Ferritin levels in the cerebrospinal fluid predict Alzheimer’s disease outcomes and are regulated by APOE

Scott Ayton1,*, Noel G. Faux2,3,*, Ashley I. Bush1,3 & Alzheimer’s Disease Neuroimaging Initiative†

Brain iron elevation is implicated in Alzheimer’s disease (AD) pathogenesis, but the impact of iron on disease outcomes has not been previously explored in a longitudinal study. Ferritin is the major iron storage protein of the body; by using cerebrospinal fluid (CSF) levels of ferritin as an index, we explored whether brain iron status impacts longitudinal outcomes in the Alzheimer’s Disease Neuroimaging Initiative (ADNI) cohort. We show that baseline CSF ferritin levels were negatively associated with cognitive performance over 7 years in 91 cognitively normal, 144 mild cognitive impairment (MCI) and 67 AD subjects, and predicted MCI conversion to AD. Ferritin was strongly associated with CSF apolipoprotein E levels and was elevated by the Alzheimer’s risk allele, APOE-ε4. These findings reveal that elevated brain iron adversely impacts on AD progression, and introduce brain iron elevation as a possible mechanism for APOE-ε4 being the major genetic risk factor for AD.
Cortical iron elevation is increasingly reported as a feature of Alzheimer’s disease (AD)\(^1\), and might contribute to the oxidative damage observed in AD brains\(^2\). A single-blind, 2-year trial of 48 AD patients with the iron chelator, deferoxamine, reported decreased cognitive decline\(^3\), but this has not been followed up. While evidence in animal models argue in favour of brain iron accumulation propelling atrophy and dementia\(^4\), prospective evidence about the link between brain iron status and clinical outcomes in AD is lacking.

CSF ferritin could be an index of brain iron load. Ferritin is the iron storage protein of the body and is elevated in AD brain tissue\(^5–8\). In cultured systems, ferritin expression\(^9,10\) and secretion\(^11\) by glia is dependent on cellular iron levels. Ferritin is the iron storage protein of the body and is elevated in AD brain tissue\(^5–8\). In cultured systems, ferritin expression\(^9,10\) and secretion\(^11\) by glia is dependent on cellular iron levels.

Here, we examined the association of baseline CSF-ferritin data with biomarker, cognitive, anatomical and diagnostic outcomes over 7 years in the Alzheimer’s disease Neuroimaging Initiative (ADNI) prospective clinical cohort. We show that CSF ferritin levels have similar utility compared with more established AD CSF biomarkers, the tau/\(\beta\)\(_{1-42}\) ratio and apolipoprotein E (ApoE) levels, in predicting various outcomes of AD. However, the nature of the relationship between CSF ferritin levels and cognitive performance was different from the other biomarkers, and, in contrast, CSF ferritin appears as a trait variable, and not a marker of disease.

**Results**

The relationship between CSF ferritin and biomarkers of AD.

In agreement with other reports\(^3,4,15\), CSF ferritin levels were not different between cognitively normal (CN; \(n = 91\)) and AD (\(n = 67\)) subjects (ANOVA: \(P = 0.591\); Table 1) in the ADNI cohort. Neither were there changes in ferritin levels when we separated the cohort according to CSF \(\beta\)\(_{1-42}\) levels (192 ng l\(^{-1}\) cutoff; as proposed previously\(^16\)) to reflect likely cerebral amyloid burden (ANOVA: \(P = 0.946\); Supplementary Fig. 1). But in multiple regression modelling of ferritin including the established CSF biomarkers of AD\(^17\), tau, p-tau, \(\beta\)\(_{1-42}\), CSF ferritin levels were predicted by \(\beta\)\(_{1-42}\) (partial \(R^2 = 0.013, P = 0.029\)) and tau (partial \(R^2 = 0.086, P < 0.001\); model 1, Supplementary Table 1), although not by p-tau. Since the apolipoprotein E gene (ApoE) alleles are the major genetic risk for AD\(^18\) and CSF apolipoprotein E protein (ApoE) levels are associated with \(\beta\)\(_{1-42}\) (refs 19,20) and tau\(^20,21\), we re-built the model to include CSF ApoE levels. This abolished the relationship between ferritin and the other biomarkers (\(\beta\)\(_{1-42}: R^2 < 0.001, P = 0.904\); tau: \(R^2 = 0.003, P = 0.219\); model 2, Supplementary Table 1). This led us to detect a surprisingly strong relationship between ApoE and ferritin (linear term partial \(R^2 = 0.243, P = 7.69 \times 10^{-22}\)), which was improved when \(\beta\)\(_{1-42}\) and tau

