Dephosphorylation of DBC1 by Protein Phosphatase 4 Is Important for p53-Mediated Cellular Functions

Citation

Published Version
doi:10.14348/molcells.2015.0066

Permanent link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:22857022

Terms of Use
This article was downloaded from Harvard University’s DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

Share Your Story
The Harvard community has made this article openly available. Please share how this access benefits you. Submit a story.

Accessibility
Dephosphorylation of DBC1 by Protein Phosphatase 4 Is Important for p53-Mediated Cellular Functions

Jihye Lee, Guillaume Adelmant1, Jarrod A. Marto1, and Dong-Hyun Lee*

Deleted in breast cancer-1 (DBC1) contributes to the regulation of cell survival and apoptosis. Recent studies demonstrated that DBC is phosphorylated at Thr454 by ATM/ATR kinases in response to DNA damage, which is a critical event for p53 activation and apoptosis. However, how DBC1 phosphorylation is regulated has not been studied. Here we show that protein phosphatase 4 (PP4) dephosphorylates DBC1, regulating its role in DNA damage response. PP4R2, a regulatory subunit of PP4, mediates the interaction between DBC1 and PP4C, a catalytic subunit. PP4C efficiently dephosphorylates pThr454 on DBC1 in vitro, and the depletion of PP4C/PP4R2 in cells alters the kinetics of DBC1 phosphorylation and p53 activation, and increases apoptosis in response to DNA damage, which are compatible with the expression of the phosphomimetic DBC1 mutant (T454E). These suggest that the PP4-mediated dephosphorylation of DBC1 is necessary for efficient damage responses in cells.

INTRODUCTION

Genotoxic stress inducing DNA breaks and replication stress stimulates genomic instability and cellular transformation. To prevent these detrimental consequences, eukaryotic cells have evolved an elaborate and complex response system called DNA damage response (DDR), which is mostly initiated by the phosphatidylinositol 3-kinase (PI3)-like family of kinases, including replication protein A 2 (RPA2), KAP-1, and 53BP1 after DNA damage and these dephosphorylation events were critical for the efficient repair of DNA double-strand breaks (DSBs) (Lee and Lee, 2014; Lee et al., 2010; 2012; Nakada et al., 2008; Wang et al., 2008). PP4 dephosphorylated the essential proteins, including replication protein A 2 (RPA2), KAP-1, and 53BP1 after DNA damage and these dephosphorylation events were critical for the efficient repair of DNA double-strand breaks (DSBs) (Lee and Lee, 2014; Lee et al., 2010; 2012; 2014). To study the functions of PP4 in depth, we performed tandem affinity purification followed by mass spectrometry. We identified DBC1, which was hyperphosphorylated in the absence of PP4C following DNA damage. Here we elucidated the importance of PP4C-mediated dephosphorylation of DBC1 and focused on the functional impact of DBC1 dephosphorylation on human cells.

MATERIALS AND METHODS

Cell culture, antibodies, and reagents
HeLa S3, U2OS, and RPE1 cells were grown in DMEM supplemented with 10% (v/v) FBS. In addition to U2OS, RPE1 cells contain an intact p53 checkpoint and were therefore used for study on p53. Antibodies used were against PP4R1 (Bethyl), PP4R2 (Bethyl), PP4R3a (Bethyl), PP4R3j1 (Bethyl), PP4C (Bethyl), DBC1 (Bethyl), pT454-DBC1 (Cell Signaling), p53 (Santa Cruz), p53-Ac-K382 (Cell Signaling), SIRT1 (Sigma), Flag-tag (Sigma) and Actin (Millipore). Etoposide was obtained from Sigma-Aldrich. ATM inhibitor (KU55094) was obtained from Selleckchem. The list of cell lines and reagents used in this study is provided in (Lee and Lee, 2014; Lee et al., 2010; 2012; 2014; Nakada et al., 2008; Wang et al., 2008)
from KuDOS Pharmaceuticals.

