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The Toll-Like Receptor 4 (TLR4) Variant rs2149356 and Risk of Gout in European and Polynesian Sample Sets

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Abstract

Deposition of crystallized monosodium urate (MSU) in joints as a result of hyperuricemia is a central risk factor for gout. However other factors must exist that control the progression from hyperuricaemia to gout. A previous genetic association study has implicated the toll-like receptor 4 (TLR4) which activates the NLRP3 inflammasome via the nuclear factor-κB signaling pathway upon stimulation by MSU crystals. The T-allele of single nucleotide polymorphism rs2149356 in TLR4 is a risk factor associated with gout in a Chinese study. Our aim was to replicate this observation in participants of European and New Zealand Polynesian (M ori and Pacific) ancestry. A total of 2250 clinically-ascertained prevalent gout cases and 13925 controls were used. Non-clinically-ascertained incident gout cases and controls from the Health Professional Follow-up (HPFS) and Nurses Health Studies (NHS) were also used. Genotypes were derived from genome-wide genotype data or directly obtained using Taqman. Logistic regression analysis was done including age, sex, diuretic exposure and ancestry as covariates as appropriate. The T-allele increased the risk of gout in the clinically-ascertained European samples (OR = 1.12, P = 0.012) and decreased the risk of gout...
in Polynesians (OR = 0.80, \( P = 0.011 \)). There was no evidence for association in the HPFS or NHS sample sets. In conclusion TLR4 SNP rs2143956 associates with gout risk in prevalent clinically-ascertained gout in Europeans, in a direction consistent with previously published results in Han Chinese. However, with an opposite direction of association in Polynesians and no evidence for association in a non-clinically-ascertained incident gout cohort this variant should be analysed in other international gout genetic data sets to determine if there is genuine evidence for association.

**Introduction**

Deposition of crystallized monosodium urate (MSU) in joints as a result of hyperuricemia is a central risk factor for gout. Genome-wide association studies of serum urate have confirmed uric acid transporters SLC2A9 and ABCG2 as important loci, that are also associated with gout, with weaker effects from other uric acid transporters [1–3]. Given that many individuals with hyperuricaemia do not develop gout, other factors must exist controlling the progression from hyperuricaemia to gout [4]. However, very little is known about the genetic basis of the progression from hyperuricaemia to MSU crystal deposition to symptomatic gout [5]. The auto-inflammatory nature of gout involves the activation of the innate immune response by MSU crystals. A central pathway is primability and actual activation of the NLRP3 inflammasome and subsequent release of mature interleukin-1\( \beta \) [6, 7]. An important question is whether or not genetic regulation of this pathway is a risk factor for gout.

Toll-like receptors (TLRs) are transmembrane pattern recognition receptors expressed by innate immune cells that trigger an innate immune response by controlling distinct signaling pathways [8]. In addition to microbial ligands, these receptors also trigger innate immune response against endogenous ligands including MSU crystals [9]. To date ten functional human TLRs (TLR1-10) have been identified [10] of which TLR4 is a prominent member and which has been associated with a number of auto-inflammatory conditions [11]. This receptor activates the NLRP3 inflammasome via the nuclear factor-\( \kappa \)B signaling pathway. A TLR4 single nucleotide polymorphism (SNP; rs2149356) has been reported to be associated with gout in a Han Chinese sample set [12], where the TT-genotype was associated with an increased risk of gout (OR = 1.96 [95% CI 1.40–2.74]). The same genotype was associated with increased TLR4 mRNA expression and increased IL1\( \beta \) expression [12]. Our aim was to replicate the genetic association of rs2149356 with gout in individuals of European and New Zealand Polynesian ancestry.