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Baseline characteristics of subjects from the ADNI cohort used in this study, stratified by diagnosis.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Units</td>
<td>CN</td>
</tr>
<tr>
<td>n</td>
<td>—</td>
</tr>
<tr>
<td>Age</td>
<td>Years (s.d.)</td>
</tr>
<tr>
<td>Female</td>
<td>n (%)</td>
</tr>
<tr>
<td>Education</td>
<td>Years (s.d.)</td>
</tr>
<tr>
<td>APOE-(\varepsilon )4 + ve</td>
<td>n (%)</td>
</tr>
<tr>
<td>ADAS-Cog13</td>
<td>Units (s.d.)</td>
</tr>
<tr>
<td>CSF Ferritin</td>
<td>ng ml(^{-1}) (s.d.)</td>
</tr>
<tr>
<td>CSF ApoE</td>
<td>(\mu)g ml(^{-1}) (s.d.)</td>
</tr>
<tr>
<td>CSF tau</td>
<td>(\mu)g ml(^{-1}) (s.d.)</td>
</tr>
<tr>
<td>CSF ptau</td>
<td>(\mu)g ml(^{-1}) (s.d.)</td>
</tr>
<tr>
<td>CSF (\beta)(_{1-42})</td>
<td>(\mu)g ml(^{-1}) (s.d.)</td>
</tr>
<tr>
<td>CSF tau/(\beta)(_{1-42}) Units (s.d.)</td>
<td>0.39 (0.26)</td>
</tr>
<tr>
<td>Hippocampus</td>
<td>mm(^3) (s.d.)</td>
</tr>
<tr>
<td>Lateral ventricle</td>
<td>mm(^3) (s.d.)</td>
</tr>
</tbody>
</table>

AD, Alzheimer’s disease; CN, cognitively normal; CSF, cerebrospinal fluid; MCI, mild cognitive impairment. Unadjusted unit values are presented in the table. P values presented for ANCOVA models of CSF analytes and MRI brain structure were adjusted for age, gender, years of education, BMI, ApoE genotype, CSF haemoglobin and CSF Factor H. Intracranial volume was also included in ANCOVA models of brain structure.
(non-significant terms) were removed from the model (linear term partial $R^2 = 0.341$, $P = 1.52 \times 10^{-29}$; model 3, Supplementary Table 1, Fig. 1a).

In model 3, APOE genotype strongly influenced CSF ferritin ($P = 1.10 \times 10^{-8}$), with the major AD risk allele, $e4$, inducing 22% higher levels than non-$e4$ carriers (Fig. 1b). Reciprocally, in multiple regression modelling of CSF ApoE, APOE $e4$-positive subjects had lower ApoE levels (-16%; $P = 2.50 \times 10^{-5}$) compared with non-$e4$ carriers (Fig. 1c). Plasma ferritin levels were not associated with plasma ApoE levels or APOE $e4$ allele status (Supplementary Fig. 2), but there was a modest association between plasma ferritin and CSF ferritin levels ($\beta = 0.075$, $P = 0.0002$; Supplementary Fig. 3).

Association of ferritin with neuropsychiatric assessments. Next, we explored whether CSF ferritin was related to cognitive performance in AD. Baseline ADAS-Cog13 (The Alzheimer’s Disease Assessment Scale) score was associated with CSF ferritin ($P = 0.006$; Table 2), ApoE levels ($P = 0.0003$) and tau/Ab$_{1–42}$ ratio ($P = 0.025$), independently, in a multiple regression model containing the AD biomarkers and other clinical variables. In tertile analysis, high ($>7.2 \text{ ng ml}^{-1}$), compared with low ($<5.4 \text{ ng ml}^{-1}$), levels of ferritin were associated with an $\approx 3$ point poorer ADAS-cog13 score (Fig. 2a). Similarly, in tertiles, lower levels of ApoE (Fig. 2b) were associated with a $\approx 4$ point worse ADAS-Cog13, and higher tau/Ab$_{1–42}$ ratio was associated with a $\approx 2$ point worse ADAS-Cog13 (Fig. 2c), as previously reported.$^{21,22}$

