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Cross-Sectional Associations of Computed Tomography (CT)-Derived Adipose Tissue Density and Adipokines: The Framingham Heart Study

Jane J. Lee, PhD; Alison Pedley, PhD; Udo Hoffmann, MD, MPH; Joseph M. Massaro, PhD; John F. Keaney, Jr., MD; Ramachandran S. Vasan, MD; Caroline S. Fox, MD, MPH

Background—Excess accumulation of abdominal subcutaneous (SAT) and visceral adipose tissue (VAT) is associated with adverse levels of adipokines and cardiovascular disease risk. Whether fat quality is associated with adipokines has not been firmly established. This study examined the association between abdominal SAT and VAT density, an indirect measure of fat quality, with a panel of metabolic regulatory biomarkers secreted by adipose tissue or the liver independently of absolute fat volumes.

Methods and Results—We evaluated 1829 Framingham Heart Study participants (44.9% women). Abdominal SAT and VAT density was estimated indirectly by adipose tissue attenuation using computed tomography. Adipokines included adiponectin, leptin receptor, leptin, fatty acid-binding protein 4 (FABP-4), retinol-binding protein 4 (RBP-4), and fetuin-A. Fat density was associated with all the biomarkers evaluated, except fetuin-A. Lower fat density (ie, more-negative fat attenuation) was associated with lower adiponectin and leptin receptor, but higher leptin and FABP-4 levels (all \( P < 0.0001 \)). SAT density was inversely associated with RBP-4 in both sexes, whereas the association between VAT density and RBP-4 was only observed in men (\( P < 0.0001 \)). In women, after additional adjustment for respective fat volume, SAT density retained the significant associations with adiponectin, leptin, FABP-4, and RBP-4; and VAT density with adiponectin only (all \( P < 0.0001 \)). In men, significant associations were maintained upon additional adjustment for respective fat volume (\( P < 0.005 \)).

Conclusions—Lower abdominal fat density was associated with a profile of biomarkers suggestive of greater cardiometabolic risk. These observations support that fat density may be a valid biomarker of cardiometabolic risk. (J Am Heart Assoc. 2016;5: e002545 doi: 10.1161/JAHA.115.002545)

Key Words: adipokine • adipose tissue • computed tomography • epidemiology

Prevalence of abdominal obesity has increased in the United States and currently affects more than 54% of US adults. Excess accumulation of abdominal adiposity increases the risks of type 2 diabetes, hypertension, dyslipidemia, and metabolic syndrome. Abdominal adipose tissue including subcutaneous (SAT) and visceral adipose tissue (VAT) has been widely recognized as a pathogenic phenotype. In addition, recent studies suggested that qualitative aspects of adipose tissue, such as adipocyte size, macrophage infiltration, angiogenesis, hypoxia, and fibrosis, are also associated with metabolic and cardiovascular disease (CVD).

Computed tomography (CT) imaging techniques allow for the indirect quantification of abdominal fat quality by assessing the density of adipose tissue by radiographic pixels that are denoted in Hounsfield units (HU) and referred to as attenuation. Our previous study utilized abdominal SAT and VAT density as an indirect measure of fat quality to explore the association with CVD risk factors. In that study, lower fat density (ie, more-negative CT fat attenuation) was associated with adverse cardiometabolic risk, including higher blood pressure, insulin resistance, triglycerides, and lower high-density lipoprotein levels cross-sectionally.

To further promote the mechanistic understanding of fat quality, we sought to explore the cross-sectional associations between abdominal fat density with a panel of circulating...
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biodmarks mainly released by adipose tissue (adiponectin, leptin receptor, leptin, and fatty acid-binding protein 4 [FABP-4]) and by both adipose tissue and the liver (retinol-binding protein 4 [RBP-4] and fetuin-A). On the basis of our previous findings regarding the inverse association of abdominal adipose tissue density with cardiometabolic risk, we hypothesized that lower SAT and VAT density would be associated with adverse levels of adipokines, even after adjusting for generalized adiposity (body mass index [BMI]), central obesity (waist circumference), and respective abdominal adipose tissue volumes.

