The Program of Gene Transcription for a Single Differentiating Cell Type During Sporulation in Bacillus Subtilis

The Harvard community has made this article openly available. Please share how this access benefits you. Your story matters.

<table>
<thead>
<tr>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Published Version</td>
<td><a href="https://doi.org/10.1371/journal.pbio.0020328">doi:10.1371/journal.pbio.0020328</a></td>
</tr>
<tr>
<td>Accessed</td>
<td>June 10, 2017 8:51:25 AM EDT</td>
</tr>
<tr>
<td>Citable Link</td>
<td><a href="http://nrs.harvard.edu/urn-3:HUL.InstRepos:2967528">http://nrs.harvard.edu/urn-3:HUL.InstRepos:2967528</a></td>
</tr>
<tr>
<td>Terms of Use</td>
<td>This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Open Access Policy Articles, as set forth at <a href="http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#OAP">http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#OAP</a></td>
</tr>
</tbody>
</table>

(Article begins on next page)
Asymmetric division during sporulation by Bacillus subtilis generates a mother cell that undergoes a 5-h program of differentiation. The program is governed by a hierarchical cascade consisting of the transcription factors: $\sigma^E$, $\sigma^K$, GerE, GerR, and SpoIIID. The program consists of the activation and repression of 383 genes. The $\sigma^E$ factor turns on 262 genes, including those for GerR and SpoIIID. These DNA-binding proteins downregulate almost half of the genes in the $\sigma^E$ regulon. In addition, SpoIIID turns on ten genes, including genes involved in the appearance of $\sigma^K$. Next, $\sigma^K$ activates 75 additional genes, including that for GerE. This DNA-binding protein, in turn, represses half of the genes that had been activated by $\sigma^K$ while switching on a final set of 36 genes. Evidence is presented that repression and activation contribute to proper morphogenesis. The program of gene expression is driven forward by its hierarchical organization and by the repressive effects of the DNA-binding proteins. The logic of the program is that of a linked series of feed-forward loops, which generate successive pulses of gene transcription. Similar regulatory circuits could be a common feature of other systems of cellular differentiation.

Introduction

A fundamental challenge in the field of development is to understand the entire program of gene expression for a single differentiating cell type in terms of an underlying regulatory circuit. This challenge can be met in part through recent advances in transcriptional profiling, which have made it possible to catalog changes in gene expression on a genome-wide basis (Brown and Botstein 1999). However, most systems of development involve multiple differentiating cell types, complicating the challenge of deciphering the program of gene expression for individual cell types. Also, many developmental systems are insufficiently accessible to genetic manipulation to allow genome-wide changes in gene expression to be understood in detail in terms of an underlying regulatory program. An understanding of how a cell differentiates from one type into another requires both a comprehensive description of changes in gene expression and an elucidation of the underlying regulatory circuit that drives the program of gene expression. Here we report our efforts to comprehensively catalog the program of gene expression in a primitive system of cellular differentiation, spore formation in the bacterium Bacillus subtilis, and to understand the logic of this program in terms of a simple regulatory circuit involving the ordered appearance of two RNA polymerase sigma factors and three positively and/or negatively acting DNA-binding proteins.

Spore formation in B. subtilis involves the formation of an asymmetrically positioned septum that divides the developing cell (sporangium) into unequal-sized progeny that have dissimilar programs of gene expression and distinct fates (Piggot and Coote 1976; Stragier and Losick 1996; Piggot and Losick 2002; Errington 2003). The two progeny cells are called the forespore (the smaller cell) and the mother cell. Initially, the forespore and the mother cell lie side by side, but later in development the forespore is wholly engulfed by the mother cell, pinching it off as a cell within a cell. The forespore is a germ cell in that it ultimately becomes the spore and, upon germination, gives rise to vegetatively growing cells. The mother cell, on the other hand, is a terminally differentiating cell type that nurtures the developing spore but eventually undergoes lysis to liberate the fully ripened spore when morphogenesis is complete. The entire process of spore formation takes 7–8 h to complete with approximately 5 h of development taking place after the sporangium has been divided into forespore and mother-cell compartments.
Much is known about the transcription factors that drive the process of spore formation, and in several cases transcriptional profiling has been carried out to catalog genes switched on or switched off by individual sporulation regulatory proteins (Fawcett et al. 2000; Britton et al. 2002; Eichenberger et al. 2003; Feucht et al. 2003; Molle et al. 2003a). Here we have attempted to go a step further by comprehensively elucidating the program of gene expression for a single cell type in the developing sporangium. For this purpose we focused on the mother cell and its 5-h program of gene expression. Gene expression in the mother cell is governed by five positively and/or negatively acting transcription factors. These are the sigma factors $\sigma^K$ and $\sigma^K$ and the DNA-binding proteins GerE, GerR (newly characterized in the present study), and SpoIIID.

The appearance of these regulatory proteins is governed by a hierarchical regulatory cascade of the form: $\sigma^K \xrightarrow{\text{SpoIIID/ GerR}} \sigma^K \xrightarrow{\text{GerE}}$ (Figure 1A) in which $\sigma^K$ is the earliest-acting factor specific to the mother-cell line of gene expression (Zheng and Losick 1990; results presented herein). The $\sigma^K$ factor is derived from an inactive proprotein, pro-$\sigma^K$ (LaBell et al. 1987), whose synthesis commences before asymmetric division (Satola et al. 1992; Baldus et al. 1994), but whose continued synthesis becomes strongly biased to the mother cell after asymmetric division (Fujita and Losick 2002 2003). Proteolytic conversion to mature $\sigma^K$ takes place just after asymmetric division (Stragier et al. 1988) and is triggered by an intercellular signal transduction pathway involving a secreted signaling protein that is produced in the forespore under the control of the forespore-specific transcription factor $\sigma^E$ (Hofmeister et al. 1995; Karow et al. 1995; Londono-Vallejo and Stragier 1995). Transcriptional profiling has established that $\sigma^K$ turns on an unusually large regulon consisting of 262 genes, which are organized into 163 transcription units (Eichenberger et al. 2003; results presented herein). Among the targets of $\sigma^K$ are the genes for the DNA-binding proteins SpoIIID and GerR (Kunkel et al. 1989; Stevens and Errington 1990; Tatti et al. 1991; Wu and Errington 2000; results presented herein). SpoIIID is both a negatively acting protein that switches off the transcription of certain genes that have been activated by $\sigma^K$ and a positively acting protein that acts in conjunction with $\sigma^K$-containing RNA polymerase to switch on additional genes, including genes involved in the appearance of $\sigma^K$ (Kroos et al. 1989).

The appearance of $\sigma^K$ is a critical control point that involves multiple levels of regulation: transcription, DNA recombination, and proprotein processing. SpoIIID both activates the transcription of the 5′ coding region for $\sigma^K$ (spoIVCB) and that for a site-specific DNA recombinase (spoIVCA) (Kunkel et al. 1990; Halberg and Kroos 1994) that joins the 5′ coding sequence to the 3′ coding region by the excision of an intervening sequence of 48 kb called skin (Stragier et al. 1989). Finally, the product of the intact coding sequence is an inactive proprotein, pro-$\sigma^K$ (Kroos et al. 1989), whose conversion to mature $\sigma^K$ (as in the case of pro-$\sigma^K$) is governed by a complex, intercellular signal transduction pathway involving a secreted signaling protein that is produced in the forespore under the control of the forespore-specific transcription factor $\sigma^E$ (Cutting et al. 1990, 1991a; Lu et al. 1990). The signal transduction pathway helps to coordinate the appearance of $\sigma^K$ in the mother cell with the timing of events taking place in the forespore. The $\sigma^K$ factor turns on an additional gene set that includes the gene for GerE (Cutting et al. 1989), a DNA-binding protein that is responsible for activating the final temporal class of genes in the mother-cell line of gene expression (Zheng et al. 1992).

Other than the case of $\sigma^K$, little was previously known about the full set of genes, whose transcription is governed by the five regulators in the mother-cell line of gene expression—indeed, nothing at all in the case of GerR, whose function had previously been uncharacterized. Here we...
present evidence indicating that the program of mother-cell-
specific gene transcription involves the activation of at least 383 genes (242 transcription units), representing 9% of the
genes in the B. subtilis genome. We explain the pattern of
transcription of each of these genes in terms of the action of
the five regulatory proteins that govern the mother-cell
program of gene transcription. Our results reveal that the
program chiefly consists of a series of pulses in which large
numbers of genes are turned on and are then turned off
shortly thereafter by the action of the next regulatory protein
in the hierarchy. Evidence is also presented that this
repression is critical for proper morphogenesis. Finally, we
show that the mother-cell program of gene transcription can
be understood in terms of a simple regulatory circuit
involving a linked series of feed-forward loops (FFLs) that
are responsible for generating pulses of gene transcription.
We propose that this regulatory circuit will serve as a model
for understanding other programs of cellular differentiation.

Results
Transcriptional Profiling

Our strategy for elucidating the mother-cell program of
gene transcription was to carry out transcriptional profiling
at hourly intervals during sporulation at 37 °C, starting just
after asymmetric division and ending before the time at
which lysis of the mother cell had commenced. At each time
point, RNA from cells mutant for the transcriptional
regulator that was maximally active at that time interval
was compared against RNA from cells mutant for the next
transcription factor in the hierarchy or, in the case of the last
regulatory protein in the hierarchy, GerE, against RNA from
wild-type cells. Thus, at hour 2.5, RNA from cells mutant for
σ^E (strain PE437) was compared against RNA from cells
(strain PE436) that were wild type for σ^E but mutant for the
next regulatory protein in the sequence, SpoIIID. Likewise, at
hour 3.5, RNA from cells that were mutant for SpoIIID (strain
PE456) was compared against RNA from cells that were
mutant for σ^K (strain PE452). (Strains PE456 and PE452 were
additionally mutant for σ^C to eliminate indirect effects of the
presence or absence of SpoIIID on the activity of the
forespore-specific transcription factor. Although SpoIIID
has no direct effect on σ^C, the absence of negative feedback
on several σ^K-controlled genes [see below] in the strain
mutated for spoIIID could have had indirect consequences on
σ^C activity.) Likewise, at hour 4.5, RNA from cells that were
mutant for σ^K (strain PE455) was compared against RNA
from cells mutant for GerE (strain PE454). Finally, at hours
5.5 and 6.5, RNA from cells mutant for GerE was compared
against RNA from wild-type cells (PY79). Three
transcriptional-profiling analyses were carried out for each of these
time points, using three independent preparations of RNA
from each of the two cultures of cells that were being
compared against each other. The complete dataset for these
experiments is presented in Table S1, and transcriptional
profiles for representative genes are displayed in Table 1.