To determine whether baseline values of CSF ferritin predict longitudinal cognitive outcome, we constructed a mixed effects model of annual ADAS-Cog13 scores over 7 years (Table 2 for statistics, Supplementary Table 2 for patient numbers) and observed that both ApoE ($P = 0.006$) and tau/Ab$_{1–42}$ ratio (cross-sectional cognition ($MR$)) were associated with CSF ferritin ($P = 1.10 \times 10^{-8}$), with the major AD risk allele, $e4$, inducing 22% higher levels than non-$e4$ carriers (Fig. 1b). Reciprocally, in multiple regression modelling of CSF ApoE, APOE $e4$-positive subjects had lower ApoE levels (-16%; $P = 2.50 \times 10^{-5}$) compared with non-$e4$ carriers (Fig. 1c). Plasma ferritin levels were not associated with plasma ApoE levels or APOE $e4$ allele status (Supplementary Fig. 2), but there was a modest association between plasma ferritin and CSF ferritin levels ($\beta = 0.075$, $P = 0.0002$; Supplementary Fig. 3).

Figure 2 | CSF ferritin levels independently predict cognitive status. (a-c) Multiple regression of baseline ADAS-Cog13 score expressed as tertiles of CSF (a) ferritin ($L < 5.5, H > 7.3 \text{ ng ml}^{-1}$), (b) ApoE ($L < 5.8, H > 7.8 \mu g \text{ ml}^{-1}$) and (c) tau/Ab$_{1–42}$ ($L < 0.35, H > 0.76$). (d) Multiple regression of baseline RAVLT score expressed as CSF ferritin tertiles. Data are adjusted for baseline diagnosis, gender, years of education and the AD CSF biomarkers in the minimal models. Data are means $\pm$ s.e. ‘n’ is shown in graph columns. CN, cognitively normal; MCI, mild cognitive impairment.

Table 2 | Modelling the association of CSF biomarkers on AD outcomes.

<table>
<thead>
<tr>
<th>Model</th>
<th>Ferritin*</th>
<th>tau/Ab$_{1–42}$</th>
<th>ApoE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cross-sectional cognition ($MR$)</td>
<td>$\beta$ (s.e.)</td>
<td>$P$ value</td>
<td>$\beta$ (s.e.)</td>
</tr>
<tr>
<td>ADAS-Cog13$^\dagger$</td>
<td>0.139 (0.050)</td>
<td>0.006</td>
<td>0.104 (0.046)</td>
</tr>
<tr>
<td>RAVLT</td>
<td>-1.77 (0.559)</td>
<td>0.0017</td>
<td>NS</td>
</tr>
<tr>
<td>Longitudinal cognition ($MELM$)</td>
<td>$\beta$ (s.e.)</td>
<td>$P$ value</td>
<td>$\beta$ (s.e.)</td>
</tr>
<tr>
<td>ADAS-Cog13$^\dagger$</td>
<td>0.0178 (0.0051)</td>
<td>0.0005</td>
<td>0.129 (0.049)</td>
</tr>
<tr>
<td>Interaction time</td>
<td>0.0005 (0.0016)</td>
<td>0.977</td>
<td>0.081 (0.016)</td>
</tr>
<tr>
<td>RAVLT</td>
<td>-1.60 (0.63)</td>
<td>0.012</td>
<td>-0.847 (0.608)</td>
</tr>
<tr>
<td>Interaction time</td>
<td>-0.035 (0.152)</td>
<td>0.817</td>
<td>-0.610 (0.150)</td>
</tr>
<tr>
<td>MCI conversion to AD</td>
<td>Statistic$^\dagger$</td>
<td>$P$ value</td>
<td>Statistic$^\dagger$</td>
</tr>
<tr>
<td>Cox (Hazard ratio)</td>
<td>1.10 (1.01-1.19)</td>
<td>0.030</td>
<td>1.53 (1.03-2.28)</td>
</tr>
<tr>
<td>LR (Odds ratio)</td>
<td>2.32 (1.86-2.90)</td>
<td>8.001 $\times 10^{-15}$</td>
<td>1.45 (1.16-1.80)</td>
</tr>
<tr>
<td>Rate of MRI atrophy ($MELM$)</td>
<td>$\beta$ (s.e.)</td>
<td>$P$ value</td>
<td>$\beta$ (s.e.)</td>
</tr>
<tr>
<td>Hippocampus</td>
<td>-18.33 (7.86)</td>
<td>0.019</td>
<td>-35.31 (7.79)</td>
</tr>
<tr>
<td>Lateral ventricles$^\ddagger$</td>
<td>0.007 (0.003)</td>
<td>0.008</td>
<td>0.013 (0.002)</td>
</tr>
</tbody>
</table>