Methods

Study Sample

The Framingham Heart Study was initiated in 1948 as a community-based observational study to explore the association between CVDs and related risk factors with lifestyle, environment, and inheritance. The present study drew participants from the multidetector CT substudy of the Third Generation Framingham cohort who underwent imaging from 2002 to 2005. Detailed information of the Third Generation cohort of the Framingham Heart Study has been previously described elsewhere. For the present investigation, we included participants who had data on (1) abdominal adipose tissue volume and attenuation evaluated by multidetector CT; (2) circulating biomarker levels, including adiponectin, leptin receptor, leptin, FABP-4, RBP-4, and fetuin-A; and (3) covariates. Among 2117 participants from the initial recruitment of the multidetector CT cohort, 250 participants were excluded because of a lack of abdominal fat and biomarker values. An additional 38 participants were excluded because of missing covariates, resulting in a total of 1829 participants (86.4% of those eligible) in this investigation. The study protocol was approved by the institutional review boards of the Boston University Medical Center and Massachusetts General Hospital. All participants provided written informed consent.

Abdominal Subcutaneous and Visceral Adipose Tissue Volume and Density

A total of 25 consecutive slices of abdomen were obtained while participants lay in a supine position by an 8-slice multidetector CT scanner (LightSpeed Ultra; General Electric, Milwaukee, WI) with a thickness of 5 mm, tube voltage of 120 kVp, and radiation dose of 3 to 5 mSv. The assessment of abdominal adipose tissue quantity and density was performed by evaluating the CT slides with a three-dimensional (3D) workstation tool (Aquarius 3D Workstation; TeraRecon Inc, San Mateo, CA). The trained technicians designated the region of interest by manually outlining the abdominal muscular wall. Subsequently, the region of SAT and VAT compartments were automatically identified based on the radiographic pixel threshold between −195 and −45 HU with center attenuation of −120 HU. Mean SAT and VAT volumes in cm³ and attenuation in HU were recorded. Previously, high reproducibility of these abdominal CT measurements were confirmed, with inter- and intrareader reliability greater than 0.99.

Adipokines

The group of circulating biomarkers produced by adipose tissue only (adiponectin, leptin receptor, leptin, and FABP-4) and by both adipose tissue and the liver (RBP-4 and fetuin-A) were evaluated. Blood samples were collected after a minimum of 8 hours of overnight fasting and analyzed following standard protocols. Plasma levels of adiponectin, leptin receptor, leptin, and RBP-4 levels were determined by the ELISA method (R&D Systems, Minneapolis, MN) with mean interassay coefficients of variation of 2.23% for adiponectin, 4.01% for leptin receptor, 4.97% for leptin, and 2.18% for RBP-4. Plasma levels of FABP-4 and fetuin-A were assessed by the sandwich ELISA method (BioVendor Research and Diagnostic Products, Candler, NC) with mean interassay coefficients of variation of 2.38% for FABP-4 and 2.52% for fetuin-A.

Measurement of Covariates

BMI was computed as body weight in kilograms divided by height in meters squared. Waist circumference was assessed by a measuring tape at the horizontal level of the umbilicus to the closest 0.25 inch. A series of questionnaires were administered to document the clinical history and lifestyle patterns of the participants, including current use of hormone replacement therapy, current smoking, alcohol use, and physical activity level. Current smoking was specified as those who smoked at least 1 cigarette per day within the previous year. Participants were considered moderate-to-heavy drinkers based on the consumption of >7 drinks/week for women and >14 drinks/week for men. Physical activity was evaluated using a questionnaire-derived physical activity score, which incorporated the time individuals participated in different levels of physical activity, taking into account the required oxygen consumption for each of the activities.

Statistical Analysis

We conducted a sex-specific analysis because of the differences in women and men regarding fat distribution and the circulating biomarker levels. All the adipokines were
natural logarithmically transformed to normalize their skewed distributions. Age-adjusted partial Pearson correlation coefficients were computed to examine the association between abdominal CT fat density and natural log-transformed biomarkers. Multivariable-adjusted linear regressions were performed to assess the association between fat density (independent variable) and each of the natural log-transformed biomarkers (dependent variable) with a separate model performed for each association tested. Multivariable adjustment included age, hormone replacement therapy (women only), current smoking, alcohol use, and physical activity score. We also examined whether the associations were independent of measures of obesity by additionally adjusting for BMI, waist circumference, or respective fat volumes. The β-coefficients computed in these models describe the estimated association in the natural log-transformed biomarkers for a 5-unit (±1-SD) decrement in fat density.

