Emerging nanotechnology approaches for HIV/AIDS treatment and prevention

Tewodros Mamø1,2, E Ashley Moseman3, Nagesh Kolishetti1,4, Carolina Salvador-Morales1,4, Jinjun Shi1,4, Daniel R Kuritzkes5, Robert Langer4,6, Ulrich von Andrian3, and Omid C Farokhzad1,4

Omid C Farokhzad: ofarokhzad@zeus.bwh.harvard.edu

1 Laboratory of Nanomedicine and Biomaterials, Brigham and Women’s Hospital, 75 Francis Street, Boston, MA, 02115, USA
2 Nanotechnology Engineering, University of Waterloo, Canada
3 Department of Pathology, Harvard Medical School, USA
4 Harvard-MIT Center for Cancer Nanotechnology Excellence, Massachusetts Institute of Technology, MA, USA
5 Section for Retroviral Therapeutics, Brigham and Women’s Hospital, Harvard Medical School, USA
6 Department of Chemical Engineering, Massachusetts Institute of Technology, USA

Abstract

Currently, there is no cure and no preventive vaccine for HIV/AIDS. Combination antiretroviral therapy has dramatically improved treatment, but it has to be taken for a lifetime, has major side effects and is ineffective in patients in whom the virus develops resistance. Nanotechnology is an emerging multidisciplinary field that is revolutionizing medicine in the 21st century. It has a vast potential to radically advance the treatment and prevention of HIV/AIDS. In this review, we discuss the challenges with the current treatment of the disease and shed light on the remarkable potential of nanotechnology to provide more effective treatment and prevention for HIV/AIDS by advancing antiretroviral therapy, gene therapy, immunotherapy, vaccinology and microbicides.

Keywords

AIDS; antiretroviral therapy; gene therapy; HIV; immunotherapy; microbicides; nanomedicine; nanoparticles; nanotechnology; vaccines

The emergence of AIDS was first reported in 1981 followed by the identification of HIV as the cause of the disease in 1983 [1–4]. HIV/AIDS is now a global pandemic that has become...
the leading infectious killer of adults worldwide [5]. By 2006, more than 65 million people had been infected with the HIV virus worldwide and 25 million had died of AIDS [6]. At the end of 2007, around 33 million people were living with the virus, with 2.7 million new infections and 2 million deaths each year [7]. This has caused tremendous social and economic damage worldwide, with developing countries, particularly Sub-Saharan Africa, heavily affected.

A cure for HIV/AIDS has been elusive in almost 30 years of research. Early treatments focused on antiretroviral drugs that were effective only to a certain degree. The first drug, zidovudine, was approved by the US FDA in 1987, leading to the approval of a total of 25 drugs to date, many of which are also available in fixed-dose combinations and generic formulations for use in resource-limited settings (to date, only zidovudine and didanosine are available as true generics in the USA) [8,9]. However, it was the advent of a class of drugs known as protease inhibitors and the introduction of triple-drug therapy in the mid-1990s that revolutionized HIV/AIDS treatment [10,11]. This launched the era of highly active antiretroviral therapy (HAART), where a combination of three or more different classes of drugs are administered simultaneously [11]. The use of the HAART regimen, particularly in the developed world, has resulted in tremendous success in improving the expectancy and quality of lives for patients [12]. However, some HAART regimens have serious side effects and, in all cases, HAART has to be taken for a lifetime, with daily dosing of one or more pills. Some patients also develop resistance to certain combinations of drugs, resulting in failure of the treatment. The absence of complete cure under current treatment underscores the great need for continued efforts in seeking innovative approaches for treatment of HIV/AIDS.

In addition to treatment, the best way to fight global infections is through preventive strategies, vaccines being the most effective agents. Vaccines have historically been very effective at controlling other major infectious diseases such as measles, mumps, rubella and polio, with smallpox completely eradicated. There have been enormous efforts to develop a safe and effective vaccine for HIV/AIDS. However, the pursuit has been very daunting so far, with recent failures of clinical trials for major candidate vaccines [13–15]. This has raised a debate over which path to take in HIV/AIDS vaccine research. Despite this debate, it is clear that novel approaches for identifying new antigens and adjuvants as well as better delivery systems are necessary. Another preventive strategy that has been under investigation is the development of effective intravaginal microbicides that can be used by women. There has been remarkable progress in the understanding and design of technologies for microbicide development. However, recent clinical trials failed to show efficacy, indicating the need for more research and development to design better systems [16,17].

Nanotechnology is a new discipline of science and engineering that is advancing many areas of medicine. It involves the understanding, design, engineering and fabrication of materials at the atomic and molecular level. The National Nanotechnology Initiative defines nanotechnology as the study of structures with roughly 1–100 nm in size in at least one dimension but structures up to several hundred nanometers are also considered under nanotechnology applications [18]. The application of nanotechnology to medicine, commonly referred to as nanomedicine, involves the use of nanoscale materials for preventive, therapeutic and diagnostic purposes [19]. There have been major advances in nanomedicine over the last few decades, particularly in cancer diagnosis and therapy [20–22]. Although at an earlier stage, applications of nanotechnology for prevention and treatment of HIV/AIDS have also gained attention in recent years. There are emerging novel approaches in which nanotechnology can enhance current treatment as well as advance new therapeutic strategies, such as gene therapy and immunotherapy. Moreover, some nanomaterials have therapeutic effects by themselves. Nanotechnology can also play a major role in preventive strategies for developing vaccines and microbicides. In this review, we discuss the potential of nanotechnology in improving the
current treatment, advancing new therapeutic strategies as well as providing alternatives in the quest for vaccine and microbicide developments for HIV/AIDS.

**Current HIV/AIDS treatment**

The current state-of-the-art treatment modality for HIV/AIDS is HAART, where three or more antiretroviral drugs are given to patients simultaneously. The drugs used in combination are in most cases from different classes that work based on different mechanisms. Despite the remarkable successes with the current HAART treatment for HIV/AIDS, there are still various challenges remaining. The major difficulty has been the failure of the treatment, typically due to poor patient compliance [23]. Due to the need to take the medication daily for a lifetime, patients fail to adhere to the treatment schedule, leading to ineffective drug levels in the body and rebound of viral replication [11,24,25].

Moreover, in some patients, the virus develops resistance to particular combinations of drugs even with good adherence. Drug resistance is mainly caused by the high genetic diversity of HIV-1 and the continuous mutation it undergoes [26]. This problem is being addressed with individualized therapy, whereby resistance testing is performed to select a combination of drugs that is most effective for each patient [26]. In addition, side effects due to toxicities of the drugs are also a concern. There are reports that patients taking HAART experience increased rates of heart disease, diabetes, liver disease, cancer and accelerated aging [11]. Most experts agree that these effects could be due to the HIV infection itself or co-infection with another virus, such as co-infection with hepatitis C virus resulting in liver disease. However, the toxicities resulting from the drugs used in HAART could also contribute to these effects.