**Subjects and Methods**

**Subjects**

Demographic and clinical data are presented in Table 1. All gout cases were clinically ascertained according to the 1977 American Rheumatism Association (ARA) classification criteria [13]. European cases (\( n = 1614 \)) were recruited from New Zealand (\( n = 647 \)), by the Eurogout consortium within the European Crystal Network (\( n = 779 \)) [14] and by the Arthritis Genomics Recruitment Initiative in Australasia (AGRIA; \( n = 188 \)). These cases were 84.1% male with average age of 63.0 (20–97) years. European non-gouty controls (\( n = 13005 \)), after exclusion criteria applied as described below, were recruited from NZ (\( n = 875 \)) and sourced from the Atherosclerosis Risk in Communities (ARIC; \( n = 8781 \)) and Framingham Heart...
(FHS; n = 3349 Generation 3 only) studies. The controls were 46.5% male with average age of 50.2 (17–95) years. There were 636 New Zealand Māori and Pacific Island (Polynesian) cases (87.5% male with average age of 49.9 (17–100) years) and 920 controls (46.7% male with average age of 41.1 (17–85) years). Individuals who ever self-reported as having gout or taking urate-lowering medication were excluded from the ARIC and FHS sample sets. The New Zealand Multi-Region Ethics Committee (MEC/105/10/130) and these institutional committees in Europe and Australia granted ethical approval: Research and Ethics Committee, Repatriation General Hospital, South Australia (32/08); Research Ethics Committee, University of New South Wales; Ethikkommission, Technische Universität Dresden (EK 8012012); South East Scotland Research Ethics Committee (04/S1102/41); Commission Cantonale (VD) D’éthique de la Recherche sur l’être Humain, Université de Lausanne; Commissie Mensgebonden Onderzoek regio Arnhem—Nijmegen; Partners Health Care System Institutional Review Board. All subjects gave written informed consent. The Database of Genotype and Phenotype (www.ncbi.nlm.nih.gov/gap) approval number was #834 for accessing data from the ARIC and FHS studies.

A separate data set, previously described in ref [15], of European Caucasian individuals was investigated consisting of 726 incident male cases and 3445 controls from the Health Professionals Follow-up Study (HPFS) and 351 incident female cases and 6317 controls from the Nurses Health Study (NHS) (Table 1). All cases were ascertained according to 1977 ARA classification criteria [13] using a self-administered gout questionnaire as previously described [16]. As summarised in ref [15] evaluation of medical records by two board-certified rheumatologists in a random audit set of 50 HPFS men demonstrated 94% (47/50) concordance of the diagnosis of gout between the self-administered questionnaire and the review of medical records and 91% (51/56) concordance in an audit sample of 56 NHS women.

### Table 1. Demographic and clinical data of participants.

<table>
<thead>
<tr>
<th></th>
<th>European</th>
<th>NZ Polynesian</th>
<th>HPFS</th>
<th>NHS</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Gout</td>
<td>Control</td>
<td>Gout</td>
</tr>
<tr>
<td><strong>Number</strong></td>
<td>13005</td>
<td>1614</td>
<td>920</td>
<td>636</td>
</tr>
<tr>
<td><strong>Male (%)</strong></td>
<td>6049 (46.52)</td>
<td>1332 (84.14)</td>
<td>419 (45.89)</td>
<td>546 (87.50)</td>
</tr>
<tr>
<td><strong>Age</strong></td>
<td>50.23±10.08</td>
<td>62.95±13.24</td>
<td>41.12±14.45</td>
<td>49.88±13.18</td>
</tr>
<tr>
<td><strong>BMI</strong></td>
<td>26.82±5.00 (12491)*</td>
<td>30.04±6.60 (1323)</td>
<td>32.67±7.26 (813)</td>
<td>35.79±7.69 (570)</td>
</tr>
<tr>
<td><strong>Serum Urate (mmol/L)</strong></td>
<td>0.336±0.087 (12483)*</td>
<td>0.399±0.136 (1118)</td>
<td>0.373±0.088 (680)</td>
<td>0.436±0.119 (484)</td>
</tr>
<tr>
<td><strong>Gout Duration (Years)</strong></td>
<td>-</td>
<td>15.19±12.56 (1289)</td>
<td>-</td>
<td>13.21±11.09 (548)</td>
</tr>
</tbody>
</table>

All values aside from sex are expressed as mean ± SD.

* For BMI, SU and gout duration, the figure inside the brackets represents the number of subjects with available data.

** Obtained at the time of gout onset among cases and at the mid-point time of cohort follow-up among controls.

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**Genotyping**

Taqman genotyping for rs2149356 was performed for the sample sets excepting ARIC, FHS, HPFS and NHS using a Lightcycler 480 Real-Time Polymerase Chain Reaction System (Roche Applied Science, Indianapolis, USA) in 384-well plates. The FHS cohort had been genotyped by the Affymetrix SNP 5 platform and a custom-designed gene-centric 50K SNP platform and
rs2149356 genotype was imputed using MACH1 v1.0.15 with the HapMap CEU sample set as reference haplotypes. In the ARIC sample set rs2149356 had been genotyped on the Affymetrix SNP 6 platform. The HPFS and NHS sample sets were genotyped using the Illumina Infinium OmniExpress and genotypes were imputed using MACH (imputation quality Rsq = 0.989). This resulted in N = 10807 non-missing genotype calls (1.5% missing). There was no evidence for departure from Hardy Weinberg equilibrium in any of the sample sets presented in Tables 2 and 3 (\(P_{\text{HWE}} > 0.01\)).

