1,4-Disubstituted-[1,2,3]triazolyl-Containing Analogues of MT-II: Design, Synthesis, Conformational Analysis, and Biological Activity

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Supporting Information

ABSTRACT: Side chain-to-side chain cyclizations represent a strategy to select a family of bioactive conformations by reducing the entropy and enhancing the stabilization of functional ligand-induced receptor conformations. This structural manipulation contributes to increased target specificity, enhanced biological potency, improved pharmacokinetic properties, increased functional potency, and lowered metabolic susceptibility. The CuI-catalyzed azide−alkyne 1,3-dipolar Huisgen’s cycloaddition, the prototypic click reaction, presents a promising opportunity to develop a new paradigm for an orthogonal bioorganic and intramolecular side chain-to-side chain cyclization. In fact, the proteolytic stable 1,4- or 4,1-disubstituted [1,2,3]triazolyl moiety is isosteric with the peptide bond and can function as a surrogate of the classical side chain-to-side chain lactam forming bridge. Herein we report the design, synthesis, conformational analysis, and functional biological activity of a series of i-to-i+5 1,4- and 4,1-disubstituted [1,2,3]triazole-bridged cyclopeptides derived from MT-II, the homodetic Asp5 to Lys10 side chain-to-side chain bridged heptapeptide, an extensively studied agonist of melanocortin receptors.

INTRODUCTION

Melanocortins are a group of peptide hormones including adrenocorticotropic (ACTH) and α-, β-, and γ-melanocyte stimulating hormones (MSHs). To date, five human melanocortin receptors (MC1R-MC5R) have been characterized as belonging to group α of the rhodopsin family of G-protein coupled receptors (GPCR).1,2 The MCRs mediate a plethora of biological functions that include sexual function,3 feeding behavior,4,5 pain modulation,6 thermoregulation, energy homeostasis,7,8 cardiovascular effects, and skin pigmentation,9,10 making them potential drug targets for treating pain, food intake, and body weight as well as erectile dysfunction. Therefore, development of an expansive toolbox of structural modifications that can be used to fine-tune the predominant conformations to achieve modulation of specificity toward...
receptor subtypes, physicochemical and pharmacological properties continues to be of great interest in the development of peptide-based drugs in general and melanocortinergic drugs in particular. Recently, the growing interest in designing highly selective and potent superagonist of the human melanocortin receptors hMC3R and hMC4R has been driven by their prospects of becoming effective drugs for treating feeding disorders such as the respective obesity and energy homeostasis. Structure−activity relationship studies (SAR) of α-MSH,11 a post-translationally generated tridecapeptide fragment originating from proopiomelanocortin (POMC),12 led to the development of MT-II,

\[ \text{N}^\alpha\text{Ac[Nle}^4\text{,Yaa}^5\text{,D-Phe}^7\text{,Xaa}^{10}\text{]αMSH(4-10)NH}_2 \]

which stabilized the pharmacophore containing sequence His\(^\beta\)-Phe\(^\beta\)-Arg\(^\beta\)-Trp\(^\beta\) in a type-II β-turn, resulting in a potent long acting nonselective superagonist of MCRs.16 Evidently, the plethora of melanocortin-mediated activities, the five melanocortin receptor subtypes, and the high profile of potential therapeutic targets associated with the melanocortin system underscores the unmet need for highly selective, pharmacokinetically diverse, and bioavailable agonists and antagonists. Intramolecular side chain-to-side chain cyclization of linear peptides is an established approach to achieve stabilization of specific conformations and a recognized strategy to improve resistance toward proteolytic degradation, increasing the metabolic stability in vitro and in vivo.17,18 We have previously reported a novel intramolecular i-to-i+4 side chain-to-side chain cyclization of linear peptides is an established approach to achieve stabilization of specific conformations and a recognized strategy to improve resistance toward proteolytic degradation, increasing the metabolic stability in vitro and in vivo.17,18 We have previously reported a novel intramolecular i-to-i+4 side chain-to-side chain cyclization of linear peptides is an established approach to achieve stabilization of specific conformations and a recognized strategy to improve resistance toward proteolytic degradation, increasing the metabolic stability in vitro and in vivo.17,18
to the peptide backbone, and the predominant conformations displayed by these heterodetic cyclic peptides.

In the current study, we explored the potential of the i-to-i+5 side chain-to-side chain 1,4- and 4,1-disubstituted [1,2,3]-triazoly-containing bridges to stabilize β-turn conformations in the context of the MT-II scaffold and evaluate its potential as a modulator of receptor subtype selectivity. We synthesized a series of heterodetic MT-II related cyclo-heptapeptides that varied in the size of the disubstituted [1,2,3]triazoly-containing bridge connecting Cαs of residues 4 and 10 and in the location and orientation of the [1,2,3]triazol moiety within the bridge (I–X, Schemes 1 and 2). The [1,2,3]triazol moiety was flanked on each side by 1 to 4 methylenes totaling in 4 or 5 methylenes. The biological activity of all the 1,4- and 4,1-disubstituted-[1,2,3]triazoly-containing cyclopeptides (1–X, Table 3) is compared to the prototypic lactam-containing peptide MT-II, as well as to the linear control precursors (1–X', Tables 2 and 3).

**SYNTHETIC STRATEGY**

In the current study, we explored the potential of the i-to-i+5 side chain-to-side chain 1,4- and 4,1-disubstituted [1,2,3]-triazolyl-containing bridges to stabilize β-turn structure, which has been identified as the MT-II bioactive conformation, responsible for its protracted, nonselective, and superagonist activity. To this end, we designed a series of [1,2,3]triazoly-containing cyclo-heptapeptides, derived from the model lactam peptide, MT-II Nω−Fmoc[Nleω-c,Aspφ-D-Pheφ,Lysφ(ω)]αMSH(4–10)NH₂, that vary in the size of the bridge and the location and orientation of the [1,2,3]triazol moiety within this bridge (Schemes 1 and 2).