In addition to the four previously known members of the
hierarchical regulatory cascade, one of the genes in the σ^E
regulon is inferred to encode a previously uncharacterized
DNA-binding protein YlbO (Wu and Errington 2000; Eichen-
berger et al. 2003). Additional transcriptional-profiling

<table>
<thead>
<tr>
<th>Table 1. Transcriptional Profile of Representative Genes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gene</td>
</tr>
<tr>
<td>------</td>
</tr>
<tr>
<td>asnO</td>
</tr>
<tr>
<td>cotA</td>
</tr>
<tr>
<td>cotB</td>
</tr>
<tr>
<td>cotD</td>
</tr>
<tr>
<td>cotE</td>
</tr>
<tr>
<td>cotH</td>
</tr>
<tr>
<td>cwiH</td>
</tr>
<tr>
<td>glnQ</td>
</tr>
<tr>
<td>spollD</td>
</tr>
<tr>
<td>spollM</td>
</tr>
<tr>
<td>spollAAA</td>
</tr>
<tr>
<td>spolVCA</td>
</tr>
<tr>
<td>spolVCB</td>
</tr>
<tr>
<td>spsG</td>
</tr>
</tbody>
</table>

^a Ratios of relative RNA levels in sigE mutant versus sigE mutant.
^b Ratios of relative RNA levels in spollD versus spollD mutant.
^c Ratios of relative RNA levels in gerR versus gerR mutant.
^d Ratios of relative RNA levels in sigK mutant versus sigK mutant.
^e Ratios of relative RNA levels in wild type versus gerE mutant.
DOI: 10.1371/journal.pbio.0020328.e001
experiments were carried out to assess the function of this putative regulatory protein.

**Updating the σ^E Regulon**

We previously reported that the σ^E regulon is composed of 253 genes, organized in 157 transcription units. Since then two additional σ^E-controlled genes, yjC4 (Kuwana et al. 2003) and cfpB (yjB) (Pan et al. 2003), have been identified. These genes were found to be transcribed in a σ^E-dependent manner during sporulation in our previous analysis, but they were not significantly induced in cells engineered to produce σ^E during growth and hence had not been included in our original list of σ^E-controlled genes. In addition, results presented here (see below) show that one gene, ypqA, and two operons, yheOP and yitCD, that are chiefly under the control of σ^E, are also transcribed, albeit at a low level, in a σ^E-dependent manner. These and other considerations (see below) bring the current total number of genes in the σ^E regulon to 262 and the total number of transcription units to 163 (Table 2).

This updated description of the σ^E regulon does not include genes and transcription units that are additionally strongly dependent upon SpoIIID for their transcription because our previous transcriptional-profiling experiments were performed with a strain that was mutant for SpoIIID. SpoIIID is a DNA-binding protein that acts in conjunction with σ^E-containing RNA polymerase (Kroos et al. 1989; Kunkel et al. 1989; Halberg and Kroos 1994). Therefore, as a starting point for the present study, we investigated the influence of SpoIIID on the global pattern of σ^E-directed transcription. As we shall see, this analysis revealed ten genes (representing eight transcription units) that were strongly dependent upon SpoIIID for expression and were not expressed under the control of σ^E alone, bringing the present total number of genes in the σ^E regulon to 272 and the total number of transcription units to 171 (Table 2).

**SpoIIID Is Both a Repressor and an Activator of Genes Whose Transcription Is Dependent Upon σ^E**

Transcriptional profiling revealed that SpoIIID had profound effects on the global pattern of σ^E-directed gene transcription. As many as 181 genes were found to be downregulated in the presence of SpoIIID. Of these, 148 had previously been identified as being activated in a σ^E-dependent manner, at least 112 of which (representing 62 transcription units) were bona fide members of the σ^E regulon (that is, they met multiple criteria for being under the direct control of σ^E) (see Table S2). Therefore, a principal function of SpoIIID is to inhibit the transcription of a substantial proportion (greater than 40%) of the genes whose transcription had been activated by σ^E prior to the appearance of SpoIIID. Members of the σ^E regulon that are downregulated by SpoIIID are colored green in Figure 2A.

SpoIIID not only repressed many genes in the σ^E regulon but also stimulated or activated the transcription of many others. At least 70 genes were identified whose transcription was upregulated by SpoIIID (Table S2), but in many cases these genes were not members of the σ^E regulon, and the effect of SpoIIID could have been indirect. Examples are seven genes (cysK, cysH, cysF, sat, cysG, yoaD, and yoaB) from the S-box regulon (Grundy and Henkin 1998) and two genes (argC and argF) from the arginine biosynthesis operon (Smith et al. 1989). In other cases, however, SpoIIID stimulated or activated the transcription of genes that had been reported to be under the control of σ^E. Thus, 13 (asnO, cwlJ, proH, proJ, spoIVCA, spoIVCB, spoVK, yheB, yheC, yheD, yknT, yknU, and yknV) of the genes whose transcription was upregulated by SpoIIID had previously been assigned to the σ^E regulon, and the dependence on SpoIIID was almost complete, whereas in the other 15 the dependence was partial.

One additional eight genes (cotF, cotT, cotV, cotW, lip, ydeI, yheL, and yheH) that were almost completely dependent on SpoIIID for their transcription and that are likely to be under the dual control of σ^E and SpoIIID. Thus, in addition to repressing at least 112 members of the σ^E regulon, SpoIIID activates the transcription of 25 other members of the regulon, representing 19 transcription units. The 15 σ^E-transcribed genes (11 transcription units) whose expression was partially dependent upon SpoIIID are indicated in orange in Figure 2B, and those whose expression was completely dependent on the DNA-binding protein are indicated in red (ten genes; eight transcription units).

Evidently, then, SpoIIID plays a pivotal role in the mother-cell line of gene expression, negatively or positively affecting the transcription of many members of the σ^E regulon. It was

---

**Table 2. Genes Activated in the Mother-Cell Line of Gene Expression**

<table>
<thead>
<tr>
<th>Category (Transcription Units)</th>
<th>Total Genes (Transcription Units)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>σ^E-Controlled Genes</strong></td>
<td>272 (171)</td>
</tr>
<tr>
<td>Activated by σ^E in the absence of SpoIIID</td>
<td>262 (163)</td>
</tr>
<tr>
<td>Repressed by GerR^a^</td>
<td>14 (10)</td>
</tr>
<tr>
<td>Repressed by SpoIIID</td>
<td>112 (62)</td>
</tr>
<tr>
<td>or GerR^B^</td>
<td>126 (90)</td>
</tr>
<tr>
<td>Activated by GerR^B^</td>
<td>0</td>
</tr>
<tr>
<td>Activated by SpoIIID^C^</td>
<td>10 (8)</td>
</tr>
<tr>
<td><strong>σ^K-Controlled Genes</strong></td>
<td>111 (71)</td>
</tr>
<tr>
<td>Activated by σ^K in the absence of GerE</td>
<td>75 (44)</td>
</tr>
<tr>
<td>Repressed by GerE^C^</td>
<td>55 (36)</td>
</tr>
<tr>
<td>Unaffected by GerE^C^</td>
<td>17 (12)</td>
</tr>
<tr>
<td>Activated by GerE^C^</td>
<td>36 (27)</td>
</tr>
<tr>
<td>Total</td>
<td>383 (242)</td>
</tr>
</tbody>
</table>

^a^ Includes four genes (two transcription units) that were also repressed by SpoIIID and one gene (one transcription unit) that was also activated by SpoIIID.

^b^ Only includes genes that were strongly dependent upon SpoIIID for expression. Fifty-seven genes (11 transcription units) that were partially dependent on SpoIIID are in the “activated by σ^K” category.

^c^ Only includes genes that were strongly dependent upon GerE for expression. Twenty-eight genes (12 transcription units) that were partially dependent on GerE for expression are in the “activated by σ^K” category.

^d^ Numbers include genes and transcription units that were transcribed under the control of σ^K as well as σ^E.

DOI: 10.1371/journal.pbio.0020328.002

---

PLOS Biology | www.plosbiology.org | October 2004 | Volume 2 | Issue 10 | e3281667
Figure 2. Location of Genes in the σ^E and σ^K Regulons and Their Regulation by DNA-Binding Proteins

(A) The σ^E regulon and its modulation by SpoIIIΔ and GerR. The first gene of each σ^E-controlled transcription unit identified by transcriptional profiling is indicated. In the inner circle, genes repressed by SpoIIIΔ are green, and genes repressed by GerR are blue. In the outer circle, genes partially dependent on SpoIIIΔ for expression are orange, and genes strongly dependent on SpoIIIΔ are red. Underlined are SpoIIIΔ-controlled genes for which SpoIIIΔ binding to their upstream sequences has been demonstrated biochemically. Genes unaffected by SpoIIIΔ or GerR are indicated in black.

(B) The σ^K regulon and its modulation by GerE. The first gene of each σ^K-controlled transcription unit identified by transcriptional profiling is indicated. In the inner circle, genes repressed by GerE are green. In the outer circle, genes partially dependent on GerE for expression are orange, and genes strongly dependent on GerE are red. Genes unaffected by GerE are indicated in black.

DOI: 10.1371/journal.pbio.0020328.g002
Biochemical Identification of SpoIIID-Binding Sites

We selected 18 of the newly identified SpoIIID-regulated genes for EMSA analysis, mostly on the basis of the importance of their role in sporulation. As positive controls, we subjected two previously known targets of SpoIIID, \( spoIID \) and \( spoIVCA \) (Halberg and Kroos 1994), to EMSA analysis, and as negative controls three Spo0A-regulated genes (Molle et al. 2003a), \( abrB \) and \( racA \), and \( spoIIA \) (Figure 3A). SpoIIID exhibited binding to the upstream sequence of all 18 of the selected genes (Figure 3B). In some cases (those of \( annO \), \( gerM \), \( spoIVA \), \( spoIVFA \), \( ybaN \), \( ycgF \), \( yitE \), \( ykoU \), and \( ylfy \)) additional shifted bands were detected at high concentrations of SpoIIID, which may indicate the presence of two or more SpoIIID-binding sites with distinct binding affinities.

