Belgorod, 308015, Russia ³³Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, Ontario, M5G 1L5, Canada
³⁴Servicio de Huellas Digitales Genéticas, School of Pharmacy and Biochemistry, Universidad de Buenos Aires, 1113 CABA, Argentina ³⁵Institute of Cytology and Genetics, Siberian Branch of Russian Academy of Sciences, Novosibirsk, 630090, Russia
³⁶Institute of Linguistics, University of Bern, Bern, CH-3012, Switzerland ³⁷Laboratory of Human Molecular Genetics, Institute of Molecular and Cellular Biology, Russian Academy of Science, Siberian Branch, Novosibirsk, 630090, Russia
³⁸Anthropologie Moléculaire et Imagerie de Synthèse, CNRS UMR 5288, Université Paul Sabatier Toulouse III, Toulouse, 31000, France ³⁹Yakut Research Center of Complex Medical Problems and North-Eastern Federal University, Yakutsk, 677010, Russia
⁴⁰Department of Human Genetics, University of Chicago, Chicago, IL, 60637, USA ⁴¹ARL Division of Biotechnology, University of Arizona, Tucson, AZ, 85721, USA ⁴²Department of Ecology and Evolution, Stony Brook University, Stony Brook, NY, 11794, USA
⁴³Department of Clinical Science, University of Bergen, Bergen, 5021, Norway ⁴⁴NextBio, part of Illumina, Santa Clara, CA, USA 95050 ⁴⁵Dept. of Medical Genetics, National Human Genome Center, Medical University Sofia, Sofia, 1431, Bulgaria
⁴⁶Institute of Biochemistry and Genetics, Ufa Research Centre, Russian Academy of Sciences, Ufa, 450054, Russia ⁴⁷Department of Genetics and Fundamental Medicine, Bashkir State University, Ufa, 450074, Russia
⁴⁸College of Medicine, University of Illinois at Chicago, Chicago, IL, 60607, USA ⁴⁹Division of Biological Anthropology, University of Cambridge, Cambridge, United Kingdom CB2 1QH ⁵⁰Department of Human and Medical Genetics, Vilnius University, Vilnius, LT-08661, Lithuania
⁵¹Estonian Biocentre, Evolutionary Biology group, Tartu, 51010, Estonia ⁵²Translational Medicine and Neurogenetics, Institut de Génétique et de Biologie Moléculaire et Cellulaire, Illkirch, 67404, France
⁵⁴Gladstone Institutes, San Francisco, CA, 94158, USA ⁵⁵Department of Evolutionary Biology, University of Tartu, Tartu, 51010, Estonia
⁵⁶Centro de Investigaciones Biomédicas de Guatemala, Ciudad de Guatemala, Guatemala ⁵⁷Research Department, 23andMe, Inc. Mountain View, CA, 94043, USA
⁵⁸Cultural Anthropology Program, University of Oulu, Oulu, 90014, Finland ⁵⁹Department of Biochemistry, Muhimbili University of Health and Allied Sciences, Dar es Salaam, Tanzania
⁶⁰Research Institute of Health, North-Eastern Federal University, Yakutsk, 677000, Russia ⁶¹Dipartimento di Fisica e Chimica, Università di Palermo, Palermo, 90128, Italy ⁶²Instituto de Alta Investigación,