Tests for sex interaction were also conducted using multivariable-adjusted linear regression. As a secondary analysis, the multivariable-adjusted least-square means for each of the biomarker levels by sex-specific tertiles of CT fat volume within tertiles of CT fat density were examined. A 2-tailed P value less than 0.05 was considered statistically significant. We did not further adjust for multiple testing because the purpose of this investigation was principal hypothesis generating (ie, to identify potential associations between abdominal fat density and a panel of adipokines). All statistical analysis was performed using SAS software (version 9.2; SAS Institute Inc., Cary, NC).

### Results

#### Descriptive Characteristics

Clinical, adiposity, biomarker, and lifestyle characteristics of the 821 women and 1008 men (overall mean age, 45 years) included in the study are shown in Table 1. Median SAT and VAT attenuations were −103.3 and −91.5 HU in women and −100.7 and −91.5 HU in men, respectively.

#### Age-Adjusted Correlations With Abdominal Fat Density and Adipokines

Age-adjusted, sex-stratified Pearson correlation coefficients between abdominal fat density and natural log-transformed biomarkers are given in Table 2. Higher (ie, more-positive CT fat attenuation) SAT and VAT density was correlated with higher levels of adiponectin and leptin receptor (r values

**Table 1. Characteristics of Study Population**

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Women (n=821)</th>
<th>Men (n=1008)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y</td>
<td>46.1 (5.7)</td>
<td>44.1 (6.3)</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>26.5 (6.0)</td>
<td>28.1 (4.4)</td>
</tr>
<tr>
<td>Waist circumference, cm</td>
<td>90.3 (15.5)</td>
<td>99.4 (11.8)</td>
</tr>
<tr>
<td>Subcutaneous adipose tissue, cm³</td>
<td>2985 (1565)</td>
<td>2588 (1226)</td>
</tr>
<tr>
<td>Visceral adipose tissue, cm³</td>
<td>1127 (729)</td>
<td>1986 (870)</td>
</tr>
<tr>
<td>Subcutaneous adipose tissue, HU</td>
<td>−101.9 (5.5)</td>
<td>−99.9 (4.5)</td>
</tr>
<tr>
<td>Visceral adipose tissue, HU</td>
<td>−92.0 (4.3)</td>
<td>−95.5 (4.5)</td>
</tr>
<tr>
<td>Adiponectin*, μg/mL</td>
<td>10.3 (6.6, 15.5)</td>
<td>5.1 (3.4, 8.0)</td>
</tr>
<tr>
<td>Leptin receptor*, ng/mL</td>
<td>18.2 (11.8, 24.8)</td>
<td>17.8 (11.7, 23.5)</td>
</tr>
<tr>
<td>Leptin*, ng/mL</td>
<td>13.5 (7.1, 26.6)</td>
<td>4.5 (2.7, 7.8)</td>
</tr>
<tr>
<td>FABP-4*, ng/mL</td>
<td>0.02 (0.01, 0.03)</td>
<td>0.02 (0.01, 0.02)</td>
</tr>
<tr>
<td>RBP-4*, μg/mL</td>
<td>37.2 (30.8, 44.6)</td>
<td>43.4 (37.2, 50.5)</td>
</tr>
<tr>
<td>Fetuin-A*, mg/L</td>
<td>418.8 (314.7, 541.9)</td>
<td>398.9 (312.8, 509.4)</td>
</tr>
<tr>
<td>Current hormone replacement therapy, %</td>
<td>9.0% (74)</td>
<td>N/A</td>
</tr>
<tr>
<td>Postmenopausal, %</td>
<td>24.7% (203)</td>
<td>N/A</td>
</tr>
<tr>
<td>Current smoking¹, %</td>
<td>14.1% (116)</td>
<td>14.8% (149)</td>
</tr>
<tr>
<td>Moderate to heavy alcohol use², %</td>
<td>14.4% (118)</td>
<td>16.0% (161)</td>
</tr>
<tr>
<td>Physical activity score</td>
<td>36.4 (6.1)</td>
<td>38.1 (8.9)</td>
</tr>
</tbody>
</table>

Data on means (SDs) or proportions (counts) are shown. FABP-4, fatty acid-binding protein 4; RBP-4, retinol-binding protein 4; HU, Hounsfield unit.

