Trapping Hydrogen Sulfide (H2S) with Diselenides: The Application in the Design of Fluorescent Probes

Citation

Published Version
doi:10.1021/acs.orglett.5b00431

Permanent link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:25658390

Terms of Use
This article was downloaded from Harvard University’s DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

Share Your Story
The Harvard community has made this article openly available. Please share how this access benefits you. Submit a story.

Accessibility
Trapping Hydrogen Sulfide (H$_2$S) with Diselenides: The Application in the Design of Fluorescent Probes

Bo Peng, Caihong Zhang, Eizo Marutani, Armando Pacheco, Wei Chen, Fumito Ichinose, and Ming Xian

†Department of Chemistry, Washington State University, Pullman, Washington 99164, United States
‡School of Chemistry and Chemical Engineering, Center of Environmental Science and Engineering Research, Shanxi University, Taiyuan, China
§Department of Anesthesia, Critical Care and Pain Medicine, Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts 02114, United States

Supporting Information

ABSTRACT: Here we report a unique reaction between phenyl diselenide-ester substrates and H$_2$S to form 1,2-benzothiaselenol-3-one. This reaction proceeded rapidly under mild conditions. Thiols could also react with the diselenide substrates. However, the resulted S–Se intermediate retained high reactivity toward H$_2$S and eventually led to the same cyclized product 1,2-benzothiaselenol-3-one. Based on this reaction two fluorescent probes were developed and showed high selectivity and sensitivity for H$_2$S. The presence of thiols was found not to interfere with the detection process.

Hydrogen sulfide (H$_2$S), traditionally known as a venomous gas with a stinky smell, has been recently recognized as an important signaling molecule. H$_2$S is generated in mammalian cells by enzymes including cystathionine β-synthase (CBS), cystathionine γ-lyase (CSE), and 3-mercaptopropionate sulfurtransferase (3-MST), and cysteine and derivatives serve as the substrates. Recent studies have revealed a variety of functions of H$_2$S in physiological and pathological processes. However, the mechanisms behind those roles are still poorly understood. H$_2$S is a highly reactive molecule that can react with many biological targets such as hemoglobin and phosphodiesterase. These reactions are responsible for the biological functions of H$_2$S. On the other hand, these complicated reactions also make the detection of H$_2$S in biological systems very difficult. Early reports claim physiological H$_2$S concentrations can be as high as 20–80 μM in plasma. However, it is believed now that the concentrations of free H$_2$S are low, at submicromolar or nanomolar levels.

In the past several years fluorescence based assays have received considerable attention in this field and a number of fluorescent probes for H$_2$S have been reported. All of these probes utilize reaction-based fluorescence change strategies, i.e. using certain H$_2$S-specific reactions to convert nonfluorescent substrates to materials with strong fluorescence or to change the fluorescent properties of the probes (for ratiometric probes). Currently three types of reactions are normally used in the design of probes: (1) H$_2$S-mediated reductions, often using azide (–N$_3$) substrates; (2) H$_2$S-mediated nucleophilic reactions; and (3) H$_2$S-mediated metal–sulfide formation. In 2011 our laboratory developed the first nucleophilic reaction based strategy for the design of probes (i.e., WSP series). The strategy is described in Scheme 1. The probe (WSP) contains two electrophilic centers. H$_2$S reacts with the pyridyl disulfide first to give a persulfide intermediate 1, which precedes a fast cyclization to release the fluorophore and 1,2-benzodithiol-3-one 2. It is anticipated that the probe can also react with biothiols to form 3. Theoretically 3 could further react with H$_2$S to form 1 and then turn on the fluorescence. However, this reaction is somewhat slow, especially when the concentration of H$_2$S is low (under physiological conditions). In addition, H$_2$S may react with both sulfurs of the disulfide of 3, which makes this pathway not productive. It should be noted that the reaction between WSP and biothiols cannot turn the fluorescence on; therefore, the selectivity of WSP is found to be excellent. We were also able to optimize the detection conditions and found fluorescence “turn-on” could be achieved in a few minutes. Such a fast reaction allows effective detection of H$_2$S even when high concentrations of biothiols are presented, albeit the fluorescence signals are significantly decreased.

Received: February 10, 2015
Published: February 27, 2015

DOI: 10.1021/acs.orglett.5b00431
Org. Lett. 2015, 17, 1541−1544
As described above, the possible consumption of WSP probes by biothiols is a weakness. To solve this problem two strategies may be applied: (1) a H2S-specific “trapper” should be used to replace the pyridyl-disulfide. This might be difficult as intracellular concentrations of biothiols are much higher than H2S; (2) a nonconsuming trapper with biothiols should be identified and used. Such a trapper may react with biothiols, but the reaction should not lead to the release of the fluorophore. Moreover, ideally the reaction product or intermediate should maintain high reactivity toward H2S which therefore can lead to fluorescence “turn-on” by H2S. Herein we report our progress in pursuing the latter strategy. Diselenide-based substrates were found to be suitable for this goal.