In addition, we also subjected the upstream region of \( cotE \) to EMSA analysis (Figure 3C). The \( cotE \) gene is transcribed from two promoters: a \( \sigma^E \)-controlled promoter called P1 and a second promoter called P2 that strongly depends on SpoIIID (Zheng and Losick 1990). It had been assumed that transcription from P2 is under the dual control of \( \sigma^E \) and SpoIIID, but EMSA analysis failed to reveal a binding site for SpoIIID, and other work presented below indicates that transcription from \( cotE \) P2 is governed by \( \sigma^E \) rather than by \( \sigma^E \). We conclude that the SpoIIID dependence of \( cotE \) P2 is an indirect consequence of the dependence of \( \sigma^E \) synthesis on SpoIIID.

To obtain further evidence for direct interaction by SpoIIID and to investigate the mechanism by which SpoIIID inhibits transcription, we subjected the promoter regions of three genes (\( spoIID \), \( spoIIAA \), and \( spoVE \)) identified as being under the negative control of SpoIIID to DNase I footprinting analysis. SpoIIID protected two regions in the upstream sequence of \( spoIID \) from DNase I digestion (Figure S1). One region (extending from positions –10 to –28 on the top strand and from –18 to –35 on the bottom strand) overlapped with the –10 element of the \( \sigma^E \) promoter, and the other (extending from –33 to –52 on the top strand) overlapped with the –35 element. The binding site for SpoIIID also overlapped with the promoter in the case of \( spoIIAA \), in this case protecting a single sequence that included the –35 element (extending from –21 to –45 on the top strand and from –30 to –48 on the bottom strand). Finally, the regulatory sequence of \( spoVE \) exhibited two binding sites, one (extending from +16 to –1 on the bottom strand) that was located in the vicinity of the predominant \( \sigma^E \)-controlled promoter (P2) for this gene and another further upstream, overlapping with a secondary promoter (P1) (extending from +13 to –7 on the top strand). Thus, repression of the promoters of \( spoIID \), \( spoIIAA \), and \( spoVE \) by SpoIIID is likely to be a direct consequence of the binding of the sporulation regulatory protein to the promoter in such a way as to compete with binding by \( \sigma^E \)-RNA polymerase.

SpoIIID Binds to Some Sites that Do Not Correspond to Genes under Its Control

ChiP-on-chip analysis was carried out as described in Materials and Methods and previously (Molle et al. 2003a, 2003b), using DNA–protein complexes from formaldehyde-treated cells at hour 3 of sporulation. After sonication, SpoIIID–DNA complexes were precipitated with antibodies against SpoIIID. Next, after reversal of the cross-links, the precipitated DNAs were amplified by PCR in the presence of cyanine 5-dUTP. In parallel, total sonicated DNA from the formaldehyde-treated cells (i.e., DNA that had not been subjected to immunoprecipitation) was similarly amplified, but in the presence of cyanine 3-dUTP. The two differentially labeled DNAs were combined and hybridized to the same batch of DNA microarrays that were used for the transcriptional-profiling experiments. Transcriptional profiling was carried out with three independent preparations of formaldehyde-treated cells, twice with two of the preparations and once with the third, for a total of five analyses. An enrichment factor was calculated for each gene, representing the enrichment of that gene by immunoprecipitation relative to DNA that had not been subjected to immunoprecipitation, and the entire dataset is displayed in Table S3.

Thirty-one genes, corresponding to 26 regions of the chromosome, were found to be enriched by immunoprecipitation by a factor of two or greater. Only seven of the regions (\( cotF \), \( lip \), \( spoIIAF \), \( spoVD \), \( ycgF \), \( yhbH \), and \( ykcl \)) identified by the ChiP-on-chip analysis were in close proximity to a gene that was differentially expressed in the SpoIIID transcriptional-profiling experiments. Thus, in only a small number of cases did ChiP-on-chip analysis support the idea that a gene under SpoIIID control was a direct target of the DNA-binding protein. Our interpretation of these findings is that ChiP-on-chip is less sensitive for detecting SpoIIID-binding sites than it is for the B. subtilis DNA-binding proteins CodY (Molle et al. 2003b), Spo0A (Molle et al. 2003a), and RacA (Ben-Yehuda et al. 2003). Likely contributing to this decreased sensitivity is the fact that SpoIIID is present in only one of the two chromosome-containing compartments (the mother cell) of the sporangium and that its concentration is low (~1 mM; Zhang et al. 1997).

While providing support for only a small proportion of the herein identified targets of SpoIIID regulation, ChiP-on-chip analysis, nonetheless, proved to be revealing. Specifically, we found that SpoIIID bound to many regions of the chromosome that did not correspond to genes under its negative or positive control. Were these regions bona fide SpoIIID-binding sites? To address this question, we subjected five regions that were most enriched for SpoIIID-binding (\( abfE–abfF \), \( detR–detP \), \( tenL–gexB–thiS \), \( treA–treR–yfhO \), and \( yfmC–yfmD \)) to EMSA analysis (Figure 3D). Given that SpoIIID was not exerting a transcriptional effect in these regions, we reasoned that the sites to which SpoIIID was binding might not reside in upstream regulatory regions and could instead be located in coding sequences. We therefore scanned across each of the five chromosomal regions by EMSA using successive DNA fragments of about 400 bp in length. The results showed that each of the five regions contained more than one binding site for SpoIIID and that some of these binding sites were indeed located within protein-coding sequences. (The presence of more than one binding site in each region may have
facilitated their detection by the ChIP-on-chip analysis.) We conclude that SpoIIID binds to some sites on the chromosome at which it does not function as a transcriptional regulator. Conceivably, it plays an architectural role in the folding of the chromosome in the mother cell in addition to its role as a transcriptional regulator. Moqtaderi and Struhl (2004) have similarly found that in Saccharomyces cerevisiae the RNA polymerase III transcription factor TFIIIC binds to sites where binding of other components of the RNA polymerase III machinery is not detected and where the transcription factor does not activate transcription.

Identification of Putative SpoIIID-Binding Sites by Bioinformatics

As a final, computational approach to identifying direct targets of SpoIIID, we used the Gibbs sampling algorithm BioProspector to identify conserved motifs in sequences upstream of genes under the control of SpoIIID (Liu et al. 2001). Initially, we limited our search to 40 regions where SpoIIID binding had been confirmed by biochemical analysis. BioProspector was used to find the best 35 motifs across several different widths (6–12 bp) under the restriction that every sequence had to contain at least one site. Each of these motifs was separately used as a starting point for BioOptimizer (Jensen and Liu 2004) and applied to an expanded dataset that included the 89 upstream sequences for all SpoIIID-controlled genes (not just those analyzed by EMSA or footprinting). BioOptimizer optimized both the set of predicted sites and the motif width, as detailed in the Materials and Methods section. BioOptimizer was required to identify at least one binding site in the sequences that had been confirmed by EMSA but was unrestricted for the sequences for which a binding site had not been confirmed biochemically. The optimized motif was 8 bp in length and identified at least one putative SpoIIID-binding site in 60 of the 89 upstream sequences that were analyzed (see Table S2). Figure 4 shows that the logo for the optimized motif (B) was similar to a consensus sequence (A) that was derived independently using 12 previously reported binding sites (for the genes bofA, cotD, spoIVCA, and racA; Halberg and Kroos 1994; Zhang et al. 1997) and five sites herein identified by DNAase I footprinting.

Figure 3. Gel Electrophoretic Mobility-Shift Analysis of SpoIIID Binding

DNA fragments of interest were amplified by PCR, gel-purified, and end-labeled using [γ-32P]-ATP and polynucleotide kinase. Purified SpoIIID was added at increasing concentrations (0 nM for lanes 1 and 5, 50 nM for lane 2, 100 nM for lane 3, and 200 nM for lane 4) and incubated at room temperature for 30 min before loading on to a non-denaturing gel containing 6% polyacrylamide. With the exception of (D), the DNA fragments corresponded to the upstream regions of the indicated genes. See Materials and Methods for the identity (coordinates) of the specific DNA sequences used in the analyses.

(A) Gel shifts for known targets of SpoIIID (bofA and spoIVCA), representing positive controls, and genes (abrB, spoIIGA, and racA) under the control of another DNA-binding protein (Spo0A), representing negative controls.

(B) Gel shifts for genes identified as possible targets of SpoIIID by transcriptional profiling.

(C) Gel shift for cotE. Expression of cotE from its P2 promoter is strongly dependent on SpoIIID. No binding of SpoIIID to the upstream sequence for cotE is observed, suggesting that the effect of SpoIIID on transcription from the P2 promoter is indirect.

(D) Gel shifts for chromosomal regions strongly enriched for SpoIIID binding as judged by ChIP-on-chip analysis. For each region, four consecutive DNA fragments of approximately 400 nucleotides in length were analyzed.

DOI: 10.1371/journal.pbio.0020328.g003
identified by ChIP-on-chip analysis, which likely represent the strongest binding sites for SpoIIID. We used Motif Discovery scan (MDscan) (Liu et al. 2002) for this analysis, which is designed to identify conserved motifs in sequences that have been ranked according to their enrichment factor in ChIP-on-chip experiments. The resulting sequence logo is displayed in Figure 4C. Whereas it is largely similar to that obtained from the BioProspector/BioOptimizer analysis (Figure 4B), there is one notable difference: The first position of the binding motif corresponds almost exclusively to a guanine in the sites identified by ChIP-on-chip analysis. The presence of a guanine at this position could be characteristic of high-affinity sites for SpoIIID binding.

In conclusion, SpoIIID negatively or positively influences the transcription of over half of the members of the $\sigma^E$ regulon, and a combination of complementary approaches leads us to believe that it does so for many of the genes so identified by direct interaction with their promoter regions. In the case of genes under the negative control of SpoIIID, the mechanism of this repression probably involves steric interference as the inferred binding sites for SpoIIID were generally found to overlap with the expected binding sites for RNA polymerase. No such overlap was generally observed in the case of genes under the positive control of SpoIIID.