Under current treatment, complete eradication of the virus from the body has not been possible. The major cause for this is that the virus resides in ‘latent reservoirs’ within memory CD4+ T cells and cells of the macrophage–monocyte lineage [11,25]. A major study recently found that, in addition to acting as latent reservoirs, macrophages significantly contribute to the generation of elusive mutant viral genotypes by serving as the host for viral genetic recombination [27]. The cells that harbor latent HIV are typically concentrated in specific anatomic sites, such as secondary lymphoid tissue, testes, liver, kidney, lungs, gut and the CNS [11,28–31]. The eradication of the virus from such reservoirs is critical to the effective long-term treatment of HIV/AIDS patients. Therefore, there is a great need to explore new approaches for developing nontoxic, lower-dosage treatment modalities that provide more sustained dosing coverage and effectively eradicate the virus from the reservoirs, avoiding the need for lifetime treatments.

**Nanotechnology for HIV/AIDS treatment**

**Nanotechnology for antiretroviral drug delivery**

The use of nanotechnology platforms for delivery of drugs is revolutionizing medicine in many areas of disease treatment [32]. Cancer patients have been the biggest beneficiaries of this revolution so far, with significant advances in the last few decades. Many nanoscale systems for systemic cancer therapy are either FDA approved or in clinical trials [19,33]. This tremendous success has been due to the unique features that nanotechnology imparts on drug delivery systems. Using nanotechnology, it has become possible to achieve improved delivery of poorly water-soluble drugs, targeted delivery of drugs to specific cells or tissues and intracellular delivery of macromolecules [18,32].

Nanotechnology-based platforms for systemic delivery of antiretroviral drugs could have similar advantages. Controlled-release delivery systems can enhance their half-lives, keeping them in circulation at therapeutic concentrations for longer periods of time. This could have
major implications in improving adherence to the drugs. Nanoscale delivery systems also enhance and modulate the distribution of hydrophobic and hydrophilic drugs into and within different tissues due to their small size. This particular feature of nanoscale delivery systems appears to hold the most promise for their use in clinical treatment and prevention of HIV. Specifically, targeted delivery of antiretroviral drugs to CD4+ T cells and macrophages as well as delivery to the brain and other organ systems could ensure that drugs reach latent reservoirs [29,34]. Moreover, by controlling the release profiles of the delivery systems, drugs could be released over a longer time and at higher effective doses to the specific targets. Various nanoscale drug delivery systems shown in Figure 1 could be explored for these purposes. The use of nanotechnology systems for delivery of antiretroviral drugs has been extensively reviewed by Nowacek et al. and Amiji et al. [29,31,34]. In this section, we only highlight a few of the most recent and significant examples of nanotechnology-based drug delivery.

In a recent study based on polymeric systems, nanosuspensions (200 nm) of the drug rilpivirine (TMC278) stabilized by polyethylene-polypropylene glycol (poloxamer 338) and PEGylated tocopheryl succinate ester (TPGS 1000) were studied in dogs and mice [35]. A single-dose administration of the drug in nanosuspensions resulted in sustained release over 3 months in dogs and 3 weeks in mice, compared with a half-life of 38 h for free drug. These results serve as a proof-of-concept that nanoscale drug delivery may potentially lower dosing frequency and improve adherence.

A series of experiments by Dou et al. showed that nanosuspension of the drug indinavir can be stabilized by a surfactant system comprised of Lipoid E80 for effective delivery to various tissues [36–38]. The indinavir nanosuspensions were loaded into macrophages and their uptake was investigated. Macrophages loaded with indinavir nanosuspensions were then injected intravenously into mice, resulting in a high distribution in the lungs, liver and spleen. More significantly, the intravenous administration of a single dose of the nanoparticle-loaded macrophages in a rodent mouse model of HIV brain infection resulted in significant antiviral activity in the brain and produced measureable drug levels in the blood up to 14 days post-treatment [38]. These studies serve as a proof of concept for indinavir delivery to the brain and the sustained drug levels for up to 14 days, which is important when considering that the half-life of indinavir in its conventional dosage form is 2 h. The demonstration that macrophages could be used to target drugs to the brain could be utilized for in vivo nanoparticle-targeted delivery of other drugs to the brain in the future.

Active targeting strategies have also been employed for antiretroviral drug delivery. Macrophages, which are the major HIV reservoir cells, have various receptors on their surface such as formyl peptide, mannose, galactose and Fc receptors, which could be utilized for receptor-mediated internalization. The drug stavudine was encapsulated using various liposomes (120–200 nm) conjugated with mannose and galactose, resulting in increased cellular uptake compared with free drug or plain liposomes, and generating significant level of the drug in liver, spleen and lungs [39–41]. Stavudine is a water-soluble drug with a very short serum half-life (1 h). Hence, the increased cellular uptake and sustained release in the tissues afforded by targeted liposomes is a major improvement compared with free drug. The drug zidovudine, with half-life of 1 h and low solubility, was also encapsulated in a mannose-targeted liposome made from stearylamine, showing increased localization in lymph node and spleen [42]. An important factor to consider here is that although most of the nucleoside drugs such as stavudine and zidovudine have short serum half-lives, the clinically relevant half-life is that of the intracellular triphosphate form of the drug. For example, despite zidovudine’s 1 h half-life in plasma, it is dosed twice daily based on intracellular pharmacokinetic and clinical efficacy data. Therefore, future nanotechnology-based delivery systems will have to focus in
showing significant increase of the half-lives of the encapsulated drugs to achieve a less frequent dosing such as once weekly, once-monthly or even less.

In separate work, a mannose-targeted poly (propyleneimine) dendrimer nanocarrier was used to deliver the drug efavirenz to human monocytes/macrophages \textit{in vitro} [43]. The targeted nanocarrier resulted in 12-fold increase in cellular uptake compared with free drug. A similar system was used to deliver the drug lamivudine \textit{in vitro}, resulting in significantly higher anti-HIV activity for the targeted and nontargeted dendrimer systems compared with free drugs [44]. In a more recent study, the tetra-peptide tuftsin (Thr-Lys-Pro-Arg) was conjugated to the same dendrimer to target the drug efavirenz to macrophages \textit{in vitro} [45]. The targeted dendrimer system resulted in sixfold prolonged release, 34-fold increased cellular uptake and sevenfold increase in anti-HIV activity compared with free drug.