In this context, Nω-Fmoc-ω-azido-α-amino acids (1–4) and Nφ-Fmoc-ω-yonoic-α-amino acids (5–8) were synthesized by diazo-transfer reaction starting from the corresponding Nω-protected αω-diamino acids (see Supporting Information). Except for the commercially available Nω-Fmoc-S-Pra-OH (8), the Nω-Fmoc-ω-alkynyl-α-amino acids (5–7) were synthesized by alkynylation of a Ni(II) complex of the Schiff base formed between glycine and (S)2-(N-benzyloxy)aminobenzophenone, as a chiral inducer, with alk-ω-ynyl bromides (see Supporting Information). Solid-phase peptide synthesis (SPPS) generated a series of linear peptide precursors (1′–X′) in which Lysω and Aspφ were replaced with ω-azido- and ω-yl-α-amino acid residues with side chains containing 1–4 methylenes. The stepwise solid-phase assembly of the linear precursors 1′–X′ was performed following Fmoc/tBu strategy on Rink-amide type resin. The chemical inertness of the ω-azido and ω-ylalkynyl functions toward the entire range of coupling and deprotection reactions and the various nucleophiles and electrophiles present during the multistep synthesis saves the need for elaborate orthogonal protection schemes. The incorporation of the building blocks Nω-Fmoc-Xaa(ω-N₃)-OH and Nφ-Fmoc-Yaa-(ω-y)-OH during the SPPS was found to be markedly advantageous to the postpeptide assembly side chain modification that introduces the ω-azido and ω-yl functions needed for the Cu'-catalyzed intramolecular azide–alkyne cycloaddition.

Solution-phase intramolecular CuAAC converted the linear precursors 1′–X′ into the 1,4- and 4,1-disubstituted-[1,2,3]triazoly-containing cyclopeptides (1–X), presenting different permutation in terms of the size of the [1,2,3]triazoly-containing bridges and the orientation and position of the [1,2,3]triazol moiety within the bridge. Depending on the positions i or i+5 of the azido and alkynyl amino acids in the peptide sequences, we generated two classes of cyclopeptides (Schemes 1 and 2). The intramolecular CuAAC of the linear precursors (1′–X′) was carried out as outlined in Scheme 3.

The first class employed linear precursors that had ω-yl- and ω-azido-α-amino acid residues in the respective positions i and i+5 (1′–IV′ and IX′) and yielded upon CuAAC the cyclopeptides presenting the [1,2,3]triazol moiety in the 4,1-orientation (1–IV and IX). The second class employed linear precursors that had ω-azido- and ω-yl-α-amino acid residues in the respective positions i and i+5 (V′–VIII′ and X′) and yielded upon CuAAC the cyclopeptides presenting the [1,2,3]triazol moiety in the 1,4-orientation (V–VIII and X). The click reaction conditions were identical to those reported by us previously and used a 10-fold molar excess of CuSO₄·H₂O and ascorbic acid in tBuOH/H₂O (1:2, v/v). Under these conditions, there was no formation of oligomeric click products resulting from intermolecular click reactions, thus suggesting formation of a Cu'/acytelylide/azide complex that has high preference for intramolecular cyclizations. Moreover, solution-phase CuAAC avoids dimerizations and macrocyclizations, which are often observed in on-resin CuAAC, resulting in crude products that are easy to purify. All copper salts were eliminated by solid-phase extraction of the crude material with water and elution of the cyclopeptides with 10–30% of ACN in water. The clicked cyclopeptides were further purified by RP-HPLC. The linear analogue of MT-II, BE124 (Ac-Nle-Asp-His-
agonist,27 the reference peptides BE124 and BE123 (Table 1), and the precursors (I′−X′), as well as the [1,2,3]triazolyl-containing cyclopeptides (I−X) (Schemes 1 and 2), was pharmacologically evaluated for functional potency (EC50) and efficacy using a reporter gene based bioassay for cAMP in HEK-293 cells stably expressing the murine (mouse) melanocortin receptor subtypes mMC1R, mMC3R, mMC4R, mMC5R. Table 1 summarizes the functional selectivity pro-

**RESULTS AND DISCUSSION**

**Biological Activity.** The potency of MT-II and of the linear peptide NDP-MSH (Table 1), the superpotent and long acting mMC1R and mMC4R, respectively. Moreover, MT-II discrim-

<table>
<thead>
<tr>
<th>Table 1. Sequences of MT-II, the Classical NDP-MSH, and the MT-II Linear Analogs BE123 and BE124</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Reference Peptides</strong></td>
</tr>
<tr>
<td>MT-II: Ac-Nle-[Asn-His-d-Phe-Arg-Trp-Lys-NH2]</td>
</tr>
<tr>
<td>NDP-MSH: Ac-Ser-Tyr-Ser-Nle-Glu-His-o-Phe-Arg-Trp-Gly-Lys-Pro-Val-NH2</td>
</tr>
<tr>
<td>BE124: Ac-Nle-Asp-His-o-Phe-Arg-Trp-Lys-NH2</td>
</tr>
<tr>
<td>BE123: Ac-Nle-Asn-His-o-Phe-Arg-Trp-Lys[N′Ac]-NH2</td>
</tr>
</tbody>
</table>

Table 2 reports the EC50 values in the adenylyl cyclase based bioassays of the linear peptides: the linear peptide precursors I′−X′, the classical NDP-MSH, and the MT-II analogues, BE123 and BE124. Peptide BE123, in which the charges on Asp and Lys present in BE124 are eliminated, reproduces closely the effect of side chain-to-side chain cyclization present in MT-II. As such, short of the conformational rigidification, BE123 is a linear version of MT-II. Table 3 summarizes the functional agonist potencies of the 1,4- and 4,1-disubstituted-[1,2,3]-triazolyl-containing cyclopeptides I′−X′ compared to MT-II. In this study, we used HEK-293 cells stably expressing the murine melanocortin receptors subtypes mMC1R, mMC3R, mMC4R, and mMC5R to assess the agonist potency and subtype selectivity profiles of these MT-II mimetic peptides.28,29