*Values are shown as medians (25th, 75th percentiles) because of the skewed distribution.

¹Defined as ≥1 cigarette per day within the previous year.

²Defined as >7 drinks per week (women) or >14 drinks per week (men).
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P was correlated with lower RBP-4 (DOI: 10.1161/JAHA.115.002545 Journal of the American Heart Association but higher leptin and FABP-4 levels in both sexes (all associated with lower adiponectin and leptin receptor levels, after covariate adjustment. Lower SAT and VAT density was ranged from 0.18 to 0.41) and lower levels of leptin and FABP-4 (r values ranged from −0.69 to −0.44) in both sexes (all P<0.001). In women, higher SAT density, but not VAT density, was correlated with lower RBP-4 (r=−0.18; P<0.001). In men, higher SAT and VAT density were correlated with lower RBP-4 (r=−0.14 for SAT, r=−0.18 for VAT; both P<0.001). In both women and men, higher VAT density, but not SAT density, was weakly correlated with a lower level of fetuin-A (both sexes r=−0.07; P<0.05). In the Context of the Current Literature

Multivariable-Adjusted Regression Models for Abdominal Fat Density and Adipokines

The association of natural log-transformed biomarkers per 5-HU (≈1-SD) decrement in fat density is shown in Table 3 after covariate adjustment. Lower SAT and VAT density was associated with lower adiponectin and leptin receptor levels, but higher leptin and FABP-4 levels in both sexes (all P<0.0001; Table 3). Lower SAT density was associated with higher RBP-4 levels in both sexes; whereas lower VAT density was associated with higher RBP-4 in men only (all P<0.0001).

Next, the regression models were further adjusted for BMI, waist circumference, or respective abdominal fat volumes (SAT volume for SAT density model and VAT volume for VAT density model; Table 3). When we additionally adjusted for BMI or waist circumference, the association with fat density generally decreased, but remained statistically significant with adiponectin, leptin, and FABP-4 in women; and with adiponectin, leptin receptor, leptin, FABP-4, and RBP-4 in men (all P<0.05). In women, after additional adjustment for the respective abdominal fat volume, SAT density maintained the associations with most of the biomarkers, as compared to VAT density. In men, further adjustment for the respective CT fat volume weakened the relationships between fat density and metabolic biomarkers; however, all of these associations remained statistically significant (all P<0.005).

Secondary Analyses

We further tested for sex interactions between abdominal fat density and the circulating biomarkers with the respective abdominal fat volumes. The multivariable-adjusted means of each biomarker are presented in Figure and Figure S1 according to the sex-specific tertiles of SAT and VAT volume by tertiles of SAT and VAT density. In general, higher tertiles of SAT and VAT HU (ie, lower fat density), within greater tertiles of respective fat volume (ie, greater fat volume), were associated with relatively lower levels of adiponectin and leptin receptor and higher levels of leptin, FABP-4, and RBP-4. However, for women with the lowest adipose tissue volume (SAT and VAT volume tertile 1), the inclusion of adipose tissue volume changed the direction of the association of fat density with leptin, leptin receptor, and RBP-4 (Figure and Figure S1).

Discussion

Principle Findings

In this cross-sectional, community-based observational study, we related CT-derived abdominal adipose tissue attenuation with a panel of circulating biomarkers released by adipose tissue or liver. Lower abdominal fat density was associated with adverse levels of circulating biomarkers; in particular, lower adiponectin and leptin receptor levels and higher leptin, FABP-4, and RBP-4 levels in both women and men. A majority of these associations remained statistically significant even after additional adjustment for generalized adiposity (BMI), central obesity (waist circumference), or respective abdominal adipose tissue volumes. Moreover, lower adipose tissue density with greater adipose tissue volumes was associated with adverse levels of circulating biomarkers consistent with greater cardiometabolic risk.