**siRNAs and plasmids**

Cells were transfected with siRNA duplexes (Invitrogen) using RNAiMAX (Invitrogen). The PP4C siRNAs were as follows: siRNA #1, sense: 5′-GCUCUAGAGGAAGUGUAUAGC-3′; antisense: 5′-UAAGUACGUGACUGGCUCUAUA-3′; siRNA #2, sense: 5′-GGACAGGUACGGGAGAGGU-3′; antisense: 5′-ACUCUCUGUGCUUCGGAGUUGCC-3′. The PP4R2 siRNAs were as follows: siRNA #1, sense: 5′-CAGAGAGGCACUGUGGUGUGCAUAU-3′; antisense: 5′-CCAGGGCCACUUAUGACGAAAAAGGGU-3′. DBC1 phosphomutants were constructed by QuikChange II XL site-directed mutagenesis kit (Stratagene) according to the manufacturer’s instructions. Primers used were the following: T454E-F: 5′-GAAGGACGCTCCACAACACAGAAGG-3′; T454E-R: 5′-CCTGCTGTCGCAAGAGG-3′; T454A-F: 5′-T454A-C-AGAGGACGCTCCACAACACAGAAGG-3′; T454A-R: 5′-CCTGCTGTCGCAAGAGG-3′.

**Co-immunoprecipitation**

HeLa S3 or U2OS cells expressing FH-DBC1 WT or phosphomutants, were lysed in buffer containing 50 mM Tris-HCl (pH 7.5), 250 mM NaCl, 5 mM EDTA, 0.5% (v/v) NP-40 and protease inhibitor cocktail (Roche). Anti-Flag-agarose (Sigma) was incubated with lysate at 4 °C for 16 h. Immunocomplexes were washed three times with buffer containing 50 mM Tris-HCl (pH 7.5), 250 mM NaCl, 5 mM EDTA and 0.5% (v/v) NP-40. The immunoprecipitated proteins were resolved by SDS-PAGE and analyzed by immunoblot.

**Immunofluorescence**

Cells plated on glass slides were fixed for 10 min with fixative (3%(w/v) PFA, 2%(w/v) sucrose and 1 X PBS) and permeabilized for 1 min with 0.2%(v/v) Triton X-100 in PBS. Cells were rinsed with PBS and incubated with primary antibody diluted in PBS with 2%(w/v) FBS for 1 h at room temperature (RT). Cells were washed three times, incubated with secondary antibody (diluted in PBS with 2%(w/v) BSA) for 30 min at RT in the dark, incubated with 4′-6-diamidino-2-phenylindole (DAPI) for 10 min and washed three times with PBS. Slides were mounted using DapiFuoromount-G (Southern Biotech) and visualized using a Zeiss Axioplan microscope. Secondary Alexa Fluor IgG antibodies used were as follows: 488 goat anti-rabbit, 594 goat anti-mouse (Invitrogen).

**In vitro dephosphorylation assay**

The in vitro dephosphorylation assay was performed as described (Lee and Lee, 2014; Lee et al., 2012). PP4C WT and DB2A mutant proteins were purified using the Bac-to-Bac Baculovirus Expression System (Invitrogen) according to the manufacturer’s manual. For the dephosphorylation assay, phosphorylated FLAG-DBC1 was prepared by immunoprecipitation of DBC1 from irradiated cells with anti-FLAG antibody-conjugated agarose. Phosphatase reactions with phosphorylated Flag-DBC1 were performed in 20 mM Tris-HCl (pH 7.4), 50 mM NaCl, 0.2 mM EDTA, 0.2% β-mercaptoethanol for 30 min at 30°C. Reactions were resolved on 4-12%(v/v) SDS-PAGE, and relative phosphatase activity was determined by loss of phospho-FLAG-DBC1 immunoreactivity, as determined by p454 antibody staining.

**Apoptosis assay**

Apoptosis was detected by using in situ cell death detection kit (Roche Applied Science, Germany), according to the manufacturer’s instructions. Briefly, U2OS cells were fixed for 10 min with fixative (4% formaldehyde) and permeabilized for 5 min with 0.2% Triton X-100. Cells were rinsed with PBS and incubated with TUNEL reaction mixture at 37°C for 1 h. Finally, cells were analyzed under Zeiss Axioplan microscope at 450-500 nm. Apoptotic index (AI) was determined as the percentage of TUNEL positive cells in 10 randomly selected fields (~200 cell counts).

**Clonogenic assay**

U2OS (0.3 × 10⁶ cells/well) were transfected with siRNAs against PP4C or PP4R2, or FH-DBC1 WT or phosphomutants. After 2 days, 1000 cells were seeded on 6-well plates in quadruplicate and incubated overnight. Cells were irradiated at indicated doses on the following day and allowed to form colonies for 2 weeks before being stained by 0.1% crystal violet solution for evaluation. Surviving colonies of < 1 mm diameter were counted.