Among the linear peptides, NDP-MSH is the most potent mMC1R agonist, showing an EC50 value that is 5- and 9-fold lower than the respective values for BE124 and BE123 (Table 2). All three linear peptides are good agonists toward the mMC4R, with very similar subnanomolar EC50 (0.20−0.31 nM), and are 4−9-fold lower than their EC50 towards mMC3R and mMC5R. Relative to the linear peptides, the cyclic MT-II is the most potent agonist toward all four receptor subtypes. It is about 2- and 4-fold more potent than NDP-MSH on the mMC1R and mMC4R, respectively. Moreover, MT-II discriminates nicely between mMC1R or mMC4R and mMC3R or mMC5R. Interestingly, both BE124 and its potential charge-bearing side chains blocked variant BE123 are almost equipotent in all four receptor subtypes. Apparently, the application of the conformational constraint in the form of i-to-i +5 side chain-to-side chain lactam bridge formation as in MT-II and not the neutralization of potential charge-bearing groups as in BE123 play a critical role in potentiation and enhancement of functional selectivity toward certain receptor subtypes.

The potencies of the linear peptide precursors I′−X′ (Table 2, Figure 1) span a range of 2 orders of magnitude between the most potent one X′ (EC50 ≈ 0.050 nM in mMC4R and mMC5R) and the least potent one II′ (EC50 = 5.33 nM in mMC5R). Analogue X′ is the most potent of all four receptor subtypes, with 5−8-fold higher potency on mMC4R and mMC5R than on mMC1R and mMC3R. The selectivity for receptor subtype of a single analogue is in the range of 5−10-fold. For example, IX′ is 10-fold more potent in mMC5R than in mMC1R and mMC3R. The detailed sequences are given in Table 1.4 The detailed sequences are given in Table 1.4 n.d. means not determined.

Table 2. Functional Potency (EC50) of Linear Peptide Precursors N′-Ac[Nle′5,Yaa5′,d-Phe′7,Xaa10′]×tMSH(4−10)NH2 (I′, II′, III′, IV′, and IX′) and N′-Ac[Nle′5′,Xaa5′,d-Phe′7,Yaa10′]×tMSH(4-10)NH2 (V′, VI′, VII′, VIII′, and X′)

| code | [Xaa′5',Yaa10'] and [Yaa′5',Xaa10'] cacy using a reporter gene based bioassay for cAMP in HEK-293 cells stably expressing the murine melanocortin receptors subtypes mMC1R, mMC3R, mMC4R, and mMC5R.28,29

The melanocortin receptor subtype related potencies of the two short linear MT-II-related peptides BE123 and BE124 is very similar to that of the linear precursors I′−X′ regardless of the nature of side-chain modifications in positions 5 and 10 (α-MSH-based numbering). Moreover, in general, most of the linear short α-MSH(4−10)-derived peptides are less potent than either MT-II or its cyclic heterodetic "clicked" MT-II-derived peptides I′−X′.

Neither MT-II nor the 1,4- and 4,1-disubstituted-[1,2,3]-triazolyl-containing cyclopeptides I−X show very high selectivity toward a specific receptor subtype, and there is no consistent trend for receptor subtype selectivity in the [1,2,3]triazolyl-containing analogue series (Table 3, Figure 2). For example, while MT-II is 3−6-fold more potent toward mMC1R and mMC4R (cf. EC50 = 0.03 and 0.06 nM on mMC3R and mMC5R, respectively).
mMC1R and mMC4R, respectively, with 0.18 and 0.19 nM on mMC3R and mMC5R, respectively), the maximal selectivity observed for the least potent analogues I and V toward the different receptor subtypes is 2−3-fold and 3−6-fold, respectively (EC50s 1.62, 3.14, and 5.18 nM of analogue I in mMC4R, mMC3R, and mMC1R, respectively, and 0.58, 1.76, and 3.70 nM of analogue V in mMC4R, mMC5R, and mMC3R, respectively). The most potent [1,2,3]triazolyl-containing analogues in this series are III, IV, VII, and X. They all have preference to the mMC4R and mMC5R (EC50s 0.051, 0.041, 0.048, and 0.067 nM, respectively, on the former receptor subtype, and 0.042, 0.035, 0.045, and 0.07 nM, respectively, on the latter one) and analogues IV and VII are active to a lesser extent on mMC1R (EC50s 0.087 and 0.074 nM, respectively).

[1,2,3]Triazolyl-containing analogues I−VIII share a common location of the bridgeheads (positions 5 and 10) and an identical size of the bridges (five methylenes in total and one amide bond). However, these heterodetic cyclopeptides differ in the location of the [1,2,3]triazolyl moiety within the bridge, so one or two methylenes are removed from the N-proximal bridgehead (analogues I and VIII and II and VII, respectively) or one or two methylenes from the C-proximal bridgehead (analogues IV and VII).