In the Context of the Current Literature

Greater accumulation of abdominal adipose tissue volumes is associated with adverse concentrations of the circulating

Table 2. Age-Adjusted Sex-Specific Pearson Correlation Coefficients Between Abdominal Fat Density and Natural Log-Transformed Adipokines

<table>
<thead>
<tr>
<th>In (Adipokines)</th>
<th>Women</th>
<th>Men</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>SAT Density</td>
<td>VAT Density</td>
</tr>
<tr>
<td>Adiponectin</td>
<td>0.30†</td>
<td>0.41†</td>
</tr>
<tr>
<td>Leptin receptor</td>
<td>0.18†</td>
<td>0.21†</td>
</tr>
<tr>
<td>Leptin</td>
<td>−0.64†</td>
<td>−0.50†</td>
</tr>
<tr>
<td>FABP-4</td>
<td>−0.44†</td>
<td>−0.48†</td>
</tr>
<tr>
<td>RBP-4</td>
<td>−0.18†</td>
<td>−0.02</td>
</tr>
<tr>
<td>Fetuin-A</td>
<td>−0.06</td>
<td>−0.07†</td>
</tr>
</tbody>
</table>

FABP-4, fatty acid-binding protein 4; RBP-4, retinol-binding protein 4; SAT, subcutaneous adipose tissue; VAT, visceral adipose tissue. †P<0.05; †P<0.001.
<table>
<thead>
<tr>
<th>Adiponectin</th>
<th>Women SAT Density</th>
<th>P Value</th>
<th>VAT Density</th>
<th>P Value</th>
<th>Men SAT Density</th>
<th>P Value</th>
<th>VAT Density</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>MV</td>
<td>-0.17 (-0.20, -0.13)</td>
<td>&lt;0.0001</td>
<td>-0.28 (-0.32, -0.24)</td>
<td>&lt;0.0001</td>
<td>-0.12 (-0.17, 0.08)</td>
<td>&lt;0.0001</td>
<td>-0.29 (-0.33, -0.25)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+BMI</td>
<td>-0.11 (-0.15, -0.08)</td>
<td>&lt;0.0001</td>
<td>-0.22 (-0.27, -0.16)</td>
<td>&lt;0.0001</td>
<td>-0.05 (-0.10, -0.01)</td>
<td>0.03</td>
<td>-0.26 (-0.31, -0.21)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+WC</td>
<td>-0.11 (-0.15, -0.07)</td>
<td>&lt;0.0001</td>
<td>-0.21 (-0.26, -0.16)</td>
<td>&lt;0.0001</td>
<td>-0.05 (-0.10, 0.00)</td>
<td>0.03</td>
<td>-0.28 (-0.32, -0.23)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+Fat volume†</td>
<td>-0.11 (-0.15, -0.07)</td>
<td>&lt;0.0001</td>
<td>-0.13 (-0.19, -0.07)</td>
<td>&lt;0.0001</td>
<td>-0.10 (-0.16, -0.05)</td>
<td>0.0001</td>
<td>-0.25 (-0.31, -0.19)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Leptin receptor</td>
<td>MV</td>
<td>-0.07 (-0.10, -0.04)</td>
<td>&lt;0.0001</td>
<td>-0.10 (-0.13, -0.06)</td>
<td>&lt;0.0001</td>
<td>-0.12 (-0.15, -0.09)</td>
<td>&lt;0.0001</td>
<td>-0.12 (-0.15, -0.09)</td>
</tr>
<tr>
<td>MV+BMI</td>
<td>-0.04 (-0.07, -0.01)</td>
<td>0.02</td>
<td>-0.04 (-0.08, 0.00)</td>
<td>0.06</td>
<td>-0.10 (-0.13, -0.06)</td>
<td>&lt;0.0001</td>
<td>-0.09 (-0.13, -0.06)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+WC</td>
<td>-0.04 (-0.07, -0.01)</td>
<td>0.02</td>
<td>-0.05 (-0.09, -0.01)</td>
<td>0.02</td>
<td>-0.09 (-0.13, -0.06)</td>
<td>&lt;0.0001</td>
<td>-0.09 (-0.12, -0.05)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+Fat volume†</td>
<td>-0.03 (-0.06, 0.00)</td>
<td>0.07</td>
<td>-0.03 (-0.08, 0.02)</td>
<td>0.28</td>
<td>-0.10 (-0.13, -0.06)</td>
<td>&lt;0.0001</td>
<td>-0.09 (-0.13, -0.04)</td>
<td>0.0005</td>
</tr>
<tr>
<td>Leptin</td>
<td>MV</td>
<td>0.53 (0.48, 0.57)</td>
<td>&lt;0.0001</td>
<td>0.52 (0.46, 0.58)</td>
<td>&lt;0.0001</td>
<td>0.63 (0.59, 0.67)</td>
<td>&lt;0.0001</td>
<td>0.54 (0.50, 0.59)</td>