**Mass spectrometry analysis of affinity-purified PP4 complexes**

Sample preparation, liquid chromatography/tandem mass spectrometry analysis, database searches and identification of proteins associated with PP4C were performed with slight modifications according to a recently described method (Wang et al., 2013). Briefly, purified protein complexes were denatured and reduced by incubation at 56°C for 30 min in 10 mM DTT and 0.1% RapiGest (Waters). Protein digestion was carried out overnight at 37°C after adding 500 ng of trypsin and adjusting the pH to 8.0. RapiGest was removed from solution following the manufacturer’s protocol and tryptic peptides were purified by batch-mode reverse-phase C18 chromatography (Porex 10R2, Applied Biosystems) using 40 μl of a 50% bead slurry in RP buffer A (0.1% trifluoroacetic acid), washed with 100 μl of the same buffer and eluted with 50 μl of RP buffer B (40% acetonitrile in 0.1% trifluoroacetic acid). After vacuum concentration, peptides were solubilized in 20 μl of 20 mM sodium phosphate (pH 7.4) and incubated with 20 μl of thiol-activated sepharose 4B (GE-healthcare) to remove excess HA peptide. Peptides were further purified by strong cation exchange SCX chromatography (Poros 20HS, Applied Biosystems) using 20 μl of a 50% bead slurry in SCX buffer A (25% ACN in 0.1% formic acid), washed with 20 μl of the same buffer and sequentially eluted with 20 μl of SCX buffer B1 (25% ACN, 30 mM KCl in 0.1% formic acid) and SCX buffer B2 (25% ACN, 300 mM KCl in 0.1% formic acid). The eluates and the SCX flow-through were concentrated in a vacuum concentrator and reconstituted with 20 μl of 0.1% TFA. Purified peptides were analyzed by LC-MS/MS (Ficarro et al., 2009) on an LTQ-Orbitrap-XL mass spectrometer (Thermo, USA) equipped with a Digital PicoView electrospray source platform (NewObjective, USA). The spectrometer was operated in data dependent mode where the 10 most abundant ions in each MS scan were subjected to CAD (35% normalized collision energy, isolation width = 2.8 Da, threshold = 20,000). Dynamic exclusion was enabled with a repeat count of 1 and exclusion duration of 40 s. ESI voltage was set to 2.2 kV. MS spectra were converted into a Mascot generic file format (.mgf) using multiplexier scripts (Parikh et al., 2009) and searched using Mascot (version 2.4) against three appended databases consisting of: i) human protein sequences (downloaded from RefSeq on 07/11/2011); ii) common lab contaminants and iii) a decoy database generated by reversing the sequences from the two databases. Precursor tolerance was set to 20 ppm and product ion tolerance to 0.6 Da. Search
**RESULTS**

**DBC1 is a bona-fide substrate of PP4**

To obtain a comprehensive understanding of PP4 functions in cells, we performed tandem affinity purification (TAP) mass spectrometry (Ikura et al., 2000; Nakatani and Ogryzko, 2003) of PP4C from HeLa S3 cells expressing epitope (FLAG/HA)-tagged protein (Fig. 1A). From the mass spectrometric data, proteins detected in a negative control were subtracted. We further analyzed data to determine a subset of PP4C-interacting proteins associated with the regulation of the response to DNA damage stimulus (GO:2001020) (Supplementary Table S1).