**Table 3. Agonist Potency (EC50) of the [1,2,3]Triazolyl-Containing Cyclopeptides**

<table>
<thead>
<tr>
<th>Code</th>
<th>mMC1R EC50 (nM) ±SEM</th>
<th>mMC3R EC50 (nM) ±SEM</th>
<th>mMC4R EC50 (nM) ±SEM</th>
<th>mMC5R EC50 (nM) ±SEM</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>5.18±0.54</td>
<td>1.54±0.25</td>
<td>1.62±0.57</td>
<td>1.93±0.07</td>
</tr>
<tr>
<td>V</td>
<td>0.85±0.07</td>
<td>3.70±0.36</td>
<td>0.58±0.14</td>
<td>1.76±0.037</td>
</tr>
<tr>
<td>II</td>
<td>0.11±0.03</td>
<td>0.29±0.1</td>
<td>0.22±0.06</td>
<td>0.34±0.09</td>
</tr>
<tr>
<td>VI</td>
<td>0.71±0.1</td>
<td>1.91±0.73</td>
<td>0.85±0.36</td>
<td>1.88±0.70</td>
</tr>
<tr>
<td>III</td>
<td>0.1±0.009</td>
<td>0.21±0.038</td>
<td>0.05±0.006</td>
<td>0.042±0.005</td>
</tr>
<tr>
<td>VII</td>
<td>0.074±0.02</td>
<td>0.20±0.006</td>
<td>0.048±0.007</td>
<td>0.045±0.004</td>
</tr>
<tr>
<td>IV</td>
<td>0.087±0.029</td>
<td>0.2±0.002</td>
<td>0.041±0.001</td>
<td>0.035±0.007</td>
</tr>
<tr>
<td>VIII</td>
<td>0.53±0.14</td>
<td>0.60±0.04</td>
<td>0.14±0.02</td>
<td>0.13±0.015</td>
</tr>
<tr>
<td>IX</td>
<td>0.32±0.097</td>
<td>0.63±0.13</td>
<td>0.14±0.029</td>
<td>0.15±0.02</td>
</tr>
<tr>
<td>X</td>
<td>0.20±0.023</td>
<td>0.29±0.07</td>
<td>0.067±0.004</td>
<td>0.07±0.004</td>
</tr>
<tr>
<td>MT-II</td>
<td>0.03±0.005</td>
<td>0.18±0.04</td>
<td>0.06±</td>
<td>0.19±0.05</td>
</tr>
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</table>

*EC50 of MT-II is reported as reference.*

**Figure 1.** Logarithmic graphical representation of functional agonist activity (EC50 in nM) of linear peptide precursors (I′−X′).

**Figure 2.** Logarithmic graphical representation of agonist potency (EC50 in nM) of the [1,2,3]triazolyl-containing cyclopeptides (I−X).
and V and III and VI, respectively). The location of the 1,2,3[triazolyl moiety within the bridge in analogues I and VIII is identical with the location of the isosteric amide within the lactam bridge in MT-II. Evidently, I and VIII are less potent than MT-II in mMC1R, mMC3R, and mMC4R (Table 3). Moreover, I, which incorporates a 4,1-[1,2,3]triazolyl moiety, is much less potent than VIII, which incorporates a 1,4-[1,2,3]triazolyl moiety. Interestingly, analogue VII, which incorporates the 1,4-[1,2,3]triazolyl, is more potent than analogue II, which incorporates the 4,1-[1,2,3]triazolyl moiety despite the [1,2,3]triazolyl moiety at the same position in the bridge. In both pairs I and VIII and II and VII, where the position of the [1,2,3]triazolyl ring is closer to the N-proximal bridgehead, the more potent analogues VII and VIII are those that incorporate the 1,4-[1,2,3]triazolyl moiety. On the contrary, in both pairs IV and V and III and VI, where the position of the [1,2,3]triazolyl ring is closer to the C-proximal bridgehead, the more potent analogues IV and V are those that incorporate the 4,1-[triazolyl] moiety. Analogues IX and X have a shorter bridge than analogues I–VIII (four methylenes in total and a 1,4-disubstituted [1,2,3]triazolyl moiety), and the [1,2,3]triazolyl moiety is flanked by two methylenes on each side in either the 4,1- or 1,4-orientation. Moreover, analogues IX and X differ in the orientation of the [1,2,3]triazolyl moiety, which is positioned equal distance from the bridgeheads, being 4,1 in the former and 1,4 in the latter. While both IX and X are more potent at mMC4R and mMC5R than on mMC1R and mMC3R, the activities of analogue IX and X differ by the latter being more potent at the two former receptor subtypes, thus being more selective than analogue IX. The potency profiles of analogues IX and X (EC_{50} 0.32, 0.63, 0.14, and 0.15 nM and 0.20, 0.29, 0.067, and 0.07 nM at mMC1R, mMC3R, mMC4R, and mMC5R, respectively) follow very closely that of the corresponding analogues VIII and III (EC_{50} 0.53, 0.60, 0.14, and 0.13 nM and 0.1, 0.21, 0.051, and 0.042 nM at mMC1R, mMC3R, mMC4R, and mMC5R, respectively).

Taken together, these results suggest that close proximity of the [1,2,3]triazolyl moiety to the N-proximal bridgehead, especially in the 4,1-orientation, is disruptive for activity at all four receptor subtypes (analogues I and VIII, respectively). Interestingly, close proximity of the [1,2,3]triazolyl moiety to the C-proximal bridgehead as in analogues V and IV, presenting the [1,2,3]triazolyl in the respective 1,4- and 4,1-orientations, results in an opposite effect in which it is very disruptive for the former and very accommodating for the latter for all four receptor subtypes. Of note is the higher activity of the linear precursors I’ and V’ on the mMC1R as compared to the activity of the corresponding [1,2,3]triazolyl-containing cyclopeptides I and V on the same receptor subtype (EC_{50} 0.46 and 0.24 nM vs 5.18 and 0.85 nM). This may be the result of the serious perturbation imposed by the 4,1-[1,2,3]triazolyl moiety when it is adjacent to the N-proximal bridgehead and to a lesser extent, regardless of the orientation of the [1,2,3]triazolyl ring, when it is adjacent to the C-proximal bridgehead.

The comparison of the activity profile of the linear peptide precursors and their corresponding [1,2,3]triazolyl-containing cyclopeptides indicates the latter, in general, to be slightly more potent. Clearly, the linear precursors are already very potent indicating that the conformational rigidification in the cyclopeptide is not critical for their high potency. Moreover, only in very few cases the linear precursor peptides are more potent than their corresponding [1,2,3]triazolyl-containing cyclopeptides (cf. I’ and V’ with I and V in mMC1R, respectively).