In a new approach to target macrophage HIV reservoirs, a peptide nanocarrier was proposed as a model where a drug is conjugated to the backbone of peptide-PEG and \textit{N}-formyl-methionyl-leucyl-phenylalanine (fMLF), a bacterial peptide sequence for which macrophages express a receptor, is attached to the PEG for targeting [46]. The study found that fMLF-targeted peptide-PEG nanocarriers show increased cellular uptake and increased accumulation in macrophages of liver, kidney and spleen compared with those which are nontargeted [30, 46].

All the aforementioned efforts are examples of the potential nanotechnology platforms hold for improving targeted delivery of antiretroviral drugs to the cellular and anatomical reservoirs of HIV. These early efforts provide evidence for the potential of nanotechnology to improve delivery of antiretroviral therapy and support ongoing efforts to initiate clinical trials. Although the early efforts have not reached clinical trials yet, the works so far provide encouraging evidence that a subset of these preclinical technologies may enter clinical evaluation in the future.

**Nanomaterials as therapeutic agents**

In addition to being used as delivery agents, nanomaterials have also been shown to have therapeutic effects of their own. Studies have shown that the capsid of HIV could be a target for structure-based drug design for inhibiting viral replication [47,48]. As a result, both computational and experimental studies have identified compounds that could inhibit the assembly of the HIV capsid. Various nanomaterials have been found to inhibit viral replication \textit{in vitro} and it is suggested that these effects are based on structural interference with viral assembly.

Various fullerene (C-60)-based structures, dendrimers and inorganic nanoparticles, such as gold and silver, have been shown to have anti-HIV activity \textit{in vitro} [49–60]. While these efforts have not yet progressed beyond \textit{in vitro} studies, they illustrate the potential of therapeutic nanomaterials to inhibit HIV replication.

**Gene therapy for HIV/AIDS**

In addition to improving existing antiretroviral therapy, there are ongoing efforts to discover alternative approaches for treatment of HIV/AIDS [25,61]. One promising alternative approach is gene therapy, in which a gene is inserted into a cell to interfere with viral infection or replication. Other nucleic acid-based compounds, such as DNA, siRNA, RNA decoys, ribozymes and aptamers or protein-based agents such as fusion inhibitors and zinc-finger nucleases can also be used to interfere with viral replication [61,62].
Early efforts in gene therapy for HIV/AIDS have been focused on viral vectors as the delivery agents with various clinical trials in progress [61,63–68]. In one of these studies, Benitec Ltd and City of Hope are collaborating in an ongoing clinical trial to study the safety and feasibility of a gene therapy strategy based on the combination of three different inhibitory genes in a single lentiviral vector that utilizes stem cells in the delivery process [65]. Recently, scientists from UCLA reported that a Phase II gene therapy clinical trial showed that cell-derived gene transfer is safe and biologically active in HIV-infected individuals [69]. These efforts are encouraging and support the growing excitement around gene therapy for the treatment of HIV/AIDS. However, lessons learned over the past two decades indicate that the use of viral vectors for gene delivery poses fundamental problems such as toxicity, immunogenicity, insertion mutagenesis and limitations with scale-up procedures [70,71]. These problems have encouraged the investigation of nonviral vectors for gene delivery, where nanotechnology platforms are showing great promise [70–72].

In recent years, the Nobel prize-winning discovery of RNA interference (RNAi) in 1998 by Fire, Mello and colleagues has gained much attention in the clinical therapeutics field and is generating billion dollar investments in therapeutic applications [73,74]. Ongoing clinical trials for the treatment of age-related macular degeneration and respiratory syncytial virus have provided data that are creating tremendous excitement in the field [74]. RNAi is also considered to have therapeutic potential for HIV/AIDS [61,62,75]. Gene silencing is induced by double stranded siRNA, which targets for destruction the mRNA of the gene of interest. For HIV/AIDS, RNAi can either target the various stages of the viral replication cycle or various cellular targets involved in viral infection such as CD4, CCR5, and/or CXCR4, the major cell surface co-receptors responsible for viral entry. HIV replicates by reverse transcription to form DNA and uses the DNA to produce copies of its mRNA for protein synthesis; siRNA therapy could be used to knock down this viral mRNA. These two mechanisms are shown in Figure 2.

As with other gene therapy techniques, delivery of siRNA to specific cells and tissues has been the major challenge in realizing the potential of RNAi [74]. New nanotechnology platforms are tackling this problem by providing nonviral alternatives for effective and safe delivery. The first nontargeted delivery of siRNA in humans via self-assembling, cyclodextrin polymer-based nanoparticles for cancer treatment have recently entered Phase I clinical trials [76]. Although at an early stage, nonviral delivery of siRNA for treatment of HIV infection is also gaining ground. A fusion protein, with a peptide transduction domain and a double stranded RNA-binding domain, was used to encapsulate and deliver siRNA to T cells in vivo [77]. CD4- and CD8-specific siRNA delivery caused RNAi responses with no adverse effects such as cytotoxicity or immune stimulation. Similarly, a protamine-antibody fusion protein-based siRNA delivery demonstrated that siRNA knockdown of the \textit{gag} gene can inhibit HIV replication in primary T cells [78].

Single-walled nanotubes were shown to deliver CXCR4 and CD4 specific siRNA to human T cells and peripheral blood mononuclear cells [79]. Up to 90% knockdown of CXCR4 receptors and up to 60% knockdown of CD4 expression on T cells was observed while the knockdown of CXCR4 receptors on peripheral blood mononuclear cells was as high as 60%. In a separate study, amino-terminated carbosilane dendrimers (with interior carbon-silicon bonds) were used for delivery of siRNA to HIV-infected lymphocytes [80].

In a recent study, peptide conjugated to an antibody was used for targeted delivery of siRNA to T cells and to suppress HIV infection in humanized mice [81]. The antibody (scFvCD7)-specific to CD7 receptor on T cells was conjugated to the oligo-9-arginine peptide resulting in the delivery system designated scFvCD7–oligo-9-arginine. CD7 is an antigen found on the majority of human T cells that is rapidly internalized on antibody binding [81]. The oligo-9-
arginine peptide is a short, positively charged cell-penetrating peptide that has been shown to be highly effective in facilitating cellular uptake of nucleic acids [82]. The peptide binds with the negatively charged siRNA, enhancing delivery to the target cells. In this study, anti-CCR5 and antiviral siRNA delivered with scFvCD7–oligo-9-arginine controlled viral replication and prevented CD4 T cell loss. Moreover, the treatment suppressed viral titers and restored CD4 T cell counts while effectively suppressing viremia in HIV-infected humanized mice. This work opens new avenues for future use of antibody-targeted nanoparticles in the delivery of siRNA to HIV-specific cells.