**Statistical analysis**

All logistic regression analysis was done using Intercooled STATA software version 8.0 (College Station, TX 77845, USA). Allelic and genotypic odds ratios for gout were calculated and all analyses were adjusted by age and sex, with the Polynesian analysis additionally adjusted by an estimate of Polynesian ancestry calculated as previously described in order to account for admixture with other ancestral groups [17]. A threshold of \(P < 0.05\) was used to declare nominal statistical significance.

**Results**

In European and Polynesian subjects, the T-allele of rs2149356 was associated with the risk of gout when compared to controls unstratified by urate level, but in an opposing direction of association (Table 2; OR\(_{\text{European}} = 1.12, P = 0.012\) and OR\(_{\text{Polynesian}} = 0.80, P = 0.011\)). The TT-genotype was also associated with risk of gout when compared to the GG genotype, again in an opposing direction of association (Table 1; OR\(_{\text{European}} = 1.32, P = 0.006\) and OR\(_{\text{Polynesian}} = 0.63, P = 0.009\)). Rs2149356 was then tested for association with gout in the HPFS and NHS studies (Table 3). There was no evidence for association in either the allelic or genotypic analyses in either of the studies (\(P > 0.31\)).

**Discussion**

Here we attempted to replicate the previously reported association with gout of TLR4 variant rs2149356 in Han Chinese in sample sets of European and Polynesian ancestry. In the NZ and Europeans recruited from Europe the association was replicated but with opposing direction in the Polynesian sample set. A similar opposing direction of association between European and Polynesian is also evident at other urate- and gout-associated loci PRKAG2 and HLF [2]. Why
this is the case is not resolved, discussed further below. The third sample set (HPFS and NHS) provided no evidence to support association of rs2149356 with gout. Two characteristics of this sample set would have reduced chances of detecting association, should genuine association exist. First, only ~90% have clinically-ascertained gout [16]. Second, the sample set are incident gout cases (with minimum age of male cases 40 years and female cases 30 years [16]). Prevalent cases are excluded—these excluded cases would be expected to have an earlier age of onset and a stronger heritable component. On balance we are unable to conclude that the TLR4 locus to be the first replicated genetic risk factor in gout outside of those that influence gout risk via modulation of serum urate levels. However, given the very strong prior functional candidacy of TLR4, we do consider that our study provides evidence consistent with a causal role for TLR4 in gout.

Regarding the inconsistent direction of association with gout between Polynesian, and Han Chinese and European, the simplest explanation is that rs2149356 is not the causal variant but in linkage disequilibrium with the causal variant, with a Polynesian-specific ancestral recombination event distinguishing the Polynesian haplotypic background around rs2149356 from European and Han Chinese, resulting in the other (G) allele of rs2149356 being on the Polynesian risk haplotype. The Polynesian population could be important in trans-ancestral genetic fine-mapping of the etiological variant in TLR4 with the variant expected to be in a genomic segment with the same alleles associated with gout between Han Chinese, European and Polynesian. Other possibilities are that this marker is subject to stratification effects that have not been adequately controlled in the Polynesian sample set, that there are non-genetic interactors (e.g. alcohol and sugar-sweetened beverage consumption [18–20]) that are unaccounted for and that would generate inconsistent rs2149356 main effects or that these are chance findings given the modest level of significance.

SNP rs2149356 is a common genetic variant of weak effect (OR≤1.4). More than 70% of genetic variants for common phenotypes identified by genome-wide association studies map to regulatory regions of the genome [21]. While it is possible that less common non-synonymous functional variants in TLR4, that have been associated with other auto-inflammatory conditions [22, 23], may also associate with gout, it is likely that the effect of rs2149356 on the risk of gout is via an influence on expression of TLR4, either the amount of TLR4 produced and/or relative levels of isoforms. Direct evidence of this was supplied by Qing et al. [12] who associated the TT-genotype rs2149356 with increased levels of TLR4 mRNA in peripheral blood mononuclear cells (PBMCs) and increased serum interleukin-1β levels in people with acute

Table 3. Rs2149356 genotype and association with risk of gout in the HPFS and NHS sample sets.