Universidad de Tarapacá, Arica, Chile ⁶³Programa de Genética Humana ICBM Facultad de Medicina Universidad de Chile, Santiago, Chile ⁶⁴Centro de Investigaciones del Hombre en el Desierto, Arica, Chile ⁶⁵Centre for Population Health Sciences, The University of Edinburgh Medical School, Edinburgh, Scotland, EH8 9AG, UK ⁶⁶Deceased: formerly of the Institute of Immunology, Academy of Science, Tashkent, 70000, Uzbekistan ⁶⁷Laboratory of Ethnogenomics, Institute of Molecular Biology, National Academy of Sciences of Armenia, Yerevan, 0014, Armenia ⁶⁸Department of Forensic Medicine, Hjelt Institute, University of Helsinki, Helsinki, 00014, Finland ⁶⁹Institute of Applied Genetics, Department of Molecular and Medical Genetics, University of North Texas Health Science Center, Fort Worth, Texas 76107, USA ⁷⁰Unidade de Xenética, Departamento de Anatomía Patolóxica e Ciencias Forenses, and Instituto de Ciencias Forenses, Grupo de Medicina Xenómica (GMX), Facultade de Medicina, Universidade de Santiago de Compostela, Galicia, 15872, Spain ⁷¹Research Fellow, Henry Stewart Group, Russell House, London WC1A 2HN, UK ⁷²Institute of Bioorganic Chemistry Academy of Sciences Republic of Uzbekistan, Tashkent, 100125, Uzbekistan ⁷³Department of Genetics and Cytology, V.N. Karazin Kharkiv National University, Kharkiv, 61077, Ukraine ⁷⁴Instituto Boliviano de Biología de la Altura, Universidad Autónoma Tomás Frías, Potosí, Bolivia ⁷⁵Inst. of Internal Medicine, Siberian Branch of Russian Acad. of Medical Sciences, Novosibirsk, 630089, Russia ⁷⁶Basic Research Laboratory, NCI, NIH, Frederick National Laboratory, Leidos Biomedical, Inc., Frederick, MD 21702, USA ⁷⁷Lebanese American University, School of Medicine, Beirut, 13-5053, Lebanon ⁷⁸Harvard School of Public Health, Boston, 02115, USA ⁷⁹Department of Medical Biology, University of Split, School of Medicine, Split, 21000, Croatia ⁸⁰Department of Genetics, Evolution and Environment, University College London, WC1E 6BT, UK ⁸¹Department of Biology and Genetics. University of Pennsylvania, Philadelphia, Pennsylvania, 19104, USA ⁸²CSIR-Centre for Cellular and Molecular Biology, Hyderabad, 500 007, India ⁸⁴Estonian Academy of Sciences, Tallinn, 10130, Estonia ⁸⁵Institut de Biologia Evolutiva (CSIC-UPF), Departament de Ciències Experimentals i de la Salut, Universitat Pompeu Fabra, Barcelona, 08003, Spain ⁸⁶Howard Hughes Medical Institute, University of Washington, Seattle, WA, USA 98195 ⁸⁷Howard Hughes Medical Institute, Harvard Medical School, Boston, MA, 02115, USA ⁸⁸Senckenberg Centre for Human Evolution and Palaeoenvironment, University of Tübingen, 72070, Germany ⁹⁰Max Planck Institut für Geschichte und Naturwissenschaften, Jena, 07745, Germany

Acknowledgments

We are grateful to Cynthia Beall, Neil Bradman, Amha Gebremedhin, Damian Labuda, Mari Nelis and Anna Di Rienzo for sharing DNA samples; to Detlef Weigel, Christa Lanz, Verena Schünemann, Peter Bauer and Olaf Riess for support and access to DNA sequencing facilities; to Philip Johnson for advice on contamination estimation; to Garrett Hellenthal for help with the ChromoPainter software; and to Pontus Skoglund for sharing graphics software. We thank Kenneth Nordtvedt for alerting us to newly discovered Y-chromosome SNPs. We downloaded the POPRES data from dbGaP at http://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000145.v4.p2 through dbGaP accession number phs000145.v1.p2. We thank all the volunteers who donated DNA. We thank the staff of the Unità Operativa Complessa di Medicina Trasfusionale, Azienda