*Ferritin values were log-transformed, excluding non-parametric Cox and LR models.
†The $k$-coefficient is for the square root of ADAS-Cog13.
‡The statistics for the conversion models were based on one interquartile range change for each analyte (ferritin: 3.3 ng ml$^{-1}$, tau/Ab$_{1–42}$: 0.67 units; ApoE: 3.1 μg ml$^{-1}$).
§For Lateral ventricles, the $k$-coefficient is for natural log of the ventricle volume.

**Cox, Cox proportional hazard model; LR, logistic regression; MELM, mixed effects linear model; MR, multiple regression; NS, not significant. All models initially contained the variables: age, gender, BMI, APOE genotype, baseline diagnosis; the MRL models additionally included intracranial volume. Minimal models for the cognition models included baseline diagnosis, gender, years of education and the AD CSF biomarkers. Minimal model for the Cox proportional hazard model (Cox) contained only the AD CSF biomarkers. Minimal models for the MRL models contained age, gender, baseline diagnosis, years of education, APOE $e4$ status and intracranial volume. All subjects with available data were included in the cross-sectional cognition models only. CN and MCI subjects were included in modelling of longitudinal cognition because short follow up of AD subjects (Supplementary Table 3). Only subjects who were classified as MCI at baseline were included in the MCI conversion models. The MRL models contained subjects who were classified as cognitively normal or MCI at baseline. AD subjects at baseline were not included because of low numbers and lack of follow-up (Supplementary Tables 3).

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§For Lateral ventricles, the $k$-coefficient is for natural log of the ventricle volume."
(P = 2.70 × 10⁻⁷) were still associated with rate of cognitive change (interacted with time), as previously reported²⁻⁴. Ferritin, however, impacted on ADAS-Cog¹³ by a constant cross-sectional decrement (P = 4.93 × 10⁻⁴ main effect only; Table 2).

We additionally modelled cognition using the Rey verbal learning test (RAVLT), which is more sensitive in distinguishing control and MCI patients²⁻⁴. In this model, only ferritin levels were associated with cross-sectional cognitive performance (P = 0.0017; Table 2, Fig. 2d), but CSF ferritin was not associated with rate of deterioration in a longitudinal model (P = 0.817; Table 2). Baseline tau/Åβ₁–₄₂ ratio was associated with rate of cognitive decline on RAVLT (P = 4.8510 × 10⁻⁴), but there was only a trend for ApoE (P = 0.066). Hence, in both cognitive scales, CSF ferritin impacted on performance by a constant amount, regardless of disease status, thus appearing as a trait variable and not a marker of disease.

We reasoned that if high ferritin levels worsened the cognitive performance by a constant value over time, then MCI individuals with high ferritin levels would satisfy the criteria for an AD diagnosis at a comparatively earlier interval. To investigate this, we employed a Cox proportional hazards model on 144 MCI subjects who had CSF ferritin, ApoE and tau/Åβ₁–₄₂ measurements. In a minimal model (containing only these CSF biomarkers; Table 2) of MCI conversion over 7 years, ferritin (P = 0.03; Fig. 3a), ApoE (P = 0.008; Supplementary Fig. 4a) and tau/Åβ₁–₄₂ (P = 0.037; Supplementary Fig. 4b) were each significant predictive variables.