These pioneering studies demonstrate that nonviral siRNA delivery is possible for HIV/AIDS treatment. However, more work needs to be done in optimizing the delivery systems and utilizing designs for efficient targeting and intracellular delivery. The recent developments in polymer- and liposome-based siRNA delivery systems could be optimized for targeting cells that are infected with HIV, such as T cells and macrophages. Moreover, since HIV mutates and has multiple strains with different genetic sequences, combination siRNA therapy targeting multiple genes should be pursued. For these applications, nanotechnology platforms with capability for co-delivery and targeting need to be developed specifically for HIV-susceptible cells. A macrophage and T-cell-targeted and nanotechnology-based combination gene therapy may be a promising platform for efficient HIV/AIDS treatment.

**Immunotherapy for HIV/AIDS**

The various treatment approaches described above focus on treating HIV/AIDS by directly targeting HIV at the level of the host cell or the virus itself. An alternative approach is immunotherapy aimed at modulating the immune response against HIV. CD8+ cytotoxic T-cell responses to acute HIV infection appear to be relatively normal, while neutralizing antibody production by B cells is delayed or even absent [83]. Over time, viral mutation leads to loss of the CD8+ T cell cytotoxic function. However, the major effect of an infection by HIV is the loss of CD4+ T cells. These ‘helper’ T cells are responsible for a number of supportive functions for other immune populations and their loss leads to profound immunosuppression, manifested by the presence of dysfunctional B-cells, natural killer cells and the macrophages in chronically HIV-infected patients [84]. In recent years, there has been increasing interest in the therapeutic use of immune responses to restore the regular function of the immune system as an effective way to treat HIV/AIDS [85–87]. There has been increasing evidence that the immune system is capable of controlling HIV in certain individuals [87]. Hence, strategies to rebuild or allow the reconstitution of immune function could be one of the best approaches for effective treatment.

Immunotherapy is a treatment approach involving the use of immunomodulatory agents to modulate the immune response against a disease. Similar to vaccines, it is based on immunization of individuals with various immunologic formulations; however, the purpose is to treat HIV-infected patients as opposed to protect healthy individuals (preventive vaccines will be discussed in an upcoming section). The various immunotherapy approaches for HIV/AIDS could be based on delivering cytokines (such as IL-2, IL-7 and IL-15) or antigens [84, 85]. The development of cellular immunity, and to a large degree humoral immunity, requires antigen-presenting cells (APCs) to process and present antigens to CD4+ and CD8+ T cells. Dendritic cells (DCs) are the quintessential professional APCs responsible for initiating and orchestrating the development of cellular and humoral (antibody) immunity [88,89]. Protein/peptide antigens or DNA immunogens (which lead to endogenous protein expression) could then be delivered through viral vectors to endogenous or *ex vivo*-generated DCs.

Since the first immunotherapeutic clinical trial for HIV/AIDS in 1983, various trials have been carried out and are still ongoing [90–93]. Unfortunately, despite the preclinical studies that
showed enhanced immune responses, most of the clinical studies have consistently failed to provide clinical improvements for patients [90]. Most of these clinical trials have been based on the delivery of the immunogenic factors through viruses or \textit{ex vivo} DCs. As discussed under the gene therapy section above, delivery through viral vectors involves various risks. In addition, \textit{ex vivo} generation of autologous DCs is a difficult therapeutic strategy to utilize widely as it involves a very labor-intensive procedure with high costs and multiple procedures for product control at different sites [94]. Hence, new approaches using targeted nanotechnology platforms for delivery of immunomodulatory factors and targeting antigens to DC surface receptors \textit{in vivo} provide immense opportunities [87]. The rationale for nanotechnology-based DC targeting and vaccine delivery will be discussed in more detail in the upcoming section. Here, the most important advances of nanotechnology-based immunotherapy will be described.

Various polymeric systems have been explored for \textit{in vivo} targeting of DCs and delivery of small molecules, proteins or DNAs showing potential for immunotherapy. Poly(ethylene glycol) (PEG) stabilized poly(propylene sulfide) polymer nanoparticles accumulated in DCs in lymph nodes [95]. Following nanoparticle injection, DCs containing nanoparticles accumulated in lymph nodes, peaking at 4 days with 40–50% of DCs and other APCs having internalized nanoparticles. In a separate study, cross-linked polymer nanoparticles with a pH-responsive core and hydrophilic charged shell were used for delivery of proteins and small molecules to DCs [96]. The cross-linked polymers were composed of 2-diethylamino ethyl methacrylate, PEG dimethacrylate and 2-aminoethyl methacrylate. Fluorescence microscopy showed delivery of the model protein antigen ovalbumin to the cytosol of bone marrow-derived DCs, where antigens can potentially be processed for cross-presentation to CD8$^+$ T cells, in addition to more classical exogenous antigen presentation to CD4$^+$ T cells.

In another study, nanoparticles of the copolymer poly(D,L-lactide-co-glycolide) (PLGA) showed efficient delivery of antigens to murine bone marrow-derived DCs \textit{in vitro}, suggesting their potential use in immunotherapy [97]. More recently, a very interesting work showed that HIV p24 protein adsorbed on the surface of surfactant-free anionic poly(D,L-lactide) (PLA) nanoparticles were efficiently taken-up by mouse DCs, inducing DC maturation [98]. The p24-nanoparticles induced enhanced cellular and mucosal immune responses in mice. Although this targeting is seen in \textit{ex vivo}-generated DCs and not \textit{in vivo} DCs, the efficient delivery of the antigen to DCs through the nanoparticles is an important demonstration that may eventually be applied to \textit{in vivo} DC targeting.

The most clinically advanced application of nanotechnology for immunotherapy of HIV/AIDS is the DermaVir patch that has reached Phase II clinical trials [99]. DermaVir is a targeted nanoparticle system based on polyethyleneimine mannose (PEIm), glucose and HIV antigen coding DNA plasmid formulated into nanoparticles (~100 nm) and administered under a patch after a skin preparation. The nanoparticles are delivered to epidermal Langerhans cells that trap the nanoparticles and mature to become highly immunogenic on their way to the lymph nodes. Mature DCs containing the nanoparticles present antigens to T cells inducing cellular immunity. Preclinical studies and Phase I clinical trials showed safety and tolerability of the DermaVir patch, which led the progression to Phase II trials. This is the first nanotechnology-based immunotherapy for HIV/AIDS that has reached the clinic and encourages further work in this area. A selection of nanotechnology platforms used for antiretroviral drug delivery, gene therapy and immunotherapy are listed in Table 1.
Nanotechnology for HIV/AIDS prevention

Vaccine delivery

The search for a safe and effective HIV/AIDS vaccine has been challenging in the almost three decades since the discovery of the disease. Recently, high-profile clinical trial failures have prompted great debate over the vaccine research, with some suggesting the need for a major focus on fundamental research, with fewer efforts on clinical trials [13–15,100,101].