Ospedaliera Umberto I, Siracusa, Italy for assistance in sample collection; and The National Laboratory for the Genetics of Israeli Populations for facilitating access to DNA. We thank colleagues at the Applied Genomics at the Children's Hospital of Philadelphia, especially Hakon Hakonarson, Cecilia Kim, Kelly Thomas, and Cuiping Hou, for genotyping samples on the Human Origins array. JKr is grateful for support from DFG grant # KR 4015/1-1, the Carl-Zeiss Foundation and the Baden Württemberg Foundation. SP, GR, QF, CF, KP, SC and JKe acknowledge support from the Presidential Innovation Fund of the Max Planck Society. GR was supported by an NSERC fellowship. JGS acknowledges use of the Extreme Science and Engineering Discovery Environment (XSEDE), which is supported by NSF grant number OCI-1053575. EB and OB were supported by RFBR grants 13-06-00670, 13-04-01711, 13-04-90420 and by the Molecular and Cell Biology Program of the Presidium, Russian Academy of Sciences. BM was supported by grants OTKA 73430 and 103983. ASaj was supported by a Finnish Professorpool (Paulo Foundation) Grant. The Lithuanian sampling was supported by the LITGEN project (VP1-3.1-ŠMM-07-K-01-013), funded by the European Social Fund under the Global Grant Measure. AS was supported by Spanish grants SAF2011-26983 and EM 2012/045. OU was supported by Ukrainian SFFS grant F53.4/071. SAT was supported by NIH Pioneer Award 8DP1ES022577-04 and NSF HOMINID award BCS-0827436. KT was supported by an Indian CSIR Network Project (GENESIS: BSC0121). LS was supported by an Indian CSIR Bhatnagar Fellowship. RV, MM, JP and EM were supported by the European Union Regional Development Fund through the Centre of Excellence in Genomics to the Estonian Biocentre and University of Tartu and by an Estonian Basic Research grant SF0270177As08. MM was additionally supported by Estonian Science Foundation grant #8973. JGS and MS were supported by NIH grant GM40282. PHS and EEE were supported by NIH grants HG004120 and HG002385. DR and NP were supported by NSF HOMINID award BCS-1032255 and NIH grant GM100233. DR and EEE are Howard Hughes Medical Institute investigators. This project has been funded in part with federal funds from the National Cancer Institute, National Institutes of Health, under contract HHSN26120080001E. The content of this publication does not necessarily reflect the views or policies of the Department of Health and Human Services, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. Government. This Research was supported in part by the Intramural Research Program of the NIH, National Cancer Institute, Center for Cancer Research.

References

1. Keller A, et al. New insights into the Tyrolean Iceman's origin and phenotype as inferred by whole-genome sequencing. *Nat Commun.* 2012; 3:698. [PubMed: 22426219]
2. Olalde I, et al. Derived immune and ancestral pigmentation alleles in a 7,000-year-old Mesolithic European. *Nature.* 2014; 507:225–228. [PubMed: 24463515]
3. Raghavan M, et al. Upper Palaeolithic Siberian genome reveals dual ancestry of Native Americans. *Nature.* 2014; 505:87–91. [PubMed: 24256729]
4. Skoglund P, et al. Origins and genetic legacy of Neolithic farmers and hunter-gatherers in Europe. *Science.* 2012; 336:466–469. [PubMed: 22539720]
5. Bramanti B, et al. Genetic discontinuity between local hunter-gatherers and Central Europe's first farmers. *Science.* 2009; 326:137–140. [PubMed: 19729620]
6. Haak W, et al. Ancient DNA from European early Neolithic farmers reveals their Near Eastern affinities. *PLoS Biol.* 2010; 8:e1000536. [PubMed: 21085689]
7. Lipson M, et al. Efficient moment-based inference of admixture parameters and sources of gene flow. *Mol Biol Evol.* 2013; 30:1788–1802. [PubMed: 23709261]
8. Patterson N, et al. Ancient admixture in human history. *Genetics.* 2012; 192:1065–1093. [PubMed: 22960212]
9. Krause J, et al. A complete mtDNA genome of an early modern human from Kostenki, Russia. *Curr Biol.* 2010; 20:231–236. [PubMed: 20045327]
10. Sawyer S, Krause J, Guschanski K, Savolainen V, Pääbo S. Temporal patterns of nucleotide misincorporations and DNA fragmentation in ancient DNA. *PLoS ONE.* 2012; 7:e34131. [PubMed: 22479540]
11. Haak W, et al. Ancient DNA from the first European farmers in 7500-Year-old Neolithic sites. *Science.* 2005; 310:1016–1018. [PubMed: 16284177]
12. Perry GH, et al. Diet and the evolution of human amylase gene copy number variation. *Nat Genet.* 2007; 39:1256–1260. [PubMed: 17828263]
13. Alexander DH, Novembre J, Lange K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res.* 2009; 19:1655–1664. [PubMed: 19648217]
14. Patterson N, Price AL, Reich D. Population structure and eigenanalysis. *PLoS Genet.* 2006; 2:e190. [PubMed: 17194218]