</tr>
<tr>
<td>MV+BMI</td>
<td>0.34 (0.30, 0.37)</td>
<td>&lt;0.0001</td>
<td>0.12 (0.07, 0.18)</td>
<td>&lt;0.0001</td>
<td>0.42 (0.39, 0.45)</td>
<td>&lt;0.0001</td>
<td>0.28 (0.23, 0.32)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+WC</td>
<td>0.32 (0.29, 0.36)</td>
<td>&lt;0.0001</td>
<td>0.14 (0.08, 0.20)</td>
<td>&lt;0.0001</td>
<td>0.37 (0.34, 0.41)</td>
<td>&lt;0.0001</td>
<td>0.24 (0.20, 0.28)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+Fat volume†</td>
<td>0.25 (0.22, 0.29)</td>
<td>&lt;0.0001</td>
<td>-0.04 (-0.11, 0.04)</td>
<td>0.34</td>
<td>0.32 (0.28, 0.36)</td>
<td>&lt;0.0001</td>
<td>0.15 (0.09, 0.22)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>FABP-4</td>
<td>MV</td>
<td>0.19 (0.17, 0.22)</td>
<td>&lt;0.0001</td>
<td>0.27 (0.23, 0.30)</td>
<td>&lt;0.0001</td>
<td>0.23 (0.20, 0.26)</td>
<td>&lt;0.0001</td>
<td>0.24 (0.21, 0.27)</td>
</tr>
<tr>
<td>MV+BMI</td>
<td>0.10 (0.08, 0.13)</td>
<td>&lt;0.0001</td>
<td>0.10 (0.07, 0.14)</td>
<td>&lt;0.0001</td>
<td>0.13 (0.11, 0.16)</td>
<td>&lt;0.0001</td>
<td>0.14 (0.11, 0.16)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+WC</td>
<td>0.09 (0.07, 0.12)</td>
<td>&lt;0.0001</td>
<td>0.11 (0.07, 0.14)</td>
<td>&lt;0.0001</td>
<td>0.11 (0.08, 0.14)</td>
<td>&lt;0.0001</td>
<td>0.12 (0.09, 0.15)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+Fat volume†</td>
<td>0.07 (0.04, 0.09)</td>
<td>&lt;0.0001</td>
<td>0.00 (-0.04, 0.05)</td>
<td>0.84</td>
<td>0.10 (0.07, 0.13)</td>
<td>&lt;0.0001</td>
<td>0.06 (0.02, 0.10)</td>
<td>0.004</td>
</tr>
<tr>
<td>RBP-4</td>
<td>MV</td>
<td>0.05 (0.03, 0.06)</td>
<td>&lt;0.0001</td>
<td>0.02 (0.00, 0.04)</td>
<td>0.09</td>
<td>0.04 (0.02, 0.05)</td>
<td>&lt;0.0001</td>
<td>0.05 (0.03, 0.06)</td>
</tr>
<tr>
<td>MV+BMI</td>
<td>0.05 (0.03, 0.06)</td>
<td>&lt;0.0001</td>
<td>0.01 (-0.01, 0.03)</td>
<td>0.45</td>
<td>0.05 (0.03, 0.06)</td>
<td>&lt;0.0001</td>
<td>0.06 (0.04, 0.08)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+WC</td>
<td>0.04 (0.03, 0.06)</td>
<td>&lt;0.0001</td>
<td>0.00 (-0.02, 0.03)</td>
<td>0.82</td>
<td>0.05 (0.03, 0.06)</td>
<td>&lt;0.0001</td>
<td>0.06 (0.04, 0.08)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>MV+Fat volume†</td>
<td>0.05 (0.03, 0.07)</td>
<td>&lt;0.0001</td>
<td>-0.01 (-0.04, 0.02)</td>
<td>0.51</td>
<td>0.06 (0.04, 0.08)</td>
<td>&lt;0.0001</td>
<td>0.06 (0.03, 0.08)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Fetuin-A</td>
<td>MV</td>
<td>0.02 (-0.00, 0.05)</td>
<td>0.12</td>
<td>0.03 (0.00, 0.07)</td>
<td>0.04</td>
<td>0.01 (-0.02, 0.03)</td>
<td>0.68</td>
<td>0.03 (0.00, 0.06)</td>
</tr>
<tr>
<td>MV+BMI</td>
<td>0.00 (-0.02, 0.03)</td>
<td>0.80</td>
<td>0.00 (-0.03, 0.04)</td>
<td>0.83</td>
<td>-0.01 (-0.04, 0.02)</td>
<td>0.49</td>
<td>0.02 (-0.01, 0.05)</td>
<td>0.21</td>
</tr>
<tr>
<td>MV+WC</td>
<td>0.00 (-0.03, 0.03)</td>
<td>0.87</td>
<td>0.01 (-0.03, 0.05)</td>
<td>0.71</td>
<td>-0.02 (-0.05, 0.01)</td>
<td>0.26</td>
<td>0.02 (-0.01, 0.04)</td>
<td>0.31</td>
</tr>
<tr>
<td>MV+Fat volume†</td>
<td>0.00 (-0.03, 0.03)</td>
<td>0.97</td>
<td>-0.01 (-0.05, 0.04)</td>
<td>0.77</td>
<td>-0.01 (-0.04, 0.02)</td>
<td>0.54</td>
<td>0.00 (-0.04, 0.04)</td>
<td>0.95</td>
</tr>
</tbody>
</table>