The major challenges in the development of a preventive HIV/AIDS vaccine have been the extensive viral strain and sequence diversity, viral evasion of humoral and cellular immune responses, coupled with the lack of methods to elicit broadly reactive neutralizing antibodies and cytotoxic T cells [102]. In order to generate T cell responses, protein antigens must enter APCs (such as DCs) where peptides are processed and loaded into MHC molecules for presentation to CD4+ T cells (extracellular antigen in MHC class II) and CD8+ T cells (intracellular antigen in MHC class I) [103]. The challenge associated with delivery of any exogenous antigen (such as nanoparticles) to APCs, is that exogenous antigens require specialized ‘cross-presentation’ in order to be presented by MHC class I and activate CD8+ cytotoxic T cells [104]. This requirement for cytosolic delivery of antigens and cross-presentation represents yet another hurdle for HIV intracellular antigen vaccine, but potentially an advantage of nanodelivery. Humoral responses (neutralizing antibodies produced by B cells) are generated to intact antigen presented on the surface for the virus, or nanoparticles, but these humoral responses typically require ‘help’ from CD4+ T cells [105], and thus the challenge is not to achieve either a single cellular or humoral response, but rather both.

Nanoparticles have potential as adjuvants and delivery systems for vaccines. Over the past few decades, controlled-release systems have been used for sustained release of various agents. This has great advantages for vaccine delivery since the release of antigens in a controlled manner could lead to a prolonged and stronger initiation of the immune response. By protecting the delivered antigen from body fluids (e.g., lymph, serum and mucus) nanoparticle antigen encapsulation can increase the half-life of an immunizing antigen. Nanoparticles can also be designed to effectively target APCs [106]. Targeting antigen delivery to DCs with surface-functionalized nanoparticles presents a major opportunity for delivery of antigen and initiation of immune responses. Another major benefit of nanoparticle vaccines is that they can be optimized for various routes of administration. Conventional vaccines are mostly administered intramuscularly, but nanoparticles provide expanded opportunity for oral and nasal vaccinations where mucosal immunity could be induced [107].

To deliver antigens, nanoparticles can be used either to encapsulate antigens in their core or absorb the antigens on their surfaces. Both strategies present advantages depending on whether the humoral or cellular immune response is desired. For efficient antigen presentation using DCs, encapsulating antigens can lead to the efficient delivery of an extracellular antigen (nanoparticle) into the cytoplasm [106]. Recently, there has been considerable progress in the field of nanoparticle design for efficient intracellular delivery of various agents. On the other hand, the ability to functionalize the surface of nanoparticles provides the capability to put antigens on the surface of the particles for direct presentation to B cells for antibody-specific immune responses. In order to achieve the coordinated effort of humoral and cellular responses that are essential for the prevention of HIV/AIDS, rationally designed nanoparticles will have the capacity to present antigens to both DCs (encapsulated) and B cells (surface absorbed).

Although the development of nanoparticle-based vaccines for HIV/AIDS is at an early stage, there has been recent progress. Here, we discuss the early developments of nanoparticles based on polymers and liposomes for HIV/AIDS vaccine.
Various lipid-based systems have been investigated for HIV/AIDS vaccine delivery. In an earlier study, nasal immunization of mice with the HIV gp160 protein encapsulated in a liposome induced high titers of gp160-specific neutralizing antibody responses [108]. The liposomes were made from a mixture of cholesterol, sphingomyelin, phosphatidylethanolamine, phosphatidylcholine and phosphatidylserine. The HIV gp 41 protein was also delivered through a variety of liposomes (110–400 nm) eliciting strong antibody responses in mice and rabbits [109–111]. To date, there have been no vaccines that can elicit a broadly neutralizing antibody response to HIV; therefore, it is doubtful that an antibody response to a monovalent encapsulated antigen alone will be enough for a sterilizing HIV vaccine [112,113]. These nanodelivery systems therefore need to be improved to include a variety of HIV epitopes and potentially even epitopes engineered to enhance access to and generation of antibody responses to areas of the HIV glycoproteins not elicited by acute HIV infection [112,113].

Dendritic cells express a variety of microbial pattern recognition receptors such as Toll-like receptors, one of which, TLR9, binds CpG oligonucleotides (CpG ODN) [114]. CpG ODNs are under intense investigation for DC-targeted vaccines, as CpG stimulation leads to activation and maturation of DCs into potent initiators of cellular immune responses. Immunization with CpG ODN delivered through liposomes (made of 1-(2-[oleoyloxy] ethyl)-2-oleyl-3-(2-hydroxy-ethyl) imidazolinium chloride (DOTIM) and cholesterol) led to stronger simian immunodeficiency virus (SIV)-specific T and B cell responses in rhesus macaques compared with control immunization [115]. Although cationic lipids such as DOTIM are not preferred in drug delivery, they present an opportunity in vaccine application because their immunogenic properties can be utilized for increased adjuvanticity.

MF59 is a nanosized (<250 nm) oil-in-water emulsion composed of squalene and the surfactants polysorbate 80 and sorbitan trioleate [116,117]. It has been shown to be safe in human trials and approved for use in over 20 countries as an adjuvant for an influenza vaccine [118]. Immunization of baboons with the HIV env and gag DNA followed by booster immunizations with oligomeric Env protein in MF59 elicited stronger antibodies and cellular responses compared with immunizations with DNA alone [119]. Another study found that priming rabbits with modified HIV env DNA, followed by boosting with oligomeric protein in MF59, elicited higher titers of env-binding and autologous neutralizing antibodies than priming with DNA alone. In a more recent study, immunization of mice with DNA plasmids encoding for the proteins gp140, Gag and Tat formulated in MF59 containing CpG ODN also elicited higher titers of binding and neutralizing antibodies compared with immunization with MF59 alone [121].

Another oil-in-water nanoemulsion (~350 nm) prepared from cetlypyridinium chloride, a nonionic surfactant and soybean oil was used for delivering gp120 protein through the nasal route, eliciting antibody and cellular immune responses in mice and guinea pigs [122]. The potential for nasal administration of vaccine represents an alternative to conventional intramuscular delivery, especially as it regards generation of mucosal immunity.

Nanoparticles based on the polymers PLA and PLGA have also shown potential for delivery of protein- and DNA-based HIV/AIDS vaccines. Surfactant-free PLA nanoparticles (300–600 nm) coated with HIV p24 protein induced high antibody titers in mice, rabbits and macaques, and elicited strong cytotoxic T cell responses in mice [123]. Similarly, co-adsorption of p24 and gp120 proteins onto PLA nanoparticles (185–250 nm) elicited high antibody titers in mice, illustrating the potential for PLA nanoparticle use as multivalent vaccines [124]. Moreover, immunization of rabbits with any of the three HIV antigens (p24, WT Tat and mutated, detoxified Tat) mixed with either PLA nanoparticles or the emulsion MF59 led to the induction
of similar level of antibody titers [125]. The ability of PLA to surface adsorb antigens and induce both antibody and cellular responses appears to be an asset in its continued use in nanoparticle vaccines.