GerR ($ylbO$), a Second Negative Regulator of the $\sigma^E$ Regulon

The spoIIID gene is not the only member of the $\sigma^E$ regulon that appears to encode a DNA-binding protein. The inferred product of $ylbO$ exhibits significant similarity to members of the basic leucine zipper family of transcription factors and is, in particular, 52% similar to RsfA (Wu and Errington 2000), a regulator of $\sigma^E$-controlled genes in the forespore line of gene expression. To study a possible role for $ylbO$ we investigated the effect of a null mutation of the gene on sporulation and on $\sigma^E$-directed gene expression. As noted previously, the mutation has no effect on the production of heat-resistant spores, but we have now discovered that the mutation causes a conspicuous defect in the capacity of the spores to germinate, as judged by their impaired ability to reduce 2,3,5-triphenyltetrazolium chloride (see Materials and Methods). We therefore rename $ylbO$ as gerR (in keeping with the nomenclature for germination genes in B. subtilis [Setlow 2003]). We also carried out transcriptional profiling using RNA collected at hour 3.5 of sporulation from cells of a strain (PE454) that was wild type for GerR and from cells of a newly constructed strain (SW282) that was mutant for GerR. Both strains were also mutant for the next transcription factor in the hierarchical cascade, $\sigma^R$. No genes were identified whose transcription was dependent on GerR, but 139 genes were found that were downregulated in a GerR-dependent manner by a factor of two or greater (see Table S1). Among the downregulated genes were 14 members of the $\sigma^E$ regulon. Nine of these members (colored blue in Figure 2A) were known not to be under SpoIIID control (cypA, kapD, spoIIA, spoIII, spoIIP, ybaS, yfnE, yfnD, yhfJ, and yhfV), whereas the remaining five (phoB, spoHAA, spoIIAB, spoIVC, and ylbO) were also under the control of SpoIIID.

We selected three of the putative targets of GerR for further analysis. The promoter sequences of spoIIA and yhfV were fused to the coding sequence of $\beta$-galactosidase and introduced into the chromosome at the amyE locus and a previously constructed fusion of lacZ to spoIIP (amyE::spoIIP-lacZ) was obtained from P. Stragier (Institut de Biologie Physico-Chimique, Paris). The results, shown in Figure 5, confirmed that GerR had a pronounced negative effect on the level of expression of all three fusions.

An example of $\sigma^E$-controlled genes that are under the dual negative control of GerR and SpoIIID is the eight-cistron spoIII operon (Illing and Errington 1991). As we have demonstrated, GerR is responsible for repressing yqhV, which is located just upstream of the spoIII operon. Given the absence of an apparent transcriptional terminator at the end of the gene, $\sigma^E$-directed transcription from yqhV is likely to read into spoIII, which is also transcribed from its own $\sigma^E$-controlled promoter located in the intergenic region between yqhV and the operon. Thus, by repressing yqhV, GerR would inhibit read-through transcription into spoIII. Indeed, our transcriptional-profiling analysis revealed a small negative effect of GerR on spoIII transcription. Meanwhile, SpoIIID acts at the promoter for the spoIII operon to inhibit it from being used by $\sigma^E$-RNA polymerase. Thus, maximum repression of spoIII is evidently achieved by the combined action of GerR and SpoIIID, each acting to block different promoters.

Finally, we note that GerR inhibited the expression of a large number of genes that do not belong to the $\sigma^E$ regulon. Interestingly, many of these genes are organized in large clusters, such as aclB–aclC–aclD–bmrQ–yrdK–gilR, albA–albB–albC–albD, yefA–yefB–yefC–yepA–yepB–yepC, yncM–ynpN–ynpO, and yydB–yydC–yydD–yydG–yydH–yydJ. The genes found in these clusters rarely belong to a single transcription unit and are sometimes transcribed in opposite directions (either convergently or divergently).

In summary, transcription of genes in the $\sigma^E$ regulon is in part self-limiting. The $\sigma^E$ factor induces the synthesis of two proteins, GerR and SpoIIID, that act to switch off other genes in the regulon, thereby preventing their continued transcription during the next stage of the mother-cell line of gene expression.

The $\sigma^K$ Regulon

Next, we used two complementary transcriptional-profiling approaches to identify genes under the control of $\sigma^K$, an RNA polymerase sigma factor that follows SpoIIID in the hierarchical regulatory cascade. In one approach, we sought to identify genes that were upregulated during sporulation in a $\sigma^K$-dependent (but not a GerE-dependent) manner. In the other approach, we sought to identify genes whose transcription was artificially activated in cells engineered to produce $\sigma^K$ during growth. For this approach we used a strain in which the coding sequence for the mature form of the transcription factor ($\sigma^K$ is normally derived by proteolytic processing from an inactive proprotein [Kroos et al. 1989]) was under the control of an inducible promoter (see Materials and Methods). Ninety-five genes were identified that were induced both during growth and sporulation in a $\sigma^K$-dependent manner. Eight additional genes (cotA, cotE, cotM, gerE, gerPA, yfhP, yjz, and ykuD) that had previously been assigned to the regulon on the basis of gene-specific analysis were added to the tally, bringing the total to 103 (and representing 63 transcription units). These eight genes were cases in which we did not obtain a statistically significant score in one or the other of the two transcriptional-profiling approaches or for which a signal was not obtained for technical reasons (e.g., the strain used was mutant for gerE and
yjcZ had not been annotated when the arrays were built). The list of 103 did not include σK-controlled genes whose transcription additionally and strongly required the DNA-binding protein GerE. Some (28) of these 103 genes were also transcribed under the control of σE (see Table 2), leaving a total of 75 genes that were newly activated during sporulation under the control of σK. As we shall see, when genes that were strongly dependent on GerE are included (41 genes, five of which were also expressed under the control of σE), the size of the regulon increases to 144 genes (103 + 41) organized in 94 transcription units (Table 2). A map of the σK regulon is displayed in Figure 2B and a detailed list of the genes in the regulon is presented in Table S4.

Identification of Promoters Controlled by σK Using Bioinformatics and Transcriptional Start Site Mapping

As a further approach to assessing our assignments to the σK regulon, we used BioProspector and BioOptimizer to obtain a consensus sequence for promoters under the control of the sporulation transcription factor. The computational approach was complicated by the fact that the program had to find a two-block motif, with the first block corresponding to the −35 element and the second block to the −10 element separated by a gap of fixed length (±1 bp). The optimal spacing between the two regions is 15 bp (±1 bp). The consensus binding sequence for σK-containing RNA polymerase obtained from the compilation of 58 sequences identified as common motifs in regions upstream of σK-regulated genes by a BioProspector/BioOptimizer computational approach (Jensen and Liu 2004). Positions 1–5 on the horizontal axis correspond to the −35 element and positions 21–30 to the −10 element. (E) Consensus promoter sequence for σK-containing RNA polymerase obtained from the compilation of 23 previously mapped (http://dbtbs.hgc.jp; Helmann and Moran 2002) and 18 newly identified σK-controlled promoters identified by transcription start site mapping.

DOI: 10.1371/journal.pbio.0020328.g004

Figure 4. Consensus Sequences for SpoIIID, σK, and σE

Consensus sequences are displayed as sequence logos (Schneider and Stephens 1990). The height of the letters in bits represents the information content at each position (the maximum value is two bits).

(A) Consensus binding sequence for SpoIIID as derived from 17 SpoIIID-binding sites mapped by DNAase I footprinting (Halberg and Kroos 1994; Zhang et al 1997; results presented herein).

(B) Consensus binding sequence for SpoIIID obtained by compilation of 68 putative SpoIIID-binding sites identified as common motifs by BioProspector and BioOptimizer analysis in sequences upstream of genes identified by transcriptional profiling or within regions identified by ChiP-on-chip analysis.

(C) Consensus binding sequence for SpoIIID obtained by MDiscan analysis of the sequences of 26 SpoIIID-binding regions identified by ChiP-on-chip analysis.

(D) Consensus promoter sequence for σK-containing RNA polymerase obtained from the compilation of 58 sequences identified as common motifs in regions upstream of σK-regulated genes by a BioProspector/BioOptimizer computational approach (Jensen and Liu 2004). Positions 1–5 on the horizontal axis correspond to the −35 element and positions 21–30 to the −10 element. The optimal spacing between the two regions is 15 bp (±1 bp).

(E) Consensus promoter sequence for σK-containing RNA polymerase obtained from the compilation of 23 previously mapped (http://dbtbs.hgc.jp; Helmann and Moran 2002) and 18 newly identified σK-controlled promoters identified by transcription start site mapping.

(F) Consensus promoter sequence for σE-containing RNA polymerase obtained from the compilation of 62 σE-controlled promoters identified by transcription start site mapping (Eichenberger et al. 2003). Positions 1–8 on the horizontal axis correspond to the −35 element, and positions 21–30 to the −10 element. The optimal spacing between the two regions is 12 bp (±1 bp).
The Mother-Cell Line of Gene Expression

Figure 5. Repression of σ^k-CONTROLLED Genes by GerR
Culture samples from strains PE551 (solid triangles, amyE::P_{amyR}::lacZ), SW312 (open triangles, amyE::P_{amyR}::lacZ, ΔgerR), PE511 (solid squares, amyE::P_{amyR}::lacZ), PE568 (open squares, amyE::P_{amyR}::lacZ, ΔgerR), PE553 (solid diamonds, amyE::P_{amyR}::lacZ), and PE558 (open diamonds, amyE::P_{amyR}::lacZ, ΔgerR) were collected at indicated intervals after the start of sporulation in Sterlini–Mandelstam medium and analyzed for β-galactosidase activity.
DOI: 10.1371/journal.pbio.0020328.g005

Consensus sequence corresponding to a total of 41 promoters (Figure 4E). The logo for σ^k promoters whose start sites had been mapped was very similar to the logo obtained by the BioProspector/BioOptimizer procedure (see Figure 4D). Moreover, out of the 41 confirmed σ^k promoters, the correct promoter was identified in 24 cases, with no prediction being made in 15 cases and an incorrect prediction in just two cases. All of the predicted sites are listed in Table S4.