Conformational Analysis. To gain insight on the relationship among the specific features of the [1,2,3]triazolyl-containing bridge, the potency and receptor subtype selectivity, and the conformation, we selected to study in detail the conformations of three representative heterodetic [1,2,3]triazolyl-containing cyclopeptides, I, IV, and V. The selection includes the least potent analogue I, one of the most potent analogues IV, and analogue V of intermediate potency. In addition, while analogues I and IV incorporate the 4,1-[1,2,3]triazolyl moiety in proximity to the N-proximal- and C-proximal bridgeheads, respectively, both analogues IV and V incorporate the [1,2,3]triazolyl moiety at the C-proximal bridgehead but at opposite orientations, 4,1- and 1,4-, respectively. We anticipated that the opposing location of the [1,2,3]triazolyl ring in the bridge will affect the conformation to the extent that will explain the large difference in activity between analogues I and IV. On the other hand, we speculate that the small difference in biological activity between analogues IV and V, which share the same C-proximal bridgehead location of the [1,2,3]triazolyl moiety, may result in similar conformations in spite of the opposite orientations of the [1,2,3]triazolyl ring within the bridge.

To allow comparison with the reported “bioactive” NMR solution structure(s) of the reference homodetic cyclopeptide MT-II, the NMR spectra of the heterodetic cyclopeptides I, IV, and V were acquired in DMSO-d_6. Moreover, this polar, basic, and aprotic solvent was shown to support intramolecular hydrogen bonds and prevent intermolecular aggregations.

To exclude potential aggregation, we recorded the 1D-^1H NMR spectra of the clicked-cyclopeptides at a concentration range spanning 1–0.1 mM. Importantly, at a concentration of 0.1 mM, there was no indication for aggregation. Nevertheless, at 0.1 mM, cyclopeptide I presented double signal pattern spectra indicating the presence of two distinct populations of slow exchanging conformations. Chemical shift assignments of the proton spectra were achieved via the standard systematic application of DQF-COSY, TOCSY, and NOESY experiments, using the SPARKY software package according to Wüthrich’s procedure.

Figure 3 reports sequential and medium range NOEs collected in NOESY spectra of cyclopeptides I, IV, and V in DMSO-d_6 confirming the preliminary analysis of 1D-^1H NMR spectrum. Two NOEs bar diagrams are reported for [1,2,3]-triazolyl-containing cyclopeptide I, corresponding to the observed double signal pattern in the respective NOESY spectrum (Figure 3A,B). The conformer IA is characterized by a more complete sequential and medium range NOE pattern, while conformer IB shows less abundant and, whenever observable, weak, sequential, and medium range connectivities. This observation is consistent with the presence of the two predominant low energy conformational ensembles IA and IB, undergoing slow dynamic interchange. We suggest that the markedly low potency of cyclopeptide I compared to cyclopeptides IV and V in all melanocortin receptor subtypes (EC_{50} 5.18, 3.14, 1.62, and 1.93 nM for mMC1R, mMC3R, mMC4R, and mMC5R, respectively) can be attributed to this conformational heterogeneity. NOE bar diagrams relative to cyclopeptides IV and V (parts C and D of Figure 3, respectively), confirm the presence of numerous and diagnostic sequential and medium range i,i+2 and i,i+3 NOE data in the corresponding NOESY spectra, although a more regular and complete NOE patterns is observed for cyclopeptide V than for cyclopeptide IV. NMR-derived models of cyclopeptides IA, IV,
and V were calculated using simulated annealing procedures imposing NOE-based interprotonic distances as constraints. Figure 4 shows the NMR-derived bundles of 20 low energy conformations of cyclopeptides IA, IV, and V overlapping with the best calculated structure of MT-II. The NMR structure bundles of cyclopeptides IA and V show high structural agreement with RMDS $\leq$ 0.50 Å. In agreement with the NOE bar diagrams reported in Figure 3, the structure bundle of cyclopeptide IV is characterized by slightly higher RMSD value (0.80 Å), indicating that cyclopeptide IV is more flexible than cyclopeptides IA and V.

Quantitative analysis of $\phi$ and $\psi$ (Supporting Information, Table 1S) dihedral angles of low-energy structures of cyclopeptides IA, IV, and V, using the PROMOTIF program (PROMOTIF, a program to identify and analyze structural motifs in proteins)$^{38}$ highlights the preponderance of a type I $\beta$-turn conformation centered on residues His$^6$-Trp$^9$. In agreement with the preliminary analysis shown in the NOEY connectivity bar diagram (Figure 3), cyclopeptide IB appears to be disordered, presenting many irregular secondary structures including some transient $\alpha$-helical segments.

The comparison of the 20 lowest-energy NMR structures of cyclopeptides IA, IV, and V (Figure 4) suggests good overlap of the backbone atoms and a common orientation of His and D-Phe side chains. Cyclopeptides IV and V show also a common orientation of the indole ring of Trp.

As shown in Figure 4a the Trp side chain of cyclopeptide IA points to an opposite direction as compared to cyclopeptide IV and V. Moreover, the three cyclopeptides differ in the orientation of the Arg side chain. In cyclopeptides IV and V, as documented by the relative NOE effects, Arg side chains point to [1,2,3]triazolyl ring and D-Phe moiety, respectively (Figure 4a). Consistent with the absence of NOEs suggesting proximity to other side chains, Arg side chain in cyclopeptide IA, displays significantly higher flexibility than in analogues IV and V (Figure 4a,b).

Stabilization of a type II $\beta$-turn because of D-Phe and the close proximity of His, D-Phe, and Trp side chains on one of the homodetic cyclopeptide surface and Arg side chain on the opposite surface in MT-II can explain the high potency and affinity to the melanocortin receptors. It is known that a correct spatial orientation of the hydrophobic aromatic residues and the basic, “arginine-like” moiety, are important for an effective interaction of MT-II and MT-II analogues at the different MCR receptors. In summary, conformational analysis of the three selected heterodetic [1,2,3]triazolyl-containing MT-II analogues suggests that (i) the conformational polymorphism of cyclopeptide I contributes to its overall low potency in all receptor subtypes as compared to the more conformationally defined cyclopeptides IV and V, and (ii) the presence, location, and orientation of the [1,2,3]triazolyl ring in the bridge does affect the spatial location of the side chains in general and of the Arg side chain in particular. It appears that biological activity is derived from indirect contribution to the spatial orientations of side chains more then by the direct interaction of the bridge with the target receptor.