Using this model we estimated how many months was required for 50% survivorship for each quintile of each biomarker. We then gated whether ferritin levels associate with neuroanatomical changes of the hippocampus and lateral ventricular area in yearly intervals over a 6-year period for CN and MCI subjects (Supplementary Table 3 for patient numbers). We explored the impact of CSF ferritin when the other biomarkers were also included in modelling, whereas CSF ferritin has previously been included in modelling, whereas CSF ferritin has previously been

In separate adjusted logistic regression models, an increase in the baseline concentration of each biomarker by its interquartile range increased the odds of converting to AD for ferritin (OR: 1.36, 95%CI: 1.17–1.58) and tau/Åβ₁–₄₂ ratio (OR: 1.13, CI: 0.95–1.35), and decreased the odds for ApoE (OR: 0.72, CI: 0.61–0.85). Including all three analytes into the one model increased the predictive value of each analyte (OR (CI): ferritin = 2.32 (1.86–2.9), tau/Åβ₁–₄₂ = 1.45 [1.16–1.8], ApoE = 0.38 [0.3–0.48]; Table 2).

Receiver-operating curves based on the logistic regression models determined the accuracy of these analytes to predict conversion to AD. The area under the curve (AUC) of the base model (age, gender, years of education, BMI, APOE ε4 genotype) was 0.6079 (Fig. 3c), which was increased by the singular inclusions of either ferritin (AUC: 0.6321; Supplementary Fig. 5b), ApoE (0.6311; Supplementary Fig. 5c) or marginally by tau/Åβ₁–₄₂ (0.6177; Supplementary Fig. 5d). When the tau/Åβ₁–₄₂ was included in the model containing ApoE, the AUC increased slightly (from 0.6311 to 0.6483; Fig. 3d). This performance, which combined the established CSF biomarkers for AD, was improved markedly by adding ferritin values (from 0.6483 to 0.6937 Fig. 3c).

**Association of ferritin with brain atrophy.** Finally, we investigated whether ferritin levels associate with neuroanatomical changes to the hippocampus and lateral ventricular area in yearly intervals over a 6-year period for CN and MCI subjects (Supplementary Table 3 for patient numbers). We explored the impact of CSF ferritin when the other biomarkers were also included in modelling, whereas CSF ferritin has previously been

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**Figure 3 | Conversion from MCI to dementia as predicted by baseline CSF biomarkers.** (a) MCI survival based on the minimal Cox proportional hazards model (Table 2), the conversion is plotted for each quintile of ferritin (applying mean values for the cohort: ApoE = 7.2 μg ml⁻¹, tau/Åβ₁–₄₂ = 0.69 units). The numbers on the right side of the graphs indicate the quintile boundaries. This minimal model contained only the CSF biomarkers. (b) Change in mean age of diagnosis according to CSF biomarkers. The months taken for ~50% survival of each quintile boundary in the Cox models were graphed against the unit values of those boundaries. The gradient of the linear model was used to estimate change in age of conversion for each unit change in analyte. (Refer to Fig. 3a, Supplementary Fig. 4). (c-e) Receiver operating curves of logistic regression modelling of MCI conversion to AD (refer to Table 2, Supplementary Fig. 5). (c) Base model controlling for age, gender, BMI, years of education and APOE ε4 status. (d) Base model plus ApoE and tau/Åβ₁–₄₂. (e) Base model plus ApoE, tau/Åβ₁–₄₂ and ferritin. AUC, area under curve.
shown to predict atrophy of various brain structures when considered in isolation. Baseline APOE, ferritin and tau/Aβ1–42 values each independently predicted hippocampal volume in an adjusted longitudinal model (Table 2). The rate of atrophy of the hippocampus was greater in individuals with high CSF ferritin (P = 0.02; Fig. 4a). Low CSF APOE (P = 0.008; Fig. 4b) or high tau/Aβ1–42 (P = 6.80 × 10−6; Fig. 4c) also predicted atrophy, as previously reported. Lateral ventricular enlargement over time was similarly associated independently with high-CSF ferritin (P = 0.008; Fig. 4d), low-CSF APOE (P = 0.0002; Fig. 4e), or high tau/Aβ1–42 (P = 4.19 × 10−8, Fig. 4f).