Polystyrene nanospheres (~350 nm) coated with inactivated HIV particles were used for nasal immunization, effectively inducing mucosal antibody responses, detected as the HIV-specific vaginal antibody response in mice [126]. The nanospheres contain poly(methacrylic acid) branches with concanavalin A immobilized on their surface. In a more promising study, immunization with the polystyrene nanospheres induced DC-mediated immune responses in mice, characterized by production of efficient antibodies against HIV detectable in the genital tract as well as specific cytotoxic T cells in the spleen [127]. This study is unique because it introduces a novel approach that combines both the older technique of using inactivated viral particles as well as the newer approaches utilizing polymeric nanoparticles for delivery of vaccines.

Nanoparticles (200 nm) composed of hydrophobically modified poly (gamma-glutamic acid) (gamma-hPGA) encapsulating gp120 protein strongly induced antigen-specific cellular immunity when targeted to DCs in Balb/c mice [128]. These nanoparticles were also shown to induce significant CD8+ T cell responses in mice compared with gp120 alone [129]. In another study, immunization of mice with the gamma-hPGA nanoparticles encapsulating the p24 protein activated antigen-specific CD8+ T cells in spleen and induced p24-specific serum antibodies, as compared with immunization with p24 alone [130]. The nanoparticles induced comparable p24-specific serum antibody to that of the complete Freund’s adjuvant, a potent immunomodulator.

The above works indicate some of the major efforts and progress made using nanoparticle systems for HIV/AIDS vaccine delivery. A summary of the major antigen candidates, the nanotechnology platforms used and the development stage for each system is provided in Table 2. Although the results so far are encouraging, future works need to focus on design and specific targeting of the necessary cells of the immune system. Optimization of different parameters such as the size, targeting ligand, surface density, encapsulation efficiency and route of administration would lead to better and more efficient vaccine delivery systems.

**Intravaginal microbicides**

Although vaccines that induce sterilizing immunity are the most ideal way to prevent the spread of HIV/AIDS, other approaches are also being pursued until a safe and effective vaccine is developed. Since sexual transmission is the major route of infection, prevention methods aimed at behavioral changes as well as the use of personal protection such as condoms have helped in reducing the spread of the disease in some countries [131]. However, effective protection methods that can be utilized by women have not been readily available, making women more vulnerable to the disease. Among people infected with HIV/AIDS, women account for nearly 50% of infections worldwide and 60% in Sub-Saharan Africa [132,133]. As a result, there are major efforts focused on developing effective microbicides for HIV/AIDS prevention. Microbicides are preventive agents that are topically applied into the vagina to prevent the transmission of HIV/AIDS or other sexually transmitted diseases.

There are currently over 50 drug candidates in preclinical development and 12 candidates in clinical trials for use as microbicides [132]. These microbicide candidates work by different mechanisms that either target the virus or inhibit viral binding to the target cell. Most of the current microbicides in development are based on gels formed from anionic polymers and polysaccharides, whereas microbicides based on antiretroviral drugs have also gained attention [132,134,135]. However, similar to the difficulties faced in vaccine development, major candidate microbicides for HIV/AIDS failed in recent efficacy clinical trials suggesting the
need for further studies and new approaches [16,17]. Here, we discuss the most recent nanotechnology-based approaches that focus on using dendrimers, siRNA and nanoparticles in microbicides for HIV/AIDS.

VivaGel is a microbicide gel formed from the L-lysine dendrimer that has a polyanionic outer surface developed by the company Starpharma [136]. After comparing the antiviral effects of various dendrimers based on L-lysine, poly(amido amine) and poly(propylene imine), the product SPL7013, based on the dendrimer L-lysine, was identified as the best microbicide candidate [137]. In earlier works, SPL7013 applied as a topical microbicide in female pigtailed macaques showed a dose-dependent resistance to viral challenge [138]. Recently, it was shown in a Phase I safety trial that the dendrimer solution is safe in humans [136].

Polymer-based nanoparticles are also being considered in the development of microbicides. PLGA nanoparticles (~260 nm), in which the protein PSC-RANTES (an amino terminus-modified analog of the chemokine RANTES, a ligand for HIV co-receptor CCR5) was encapsulated, maintained comparable anti-HIV activity compared with unformulated PSC-RANTES in HeLa cells [139]. In an ex vivo cervical tissue model, the PSC-RANTES-encapsulated nanoparticles displayed a fivefold increase in tissue uptake, enhanced tissue permeation, and significant localization at the basal layers of the epithelium over unformulated PSC-RANTES. A major study has shown that PSC-RANTES provides potent protection against vaginal challenge in rhesus macaques, indicating its potential for use in microbicides [140]. This protection is based on the inhibition of the CCR5 co-receptor, which prevents HIV entry into cells.

Another novel approach is the use of siRNA as an intravaginal viral microbicide. As discussed above, siRNA are small single-stranded RNA molecules that are complementary to the gene targeted for knockdown [74]. The first study reporting the use of siRNA for intravaginal microbicide-delivered siRNA using lipids and protected mice from lethal Herpes simplex virus 2 infection [141,142]. The efficacy of siRNA gene knockdown was further improved by using cholesterol-conjugated siRNAs to silence gene expression in the vagina without causing inflammation or inducing interferons [143]. More recently, it was reported that biodegradable polymer nanoparticles are effective delivery vehicles for siRNA to the vaginal mucosa [144]. PLGA nanoparticles (<200 nm) mixed with the polyamines spermidine or putrescine delivered siRNA to the vaginal mucosa, leading to sustained gene silencing for up to 14 days. In vivo studies in mice suggest that knockdown of gene expression was proximal (in the vaginal lumen) and distal (in the uterine horns) to the site of topical delivery and the nanoparticles also penetrated deep into the epithelial tissue. Although this work is at an early stage, it shows that siRNA-based microbicides may be a tool for HIV/AIDS prevention in the near future.
dendrimers and inorganic nanoparticles such as silver have antiviral effects or improve antiviral effects of other molecules, as in the case of gold nanoparticles.

Newer treatment approaches, such as gene therapy and immunotherapy, can be enhanced with nanotechnology. Nonviral delivery of siRNA is one of the most active areas of research in nanotechnology. Delivery of siRNA to HIV-specific cells has been demonstrated; however, work remains in this area since safe and effective nanotechnology for RNAi has yet to be developed for HIV/AIDS applications. Immunotherapy is another major area where nanotechnology could play a significant role. The movement of the DermaVir Patch into Phase II trials is an indication that nanoimmunotherapy may be the earliest form of nanotechnology-based treatment that will be available for HIV/AIDS.