### CONCLUSIONS

In the present paper, we report for the first time a systematic structure–activity conformation relationship study in which a i-to-i+5 side chain-to-side chain Cu$^+$-catalyzed azido-to-alkyne 1,3-dipolar cycloaddition (CuAAC) was performed in an attempt to reproduce a type I $\beta$-turn, which is considered to be a critical feature in the putative bioactive conformation of MT-II. We demonstrate that (1) i-to-i+5 side chain-to-side chain CuAAC yields potent and interesting melanocortin receptor agonists; (2) side chain-to-side chain conformational stabilization of type I $\beta$-turn by i-to-i+5 side chain-to-side chain [1,2,3]triazolyl-containing bridges impacts the in vitro potency and selectivity of these new stapled peptides at the different MCR receptor subtypes. Relative to MT-II, we frequently observe enhancement of potency at the mMCSR and loss of potency at mMC1R. (3) Similar to the general enhancement of potency as a result of homodetic side chain-to-side chain cyclization (cf. BE124 and BE123 with MT-II, Table 2), we also observe enhancement of potency upon transforming the linear precursors I′–X′ to the heterodetic [1,2,3]triazolyl-containing cyclopeptides. Interestingly, similar observation was reported by Cho et al. in i-to-i+5 disulfide-bridged cyclic MSH peptides. (4) Regardless of the nature of i-to-i+5 side chain-to-side chain cyclizations, their contribution to the stability of the critical type I $\beta$-turn contributes to the enhancement of their agonist potency at all MCRs. Our observations concur with those that propose side chains orientation of residues within the cyclic segment and those that are flanking it to play a predominant role in determining receptor subtype selectivity. As such, the i-to-i+5 side chain-to-side chain CuAAC offers obvious advantages over the more traditional side chain-to-side chain cyclizations. These heterodetic [1,2,3]triazolyl-containing MT-II mimetics can be
constructed by taking advantage of the chemical orthogonality of the azido and alkynyl functions, which reduces synthetic complexity. Moreover, our approach offers access to a vast structural diversity presented by the size of the bridge, the location, and the orientation of the [1,2,3]triazolyl moiety within the ring. In the future, we will seek to reduce the size of the [1,2,3]triazolyl-containing bridge to achieve greater structural rigidity and diversify the nature of the residues flanking the cyclic segment in order to achieve greater receptor subtype selectivity.

**EXPERIMENTAL SECTION**

**Materials and Methods.** The ω-alkynyl-alcohols were purchased from Alfa Aesar; Fmoc-ω-Arg-(Boc)-OH, Fmoc-ω-Phe-OH, and Fmoc-ω-Pra-OH amino acids, Fmoc-Rink amide resin, and HOBT were purchased from Iris Biotech GmbH (Markredwitz, Germany); Fmoc-ω-Arg(Pbf)-OH and Fmoc-ω-His(Trt)-OH amino acids were purchased from CBL (Patras, Greece); Fmoc-ω-Nle was purchased from Acros Organics (Geel, Belgium), and Carlo Erba (Milan, Italy). The scavengers for cleavage of peptides from resin, 1,2-ethanedithiol (EDT), thioanisole, and phenol (PhOH), were purchased from Janssen Chimica (Beerse, Belgium), and Carlo Erba (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy). Peptide-synthesis grade N,N-dimethylformamide (DMF) was purchased from NeoMPS (Strasbourg, France); TBTU from Advanced Biotech Italy (Milan, Italy).

The purity of the final linear precursors (I′−X′) and the [1,2,3]triazolyl-containing MT-II analogues (I−X) was established by analytical RP-HPLC and exceeded 95%, their structural integrity was established by ESI-MS.

### Synthesis of N^ω^-Fmoc-Xaa(ω-N$_2$)-OH and N^ω^-Fmoc-Yaa(ω-ynyl)-OH.

N^ω^-Fmoc-ω-azido-ω-amino- and N^ω^-Fmoc-ω-ynoic-ω-amino acids (1−4 and 5−7, respectively), with (CH$_2$)$_n$/m = 1−4 in the side chain, were synthesized as described before.$^{21,22}$ In particular, N^ω^-Fmoc-ω-azido-ω-amino acids were obtained by diazo-transfer from correspondent N^ω^-Fmoc-ω-amino-ω-amino acids; N^ω^-Fmoc-ω-ynoic-ω-amino acids were obtained by alkylation of a Ni(II) complex of the Schiff base derived from glycine and the chiral inducer (S)-2-(N-benzylpropyl)amino benzophene with alk-ω-ynyl bromides.