DBC1, a nuclear protein, is phosphorylated upon DNA damage at Thr454 by ATM/ATR and Chk2, inhibits NAD-dependent deacetylase sirtuin-1 (SIRT1), and promotes p53 activation (Magni et al., 2014; Zannini et al., 2012). To validate the interaction between PP4 and DBC1, we analyzed the association of these proteins by endogenous and reciprocal immunoprecipitation/immunoblot assays using lysates from HeLa cells and observed that DBC1 interacts with PP4C and PP4R2, but not with other subunits, including PP4R1, PP4R3α, or PP4R3β (Fig. 1B). To examine whether PP4C and PP4R2 are independently associated with DBC1, we silenced PP4R2 and observed that the interaction of PP4C with DBC1 is dramatically reduced in the
Fig. 2. DBC1 phosphorylation is regulated by PP4C/PP4R2 complex. (A) Depletion of PP4C or PP4R2 induces hyperphosphorylation of DBC1 on T454. U2OS cells transfected with siRNAs against PP4C or PP4R2 were irradiated and harvested at the indicated times and phospho-DBC1 was assessed by immunoblot using phospho-DBC1 antibody (p-T454). The kinetics of pT454-DBC1 formation was monitored after irradiation. (B) PP4C or PP4R2 depletion attenuates pT454-DBC1 turnover after IR. U2OS cells were transfected with control, PP4R2, or PP4C siRNAs. After 72 h, cells were irradiated, fixed at the indicated times, and immunostained for pT454-DBC1 (green), γH2AX (red) and DAPI (blue). γH2AX served as control showing the DNA damage (DSBs) in cells. Cells expressing PP4C D82A mutant or without transfection were used as control. (C) The average pT454-DBC1 signal intensity per nucleus was quantified using ImageJ software. Data are expressed as mean ± SD; n = 3. *P = 0.0015 (PP4C siRNA), **P = 0.0034 (PP4R2 siRNA), ***P = 0.0067 (PP4C D82A).
PP4 has an impact on p53 activity through the dephosphorylation of DBC1. (A) Depletion of PP4C or PP4R2 enhances p53 activity, compatible with DBC1 T454E expression. RPE1 cells transfected with siRNAs against PP4C or PP4R2 were irradiated and harvested at the indicated times (Left panel) and U2OS cells expressing Flag-DBC1 WT, T454E, or T454A were irradiated and harvested at the indicated times (Right panel). Ac-p53 was assessed by immunoblot using acetylated p53 antibody (Ac-K382). (B) Depletion of PP4R2 enhances the interaction between DBC1 and SIRT1. RPE1 cells expressing empty vector (FH-Empty), FH-tagged DBC1 were subjected to immunoprecipitation using anti-FLAG beads after IR. PP4R2 siRNAs or control siRNAs were transfected in cells 72 h prior to immunoprecipitation. The immunoprecipitate was probed with antibodies against SIRT1 and PP4R2 as indicated. (C) In vitro dephosphorylation assay. PP4C WT, PP4C D82A, and PP4R2 were purified using the baculoviral system and were serially diluted in the phosphatase reaction. λ phosphatase served as positive control for the reaction. PP4C dephosphorylates phospho-DBC1 on T454 in a dose-dependent manner. Phosphatase reactions were probed with indicated antibodies.

PP4 has an impact on p53 activity through the dephosphorylation of DBC1

Next, we examined the impact of PP4 on p53 activation mediated by DBC1. Previous studies showed that the phosphorylation of DBC1 is crucial for acetylation-mediated p53 activation through its interaction with SIRT1 (Park et al., 2014; Zannini et al., 2012). Therefore, we reasoned that PP4-mediated dephosphorylation of DBC1 acts on p53 activation. We assessed acetylated p53 on K382 (Ac-p53) as a maker of p53 activation. RPE1 cells depleted of either PP4C or PP4R2 were irradiated with 10 Gy IR and harvested at various time points. In response to IR, there is a great induction of Ac-p53 at 2 h after IR, which significantly drops by 4 h. However, in the absence of either PP4C or PP4R2, there is a significantly higher amount of Ac-p53 by 4 h (Fig. 3A, Left panel). A compatible result was obtained from cells expressing DBC1 T454E, whereas the expression of DBC1 T454A had no significant impact on p53 acetylation (Fig. 3A, Right panel). Interestingly, the depletion of either PP4R2 or PP4C increases the p53 expression even in unperurbed cells, which is not the case in DBC1 T454E expression. This suggests that in addition to the regulation of DBC1 phosphorylation, PP4 contributes to the regulation of p53 activity by an unknown mechanism. Previous reports described that the association of DBC1 with SIRT1 was increased upon DNA damage, whereas the expression of DBC1 T454A significantly decreased the interaction with SIRT1 (Kim et al., 2009b; Zannini et al., 2012). Moreover, both DBC1 phosphorylation

ished rapidly after ATMi treatment (Supplementary Fig. 1A). Silencing PP4C or PP4R2 arouses the persistence of pT454-DBC1 several hours after IR even in the presence of ATM (Supplementary Fig. 1B). A PP4 complex including PP4C and PP4R2 dephosphorylates γH2AX (Chowdhury et al., 2008; Nakada et al., 2008). Therefore, it is feasible that silencing PP4C and PP4R2 affects DBC1 phosphorylation via H2AX. To evaluate this possibility, we silenced PP4R2 or PP4C in h2ax-deleted cells and assessed the phosphorylation status of DBC1. The absence of H2AX did not alter the impact of PP4 on DBC1 phosphorylation (Supplementary Fig. 2). Together, these results suggest that PP4C/PP4R2 dephosphorylates DBC1.
PP4-Mediated Dephosphorylation of DBC1
Jihye Lee et al.