Nanotechnology-based vaccines for HIV/AIDS are also showing potential in early studies. Their ability to target specific cells and release antigens in a controlled and sustained manner makes nanoparticles a great alternative to viral vectors. Lipid- and polymer-based nanoparticles have been shown to induce HIV-specific antibody and cellular immune responses in animal studies. Despite the progress so far, preclinical studies remain important to ensure a clear understanding and optimization of mechanisms involved in the nanoparticle induction of strong humoral and cellular immunity. In addition to continued efforts in vaccine development, research into microbicide development remains important. Nanotechnology can play a major role in microbicide development by providing innovative strategies for nanoparticle-based delivery of therapeutic molecules or RNAi.

While there is exciting potential for nanomedicine in the treatment of HIV/AIDS, challenges remain to be overcome before the potential is realized. These include toxicity of nanomaterials, stability of nanoparticles in physiological conditions and their scalability for large-scale production. These are challenges general to all areas of nanomedicine and various works are underway to tackle them. Moreover, most of the nanotechnology-based studies undertaken so far for the treatment and prevention of HIV/AIDS are in the preclinical stages. This implies that many hurdles remain before there is widespread use of these technologies in the clinic. The \textit{in vitro} and \textit{in vivo} studies performed so far are preliminary in most cases and more work needs to be done with clear therapeutic end points that will indicate the efficacy of the nanotechnology-based systems. One of the major challenges to animal studies for HIV/AIDS treatment is the lack of a suitable animal model. Nonhuman primate models utilize the HIV relative SIV, which has similarities but ultimately differs from human HIV pathogenesis. The development of humanized mice, mice that possess a human immune compartment, and therefore can support HIV infection, has allowed the study of human T cells \textit{in vivo} [149, 150]. This has allowed the study and manipulation of HIV interaction with human T cells \textit{in vivo}. A study that investigated the targeted delivery of siRNA to T-cells was able to use humanized mice to suppress endogenous viral replication and restore CD4$^+$ T cell counts, while effectively suppressing viremia in HIV-infected mice [81]. The use of humanized mice will provide the means for direct experimentation with human cells, and hasten the pace of translational research.

Another important aspect to consider in the application of nanotechnology-based delivery systems for HIV/AIDS treatment and prevention is the route of administration. Most of the studies illustrated so far were based on parenteral administration of the formulations. Recently, a lot of effort has been directed towards needle-free delivery methods, mainly oral, transdermal, nasal and pulmonary routes, in addition to injectable administration [151–154]. In the context of drug delivery for HIV/AIDS, nonparenteral routes could be preferred considering the ease of administration and user-friendliness (e.g., oral vaccine delivery). These also have great advantages in improving patient adherence to the regimens. However, nonparenteral routes have limitations in their efficiency and bioavailability, and their long-term application could
have negative effects (e.g., potential irritation by transdermal or pulmonary route). To overcome these obstacles, further research efforts would be required to investigate highly efficient nanoparticle delivery systems for the replacement of injection/infusion of HIV/AIDS drugs.

Another important consideration in investigating nanotechnology-based systems for HIV/AIDS is the economic aspect, as the hardest hit and most vulnerable populations reside in underdeveloped and economically poor countries. In the case of antiretroviral therapy, nanotherapeutics may increase the overall cost of treatment, reducing the overall value. However, if the nanotherapeutics could improve patient adherence by reducing dosing frequency as expected, and furthermore, if they can eradicate viral reservoirs leading to a sterile immunity, these advantages may effectively offset the added cost. In the case of new treatment approaches such as gene therapy and immunotherapy, the use of nanotherapeutics could even reduce the cost when compared with the other alternative approaches being considered. Although no data exist for such comparisons, it can be assumed that nanotherapeutics might have advantages for large-scale productions when compared with viral or ex vivo DC-based gene therapy or immunotherapy. Similarly, in the case of the prevention methods, the nanotherapeutics should not be more costly compared with existing systems. Since bulk quantities of vaccines are widely distributed through government agencies, development of nanotechnology-enabled vaccines would be one of the most cost effective approaches to address the HIV/AIDS epidemic worldwide. Nanotechnology-enabled microbicides may also need to rely on support from governmental or nongovernmental organizations for effective distribution to economically disadvantaged countries.

Overall, these are exciting times for nanotechnology research and the pace of scientific discovery is beginning to gain momentum. It is widely accepted that with continued support, medicine and the field of HIV/AIDS will be important beneficiaries of nanotechnology for years to come.

### Executive summary

**Challenges of HIV/AIDS treatment**

- HIV resides in latent cellular and anatomical reservoirs where current drugs are unable to completely eradicate the virus.
- Macrophages are major cellular reservoirs, which also contribute to the generation of elusive mutant viral genotypes by serving as the host for viral genetic recombination.
- Anatomical latent reservoirs include secondary lymphoid tissue, testes, liver, kidney, lungs, the gut and the brain.
- The major challenge facing current drug regimens is that they do not fully eradicate the virus from these reservoirs; requiring patients take medications for life. Under current treatment, pills are taken daily, resulting in problems of patient adherence. The drugs also have side effects and in some people the virus develops resistance against certain drugs.

**Nanotechnology-based drug delivery for HIV/AIDS treatment**

- Nanotechnology-based targeted delivery of antiretroviral drugs to CD4+ T cells and macrophages, as well as delivery to the brain and lymphocytes, could ensure that drugs reach latent reservoir.
Various nanotechnology-based drug delivery systems have been successfully used to deliver various antiretroviral drugs in vitro and in vivo.

Nanosuspensions that were used to deliver the drug rilpivirine (TMC278) resulted in sustained release over 3 months in dogs and 3 weeks in mice. This result is a major indication of how nanotechnology-based drug delivery could improve antiviral treatment.

Nanomaterials as therapeutic agents

- Various nanomaterials such as fullerenes, dendrimers, silver and gold nanoparticles have shown anti-HIV effects in vitro.
- Silver nanoparticles showed size-dependent interaction with HIV, inhibiting the virus from binding to CD4+ T cells while gold nanoparticles conjugated to the molecule SDC-1721 (a segment of the CCR5 inhibitor TAK-779) showed strong anti-HIV activity compared with free SDC-1721.

Gene therapy for HIV/AIDS treatment

- Gene therapy based on siRNA has shown promise for HIV/AIDS treatment. Nanotechnology platforms for delivery of siRNA for HIV/AIDS treatment are in their early stages but recent work has been met with optimism.
- Single-walled nanotubes, dendrimers, fusion proteins, peptide–antibody conjugates have all been used for delivery of siRNA to HIV-specific cells.