15. Reich D, Thangaraj K, Patterson N, Price AL, Singh L. Reconstructing Indian population history. *Nature*. 2009; 461:489–494. [PubMed: 19779445]
16. Moorjani P, et al. Genetic evidence for recent population mixture in India. *Am J Hum Genet*. 2013; 93:422–438. [PubMed: 23932107]
17. Reich D, et al. Reconstructing Native American population history. *Nature*. 2012; 488:370–374. [PubMed: 22801491]
18. Botigué LR, et al. Gene flow from North Africa contributes to differential human genetic diversity in southern Europe. *Proceedings of the National Academy of Sciences*. 2013
19. Cerezo M, et al. Reconstructing ancient mitochondrial DNA links between Africa and Europe. *Genome Res*. 2012; 22:821–826. [PubMed: 22454235]
20. Moorjani P, et al. The history of African gene flow into southern Europeans, Levantines, and Jews. *PLoS Genet*. 2011; 7:e1001373. [PubMed: 21533020]
21. Pickrell JK, Pritchard JK. Inference of population splits and mixtures from genome-wide Allele frequency data. *PLoS Genet*. 2012; 8:e1002967. [PubMed: 23166502]
22. Fu Q, et al. DNA analysis of an early modern human from Tianyuan Cave, China. *Proc Natl Acad Sci USA*. 2013; 110:2223–2227. [PubMed: 23341637]
23. Bar-Yosef, O. *The chronology of the Middle Paleolithic of the Levant*. New York: Plenum Press; 1998. p. 39–56.
24. Armitage SJ, et al. The southern route “Out of Africa”: evidence for an early expansion of modern humans into Arabia. *Science*. 2011; 331:453–456. [PubMed: 21273486]
25. Rose JI, et al. The Nubian Complex of Dhofar, Oman: an African middle stone age industry in Southern Arabia. *PLoS ONE*. 2011; 6:e28239. [PubMed: 22140561]
26. Brace CL, et al. The questionable contribution of the Neolithic and the Bronze Age to European craniofacial form. *Proc Natl Acad Sci U S A*. 2006; 103:242–247. [PubMed: 16371462]
27. Browning BL, Browning SR. Improving the Accuracy and Efficiency of Identity-by-Descent Detection in Population Data. *Genetics*. 2013; 194:459–471. [PubMed: 23535385]
28. Ralph P, Coop G. The geography of recent genetic ancestry across Europe. *PLoS Biol*. 2013; 11:e1001555. [PubMed: 23667324]
29. Lawson DJ, Hellenthal G, Myers S, Falush D. Inference of Population Structure using Dense Haplotype Data. *PLoS Genet*. 2012; 8:e1002453. [PubMed: 22291602]
30. Brandt G, et al. Ancient DNA reveals key stages in the formation of central European mitochondrial genetic diversity. *Science*. 2013; 342:257–261. [PubMed: 24115443]
31. Delsate D, Guinet JM, Saverwyns S. De l’ocre sur le crâne mésolithique (haplogroupe U5a) de Reuland-Loschbour (Grand-Duché de Luxembourg) ? *Bull Soc Préhist Luxembourgeoise*. 2009; 31:7–30.
32. Rohland N, Hofreiter M. Ancient DNA extraction from bones and teeth. *Nat Protocols*. 2007; 2:1756–1762.
33. Dabney J, et al. Complete mitochondrial genome sequence of a Middle Pleistocene cave bear reconstructed from ultrashort DNA fragments. *Proceedings of the National Academy of Sciences*. 2013; 110:15758–15763.
34. Stäuble, HSfV-uFdUF. Häuser und absolute Datierung der Ältesten Bandkeramik. *Habelt*: 2005.
35. Yang DY, Eng B, Wayne JS, Dudar JC, Saunders SR. Improved DNA extraction from ancient bones using silica-based spin columns. *Am J Phys Anthropol*. 1998; 105:539–543. [PubMed: 9584894]
36. Meyer M, Kircher M. Illumina sequencing library preparation for highly multiplexed target capture and sequencing. *Cold Spring Harb Protoc*. 2010; 2010 pdb prot5448.
37. Meyer M, et al. A High-Coverage Genome Sequence from an Archaic Denisovan Individual. *Science*. 2012; 338:222–226. [PubMed: 22936568]
38. Briggs AW, et al. Removal of deaminated cytosines and detection of in vivo methylation in ancient DNA. *Nucleic Acids Res*. 2010; 38:e87–e87. [PubMed: 20028723]
39. Kircher M. *Methods Mol Biol Vol. 840 Methods in Molecular Biology*. 2012:197–228.
40. Li H, Durbin R. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics*. 2009; 25:1754–1760. [PubMed: 19451168]