The σ^k and σ^K factors are highly similar to each other, and the promoters they recognize are also very similar. The availability of updated logos for both categories of promoters based on the nearly complete regulons for both regulatory proteins provided an opportunity to revisit the issue of how the two regulatory proteins discriminate between their two classes of cognate promoters. A comparison of the motif recognized by σ^k to that recognized by σ^K (Figure 4F) reveals that both classes of promoters share identical −10 sequences and that the −35 elements differ by a single base pair: a cytosine in the fourth position of σ^k-controlled promoters versus a thymine at the corresponding position in σ^K-controlled promoters. These results reinforce the findings of Tatti et al. (1995) who identified glutamine 217 of σ^k as the contact residue for the base pair at position 4. The two proteins are identical to each other in the region inferred to interact with the −35 element except for the presence of arginine instead of glutamine at the corresponding position in σ^K. Moreover, replacing glutamine 217 with arginine was found to confer on σ^k the capacity to recognize σ^K-controlled promoters (Tatti et al. 1995). The high similarity between the two classes of promoters also helps to explain why some σ^K-controlled promoters are also recognized by σ^k, but our bioinformatics analysis does not allow us to explain why some promoters are recognized exclusively by one or the other sigma factor and others are not.

GerE Is Both a Repressor and an Activator of Genes Whose Transcription Is Dependent upon σ^K

The last regulator in the mother-cell line of gene expression is the DNA-binding protein GerE (Cutting et al. 1989). Genes under GerE control were identified by transcriptional-profiling experiments carried out at two times (5.5 h and 6.5 h) late in sporulation. Strikingly, as many as 209 genes were downregulated in the presence of GerE at one or both time points, with many more genes being downregulated at the later time point (201 versus 61; see Table S1). Some of these downregulated genes (55) were members of the σ^K regulon, with 29 being downregulated at the earlier time point and an additional 26 at the later time point. Thus, GerE is responsible for inhibiting the expression of 55% of the genes in the σ^K regulon, but its repressive effects are not limited to genes under σ^K. We note that the gene coding for σ^K is itself repressed by GerE, which would be expected to curtail further synthesis of the mother-cell sigma factor late in sporulation. Thus, GerE has a wide impact in inhibiting gene transcription late in the process of spore maturation, including many genes in the preceding regulon of σ^K-activated genes.

At the same time, GerE is also an activator that stimulated or switched on the transcription of as many as 65 genes by hour 5.5 and 71 genes by hour 6.5. Of these, 41 were strongly dependent upon GerE for their expression and hence were not identified as members of the σ^K regulon. Leaving aside genes that were members of both the σ^k and σ^K regulons (five), we see that GerE is responsible for turning on an additional 36 genes (27 transcription units) in the final phase of the mother-cell line of gene expression (Table 2).

Evidence that SpoIIID-Mediated Repression Is Required for Sporulation

As we have seen, a striking feature of the mother-cell line of gene expression is that many of the genes activated by one transcription factor are turned off by the next-appearing regulatory protein in the cascade. Thus, most of the genes that are turned on by σ^k are subsequently repressed by GerR or SpoIIID. Likewise, many of the genes activated by σ^k are, in turn, downregulated by GerE. In the case of GerR, a mutant lacking the regulatory protein produced spores that were defective in germination. Hence, proper morphogenesis depends on the capacity of GerR, which appears to act exclusively as a repressor, to turn off genes under its control.

The case of SpoIIID is more complex because in addition to its role as a repressor this DNA-binding protein is also an activator of two genes, spoIVCA and spoIVCB, that are essential for sporulation because of their role in the synthesis of σ^K (Halberg and Kroos 1994). To investigate the role of SpoIIID-mediated repression in spore formation, we created a construct in which a copy of the intact pro-σ^K coding sequence, sigK, was introduced into the amyE locus, thereby bypassing the requirement for the spoIVCA-encoded recombinase, which is normally needed for creating sigK by a chromosomal rearrangement (Stragier et al. 1989), and for spoIVCB, the 5′ portion of the coding sequence that participates in the rearrangement. In our construct, the insertion of sigK at amyE was under the direction of a σ^K-controlled promoter that is not dependent upon SpoIIID for its activation (the promoter for spoVF; Cutting et al. 1991b). The amyE::P_{spoVF}-sigK construct was introduced into spoIVCB mutant cells to create strain BDR1663. Even though pro-σ^K was expected to be synthesized somewhat prematurely in BDR1663, the appearance of mature σ^K remained subject to the pathway governing the proteolytic processing of pro-σ^K and hence would have occurred at the normal time (Cutting...
et al. 1991a). Indeed, cells harboring the amyE::PspoIVF–sigK construct sporulated as efficiently as the wild type and did so in a manner that did not depend on the presence of spoIVCB (Table 3). We conclude that bypassing the requirement for SpoIIID in \( \sigma^E \) synthesis does not measurably affect sporulation efficiency.

However, when the amyE::PspoIVF–sigK construct was introduced into cells harboring a spoIID mutation (generating strain BDR1666), sporulation efficiency was still reduced by about a 100,000-fold compared to the wild type (Table 3). This result reinforces the findings of Lu and Kroos (1994), who showed that sporulation was impaired in spoIID mutant cells even in the presence of a construct that allowed pro-\( \sigma^E \) to be produced in a SpoIIID-independent manner. A possible explanation for these results is that, in addition to its role in \( \sigma^E \) synthesis, SpoIIID is required for the synthesis of some other unidentified protein or proteins that are needed for sporulation. To investigate this possibility, we systematically inactivated all of the newly identified SpoIIID-activated transcription units (Table 3). With three exceptions, those of spoVK, asnO, and ycgM, the resulting mutants sporulated at levels comparable to that of the wild type. In the case of spoVK, asnO, and ycgM, evidence suggests that each is transcribed in both a SpoIIID-dependent and a SpoIIID-independent mode. Thus, spoVK is transcribed from both a \( \sigma^E \)-controlled (P1) and a \( \sigma^K \)-controlled (P2) promoter, and it is known that P1 is dispensable for sporulation (Foulger and Errington 1991). Experiments based on the use of cells engineered to produce \( \sigma^E \) during growth indicate that asnO is capable of being transcribed under the direction of \( \sigma^K \).

Finally, it has been shown that ycgM is induced during the early stages of sporulation under the control of Spo0A (Molle et al. 2003a), and so at least some YcgM protein should be present in a spoIID mutant. Besides, complete inactivation of ycgM resulted in a sporulation defect that is less severe than that observed for strain BDR1666.

These results do not rule out the possibility that SpoIIID activates the transcription of one or more genes in addition to spoIVCA and spoIVCB that are needed for sporulation. Nevertheless, the simplest interpretation of our findings is that the strong sporulation defect of strain BDR1666 is due to a failure in gene turn off rather than gene activation.

**Discussion**

**The Mother-Cell Line of Gene Transcription Is a Hierarchical Regulatory Cascade That Is Subject to Successive Negative Regulatory Loops**

Our results reveal the almost complete program of gene transcription for a single differentiating cell type, the mother-cell compartment of the *B. subtilis* sporangium. The mother cell is a terminally differentiating cell that ultimately undergoes lysis (programmed cell death) when its contribution to the maturation of the spore is complete. Its program of transcription is played out over the course of about 5 h and, as we have shown, involves the activation in a cell-type-specific manner of 383 genes, which are grouped together in 242 transcription units. This corresponds to 9% of the 4,106 annotated protein-coding genes in the *B. subtilis* genome. The transcription of these 383 genes is orchestrated by five developmental regulatory proteins: two RNA polymerase sigma factors, \( \sigma^E \) and \( \sigma^K \); and three DNA-binding proteins, SpoIID, GerE, and a previously uncharacterized regulatory protein, GerR. The five regulatory proteins are organized in a hierarchical regulatory cascade of the form: \( \sigma^E \rightarrow \text{SpoIID}/\text{GerE} \rightarrow \sigma^K \rightarrow \text{GerE} \). The earliest-acting regulatory protein in the cascade, \( \sigma^E \), turns on the transcription of 262 genes (163 transcription units), including the genes for GerR and SpoIIID. GerR and SpoIIID, in turn, acting as repressors, downregulate further transcription of almost half of the genes in the \( \sigma^K \) regulon. In addition, however, SpoIIID, acting in conjunction with \( \sigma^K \)-containing RNA polymerase, turns on the transcription of ten genes (eight transcription units), including genes involved in the appearance of \( \sigma^K \). Next, \( \sigma^K \) activates 75 additional genes (44 transcription units). Among the members of the \( \sigma^K \) regulon is the gene for the final regulatory protein in the cascade GerE. Strikingly, GerE represses the transcription of over half of the genes that have been activated by \( \sigma^K \) while switching on 36 additional genes (27 transcription units), the final temporal class in the mother-cell line of gene transcription. Thus, the program of gene expression is driven forward by its hierarchical organization as well as by the successive, repressive effects of the DNA-binding proteins, which inhibit continued transcription of many genes that had been activated earlier in the

### Table 3. Systematic Inactivation of SpoIIID-Activated Genes

<table>
<thead>
<tr>
<th>Strain</th>
<th>Description</th>
<th>Sporulation Efficiency (Percent)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>PY79</td>
<td>wild type</td>
<td>100</td>
</tr>
<tr>
<td>RL2391</td>
<td>asnO::erm</td>
<td>0.000001</td>
</tr>
<tr>
<td>PE563</td>
<td>cotF::erm</td>
<td>83</td>
</tr>
<tr>
<td>PE549</td>
<td>cotT::erm</td>
<td>100</td>
</tr>
<tr>
<td>PE566</td>
<td>cotW::erm</td>
<td>68</td>
</tr>
<tr>
<td>RL2396</td>
<td>cwlJ::tet</td>
<td>93</td>
</tr>
<tr>
<td>PE548</td>
<td>lip::erm</td>
<td>94</td>
</tr>
<tr>
<td>PE534</td>
<td>mpr::erm</td>
<td>84</td>
</tr>
<tr>
<td>RL2559</td>
<td>proH::erm</td>
<td>75</td>
</tr>
<tr>
<td>RL2519</td>
<td>spoVK::erm</td>
<td>0.000001</td>
</tr>
<tr>
<td>RL3231</td>
<td>ycgM::erm</td>
<td>0.7</td>
</tr>
<tr>
<td>PE569</td>
<td>ydcI::erm</td>
<td>93</td>
</tr>
<tr>
<td>RL2570</td>
<td>yhbB::erm</td>
<td>83</td>
</tr>
<tr>
<td>RL2571</td>
<td>yheC::erm</td>
<td>100</td>
</tr>
<tr>
<td>PE529</td>
<td>yheD::erm</td>
<td>100</td>
</tr>
<tr>
<td>PE570</td>
<td>yhel::erm</td>
<td>95</td>
</tr>
<tr>
<td>RL2393</td>
<td>yknT::erm</td>
<td>81</td>
</tr>
<tr>
<td>RL2581</td>
<td>yknV::erm</td>
<td>100</td>
</tr>
<tr>
<td>PE538</td>
<td>yqfT::erm</td>
<td>69</td>
</tr>
<tr>
<td>PE564</td>
<td>spoIVCA::erm</td>
<td>0.000001</td>
</tr>
<tr>
<td>BDR107</td>
<td>spoIVCB::erm</td>
<td>0.000002</td>
</tr>
<tr>
<td>BDR1663</td>
<td>spoIVCA::erm, amyE::PspoIVF–sigK</td>
<td>96</td>
</tr>
<tr>
<td>BDR362</td>
<td>spoIIID::erm, amyE::PspoIVF–sigK</td>
<td>0.000014</td>
</tr>
<tr>
<td>BDR1666</td>
<td>spoIIID::erm, amyE::PspoIVF–sigK</td>
<td>0.0016</td>
</tr>
</tbody>
</table>

* Sporulation efficiency is defined as the number of heat-resistant spores in a sporulating culture of the mutant strain divided by the number of heat-resistant spores present in a sporulating culture of the wild-type (PY79) strain grown in parallel.