Immunotherapy for HIV/AIDS

- Immunotherapy for HIV/AIDS based on viral agents and administration of ex vivo-generated autologous dendritic cells involves risks due to the viral agents and the complicated procedures used in ex vivo dendritic cell generation and manipulation.
- Nanotechnology-based immunotherapy that uses DNA plasmid delivered through mannose targeted polyethyleimine has reached Phase II clinical trials.

Nanotechnology-based preventive HIV/AIDS vaccine

- Generation of an effective HIV/AIDS vaccine has been notoriously difficult and new approaches are always sought.
- Nanoparticles have various advantages in improving delivery of antigens to enhance the immune response. They can be used both for encapsulating antigens in their core from which antigen presenting cells can process and present and cross-present antigen to CD4+ and CD8+ T cells respectively, or absorbing the antigens on their surfaces, allowing B cells to generate humoral responses. Nanoparticle vaccines can also be optimized for various routes of administration.
- Various polymeric and lipid-based nanoparticles have been used to deliver DNA-, protein- or peptide-based antigens in vivo, eliciting strong cellular, humoral and mucosal immune responses.

Nanotechnology-based intravaginal microbicides

- Intravaginal microbicides are preventive agents that are topically applied to the vagina to prevent the transmission of HIV/AIDS or other sexually transmitted diseases.
- Nanotechnology-based approaches are being developed to use dendrimers, siRNA and nanoparticles for microbicidal functions.
• VivaGel is a dendrimer-based microbicide gel that has been shown to be safe in humans in Phase I clinical trials.
• Polymeric nanoparticles have been used to deliver the CCR5 inhibitor PSC-RANTES and HIV-specific siRNA as microbicides.

Acknowledgments

The authors would also like to thank Paul Sax for helpful discussions.

Bibliography


Figure 1.
Schematic representation of various nanotechnology platforms that can be used in HIV/AIDS treatment and prevention.
Figure 2. Mechanism for siRNA-based gene therapy of HIV/AIDS

The siRNA acts by degrading mRNA in at least two major ways: (A) Inhibit entry and fusion by interfering with production of receptors or co-receptors and (B) interfere with translation and transcription of viral genes preventing production of proteins and genomic RNA. (The viral entry and replication stages shown here are also the targets for the antiretroviral drugs discussed above [29]).
**Table 1**

Summary of nanotechnology-based treatment approaches for HIV/AIDS.

<table>
<thead>
<tr>
<th>Type of therapy</th>
<th>Therapeutic agent (drug or gene)</th>
<th>Nanotechnology delivery platform</th>
<th>Development stage</th>
<th>Refs.</th>
</tr>
</thead>
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<td>Antiretroviral therapy</td>
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<td>Poloxamer 338/TPGS 1000</td>
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<td>[35]</td>
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<tr>
<td></td>
<td>Indinavir</td>
<td>Liposome-laden macrophages</td>
<td>Preclinical</td>
<td>[36–38]</td>
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<td></td>
<td>Stavudine</td>
<td>Mannose- and galactose-targeted liposome</td>
<td>Preclinical</td>
<td>[39–41]</td>
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<td></td>
<td>Zidovudine</td>
<td>Mannose-targeted liposome</td>
<td>Preclinical</td>
<td>[42]</td>
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<td></td>
<td>Efavirenz</td>
<td>Mannose-targeted dendrimer</td>
<td>Preclinical</td>
<td>[43,45]</td>
</tr>
<tr>
<td></td>
<td>Lamivudine</td>
<td>Mannose-targeted dendrimer</td>
<td>Preclinical</td>
<td>[46]</td>
</tr>
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<td>Nanomaterials</td>
<td>Fullerene derivatives</td>
<td>–</td>
<td>Preclinical</td>
<td>[49–55]</td>
</tr>
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<td></td>
<td>Dendrimers</td>
<td>–</td>
<td>Preclinical</td>
<td>[56,57]</td>
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<td></td>
<td>Silver nanoparticles</td>
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<td>Preclinical</td>
<td>[58,59]</td>
</tr>
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<td></td>
<td>SDC-1721/gold nanoparticles</td>
<td>Gold nanoparticles</td>
<td>Preclinical</td>
<td>[60]</td>
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<td>Gene therapy</td>
<td>siRNA</td>
<td>Peptide fusion proteins, protamine–antibody fusion proteins, dendrimers, single walled carbon nanotubes, peptide–antibody conjugates</td>
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<td>Immunotherapy</td>
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<td>Poly (D,L-lactide) nanoparticles/dendritic cells</td>
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<td>[98]</td>
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<td>Plasmid DNA</td>
<td>Mannose-targeted polyethyleimine polymers</td>
<td>Phase II clinical trials</td>
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</table>
Table 2

Summary of nanotechnology developments for prevention of HIV/AIDS.

<table>
<thead>
<tr>
<th>Type of preventive agent</th>
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<th>Nanotechnology platform</th>
<th>Development stage</th>
<th>Refs.</th>
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</thead>
<tbody>
<tr>
<td>Protein or peptide vaccine</td>
<td>gp41, gp120, gp160, p24, Env, Gag, Tat</td>
<td>Liposomes, nanoemulsion, MF59, PLA nanoparticles, poly(γ-glutamic acid) nanoparticles</td>
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<td>[108–111]</td>
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<td></td>
<td></td>
<td></td>
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<td>[119–120]</td>
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<td></td>
<td>[128–130]</td>
</tr>
<tr>
<td>DNA vaccine</td>
<td>env, rev, gag, tat, CpG ODN</td>
<td>Liposomes, nanoemulsion, PLA nanoparticles</td>
<td>Preclinical</td>
<td>[115,121]</td>
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<td>Inactivated viral particle</td>
<td>Inactivated HIV viral particle</td>
<td>Polystyrene nanospheres</td>
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<td>[126–127]</td>
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<td>Microbicides</td>
<td>L-lysine dendrimer</td>
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<td>Phase I/II</td>
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<td></td>
<td>PLGA nanoparticles</td>
<td>PLGA nanoparticles</td>
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<td></td>
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<tr>
<td></td>
<td>PSC-RANTES</td>
<td>PLGA</td>
<td>Preclinical</td>
<td>[139]</td>
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<tr>
<td>siRNA</td>
<td>Nanoparticles, lipids, cholesterol conjugation</td>
<td></td>
<td>Preclinical</td>
<td>[141–144]</td>
</tr>
</tbody>
</table>

ODN: Oligonucleotides; PLA: Poly(D,L-lactide); PLGA: Poly(D,L-lactide-co-glycolide).