41. McKenna A, et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* 2010; 20:1297–1303. [PubMed: 20644199]
42. Maricic T, Whitten M, Pääbo S. Multiplexed DNA Sequence Capture of Mitochondrial Genomes Using PCR Products. *PLoS ONE.* 2010; 5:e14004. [PubMed: 21103372]
43. Behar, Doron M., et al. A Copernican Reassessment of the Human Mitochondrial DNA Tree from its Root. *Am J Hum Genet.* 2012; 90:675–684. [PubMed: 22482806]
44. Green RE, et al. A Complete Neandertal Mitochondrial Genome Sequence Determined by High-Throughput Sequencing. *Cell.* 2008; 134:416–426. [PubMed: 18692465]
45. Fu Q, et al. A Revised Timescale for Human Evolution Based on Ancient Mitochondrial Genomes. *Curr Biol.* 2013; 23:553–559. [PubMed: 23523248]
46. Fu Q. 2014 (in preparation).
47. Rasmussen M, et al. An Aboriginal Australian Genome Reveals Separate Human Dispersals into Asia. *Science.* 2011; 334:94–98. [PubMed: 21940856]
48. Vianello D, et al. HAPLOFIND: a new method for high-throughput mtDNA haplogroup assignment. *Hum Mutat.* 2013; 34:1189–1194. [PubMed: 23696374]
49. Tamura K, et al. MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Mol Biol Evol.* 2011; 28:2731–2739. [PubMed: 21546353]
50. Skoglund P, Storå J, Götherström A, Jakobsson M. Accurate sex identification of ancient human remains using DNA shotgun sequencing. *J Archaeol Sci.* 2013; 40:4477–4482.
51. Drummond A, Rambaut A. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evol Biol.* 2007; 7:214. [PubMed: 17996036]
52. Lippold S, et al. Human paternal and maternal demographic histories: insights from high-resolution Y chromosome and mtDNA sequences. *bioRxiv.* 201410.1101/001792
53. Green RE, et al. A Draft Sequence of the Neandertal Genome. *Science.* 2010; 328:710–722. [PubMed: 20448178]
54. Reich D, et al. Genetic history of an archaic hominin group from Denisova Cave in Siberia. *Nature.* 2010; 468:1053–1060. [PubMed: 21179161]
55. Prufer K, et al. The complete genome sequence of a Neanderthal from the Altai Mountains. *Nature.* 2014; 505:43–49. [PubMed: 24352235]
56. Li H, Durbin R. Inference of human population history from individual whole-genome sequences. *Nature.* 2011; 475:493–496. [PubMed: 21753753]
57. Hach F, et al. mrsFAST: a cache-oblivious algorithm for short-read mapping. *Nat Meth.* 2010; 7:576–577.
58. An integrated map of genetic variation from 1, 092 human genomes. *Nature.* 2012; 491:56–65. [PubMed: 23128226]
59. Danecek P, et al. The variant call format and VCFtools. *Bioinformatics.* 2011; 27:2156–2158. [PubMed: 21653522]
60. Li H. The sequence alignment/map (SAM) format and SAMtools. *Bioinformatics.* 2009; 25:2078–2079. [PubMed: 19505943]
61. Keinan A, Mullikin JC, Patterson N, Reich D. Measurement of the human allele frequency spectrum demonstrates greater genetic drift in East Asians than in Europeans. *Nat Genet.* 2007; 39:1251–1255. [PubMed: 17828266]
62. Price AL, et al. Principal components analysis corrects for stratification in genome-wide association studies. *Nat Genet.* 2006; 38:904–909. [PubMed: 16862161]
63. Purcell S, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet.* 2007; 81:559–575. [PubMed: 17701901]
64. Alexander D, Lange K. Enhancements to the ADMIXTURE algorithm for individual ancestry estimation. *BMC Bioinformatics.* 2011; 12:246. [PubMed: 21682921]
65. Jakobsson M, Rosenberg NA. CLUMPP: a cluster matching and permutation program for dealing with label switching and multimodality in analysis of population structure. *Bioinformatics.* 2007; 23:1801–1806. [PubMed: 17485429]