DOI: 10.1371/journal.pbio.0020328.t003
cascade. Indeed, evidence presented herein is consistent with the idea that repression by GerR and SpoIIID contributes to proper sporulation, modestly in the case of GerR, and perhaps more significantly in the case of SpoIIID.

**The Mother-Cell Line of Gene Transcription Is Governed by a Linked Series of Coherent and Incoherent FFLs**

Transcription networks are based on recurring circuit modules, one of the most common of which is the FFL (Milo et al. 2002; Shen-Orr et al. 2002; Mangan and Alon 2003). FFLs are simple circuits involving two regulatory proteins in which one (the primary regulatory protein) governs the expression of the other and both then control the expression of a set of target genes. Certain types of FFLs known as type I are particularly prevalent because of their favorable biological properties (Shen-Orr et al. 2002). In type-I FFLs, the primary regulatory protein acts positively on the synthesis of the second. The mother-cell line of gene transcription is based on two kinds of type-I FFLs known as a “coherent” and “incoherent.” In coherent type-I FFLs, both regulatory proteins act positively on target genes, whereas in incoherent type-I FFLs, the primary regulatory protein acts positively and the second acts negatively.

Using this nomenclature, we see that the hierarchical regulatory cascade that governs the mother-cell line of gene transcription is a circuit composed of two coherent type-I FFLs linked in series (Figure 1B). Thus, σK turns on the synthesis of SpoIIID, and both transcription factors then act jointly to turn on target genes, including genes involved in the appearance of σK. The FFL is acting by the logic of an AND gate in that both σK and SpoIIID are required for the expression of target genes. This first FFL is linked in series to a second coherent type-I FFL, in which σE turns on the synthesis of GerE, and the two transcription factors then collaborate to activate the transcription of target genes (the terminal temporal class of gene transcription in the mother cell). Once again this is an AND gate in that both σK and GerE are required for the activation of target genes. Simulation studies show that coherent type-I FFLs have the property of being persistence detectors in which the activation of target genes depends on the persistence of the primary regulatory protein (σK and σE) and “rejects” situations in which the primary regulatory protein is present only transiently in its active form (Mangan and Alon 2003).

The mother-cell line of gene transcription is also governed by three incoherent type-I FFLs, involving SpoIIID, GerR, and GerE, each acting in this context as repressors. Thus, σE turns on the synthesis of SpoIIID, which in turn represses a subset of the genes that have been turned on by the primary regulatory protein. The σE factor similarly turns on the synthesis of GerR, which then represses a largely nonoverlapping subset of the genes that have been activated by σE. Finally, the σK factor turns on the synthesis of GerE, which then acts to downregulate the transcription of many of the genes that have been switched on by σE. Simulations have shown that incoherent type-I FFLs have the property of producing a pulse of gene transcription (Mangan and Alon 2003). Incoherent type-I FFLs also operate by the logic of an AND gate in that pulses of gene transcription require the action of both the activator and the delayed appearance of the repressor.

Viewing the mother-cell line of gene transcription in terms of an interconnected series of FFLs reveals an underlying logic to the mother-cell program of gene expression. The use of coherent type-I FFLs to drive the activation of successive sets of genes and the ordered appearance of regulatory proteins may help to minimize noise and to ensure that each temporal class of gene activation is tightly tied to the persistence of the previously acting regulatory proteins in the sequence (Mangan et al. 2003). Meanwhile, the use of incoherent type-I FFLs to switch off the transcription of genes in previously activated gene sets helps to generate pulses of gene transcription in which certain genes, whose products may only be required transiently during differentiation, are transcribed over a limited period of time. Indeed, as we now consider, genes with related functions are often transcribed coordinately in a pulse, the timing of which corresponds to the function of their products.

**Coordinated Expression of Functionally Related Genes**

The mother-cell program of gene expression is characterized, as we have seen, by pulses of gene expression in which different sets of genes are successively switched on and then switched off. In some cases, these pulses correspond to the expression of genes with related functions (Table 4). This can be most clearly seen with the gene set that is activated by σK and repressed by SpoIIID or GerR, which includes genes involved in engulfment, cortex formation, and the appearance of σG and σK. Thus, three genes that are responsible for driving engulfment, spoIID (Lopez-Diaz et al. 1986), spoIIM (Smith and Youngman 1993; Smith et al. 1993), and spoIHP (Frandsen and Stragier 1995), are coordinately activated by σK and then repressed by SpoIIID (in the case of spoIID) or by GerR (in the case of the other two). Likewise, all of the σE-controlled genes that are known to be required for spore cortex formation (cadD, dacB–spmA, spoIVA, spoVB, spoVD, spoVE, yabPQ, ykvUV, yhlf, and yqfCD; Pigott and Losick 2002; Eichenberger et al. 2003) are repressed by SpoIIID. Yet another example is the eight-gene spoIHA operon, which is involved in the activation of σE in the forespore (Stragier and Losick 1996). The operon is transcribed from two σE-controlled promoters, one located immediately upstream of the operon and one preceding the next upstream gene, yqfH. As we have shown, both promoters are turned off shortly after their activation; one by SpoIIID and the other by GerR.

Particularly illuminating is the case of the five σE-controlled genes involved in the appearance of σK: bofA, spoIVCA, spoIVCB, spoIVFA, and spoIVFB. Two of these genes (spoIVCA and spoIVCB) are involved in the synthesis of the proprotein precursor, pro-σK, whereas the remaining three (bofA, spoIVFA, and spoIVFB) are involved in the conversion of the proprotein to mature σK (Cutting et al. 1991b; Ricca et al. 1992). Interestingly, bofA, bofK, spoIVFA, and spoIVFB are repressed by SpoIIID, whereas spoIVCA and spoIVCB are switched on by SpoIIID, in this context acting as an activator. Hence, and ironically, genes involved in the processing of pro-σK are expressed in a pulse that precedes the time of activation of the genes involved in the synthesis of the substrate for processing.

How can we explain these seemingly anomalous observations? BofA, SpoIVFA, and SpoIVFB are integral membrane proteins that form a complex in the mother-cell membrane that surrounds the forespore (Resnekov et al. 1996; Rudner and Losick 2002). Evidence indicates that they initially localize to the cytoplasmic membrane that surrounds the...
mother cell and then reach their final destination by diffusion to, and capture at, the outer forespore membrane (Rudner et al. 2002). Such a diffusion-and-capture mechanism requires that the synthesis of BofA, SpoIFVA, and SpoIFVB takes place prior to the completion of engulfment since the outer membrane surrounding the forespore has become topologically isolated from the cytoplasmic membrane once engulfment is complete. Conversely, no such restriction applies to pro-$\sigma^K$ (a peripheral membrane protein) whose synthesis is delayed (by virtue of being under the positive control of SpoIIIID) relative to that of the integral membrane proteins. Strikingly, and in extension of these observations, a high proportion of $\sigma^K$-controlled genes that encode proteins with predicted transmembrane segments are negatively regulated by SpoIIIID and GerR. We speculate that many of these genes encode proteins that localize to the outer forespore membrane and do so by a diffusion-and-capture mechanism.

Hence their synthesis is restricted to the time prior to the completion of engulfment. By contrast, $\sigma^K$-controlled genes that are unaffected by SpoIIIID and GerR, or are activated by SpoIIIID, rarely encode proteins with predicted transmembrane segments (see Table S2).

As a final example of the coordinate expression of genes with related function we consider the case of cwlC and cwlH, which are switched on in the terminal phase of differentiation under the positive control of GerE (Kuroda et al. 1993; Smith and Foster 1995; Nugroho et al. 1999). The cwlC and cwlH genes encode cell-wall hydrolases that are responsible for the lysis of the mother cell when morphogenesis is complete so that the mature spore can be liberated from the sporangium. It is of crucial importance that mother-cell lysis not take place prematurely, and thus it makes sense that genes involved in this process are among the last genes to be turned on in the mother-cell line of gene expression.

**Some Functionally Related Gene Classes Exhibit Heterogeneous Patterns of Gene Expression**

Many of the genes in the mother-cell line of gene expression are known or inferred to be involved in metabolism, assembly of the spore coat, or the synthesis of coat-associated polysaccharides (see Table 4). Interestingly, not all of the genes in these categories are coordinately expressed. Rather, genes in all three categories exhibit heterogeneous patterns of expression. Thus, among genes inferred to be involved in metabolism, some, such as members of the yngJHGFE operon, which are expected to govern lipid catabolism, and members of the yjmCD–uxaA–yjmF–exuTR operon, which are expected to direct hexuronate synthesis (Mekjian et al. 1999), are expressed early in development, whereas other genes, such as the members of the yitCD and yitBA–yisZ operons, which are inferred to be involved in phosphosulfolactate synthesis (Graham et al. 2002), are expressed late in development. Sulfolactate is indeed known to be a major component of the dry weight (5%) of mature spores of B. subtilis but is not found in spores of B. megaterium and B. cereus (Bonsen et al. 1969). Consistent with these observations, the genome of B. cereus lacks an ortholog of the yitCD operon. Interestingly, the gene for asmO, which encodes an asparagine synthetase (Yoshida et al. 1999), is under the positive control of three of the five mother-cell-specific transcription factors ($\sigma^K$, $\sigma^K$, and SpoIIIID) and the negative control of GerE, and hence its expression is maintained until very late in development.