Data are shown as estimated β-coefficient (95% confidence intervals). The values of estimated β-coefficient describe the association of natural log-transformed biomarkers for a 1 SD decrement (5 HU) in CT fat density. Sex interaction was tested based on the multivariable model. Significant sex interactions were identified between SAT density with leptin receptor (P=0.02) and leptin (P=0.001); and between VAT density and RBP-4 (P=0.03) only. BMI, body mass index; CT, computed tomography; FABP-4, fatty acid-binding protein 4; HU, Hounsfield unit; MV, multivariable; RBP-4, retinol-binding protein 4; SAT, subcutaneous adipose tissue; VAT, visceral adipose tissue; WC, waist circumference.

† Multivariable model (MV) adjusted for age, hormone replacement therapy (women only), current smoking, alcohol use, and physical activity score.

‡ Regression model for SAT density is adjusted for SAT volume; regression model for VAT density is adjusted for VAT volume.
biomarkers of metabolic regulation.\textsuperscript{18–20} Yet, the quality of abdominal adipose tissue assessed by radiographic imaging techniques is relatively new, because only a few human-based studies have been reported in recent years.\textsuperscript{13,17,21,22} To our knowledge, only 1 previous study explored the associations between SAT and VAT HU with a panel of adipose tissue-derived biomarkers, referred to as adipokines, in an older population (ages 65 and older) and in nonhuman primates.\textsuperscript{17} In that report, lower attenuation of CT fat was associated with larger adipocyte size, as well as lower adiponectin and higher leptin levels.\textsuperscript{17} Our study observed similar findings, and we extend the literature to a more-comprehensive panel of circulating biomarkers (including leptin receptor, FABP-4, and RBP-4) and a broader age range of individuals. In our study, a few significant sex interactions between abdominal fat density and metabolic biomarkers were detected. The most distinctive sex interactions were identified between SAT density with leptin receptors and leptin, where the associations were more pronounced in men.

Previous studies have explored differences in adipogenesis between women and men in the aspects of adipose tissue cellularity.\textsuperscript{23–25} Progression to obesity was associated with adipocyte hypertrophy in both women and men; and adipocyte hyperplasia only in women.\textsuperscript{23} These findings support the sexual dimorphism in the process of adipose tissue remodeling. In addition, the size of the adipocyte derived from SAT and VAT was larger in men than women, even after adjusting for BMI.\textsuperscript{26} In our study, CT attenuation of adipose tissue was implicated as a surrogate measure of fat quality because lower attenuation of adipose tissue assessed by CT corresponded to larger adipocyte size\textsuperscript{17} with highly vascularized tissue.\textsuperscript{27} Of note, larger adipocytes were associated with higher expression of leptin mRNA,\textsuperscript{26} which may explain our findings of a more pronounced association between fat density and leptin in men, as opposed to women. However, further studies are necessary to elucidate the differences in the mechanism in women and men linked with adipose tissue quality and a broad array of adipokines.