Discussion

Our analyses show that CSF ferritin levels were independently related to cognitive performance in the ADNI cohort and predicted MCI conversion to AD. The magnitude impact of ferritin on these outcomes was comparable to the established effect of ferritin on these outcomes was comparable to the established effect of tau/Aβ1–42 (units)*17 (refer to Table 2). This effect is constant over time, such that [x] = β1−42. Consequently, ferritin causes a shift to the left in age of conversion to AD by [6] months, where [6] = ferritin (ng ml−1)*3 (refer to Fig. 3b). Levels of tau/Aβ1–42 or APOE are associated with both baseline cognitive status [x] and the rate of cognitive deterioration, such that [x] < [y]. The effect causes a shift in age of diagnosis by [n] months where [n] = APOE (μg ml−1)*8 or tau/Aβ1–42 (units)*17 (refer to Fig. 3b).

An unresolved question arising from this study is why are CSF ferritin levels not elevated in AD, where brain iron levels are reported as elevated? We hypothesize that ferritin levels in the CSF reflect global brain iron burden, whereas iron elevation in AD has only been reported in affected regions (for example, frontal cortical tissue). Possibly, iron elevation in brain regions affected by AD is too confined regionally to be reflected in CSF. An altered relationship between tissue and CSF ferritin in AD, however, cannot yet be excluded.
Our data also provide exploratory insights into iron in AD aetiopathogenesis, identifying an unexpected interaction of ApoE with ferritin. That ferritin levels are increased by the APOE-e4 allele argues that ApoE influences ferritin levels, rather than the reverse. Our current findings indicate that APOE genotype should influence constitutive brain iron burden. However, to our knowledge, a post mortem study of iron or ferritin in brain tissue, stratified according to APOE genotype, has not been reported. Focal changes to iron and ferritin have been observed in AD brains post mortem1–4,5, and on the basis of our findings we propose that the e4 genotype raises the baseline iron load of the brain, thus lowering the threshold for iron-mediated neuronal loss in disease. This proposal awaits experimental confirmation, but it is possible that increased plaque pathology associated with the APOE e4 isoform24,25 might be a consequence of interactions between ApoE and iron26,27, leading to oxidative stress and ApE aggregation28,29. Elevated iron could likewise contribute to tau pathology by causing its aggregation40, indeed iron is co-localized in neurofibrillary tangles in AD, and such co-localization is also observed in a primary disease of brain iron overload, neurodegeneration with iron accumulation2,31. Superficial CNS siderosis is also characterized by brain iron deposition, and tau is elevated in the CSF in this condition32. A relationship between brain iron and tau is supported by the results in our study, where CSF ferritin levels correlated with tau levels (Model 1 of Supplementary Table 1: when ApoE is excluded from the analysis).

How, then, could ApoE impact on brain iron homeostasis? To our knowledge, no previous study has directly explored this, but synaptic zinc was reported to be lowered in APOE KO mice,33 and in a closed head injury model, iron accumulation was shown to be exaggerated in APOE-KO mice34. The mechanism for ApoE in iron regulation could involve the trafficking of lipoproteins by ApoE. Treatment of macrophages with high-density lipoprotein (HDL; the lipoprotein of CSF) has been shown to lower intracellular iron levels and to promote ferritin release35. APOE e4 carriers have less CSF ApoE, and the e4 isoform has comparatively lower affinity for HDL36, so reduced delivery of HDL in APOE e4 carriers could result in iron retention in the brain. Notably, the iron accumulation mutation of HFE (associated with hemochromatosis) has an epistatic interaction with APOE e4 to increase AD risk and accelerates disease onset by 5.5 years (reviewed in ref. 37). We therefore introduce the concept that APOE e4 status confers susceptibility to AD by increasing ferritin levels. The association between ApoE and brain iron status will warrant further investigation.

Methods
ADNI description. Data used in the preparation of this article were downloaded on 15 July 2014 from the Alzheimer’s Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). The ADNI study has been previously described in detail38. The ADNI was launched in 2003 by the National Institute on Aging (NIA), the National Institute of Biomedical Imaging and Bioengineering (NIBIB), the Food and Drug Administration (FDA), private pharmaceutical companies and non-profit organizations, as a $60 million, five-year public-private partnership. The primary goal of ADNI has been to test whether serial magnetic resonance imaging (MRI), positron emission tomography (PET), other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of mild cognitive impairment (MCI) and early Alzheimer’s disease (AD). Determination of sensitive and specific markers of very early AD progression is intended to aid researchers and clinicians to develop new treatments and monitor their effectiveness, as well as lessen the time and cost of clinical trials.