Fig. 4. Functional impact of PP4-mediated dephosphorylation of pT454 DBC1 (A) Hyperphosphorylation of DBC1 induced by PP4C/PP4R2 depletion increases cellular apoptosis. U2OS cells were transfected with siRNAs against PP4C or PP4R2 (Left panel), or expressed with DBC1 WT, T454A, or T454E (Right panel). The 48 h after etoposide treatment, apoptotic cells were analyzed under fluorescence microscope by counting > 200 cells. Data represent average and s. d. of three independent experiments. Immunoblots performed to confirm siRNA efficiency and expression of FH-DBC1 constructs are shown. Data are expressed as mean \( \pm \) SD; n = 3. *P = 0.0021, **P = 0.0061, ***P = 0.0083, ****P = 0.018, *****P = 0.028, ******P = 0.017. (B) Radiosensitivity of DBC1 phosphomutants. U2OS cells were transfected with siRNAs against PP4C or PP4R2 (Left panel), or expressed with DBC1 WT, T454A, or T454E (Right panel). At 72 h after transfection, cells were irradiated at the indicated doses, and viability was evaluated by clonogenic survival. Immunoblots performed to confirm siRNA efficiency and expression of FH-DBC1 constructs are shown. Data are expressed as mean \( \pm \) SD; n = 3.*P = 0.014 (3 Gy), *P = 0.0027 (6 Gy), **P = 0.0057 (3 Gy), **P = 0.0035 (6 Gy), *P = 0.019 (3 Gy), *P = 0.0082 (6 Gy), ***P = 0.039 (3 Gy), **P = 0.029 (6 Gy).

and the expression of DBC1 T454E promoted its sumoylation, which is also important for p53-mediated apoptosis (Park et al., 2014). Consistent with previous reports (Kim et al., 2009b; Zannini et al., 2012), we observed that the expression of DBC1 T454E revealed a greater induction of the interaction with SIRT1 than in DBC1 WT-expressing cells, which is compatible with that in PP4R2-depleted cells (Fig. 3B). Together, the results we obtained suggest that PP4-mediated dephosphorylation of DBC1 has a significant impact on p53 activity.

Direct in vitro dephosphorylation of DBC1 at T454 by PP4C
To determine whether PP4C can dephosphorylate pT454-DBC1 directly, we immunopurified phospho-DBC1 from IR-treated cells and performed dephosphorylation assays as described earlier (Lee et al., 2014; Smith et al., 2010). PP4C dephosphorylates pT454-DBC1 in a dose-dependent manner (Fig. 3C, Left panel). However, the addition of PP4R2 protein to the reaction has no impact on the efficiency of dephosphorylation, suggesting PP4R2, a regulatory subunit, is only required for PP4C-mediated dephosphorylation in vivo (Fig. 3C, Right panel). Catalytically inactive PP4C mutant (PP4C D82A) and λ phosphatase served as controls. Together, these results strongly suggest that PP4C directly dephosphorylates DBC1.

Functional impact of PP4-mediated dephosphorylation of T454-DBC1
As previously reported (Zannini et al., 2012), there was a greater reduction of apoptosis among cells expressing DBC1 T454A than in DBC1 WT-expressing cells and a survival assay revealed that clonogenic survival was significantly lower in cells expressing DBC1 WT than in cells expressing DBC1 T454A. Based on our results described above, we hypothesized that the depletion of either PP4C or PP4R2 is functionally equivalent to the expression of DBC1 constitutively phosphorylated at T454 (T454E). To test this, we performed the apoptosis assay (Fig. 4A, Left panel). U2OS cells, depleted of PP4C or PP4R2 by siRNA transfection, were treated with etoposide to induce apoptosis for 48 h. Etoposide treatment increased the percentage of apoptotic cells to 18.5%. In the absence of either PP4C or PP4R2, 37.2% or 41.1% of cells were detected as apoptotic.
cells. These differences are statistically significant (Fig. 4A, Middle panel). Cells expressing DBC1 T454E show a greater induction of apoptosis than in DBC1 WT-expressing cells. Even though this difference is not as great as that observed in between PP4C- or PP4R2-depleted cells and control cells, it is also statistically significant (Fig. 4A, Right panel). Consistent with a previous report (Zannini et al., 2012), we also observed that the expression of DBC1 T454A exhibits significantly decreased apoptosis, compared to that in cells expressing DBC1 WT (Fig. 4A, Right panel). The effect on apoptosis is expected to be biologically relevant, and indeed PP4C- or PP4R2-deficient cells and cells expressing DBC1 T454E have lower viability than control cells at all tested doses of IR (Fig. 4B). Depletion of PP4C/PP4R2 has an impact on p53 activation that is compatible to the phenotype induced by the expression of the phosphomimetic (T454E) DBC1 mutant. However it is unclear whether the impact of PP4 on p53 is mediated directly by DBC1. Theoretically if indeed the function of PP4 on p53 was mediated by DBC1, then phenotype induced by PP4C/PP4R2 depletion would be rescued by expressing the DBC1 T454A. Consistent with this notion, there is no significant effect on p53 activation in cells expressing DBC1 T454A when PP4C/PP4R2 was depleted (Supplementary Fig. 3A). To rule out the possibility that the altered DNA damage response by depletion of PP4C/PP4R2 may not be mediated by altered p53 response, we depleted PP4C or PP4R2 in cells where p53 was silenced. And we observed that the survival rate induced by silencing PP4C or PP4R2 was rescued by the depletion of p53 (Supplementary Fig. 3B). Together, these results show that absence of a PP4C/PP4R2 complex leads to elevated levels of hyperphosphorylated DBC1, which impacts on apoptosis and sensitizes cells to DNA damage agents.