Of special interest are genes involved in the assembly of the coat, the most conspicuous morphological feature of the mature spore. The coat is a complex, two-layered structure that creates a protective shield around the spore and is composed of at least 30 proteins (Driks 2002; Kuwana et al. 2002; Takamatsu and Watabe 2002; Lai et al. 2003). The earliest-acting protein in the formation of the coat is SpoIVA, which creates a substratum around the outer forespore membrane upon which assembly of the coat takes place (Roels et al. 1992; Stevens et al. 1992; Driks et al. 1994; Price and Losick 1999).

In keeping with its early role in the assembly process, the gene for SpoIVA is switched on early in the mother-cell line of gene expression under the control of $\sigma^K$ and is then turned off by the action of SpoIIIID. The $\sigma^K$ factor also turns on the genes for at least five other coat proteins that play important roles in coat assembly (cotE, cotH, safA, spoVM, and spoVID; Piggot and Losick 2002), but expression of these genes persists longer than that for SpoIVA as none of these is repressed by SpoIIIID. In fact, cotE and cotH continue to be expressed at even higher levels later in development under the control of $\sigma^K$, eventually being downregulated by GerE. In the case of cotE, Li and Piggot (2001) have shown that transcription from its $\sigma^K$-dependent promoter P1 ceases before the activation of $\sigma^K$. Interestingly, certain temporal classes of mother-cell-specific genes are particularly enriched in coat protein genes. For instance, almost half of the $\sigma^K$-controlled genes that are strongly or partially dependent on SpoIIIID for expression (i.e., ten out of 25; C. F., P. E., and R. L., unpublished data) code for coat proteins. Similarly, our preliminary cytological data (C. F., P. E., and R. L., unpublished data) indicate that many of the newly identified $\sigma^K$-controlled genes encode coat-associated proteins.

In addition to being composed of many different proteins, the coat is composed of polysaccharides. Playing an important role in the synthesis of these polysaccharides is the 11-gene sps operon, the longest of the 236 mother-cell-specific transcription units identified in this study. The sps operon is transcribed from a $\sigma^K$-controlled promoter, which we have mapped to a site just upstream of the first gene in the operon, spsA. Transcription from this promoter is enhanced by the appearance of GerE but is not dependent upon it. Thus, expression of genes involved in the biosynthesis of spore-coat polysaccharides persists until the very late stages of sporulation, in keeping with the idea that these polysaccharides are a component of the outer surface of the spore. Nevertheless, some genes in the sps operon are switched on early in sporulation under the control of $\sigma^K$, most likely from a second promoter located upstream of the seventh gene in the operon, spsG. Hence spsG and the genes downstream of it exhibit a protracted pattern of expression that persists throughout the entire process of differentiation.

The sps operon may not be the only set of genes involved in the synthesis of coat-associated polysaccharides. We have identified several paralogs of members of the operon that contribute to the mother-cell line of gene expression. These include genes in the yfnED operon, which is switched on by $\sigma^K$, downregulated by GerR, and turned on again by $\sigma^K$. Another example is the ynfHGF operon, which is under the positive control of $\sigma^K$ and GerE. Yet another example is a paralog of spoJ, yodU–yqpP, which is activated under the dual
control of $\sigma^K$ and GerE. Interestingly, in the strain used in this study (PY79), $yodU$ and $ypqP$ actually correspond respectively to the 5’ end and the 3’ end of a single gene. However, in strain 168, the gene formed by $yodU$ and $ypqP$ is interrupted by the prophage of the large temperate phage SPβ, thereby greatly separating $ypqP$ from the sporulation promoter that would otherwise direct its transcription. It would be interesting to investigate whether the interruption of the $yodU$–$ypqP$ gene by SPβ influences the polysaccharide composition of the spore coat.

The Mother-Cell Line of Gene Transcription in Other Endospore-Forming Bacteria

Endospore formation has been documented in many species of the low G+C group of gram-positive bacteria (Stragier 2002). Two distantly related genera in that group,

<table>
<thead>
<tr>
<th>Functional Category</th>
<th>Gene or Operon</th>
<th>$\sigma$ Factor</th>
<th>Activated by</th>
<th>Repressed by</th>
</tr>
</thead>
<tbody>
<tr>
<td>Engulfment</td>
<td>$spolIID$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$spolIM$</td>
<td>$\sigma^E$</td>
<td>$GerR$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$spolIP$</td>
<td>$\sigma^E$</td>
<td>$GerR$</td>
<td>$GerE$</td>
</tr>
<tr>
<td>Spore cortex formation</td>
<td>$cwlD$</td>
<td>$\sigma^E, \sigma^G$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$spovB$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$spovD$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$spovE$</td>
<td>$\sigma^E$ (P1, P2)</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$yibJ$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$yqfC$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td>$\sigma^K$ Activation</td>
<td>$spollIA$</td>
<td>$\sigma^E$</td>
<td></td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$yqhv-spoolIA$</td>
<td>$\sigma^E$</td>
<td></td>
<td>$GerR$</td>
</tr>
<tr>
<td>Mother-cell metabolism</td>
<td>$asnO$</td>
<td>$\sigma^E, \sigma^K$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$comER$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$glnQ$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$mpr$</td>
<td>$\sigma^K$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$yitB$</td>
<td>$\sigma^K$</td>
<td>$GerE$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$yitC$</td>
<td>$\sigma^E, \sigma^K$</td>
<td>$GerE$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$yjmC$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$yngJ$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td>$\sigma^K$ Synthesis and activation</td>
<td>$bolA$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$spoIVF$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$spoIVCA$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$spoIVCB$</td>
<td>$\sigma^E, \sigma^K$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td>Spore-coat assembly and synthesis</td>
<td>$spolIVA$</td>
<td>$\sigma^E$ (P1, P2)</td>
<td>$SpolIID$</td>
<td></td>
</tr>
<tr>
<td></td>
<td>$spovID$</td>
<td>$\sigma^E$</td>
<td></td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotB$</td>
<td>$\sigma^E, \sigma^K$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotD$</td>
<td>$\sigma^K$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotE$</td>
<td>$\sigma^E$ (P1) $\sigma^K$ (P2)</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotF$</td>
<td>$\sigma^E, \sigma^K$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotH$</td>
<td>$\sigma^E, \sigma^K$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotJ$</td>
<td>$\sigma^E$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotQ$</td>
<td>$\sigma^K$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotR$</td>
<td>$\sigma^K$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$cotT$</td>
<td>$\sigma^E, \sigma^K$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td>Polysaccharide biosynthesis</td>
<td>$spv$</td>
<td>$\sigma^E$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td></td>
<td>$yfnE$</td>
<td>$\sigma^E$</td>
<td>$GerE$</td>
<td>$GerR$</td>
</tr>
<tr>
<td></td>
<td>$yfnH$</td>
<td>$\sigma^E$</td>
<td>$GerE$</td>
<td>$SpolIID$</td>
</tr>
<tr>
<td>Germination</td>
<td>$cwlJ$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$gerM$</td>
<td>$\sigma^E$</td>
<td>$SpolIID$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$gerP$</td>
<td>$\sigma^K$</td>
<td>$GerE$</td>
<td>$GerE$</td>
</tr>
<tr>
<td></td>
<td>$spovF$</td>
<td>$\sigma^K$</td>
<td></td>
<td>$GerE$</td>
</tr>
<tr>
<td>Mother-cell lysis</td>
<td>$cwlC$</td>
<td>$\sigma^K$</td>
<td>$GerE$</td>
<td></td>
</tr>
<tr>
<td></td>
<td>$cwlH$</td>
<td>$\sigma^K$</td>
<td>$GerE$</td>
<td></td>
</tr>
</tbody>
</table>

$^a$ In cases where the transcription start site has been mapped by primer extension, the corresponding $\sigma$ factor is indicated in bold characters.

$^b$ In cases where binding of the transcription factor has been confirmed by DNAase I footprinting or EMSA analysis, the corresponding regulator is indicated in bold characters.

DOI: 10.1371/journal.pbio.0020328.T004

Table 4. Functional Categories

The Mother-Cell Line of Gene Expression
Bacillus and Clostridium, are able to sporulate, whereas several genera that are phylogenetically closer to Bacillus, such as Listeria and Staphylococcus, do not sporulate. Remarkably, in genome regions of otherwise high conservation (synteny) to corresponding regions in B. subtilis, sporulation genes are missing from Listeria (Eichenberger et al. 2003) and Staphylococcus. It is likely that the common ancestor of all of these genera was an endospore-forming bacterium and that sporulation genes were deleted over time from genera that had adapted alternative modes of survival in their ecological niche or host in a manner that did not involve the need for a robust resting state.

To investigate further the evolutionary relatedness of the mother-cell differentiation program among endospore-forming species, we searched for the presence of orthologs of B. subtilis genes in the mother-cell line of gene expression in the genome sequences of the following species: B. anthracis (Ames strain) (Read et al. 2003), B. cereus (ATCC14579) (Ivanova et al. 2003), B. halodurans (Takami et al. 2000), and Oceanobacillus iheyensis (HTE831) (Takami et al. 2002); Listeria monocytogenes and L. innocua (Glaser et al. 2001); and Clostridium acetobutylicum (ATCC 824) (Nolling et al. 2001) and C. perfringens (strain 13) (Shimizu et al. 2002) (Tables S2 and S4).