The principal investigator of this initiative is Michael W. Weiner, MD, VA Medical Center and University of California, San Francisco. ADNI is the result of efforts of many co-investigators from a broad range of academic institutions and private corporations, and subjects have been recruited from over 50 sites across the United States and Canada. The initial goal of ADNI was to recruit 800 subjects but ADNI has been followed by ADNI-GO and ADNI-2. To date these three protocols have recruited over 1,500 adults, ages 55 to 90, to participate in the research, consisting of cognitively normal older individuals, people with early or late MCI, and people with early AD. The follow-up duration of each group is specified in the protocols for ADNI-1, ADNI-2 and ADNI-GO. Subjects originally recruited for ADNI-1 and ADNI-GO had the option to be followed in ADNI-2. For up-to-date information, see www.adni-info.org.

Recruitment inclusion and exclusion criteria for ADNI 1. Inclusion criteria were as follows: (1) Hachinski ischaemic Score ≤4; (2) permitted medications stable for 4 weeks before screening; (3) Geriatric Depression Scale score <6; (4) visual and auditory acuity adequate for neuropsychological testing; good general health with no diseases precluding enrolment; (5) six grades of education or work history and language proficiency; (6) ability to speak English or Spanish; and (7) a study partner with 10 h per week of contact either in person or on the telephone who could accompany the participant to the clinic visits.

Criteria for the different diagnostic groups are summarized in Supplementary Table 1. Groups were age-matched. Cognitively normal (CN) subjects must have had no history of cognitive impairment or impaired activities of daily living. Clinically diagnosed AD patients must have had mild AD and had to meet the National Institute of Neurological and Communicative Disorders and Stroke–Alzheimer’s Disease and Related Disorders Association criteria for probable AD39, whereas mild cognitive impairment subjects (MCI) could not meet these criteria, have an intact general cognition as well as functional performance, but meet-deficit criteria for MCI.

CSF biomarker collection and analysis. CSF was collected once in a subset of ADNI participants at baseline. ApoE and ferritin protein levels were determined using a Myriad Rules Based Medicine platform (Human Discovery MAP, v1.0; see ADNI materials and methods). CSF Factor H (FH) levels were measured using a multiplex human neurodegenerative kit (HMDG1-36k; Millipore, Billerica, MA) according to the manufacturer’s overnight protocol with minor modifications. CSF was collected into polypropylene tubes or syringes provided to each site, and then be transferred into polypropylene transfer tubes without any centrifugation step followed by freezing on dry ice within 1 h after collection for subsequent shipment overnight to the ADNI Biomarker Core laboratory at the University of Pennsylvania Medical Center on dry ice. Aliquots (0.5 ml) were prepared from these samples after thawing (1 h) at room temperature and gentle mixing. The aliquots were stored in bar code-labelled polypropylene vials at −80 °C. Fresh, never before thawed, 0.5 ml aliquots for each subject’s set of longitudinal time points were analysed on the same 96-well plate in the same analytical run for this study to minimize run to run and reagent kit source of variation. Within run coefficient of variation (%CV) for duplicate samples ranged from 2.5 to 5.9% for ApoE, 2.2–6.3% for tau and the inter-run %CV for CSF pool samples ranged from 3.1 to 14% for ApoE, 2.7–11.2% for tau. Apolipoprotein E (ApoE) and ferritin protein levels were determined using Rules Based Medicine (Human Discovery MAP, v1.0).

Further information on the procedures and standard operating procedures can be found in previous publications40,41 and online (http://www.adni-info.org/).

Structural MRI acquisition and processing. Subjects with a 1.5-T MRI and a sagittal volumetric 3D MPRA GE with variable resolution around the target of 1.2 mm isotropically were included in the analysis. See (www.loni.ucla.edu/ADNI) and for detail42. The hippocampal and ventral volumes utilized were those in the ADNI MERGE primary table allowing the ADNI MERGE participants to be downloaded on the 15 July 2014. Only CN and MCI subjects were included in the MRI analysis. MRI scans were performed at baseline, 6 months, 1 year and then yearly for six years.