Our design of the diselenide-based strategy for the selective trapping of H2S (not consumed by biothiols) is shown in Scheme 1. It is known that diselenide bonds can be cleaved by sulfur-based nucleophiles very effectively (about 5 orders of magnitude faster than disulfide bonds).19,20 It is also known that diselenides can facilitate disulfide formation from thiols.21 H2S (pKₐ 6.8) is a stronger nucleophile than common biothiols such as Cys and GSH. Therefore, we expect diselenide-based reagents such as 4 should react with H2S very effectively. As such two products should be formed: a thio-benzeneselenol derivative 5 and a benzeneselenol derivative 6. As an extremely unstable intermediate, benzeneselenol 6 should be rapidly oxidized to reform the probe 4.22 In contrast, 5 should undergo a fast and spontaneous cyclization to form 1,2-benzothia-selenol-3-one 7 and release the fluorophore. We also expected the diselenide bond should be quite reactive to biothiols (RSH). Such reactions should lead to two products: 6 and a S–Se conjugate 8. Again, 6 should be oxidized to regenerate the probe. For as 8, previous studies revealed that nucleophilic attack by thiols at selenium is both kinetically much faster and thermodynamically more favorable.19 As such, the reaction between 8 and other biothiols should not change the probe’s structural template (i.e., maintaining the −S−Se− conjugation). However, if H2S is present, the reaction should lead to intermediate 5, and the following cyclization should produce 7 and the fluorophore. Overall we expected the probe would specifically react with H2S to release the fluorophore and the presence of biothiols should not interfere with the process.

With this idea in mind, a model compound 9 was prepared and tested. As shown in Scheme 2, the reaction between compound 9 and H2S (5 equiv, using Na2S as the equivalent) was found to be fast, which completed in 10 min. The desired products 7 and phenol were obtained in excellent yields. The yield of 7 was calculated based on two selenium moieties in the starting material. We did not observe the formation of the benzeneselenol product in the reaction. When Cys (10 equiv) coexisted with H2S (5 equiv) in the reaction, we obtained the same products in high yields. More interesting, even if Cys (5 equiv) was treated with 9 first for 1 h, the addition of H2S (2.5 equiv) still provided the desired products in similar yields. These results supported our hypothesis that diselenide-based substrates could selectively and effectively react with H2S to form the cyclized product 7 and this reaction was not affected by thiols.

Based on this unique reaction we synthesized two fluorescent probes SeP1 and SeP2 (shown in Figure 1). 7-Hydroxy-

---

Scheme 1. Design of the Fluorescent Probes for H2S Detection

Previous work

This work

Scheme 2. Model Reactions between Compound 9 and H2S

---

Figure 1. Structures of diselenide-based probes.
responses to H$_2$S in different solvent systems, and a mixed CH$_3$CN/PBS buffer (10 mM, pH 7.4, 1:1) solution was found to be the optimum system for the measurement. In this system the fluorescence intensity of the probe (10 μM) could reach the maximum in less than 2 min upon treatment of H$_2$S (using 50 μM Na$_2$S) as shown in Figure 2, demonstrating this was a fast process. Fluorescence increases were also found to be significant. Intensities increased 35- and 14-fold for probe SeP1 and SeP2 respectively. When a series of different concentrations of Na$_2$S were treated with the probes we observed fluorescence intensities increased in almost a linear fashion in the range 0–15 μM. Data obtained with SeP2 are shown in Figure 3. The detection limit of SeP2 was calculated to be 0.06 μM. Data of SeP1 are shown in Figure S1 in the Supporting Information.

![Figure 2](image1.png)

**Figure 2.** Time-dependent fluorescence changes of the probes (10 μM) in the presence of Na$_2$S (50 μM). The reactions were carried out for 30 min at room temperature in CH$_3$CN/PBS buffer (10 mM, pH 7.4, 1:1, v/v). Data were acquired at 455 nm with excitation at 340 nm for SeP1 (■) and at 521 nm with excitation at 498 nm for SeP2 (●).

![Figure 3](image2.png)

**Figure 3.** Fluorescence emission spectra of SeP2 (10 μM) in the presence of varied concentrations of Na$_2$S (0, 0.5, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15 μM). The reactions were carried out for 5 min at room temperature in CH$_3$CN/PBS buffer (10 mM, pH 7.4, 1:1, v/v). Data were acquired with excitation at 498 nm for SeP2.