66. Price AL, Zaitlen NA, Reich D, Patterson N. New approaches to population stratification in genome-wide association studies. *Nat Rev Genet.* 2010; 11:459–463. [PubMed: 20548291]
67. Busing FTA, Meijer E, Leeden R. Delete-m Jackknife for Unequal m. *Statistics and Computing.* 1999; 9:3–8.
68. Loh PR, et al. Inferring Admixture Histories of Human Populations Using Linkage Disequilibrium. *Genetics.* 2013; 193:1233–1254. [PubMed: 23410830]
69. Nelson MR, et al. The Population Reference Sample, POPRES: a resource for population, disease, and pharmacological genetics research. *Am J Hum Genet.* 2008; 83:347–358. [PubMed: 18760391]

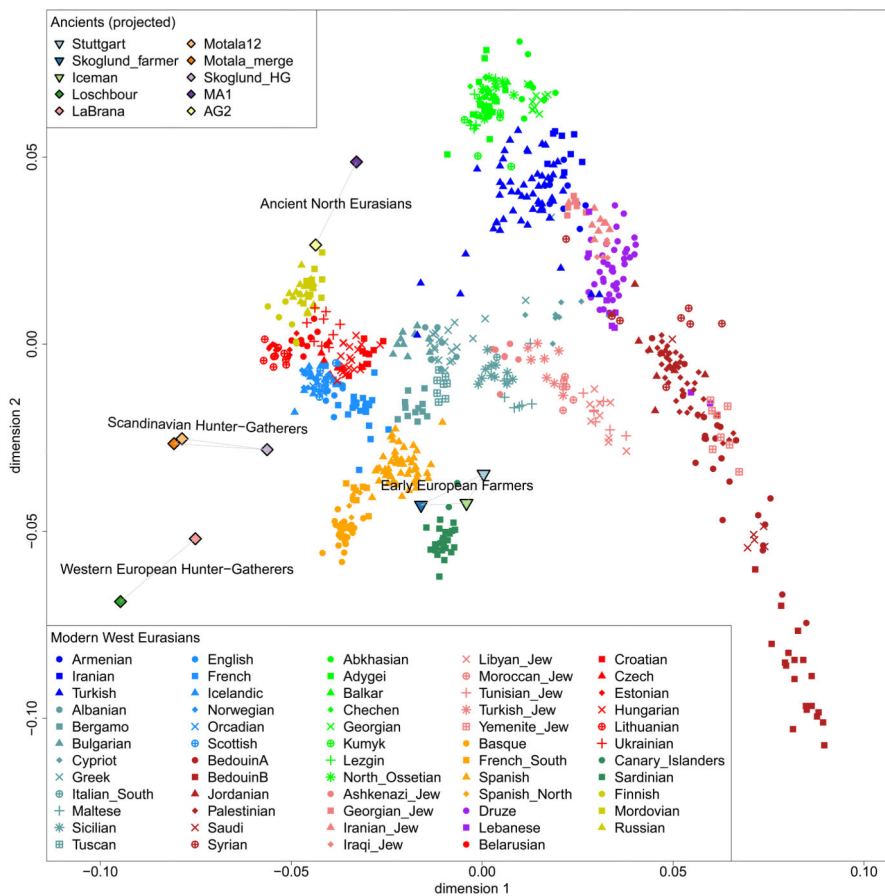
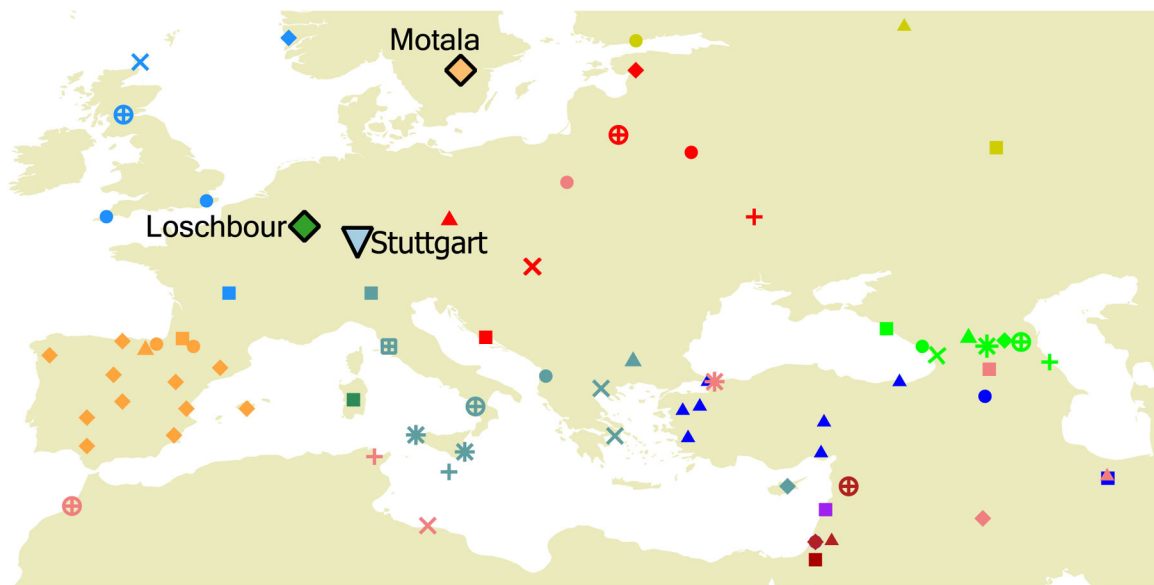