Statistical analysis. All statistical work was conducted with R (version 3.1.0)43, using packages ggplot2 (ref. 44), nlme45, car46 and Deducer47. We tested the conditions necessary to apply the regression models, normal distribution of the residuals and the absence of multicollinearity. All models satisfied these conditions. Regression models were not stratified with regard to the entry of control for the possibility of a traumatic tap introducing plasma ferritin into the CSF samples. FH was used to control for inflammation, since ferritin levels are known to be elevated in certain inflammatory conditions (for example, bacterial meningitis48).

Multiple regression models of CSF ferritin and ApoE initially contained age, gender, BMI, ApoE genotype, and levels of CSF hemoglobin (Hb) and Factor H (FH). CSF Hb was included as a proxy for blood contamination, to control for the possibility of a traumatic tap introducing plasma ferritin into the CSF samples. FH was used to control for inflammation, since ferritin levels are known to be elevated in certain inflammatory conditions (for example, bacterial meningitis48).

Multiple regression models of CSF ferritin and ApoE initially contained age, gender, BMI, ApoE genotype, and levels of CSF Hb and FH, plus various inclusions of CSF tau, ApoE and either ferritin or ApoE. The minimal models are described in the table legend.
Associations between the baseline Alzheimer’s Disease Assessment Scale Cognition (ADAS-cog13) and Rey Auditory Verbal Learning Test (RAVLT) scores with CSF ferritin, the CSF tau/\(A\beta_42\) ratio and CSF ApoE were tested with a covariate-adjusted multiple regression for each cognitive scale. For these analyses, age, gender, BMI, years of education, \(A\beta_42\) and ferritin are baseline diagnosis were initially included as covariates. To assess the association of baseline CSF ferritin values with the longitudinal clinical outcomes (ADAS-cog13 and RAVLT scores over 7 years), linear mixed effects models were used. These models were adjusted for the same variables as the baseline models of cognition, and additionally included time as interacting variable with each of the CSF biomarkers. AD subjects were excluded from the longitudinal analysis because of low rate of follow up (Supplementary Table 2). A significant value for any of these interaction terms would indicate that the variable affected the rate of cognitive change. For the ADAS-cog13, longitudinal analysis, the minimal model included years of education, gender and \(A\beta_42\)-ferritin. For the longitudinal analysis with RAVLT, the minimal model included years of education and gender. Cov proportional hazards model was used to assess the impact of CSF analytes on the time to AD conversion. The initial model contained age at baseline, gender, years of education and \(A\beta_42\)-ferritin as confounding variables together with CSF ApoE, tau/\(A\beta_42\) and ferritin. A minimal model containing only the CSF biomarkers was identified via BIC step down procedure and log likelihood test. Logistic regression analysis was used to assess the impact of CSF analytes on risk of conversion to AD. Combinations of CSF ferritin, ApoE and tau/\(A\beta_42\) analytes were included in logistic regression models of MCI conversion to AD that were adjusted for age at baseline, gender, years of education, ApoE genotype and BMI. These models determined the predictive performance of these analytes in identifying stable MCI participants from MCI participants who, up to 102 months, had a diagnosis change to AD. The receiver-operator curves and the area under the curve were derived from the predictive probabilities of the logistic regression models. The relationships between CSF ferritin, ApoE, tau/\(A\beta_42\) with longitudinal structural (MRI) changes to hippocampus and cerebral ventricles were assessed using linear mixed effects models for age, years of education, BMI, gender and ApoE genotype and intracranial volume. For all models, CSF ferritin, ApoE, tau/\(A\beta_42\) and baseline diagnosis were included as fixed effects and were not removed from a minimal model. Two random effects were also included, intercepts and slope (time). An interaction between time and diagnosis, time and CSF ferritin, time and CSF ApoE well as time and tau/\(A\beta_42\) were also included for all models. All the AD subjects were excluded from MRA analyses due to low number and short follow-up. PET imaging data from ADNI were not included in the analysis because there were too few patients who had CSF ferritin measured and who also underwent PET imaging at baseline.

References


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Author contributions

S.A. provided-scientific concept, modelling of data and wrote manuscript. N.G.F. helped in modelling of data and wrote manuscript. A.I.B. supervised modelling, wrote manuscript and funded analysis project.

Additional information

Supplementary Information accompanies this paper at http://www.nature.com/nc

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