**DISCUSSION**

DBC1 was originally identified as a nuclear protein deficient in breast cancers (Hamaguchi et al., 2002; Nakatani and Oygrzyko, 2003). There are accumulating reports focused on primary roles of DBC1 in DDR. Recent studies revealed that DBC1 is phosphorylated on T454 in response to DNA damage, which is important for sumoylation on itself, p53 activation, and p53-mediated apoptosis, but the expression of DBC1 T454A represses exactly opposite results (Kim et al., 2008; 2009b; Magni et al., 2014; Park et al., 2014; Zannini et al., 2012; Zheng et al., 2013). Therefore, the phosphorylation of DBC1 has an important role and need to be tightly regulated. With mass spectrometric data, we identified DBC1 as a potential interacting protein of PP4C and confirmed the association between them. Therefore, we hypothesized that PP4 is required for the regulation of DBC1 phosphorylation. We show that PP4C or PP4R2, but not other PP4 subunits, interacts with DBC1, independent of DNA damage and the interaction of DBC1 with PP4 is PP4R2-mediated. PP4C dephosphorylates phospho-DBC1 in vitro in a dose-dependent manner. In the absence of either PP4C or PP4R2, the signal of pT454 DBC1 was significantly enhanced and extended. By expressing the DBC1 phosphomimetic mutant (T454E), we can recapitulate the effect of silencing PP4R2 on the DNA damage response to a considerable degree. Why is it important to dephosphorylate DBC1? Consistent with earlier reports (Park et al., 2014; Zannini et al., 2012), we found that the hyperphosphorylation of DBC1 sustained the p53 activation and enhanced apoptosis in PP4-depleted cells. Dephosphorylation of DBC1 is therefore necessary for turning off DDR, and this in turn allows the cell to resume cycling. In other words, the dephosphorylation of DBC1 to recover cells to unperturbed status is as important as phosphorylation of that in response to DNA damage to turn on p53 signaling. In addition, hyperphosphorylated DBC1 in PP4R2- or PP4C-depleted cells significantly enhances apoptosis and sensitivity to DNA damaging agents. Our results now show that the phospho-signaling network centering DBC1 is initiated by ATM/ATR at one end and balanced by a PP4C/PP4R2 complex at the other end. Although it is generally accepted that the phosphorylation of DBC1 is necessary for an efficient response to genotoxic stress, our data emphasize a point that the PP4-mediated dephosphorylation of DBC1 may play an equally important role in DDR.

In conclusion, our study, for the first time, demonstrated the molecular mechanism by which the phosphorylation of DBC1 in response to DNA damage is regulated by PP4, having a significant importance in maintaining cell physiology. Since PP2A-like phosphatases have overlapping roles in DDR, future studies will clarify the possibility that other phosphatases contribute to the dephosphorylation of DBC1.

**ACKNOWLEDGMENTS**

The Flag-DBC1 construct was a gift from J.E. Kim (Kyung Hee University). This study was supported by grants from Basic Science Research Program through the National Foundation of Korea (NRF) funded by the Ministry of Science, ICT and Future Planning (NRF-2013R1A1A1061207), and Chonnam National University, 2014.

**REFERENCES**


Lee, D.H., Pan, Y., Kanner, S., Sung, P., Borowlec, J.A., and...