We also examined the selectivity of the probes for H$_2$S over other reactive sulfur species, including cysteine (Cys), glutathione (GSH), sulfite (SO$_3^{2-}$), sulfate (SO$_4^{2-}$), and thiosulfate (S$_2$O$_3^{2-}$). As shown in Figure 4, all of these species did not show a significant fluorescence increase even under much higher concentrations (up to mM). In addition, when H$_2$S (50 μM) coexisted with Cys or GSH (1 mM), we observed strong fluorescence responses that were comparable (at ~90% levels) to the signals obtained with H$_2$S only. This was a significant improvement from our pyridyl disulfide based probes, which gave much decreased fluorescence when high concentrations of biothiols were present (at 25–55% levels of the signal of H$_2$S only).$^{15b}$

Next we tested SeP2 in imaging H$_2$S in cells. Freshly cultured HeLa cells were first incubated with SeP2 (50 μM) for 30 min and then washed with DMEM to remove excess probe. We did not observe significant fluorescent cells (Figure 5). However, strong fluorescence in the cells was observed after treating with Na$_2$S (100 μM) for 30 min. These results demonstrated that SeP2 could be used for cell imaging.

Finally we wondered if SeP2 could be used to measure endogenous H$_2$S concentrations changes. To this end, human neuroblastoma cells (SH-SY5Y) were separately treated with L- and D-cysteine (which are H$_2$S biosynthestic substrates), S-
showed decreased with Na2S showed the strongest
phenyl diselenide and H2S to form 1,2-benzothiaselenol-3-one.
changes.
neuroblastoma cells (treated with H2S substrates or CBS activator showed clearly
two fluorescent probes for the detection of H2S were
prepared and evaluated. The probes showed excellent
sensitivity and selectivity.

ASSOCIATED CONTENT
Supporting Information
Detailed synthetic procedures, characterist data, and exper-
imental procedures. This material is available free of charge via
the Internet at http://pubs.acs.org.

AUTHOR INFORMATION
Corresponding Author
*E-mail: mxian@wsu.edu.

Author Contributions
%B.P. and C.Z. contributed equally to this work.
Notes
The authors declare no competing financial interest.

ACKNOWLEDGMENTS
This work is supported by an American Chemical Society Teva
USA Scholar Grant and the NIH (R01HL101930 and
R01HL116571).

REFERENCES
(2) Fukuto, J. M.; Carrington, S. J.; Tantillo, D. J.; Harrison, J. G.;
Toxicol. 2012, 25, 769.
(4) Kolluru, G. K.; Shen, X.; Bir, S. C.; Kevil, C. G. Nitric Oxide
2013, 35, 5.
(9) Kabl, O.; Motl, N.; Banerjee, R. Biochim. Biophys. Acta 2014,
1844, 1355.
(10) Ríos-González, B. B.; Román-Morales, E. M.; Pietri, R.; López-
37.
Papapetropoulos, A. Nitric Oxide 2014, DOI: 10.1016/
j.niox.2014.12.004. (b) Ono, K.; Akaite, T.; Sawa, T.; Kumagai, Y.;
Wink, D. A.; Tantillo, D. J.; Hobbs, A.; Nagy, P.; Xian, M.; Lin, J.;
Wang, B. Sensors 2012, 12, 15907. (b) Pluth, M. D.; Bailey, T. S.;
Hammers, M. D.; Montoya, L. A. Biochalcogen Chemistry: The
Biological Chemistry of Sulfur, Selenium, and Tellurium, Vol. 1152;
Baysel, C. A., Brumaghim, J. L., Eds; American Chemical Society:
Washington, DC, 2013; p 15. (c) Lin, Y. S.; Chen, W.; Xian, M.;
Predmore, B. L.; Lefer, D. J.; Wang, B. Angew. Chem., Int. Ed. 2011,
50, 9672. (c) Wan, Q.; Song, Y.; Li, Z.; Gao, X.; Ma, H. Chem. Commun.
(15) (a) Liu, C.; Pan, J.; Li, S.; Zhao, Y.; Wu, L. Y.; Berkman, C. E.;
(b) Peng, B.; Chen, W.; Liu, C.; Rosser, E. W.; Pacheco, A.; Zhao, Y.;
(16) (a) Qian, Y.; Karpus, J.; Kabl, O.; Zhang, S. Y.; Zhu, H. L.;
Banerjee, R.; Zhao, J.; He, C. Natl. Commun. 2011, 2, 495. (b) Liu, J.;
Sun, Y.-Q.; Zhang, J.; Yang, T.; Cao, J.; Zhang, L.; Guo, W. Chem.—
Eur. J. 2013, 19, 4717. (c) Chen, Y.; Zhu, C.; Yang, Z.; Chen, J.; He,
2013, 52, 1688.
(17) (a) Sasakura, K.; Hanaoka, K.; Shibuya, N.; Mikami, Y.; Kimura,
2012, 41, 5799.
(18) (a) Furne, J.; Saeed, A.; Levitt, M. D. Am. J. Physiol Regul Integr
75, 6696.
2010, 5, 177. (b) Joris Beld, J.; Woycechowsky, K. J.; Hilvert, D.
Biochemistry 2008, 47, 6985. (c) Metanis, N.; Hilvert, D. Angew. Chem.,
(22) Rasmussen, B.; Sørensen, A.; Gottfredsen, H.; Pittelkow, M.