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Total</th>
<th>Gout</th>
<th>Unadjusted RR[95% CI]</th>
<th>P</th>
<th>Adjusted RR† [95% CI]</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>HPFS</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GG</td>
<td>1920</td>
<td>336</td>
<td>1.00</td>
<td>-</td>
<td>1.00</td>
<td>-</td>
</tr>
<tr>
<td>GT</td>
<td>1824</td>
<td>315</td>
<td>0.98 [0.84–1.14]</td>
<td>0.78</td>
<td>0.99 [0.84–1.15]</td>
<td>0.84</td>
</tr>
<tr>
<td>TT</td>
<td>427</td>
<td>75</td>
<td>1.03 [0.80–1.32]</td>
<td>0.83</td>
<td>1.05 [0.82–1.35]</td>
<td>0.70</td>
</tr>
<tr>
<td>T</td>
<td>2251</td>
<td>390</td>
<td>0.99 [0.85–1.14]</td>
<td>0.85</td>
<td>1.00 [0.86–1.15]</td>
<td>0.94</td>
</tr>
<tr>
<td><strong>NHS</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GG</td>
<td>3137</td>
<td>176</td>
<td>1.00</td>
<td>-</td>
<td>1.00</td>
<td>-</td>
</tr>
<tr>
<td>GT</td>
<td>2901</td>
<td>143</td>
<td>0.88 [0.70–1.10]</td>
<td>0.25</td>
<td>0.89 [0.71–1.11]</td>
<td>0.28</td>
</tr>
<tr>
<td>TT</td>
<td>630</td>
<td>32</td>
<td>0.93 [0.64–1.36]</td>
<td>0.71</td>
<td>0.97 [0.66–1.42]</td>
<td>0.87</td>
</tr>
<tr>
<td>T</td>
<td>3531</td>
<td>175</td>
<td>0.88 [0.72–1.09]</td>
<td>0.24</td>
<td>0.90 [0.73–1.11]</td>
<td>0.31</td>
</tr>
</tbody>
</table>

1 Adjusted by age and diuretic usage.

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gout. Conversely the TT-genotype associated with reduced levels of TLR4 mRNA in PBMCs from people with intercritical gout. These important findings are consistent with the increased risk observed for the TT-genotype for gout. Understanding how the T-allele controls expression of TLR4 will be important for improved molecular understanding of the pathogenic role of the TLR4-pathway in gout. Our data, with an opposing direction of association in Polynesian, suggest that rs2149356 is not causal but in linkage disequilibrium (LD) with, and a marker for, the causal variant. Rs2149356 maps to intron 4 of TLR4 but is in strong LD with two SNPs in the promoter of TLR4 (rs2737190, $r^2 = 0.92$ in Europeans and 0.97 in Asians, ‘-2570’ [24]; rs1927914, $r^2 = 0.93$ in Europeans and 0.97 in Asians, ‘-2026’ [24]). The gout risk allele (T) of rs2149356 would be present (co-inherited) on a haplotype with the minor allele of each of these promoter SNPs. The minor allele (G) of rs2737190 is predicted to create a v-Myb transcription factor binding site and the minor allele (G) of rs1927914 is predicted to remove Oct-1 and C/EBP transcription factor binding sites [24]. When studied individually in vitro, the rs2737190 G allele had no effect, but the rs1927914 G allele significantly decreased basal expression from the TLR4 promoter and increased response to challenge with a uropathogenic Escherichia coli strain [24], which is consistent with the observed expression pattern from Qing et al. [12]. This genetic variant is therefore a candidate functional etiological variant for gout.

In conclusion, TLR4 SNP rs2143956 is associated with gout risk in prevalent clinically-ascertained gout in Europeans, in a direction consistent with previously published results in Han Chinese [12]. However, with an opposite direction of association in Polynesians and no evidence for association in a non-clinically-ascertained incident gout cohort this variant should be analysed in other international gout genetic data sets.

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Author Contributions

Conceived and designed the experiments: HR CM TRM. Performed the experiments: HR CM RKT. Analyzed the data: HR CM LL EAS HKC. Contributed reagents/materials/analysis tools: LKS ND RD DK KW MS MJ TLJ LAJ TRR PLR AKT FL HKC AS. Wrote the paper: HR CM LKS ND RKT RD DK KW MS MJ TLJ LAJ TRR PLR AKT FL LL EAS HKC AS TRM.
The TLR4 Gene and Gout

References