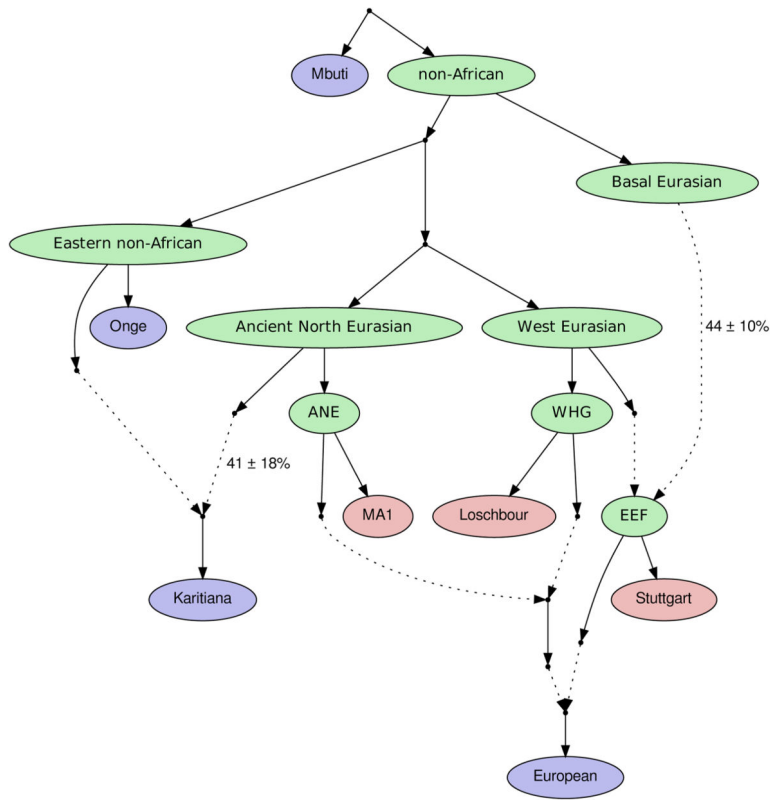


Figure 1. Map of West Eurasian populations and Principal Component Analysis
 (a) Geographical locations of analyzed samples, with color coding matching the PCA. We show all sampling locations for each population, which results in multiple points for some

(e.g., Spain). (b) PCA on all present-day West Eurasians, with ancient and selected eastern non-African samples projected. European hunter-gatherers fall beyond present-day Europeans in the direction of European differentiation from the Near East. Stuttgart clusters with other Neolithic Europeans and present-day Sardinians. MA1 falls outside the variation of present-day West Eurasians in the direction of southern-northern differentiation along dimension 2.