### Synthesis of N^ω^-Ac[Nle(4),Yaa(5),Phe(7),Xaa(10)]MSH(4−10)NH$_2$

(I′, II′, III′, IV′, IX′) and N^ω^-Ac[Nle(4),Xaa(5),Phe(7),Yaa(10)]MSH(4−10)NH$_2$ (VI′, V′, VII′, VIII′, X′). Peptides I′−X′ were synthesized as a manual batch synthesizer (PLS 4×4, Advanced ChemTech) employing Fmoc/tBu chemistry. The syntheses were performed on Rink-amide NovaSyn TGR resin (0.14 mmol/g, 300 mg). The Fmoc-Rink-amide resin was swelled with DMF (1 mL/100 mg of resin) for 20 min before use. Stepwise peptide assembly was performed by repeating for each added amino acid the following deprotection-coupling cycle. (1) Swelling: DMF (1 mL/100 mg of resin) for 5 min. (2) Fmoc-deprotection: was washed twice with 20% (v/v) piperidine in DMF (1 mL/100 mg of resin, one wash for 5 min, followed by another wash for 20 min). (3) Resin washing: DMF (3−5 min). (4) Coupling: scale employed TBTU/HOBt/NMM (2.5:2.5:3.5 equiv) as the coupling system and 2.5 equiv of the Fmoc protected amino acids, except for the unnatural amino acids N^ω^-Fmoc-Xaa(ω-N$_2$)-OH and N^ω^-Fmoc-Yaa(ω-ynyl)-OH for which 1.5 equiv were used. The coupling was carried out in DMF (1 mL/100 mg of resin) for 50 min. (5) Resin washings: DMF (3−5 min) and DCM (1−5 min). Each coupling was monitored by Kaiser test$^{12}$ and was negative, therefore recouplings were not needed. Acetylation of the amino terminus was carried out in the presence of Ac$_2$O/NMM in DCM (20 equiv, 1.6 mL of DCM). The reaction was monitored by Kaiser test.

*Deprotection, Cleavage, and Purification of Peptides: General Procedure.* Peptide cleavage from the resin and simultaneous deprotection of the amino acid side chains were carried out with a mixture of TFA/anisole/1,2-ethanediol/phenol/H$_2$O (94:1:1:1:1 v/v/v/v, 1 mL/100 mg of resin-bound peptide). The cleavage was carried out for 3 h with vigorous shaking at room temperature. Resin was filtered and washed with TFA. The filtrate was concentrated under N$_2$ stream, addition of cold diethyl ether resulted in a precipitate that was separated by centrifugation, dissolved in H$_2$O.
and lyophilized on an Edwards apparatus, model Modulyo. Lyophilized crude peptides were prepurified by solid-phase extraction with a RP-18 LiChrorep silica column from Merck (Darmstadt, Germany) using H2O/ACN as eluents. The final purification of the peptides was performed by semipreparative RP-HPLC on a Phenomenex Jupiter C-18 (250 mm × 4.6 mm) column at 28 °C using a Waters instrument (separation module 2695, detector diode array 2996) working at 4 mL/min. The solvent systems used were: A (0.1% TFA in H2O) and B (0.1% TFA in CH3CN). Final purity of all peptides was ≥95%. Peptides were characterized by RP-HPLC ESI-MS. HPLC was performed using a Phenomenex Kinetex C-18 column 2.6 μm (100 mm × 3.0 mm) working at 0.6 mL/min, with UV detection at 215 nm, coupled to a Phenomenex ESI-MS using an Alliance Chromatography (Waters) with a Phenomenex Kinetex C-18 column 2.6 μm (100 mm × 3.0 mm) working at 0.6 mL/min, with UV detection at 215 nm, coupled to a single quadrupole ESI-MS (Micromass ZQ). The solvent systems used were: A (0.1% TFA in H2O) and B (0.1% TFA in CH3CN).

Synthesis of Nα-Nle4,Yaa5,O-Phe7,Xaa10(α,β)αMSH(4-10)NH2, [8′[N1-1,2,3-triazolyl-CH2]3,β&] (I, II, III, IV, IX) and Nα-Nle4,Yaa5,O-Phe7,Xaa10(α,β)αMSH(4-10)NH2, [8′[N1-1,2,3-triazolyl-CH2]3,β&] (V, VI, VII, VIII, X). Purified linear peptide precursors I–X (>95% purity) were subjected to intramolecular Cu-catalyzed side-chain-to-side-chain azide–alkyne 1,3-dipolar Huisgen cycloaddition (CuAAC). To a solution of the purified linear peptide (4.77 μmol) in H2O/tBuOH (5 mL, 2:1 v/v) were added CuSO4·5H2O (40 μmol) and ascorbic acid (40.3 μmol). The mixture was stirred overnight at room temperature, concentrated, and lyophilized. Complete and clean conversion of all linear precursors into the desired [1,2,3]triazolyl-containing cyclopeptides I–X (Schemes 1 and 2) was observed. The cyclization was monitored by RP-HPLC ESI-MS. The crude [1,2,3]triazolyl-containing cyclopeptides were purified (>97% purity) by solid-phase extraction (SPE) on RP (C18) column, with CH3CN in H2O as eluents. The copper salts were removed by elution with H2O, whereas the desired cyclopeptides were eluted with CH3CN/H2O mixture. Peptides were characterized and semipreparative RP-HPLC and RP-HPLC ESI-MS, respectively.

Solid Phase Synthesis of MT-II: Nα-Nle4,Yaa5,c[Aspβ−O-Phe−Lysβ]αMSH(4-10)NH2. The fully protected resin-bound peptide was synthesized using a Rink-amine NovaSyn TGR resin (0.63 mmol/g, 300 mg) on a manual batch synthesizer (PLS 4 × 4, Advanced ChemTech) applying the Fmoc/tBu SPPS procedure, as previously reported. The coupling of Nα-Fmoc-amino acids was performed using TBTU/HOBt/NMM (2.5 equiv:2.5 equiv:3.5 equiv) and 2.5 equiv of each Fmoc protected amino acids. The coupling was carried out in DMF (1 mL/100 mg of resin) for 40 min. The formation of the lactam bridge between side chains of Lysβ and Aspβ was performed on the resin-bound peptide, following an orthogonal protocol of deprotection. In particular, Lysβ and Aspβ were protected on side chain respectively with 1-[(4,4-dimethyl-2,6-dioxycyclohex-1-ylidene)ethyl] group (Dde) and with β-4-{N-1,4-diacylaminocyclohexyliden-3-methylbutyl]-amino benzyl ester (ODmbb), stable under the conditions of peptide elongation. After acetylation of the N-terminal, Dde and ODmbb were removed by two consecutive steps, using hydrazine 20% in DMF (1 mL), first for 5 min, followed by an additional step with freshly prepared deprotection mixture for 15 min. After deprotection of Lysβ and Aspβ, the formation of intramolecular lactam bridge was performed using TBTU/HOBt/NMM (2.5:2.5:3.5 equiv) as coupling system in DMF for 40 min. The cyclization reaction was monitored by Kaiser test and by RP-HPLC ESI-MS analysis of MW-assisted mini-cleavage resin-bound fragment. Mini-cleavage was carried out with TFA/TIS/water solution (95:2.5:2.5 v/v/v), using a DiscoverTMS-class single-mode MW reactor equipped with an Explorer-48 autosampler (CEM). The reaction was performed at 45 °C, using 15 W for 15 min. The reaction mixture was then filtered, and the crude peptide was precipitated from the cleavage mixture by addition of ice-cold diethyl ether followed by cooling for 5 min at −20 °C. The product was collected by centrifugation and directly subjected to RP-HPLC ESI-MS analysis.