**Potential Physiological Mechanisms**

Consistent with our a priori hypothesis, lower abdominal fat density was associated with adverse concentrations of circulating biomarkers, although the underlying mechanisms,
particularly the causal association between abdominal fat density and the panel of biomarkers, remain unclear. There are several potential mechanistic explanations for our findings. One potential explanation is that lower attenuation of CT fat is an indicator of adipose tissue with dense lipid content consisting of large adipocytes that are filled with enlarged lipid droplets. Enlarged adipocytes may reflect the insufficient proliferation of adipocytes attributed to impaired adipogenesis in the state of obesity. Larger adipocyte size has been related to increased metabolic activity and greater secretion of adipocytokines, including adiponectin and leptin. Insulin-stimulated glucose uptake and protein expressions that are essential for lipid, fatty acid, and glucose metabolism appear to differ between small and large adipocytes. Collectively, dysfunctional secretion of adipokines induced by hypertrophied adipocytes may provide insight into the pathophysiological associations between hypertrophied adipocyte and adverse cardiometabolic risk factors.

Second, the low attenuation of CT fat may reflect adipose tissue that is not affected by adipose tissue fibrosis, which allows the extracellular matrix remodeling of adipose tissues to accommodate adipocyte expansion. This notion can be supported by the association between elevated urinary connective tissue growth factor (ie, a marker of systemic fibrosis) and higher CT fat attenuation. In line with this, a dose-dependent reduction of collagen type VI α3 gene expression (ie, a gene that encodes fibrotic extracellular matrix protein) by leptin support the regulatory effect of leptin on cellular fibrosis. Taken together, the higher secretion of leptin may suppress adipose tissue fibrosis and subsequently contribute to expandability of adipocytes.

Third, fat quality may be related to metabolic regulatory biomarkers by systemic inflammation. Obesity is associated with a chronic, low-grade inflammatory response with up-regulation of proinflammatory adipokines that promote systemic inflammation, such as leptin, FABP-4, RBP-4, and fetuin-A; and down-regulation of anti-inflammatory adipokines that reduce cellular inflammation, such as adiponectin. It is plausible that altered adipose tissue quality, in particular, adipocyte hypertrophy manifested by lipid overaccumulation, may trigger abnormal production of pro- and anti-inflammatory adipokines and subsequently leads to a generalized inflammatory state.

In addition, there are other potential mechanisms that may elaborate our findings. Lower attenuation of CT fat as a marker of poor vascularity and cellular hypoxia may explain the association between fat quality and adverse levels of adipokines. More specifically, cellular hypoxia developed along with adipocyte hypertrophy may be one of the key factors given that secretion of adipose tissue-derived biomarkers is considerably
modified under hypoxic conditions. Importantly, reduced vascularity is accompanied by more adipose tissue hypoxia as a consequence of the progression to obesity. Taken globally, these aforementioned findings add to the growing body of the literature for the potent mechanistic explanations between fat quality and adipokines.

Implications

Our findings suggest that abdominal SAT and VAT density, an indirect measure of abdominal fat quality, may be a potential indicator of cardiometabolic risk associated with metabolic regulatory biomarkers. Accordingly, our data raise the possibility of using abdominal adipose tissue quality assessed by CT as an indicator to identify individuals at high risk for developing cardiometabolic disease. Further studies are necessary to explore the causal association between fat density and levels of circulating biomarkers, as well as the quality of abdominal adipose tissue quality has a pathogenic impact on metabolism through its relationship with metabolic regulatory biomarkers.

Strengths and Limitations

The strengths of this study include the use of a highly reproducible and noninvasive measurement of abdominal CT fat quality in a large, community-based sample. Limitations include the cross-sectional and observational design that precludes the temporal and causal inferences between abdominal adipose tissue density and circulating adipokine levels. The majority of the participants were white; our findings cannot be generalized to other ethnicities. Serum fatty acids and triacylglycerol were not measured. Finally, we have used fat attenuation assessed by CT as a proxy for fat quality. Mechanistic studies are necessary to better understand the underlying cellular and histological characteristics of this novel imaging measurement.

Conclusions

Low abdominal SAT and VAT density is associated with adverse levels of circulating adipokines, suggestive of greater cardiometabolic risk independent of generalized adiposity (BMI), central obesity (waist circumference), and respective abdominal SAT and VAT quantity.

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Disclosures

Alison Pedley is an employee of Merck & Company, Inc.

References

Abdominal Fat Density and Adipokines  

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