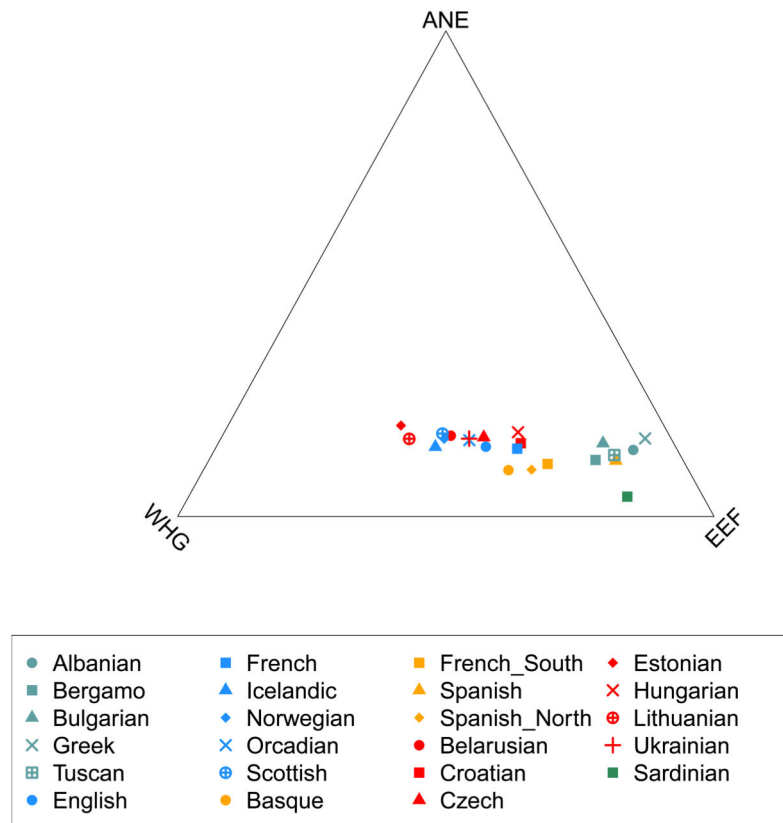


Figure 2. Modeling of West Eurasian population history

(a) A three-way mixture model that is a fit to the data for many populations. Present-day samples are colored in blue, ancient in red, and reconstructed ancestral populations in green. Solid lines represent descent without mixture, and dashed lines represent admixture. We print mixture proportions and one standard error for the two mixtures relating the highly divergent ancestral populations. (We do not print the estimate for the “European” population as it varies depending on the population). (b) We plot the proportions of ancestry from each of three inferred ancestral populations (EEF, ANE and WHG).

Table 1Lowest f_3 -statistics for each West Eurasian population

<i>Ref₁</i>	<i>Ref₂</i>	Target for which these two references give the lowest $f_3(X; Ref_1, Ref_2)$
WHG	EEF	Sardinian***
WHG	Near East	Basque, Belarusian, Czech, English, Estonian, Finnish, French_South, Icelandic, Lithuanian, Mordovian, Norwegian, Orcadian, Scottish, Spanish, Spanish_North, Ukrainian
WHG	Siberian	Russian
EEF	ANE	Abkhasian***, Albanian, Ashkenazi_Jew****, Bergamo, Bulgarian, Chechen****, Croatian, Cypriot****, Druze**, French, Greek, Hungarian, Lezgin, Maltese, Sicilian, Turkish_Jew, Tuscan
EEF	Native American	Adygei, Balkar, Iranian, Kumyk, North_Ossetian, Turkish
EEF	African	BedouinA, BedouinB [†] , Jordanian, Lebanese, Libyan_Jew, Moroccan_Jew, Palestinian, Saudi****, Syrian, Tunisian_Jew***, Yemenite_Jew***
EEF	South Asian	Armenian, Georgian****, Georgian_Jew*, Iranian_Jew***, Iraqi_Jew***

Note: WHG = Loschbour or LaBraña; EEF=Stuttgart; ANE=MA1; Native American=Piapoco; African=Esan, Gambian, or Kgalagadi; South Asian=GujaratiC or Vishwabrahmin. Statistics are negative with $Z < -4$ unless otherwise noted: [†] (positive) or *, **, ***, ****, to indicate Z less than 0, -1, -2, and -3 respectively. The complete list of statistics can be found in Extended Data Table 1.