Biological Activity. cAMP-based functional bioassay: HEK-293 cells stably expressing the mouse melanocortin receptors were transiently transfected with 4 μg of CRE/β-galactosidase reporter gene as previously described.28,29,43,44 Briefly, 5000–15000 post-transfection cells were plated into collagen treated 96-well plates (Nunc) and incubated overnight. Then 48 h post-transfection, the cells were treated with 100 μL of peptide (10−6–10−10 M) or forskolin (10−4 M) as a control in assay medium (DMEM containing 1.0 mg/mL BSA and 0.1 mM isobutylmethylxanthine) for 6 h. The assay media was aspirated, and 50 μL of lysis buffer (250 mM Tris-HCl pH = 8.0 and 0.1% Triton X-100) was added. The plates were stored at −80 °C overnight. The plates containing the cell lysates were thawed the following day. Aliquots of 10 μL were taken from each well and transferred to another 96-well plate for relative protein determination. To the cell lysate plates, 40 μL of phosphate-buffered saline with 0.5% BSA was added to each well. Subsequently, 150 μL of substrate buffer (60 mM sodium phosphate, 1 mM MgCl2, 10 mM KCl, 5 mM β-mercaptoethanol, 2 mg/mL ONPG) was added to each well, and the plates were incubated at 37 °C. The sample absorbance, OD595, was measured using a 96-well plate reader (Molecular Devices). The relative protein concentration was determined by adding 200 μL of 1:5 dilution Bio Rad G250 protein dyewater to the 10 μL cell lysate sample taken previously, and the OD595 was measured on a 96-well plate reader (Molecular Devices). Data points were normalized to the relative protein content. EC50 values represent the mean of three or more independent experiments. EC50 estimates, and their associated standard errors of the mean, were determined by fitting the data to a nonlinear least-squares analysis using the PRISM program (v4.0, GraphPad Inc.).

NMR Conformational Analysis. Samples for NMR were prepared by dissolving 1.2 mg of the lactam- and heterodetic [1,2,3]triazolyl-containing cyclopeptides in 0.5 mL of aqueous phosphate buffer (pH 6.6, 100 mM). Samples were lyophilized and dissolved in a mixture of DMSO/water (0.5 mL, 80:20, v/v). NMR spectra were recorded on a Bruker DRX-600 spectrometer. To exclude potential aggregation, one-dimensional (1D) proton spectra of the cyclopeptides at a concentration range spanning 0.1–1 mM were recorded. At a peptide concentration of 0.1 mM, the peptides did not display any noticeable effects of aggregation. 1D NMR spectra were recorded in the Fourier mode with quadrature detection. The water signal was suppressed by a low-power selective irradiation in the homogenated mode. DQF-COSY, TOCSY, and NOEY experiments were performed in the phase-sensitive mode using quadrature detection in one-dimensional (1D) proton spectra of the cyclopeptides at a concentration range spanning 0.1–1 mM. 2D spectroscopy of the precursors was performed in the phase-sensitive mode using quadrature detection in both dimensions. 2D experiments were multiplied by shifted sin2 functions in both dimensions. A mixing time of 15 or 25 ms was used, and the mixing period was selected based on the relaxation time of the signal. 3D experiments were performed in the phase-sensitive mode using quadrature detection in one-dimensional (1D) proton spectra of the precursors. 3D spectra of the precursors were multiplied by shifted sin2 functions in both dimensions. 3D experiments were performed in the phase-sensitive mode using quadrature detection in one-dimensional (1D) proton spectra of the precursors. 3D experiments were performed in the phase-sensitive mode using quadrature detection in one-dimensional (1D) proton spectra of the precursors. 3D experiments were performed in the phase-sensitive mode using quadrature detection in one-dimensional (1D) proton spectra of the precursors. 3D experiments were performed in the phase-sensitive mode using quadrature detection in one-dimensional (1D) proton spectra of the precursors.
ASSOCIATED CONTENT

Supporting Information
Product characterization and NMR data. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes
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ABBREVIATIONS USED

ACN, acetonitrile; CuAAC, Cu¹-catalyzed azido-to-alkyne 1,3-dipolar cycloaddition; DCM, dichloromethane; DMF, N,N-dimethylformamide; EDT, 1,2-ethanedithiol; ESI-MS, electron-spray ionization mass spectrometry; GPCR, G protein-coupled receptors; HOBt, N-hydroxybenzotriazole; MCR, melanocortin GPCR receptors; MSH, melanocortin stimulating hormones; MT-II, melanotropins; NMM, N-methyl morpholine; MCR, melanocortin receptors; NMM, N-methyl morpholine; MSH, melanocortin stimulating hormones; MT-II, melanotropins; NMP, N-propargylglucose; POMC, pro-opiomelanocortin prohormone; PTHrP, parathyroid hormone-related protein; SAR, structure-activity relationships; TBTU, 1-[bis(dimethylamino)-methylene]-1H-benzo[d]triazol-3-oxide tetrafluoroborate; TFA, trifluoroacetic acid; UPLC, ultra-performance liquid chromatography

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