First, we searched for orthologs of the mother-cell-specific transcription factors. Interestingly, whereas genes for $\sigma^E$, $\sigma^F$, and SpoIID were present in the Bacillus and Clostridium species, GerE (Stragier 2002) and GerR were absent from Clostridium, suggesting that significant differences exist in the mother-cell programs between the two genera, especially during the terminal (GerE-controlled) phase of gene expression. Nonetheless, in cases when a transcription factor was introduced into the pro-less derivative in the PY79 background of strain MO1027 [Karmazyn-Campelli et al. (1989)], PE454 was generated by transformation of strain PY79 to chloramphenicol resistance with chromosomal DNA from strain RL16 (gerE:cat; Cutting and Mandelstam 1986). PE455 is the result of the transformation of strain PE454 with chromosomal DNA from strain MO1027. PE456 was obtained by transformation of PE452 to spectinomycin resistance with chromosomal DNA from strain PE239 (spoIID::spc; Eichenberger et al. 2001). SW282 was generated by transformation of PE454 to spectinomycin resistance with chromosomal DNA from strain PE316 (xyoA::spc; Eichenberger et al. 2003). Strain S101, which was used for overproduction of $\sigma^D$, was created by double cross-over recombination at the amyE locus, following transformation with Xhol-digested plasmid pMFS1gk. Strains PE511, PE551, PE553, PE558, PE568, and SW312 were used for $\beta$-galactosidase activity assays. PE311 is a derivative in the PY79 background of strain MO1533 (amyE::spdIP::lacZ; Frandsen and Stragier 1995). PE531 and PE553 were obtained by double cross-over recombination of Xhol-digested plasmids PE72 and PE74, respectively, into PY79 and selection for chloramphenicol resistance and spectinomycin sensitivity. SW312, PE568, and PE558 were generated by transformation with chromosomal DNA from strain PE316 to spectinomycin resistance. Strains PE551, PE511, and PE553, respectively. Strains PE529 (xyoA::erm), PE534 (mpa::tet), PE538 (ogyT::kan), PE548 (lip::erm), PE549 (coT::spc), RL3231 (ogyM::ata), PE566 (owl::para), PE569 (ylbO::ata), and PE570 (hydA::spa) were generated with the technique of long-flanking homology PCR (Wach 1996). The sequence of the primers used for gene inactivation is available upon request. Strain PE563 (fkh::erm) was obtained by transformation of PY79 to chloramphenicol resistance with chromosomal DNA from RL654 (Cutting et al. 1991c). PE564 is the equivalent of strain MO1057 (spoIVF::erm; a gift from P. Stragier) in the PY79 background. Strains RL2901, RL2996, and RL2519 are from Eichenberger et al. 2001 and RL2393, RL2559, RL2576, RL2571, and RL2581 are from Eichenberger et al. 2003. Strain BDR107 is a derivative of PY79 harboring spoIVF::erm from strain MO1027. BDR362 corresponds to RL75 (spoIII::erm; Kunkel et al. 1989). P$_{\text{spoIII}}$::sgk spc from pDR191 was introduced into the amyE locus of BDR107 to generate BDR1663. Genomic DNA from BDR1663 was used to transform BDR362 to spectinomycin resistance to generate BDR1666.

Plasmids. Plasmid pMF20 was constructed by cloning a PstI (blunted)-EcoRII (blunted) 1.3-kb Neo' cassette from pBES1501 (Iwaya et al. 1989) and a 7.2-kb DNA fragment from plasmid pBR322 (M. F., unpublished data), amplified with primers S11 (5'-AGGATGAGC-GATGATGATCGGTAATTAAGCTGTC-3') and S12 (5'-AATTACTTCGCCGATGATTACGCTGAC-3'). The pro-less sgk gene was constructed using recombinant PCR (Wach 1996). A DNA fragment containing the

Conclusions

We have provided a comprehensive description of the program of gene transcription for a single differentiating cell type and have shown that this program is governed by a regulatory circuit involving the action of five transcriptional control proteins acting as activators or repressors or both. The underlying logic of the circuit is that of a linked series of coherent and incoherent type-1 FFLs involving two-way combinations of the five regulatory proteins. The circuit is expected to create pulse of gene transcription in which large numbers of genes are switched on and subsequently switched off. We anticipate that type-1 FFLs linked in series are likely to be a common feature of programs of cellular differentiation in a wide variety of developing systems.

Materials and Methods

Strains. All strains used here are derivatives of the wild-type strain PY79, with the exception of the $\sigma^E$ overproducing strain, which is a derivative of strain 168. Strains PE454 and PE455 (Eichenberger et al. 2001), PE452, PE454, PE455, PE456, and SW282 were used for transcriptional profiling under conditions of sporulation. PE452 was obtained by transformation of strain RL360 to MLS resistance with chromosomal DNA from strain MO1027 (spoIVC::erm, a gift from P. Stragier, Institut de Biologie Physico-Chimique, Paris) (sigG::cat; a derivative in the PY79 background of strain MO479.2 [Karmazyn-Campelli et al. 1989]). PE454 was generated by transformation of PY79 to chloramphenicol resistance with chromosomal DNA from strain RL16 (gerE:cat; Cutting and Mandelstam 1986). PE455 is the result of the transformation of strain PE454 with chromosomal DNA from strain MO1027. PE456 was obtained by transformation of PE452 to spectinomycin resistance with chromosomal DNA from strain PE239 (spoIID::spc; Eichenberger et al. 2001). SW282 was generated by transformation of PE454 to spectinomycin resistance with chromosomal DNA from strain PE316 (xyoA::spc; Eichenberger et al. 2003). Strain S101, which was used for overproduction of $\sigma^D$, was created by double cross-over recombination at the amyE locus, following transformation with Xhol-digested plasmid pMFS2gk. Strains PE511, PE551, PE553, PE558, PE568, and SW312 were used for $\beta$-galactosidase activity assays. PE311 is a derivative in the PY79 background of strain MO1533 (amyE::spdIP::lacZ; Frandsen and Stragier 1995). PE531 and PE553 were obtained by double cross-over recombination of Xhol-digested plasmids PE72 and PE74, respectively, into PY79 and selection for chloramphenicol resistance and spectinomycin sensitivity. SW312, PE568, and PE558 were generated by transformation with chromosomal DNA from strain PE316 to spectinomycin resistance. Strains PE551, PE511, and PE553, respectively. Strains PE529 (xyoA::erm), PE534 (mpa::tet), PE538 (ogyT::kan), PE548 (lip::erm), PE549 (coT::spc), RL3231 (ogyM::ata), PE566 (owl::para), PE569 (ylbO::ata), and PE570 (hydA::spa) were generated with the technique of long-flanking homology PCR (Wach 1996). The sequence of the primers used for gene inactivation is available upon request. Strain PE563 (fkh::erm) was obtained by transformation of PY79 to chloramphenicol resistance with chromosomal DNA from RL654 (Cutting et al. 1991c). PE564 is the equivalent of strain MO1057 (spoIVF::erm; a gift from P. Stragier) in the PY79 background. Strains RL2901, RL2996, and RL2519 are from Eichenberger et al. 2001 and RL2393, RL2559, RL2576, RL2571, and RL2581 are from Eichenberger et al. 2003. Strain BDR107 is a derivative of PY79 harboring spoIVF::erm from strain MO1027. BDR362 corresponds to RL75 (spoIII::erm; Kunkel et al. 1989). P$_{\text{spoIII}}$::sgk spc from pDR191 was introduced into the amyE locus of BDR107 to generate BDR1663. Genomic DNA from BDR1663 was used to transform BDR362 to spectinomycin resistance to generate BDR1666.
upstream region of the sigK gene was amplified from chromosomal DNA of strain 168 using primers S15 (5'-CCCGAATTCTTAGTATGCTGCTACCTGAAAGATGAT-3') and S14 (5'-AGAGATGTGATTTTGAGTGATCATGTCATCCCTCACAAAAGATGAT-3') (restriction site underlined; the sequence complementary to primer S15 is italicized). The amplified region (496 bp) was digested with HindIII and BamHI and ligated into the HindIII and BamHI sites of pRS426 (Invitrogen) to generate plasmid pRS426-SigK. pRS426-SigK was transformed into E. coli DH5α competent cells by CaCl2-mediated transformation. The transformants were selected on LB plates containing 50 μg/ml of ampicillin. The cells were grown by incubation at 37°C and the overnight culture was used for the preparation of total RNA. Total RNA was isolated by the method of Chomczynski and Sacchi (1987) and was subjected to Northern blot analysis with a XbaI-SalI fragment of pRS426-SigK as a probe. The obtained blots were hybridized with a [γ-32P]-ATP-labeled 5′-CCAATGAGGTATTTTGGATCC-3′ probe. The hybridized blots were visualized and quantified by a Bio-Rad imaging system. The signals were normalized by the β-2-microglobulin gene transcript, which was amplified and purified from the same RNA samples and used as a loading control.
arrays (ChIP-on-chip). Three hours after resuspension of PY79 cells in Sterlini-Mandelstam medium at 37 °C, cross-links were generated by treatment with formaldehyde (1% final concentration) for 30 min. The rest of the procedure was identical to the one described by (Molle et al. 2003a, 2003b).

The data analysis for the ChIP-on-chip experiments was carried out using the Resolver statistical package (Rosetta, Seattle, Washington, United States). Experiments were normalized and combined for enrichment-factor determination (Rosetta Resolver). An enrichment factor for a given gene represents the ratio of immunoprecipitated DNA to total DNA. It was considered significant when higher than 2 and with an associated p-value lower than 0.001.

BioProspector/BioOptimizer. BioProspector (Liu et al. 2001) is a stochastic motif-discovery program used to find conserved subsequences of fixed width in a set of DNA sequences, based on a statistical motif-discovery model reviewed in Jensen et al. (2004). The program can also be used for motifs consisting of two conserved blocks connected with a variable-length gap of unconserved nucleotides, and BioProspector can also be forced to find sites in every input sequence. Since BioProspector is a stochastic algorithm, more than one possible motif can be found, and since the program requires the motif width to be fixed, several different fixed widths should be used in the usual case where the motif width is not known. Thus, we collected the top five BioProspector motifs under a range (6–12 bps) of seven fixed widths, giving a total of 35 putative motifs.

BioOptimizer (Jensen and Liu 2004) is an optimization program designed to improve the results of each discovered BioProspector motif to search each motif so that the "best" putative motif can be selected out of the 35 we discovered. The scoring function used is the exact log-posterior density of the Bayesian motif-discovery model given in Jensen and Liu (2004). Starting from the set of sites predicted by BioProspector, the scoring function is optimized by adding the contribution of new motif sites or removal of current motif sites only if these changes increase the score. BioOptimizer also has the flexibility to allow the motif width to vary, so that the "best" width can also be determined. As well, BioOptimizer can be restricted to force particular sequences in the dataset to contain at least one site while leaving other sequences unrestricted. The property was used in our SpoIIID motif search, where a subset of sequences has additional biochemical evidence that they contain at least one SpoIIID-binding site.

Having found an optimal motif with our combined BioProspector/BioOptimizer procedure, we implemented an additional scanning procedure to find more potential SpoIIID sites. Using the estimated proportion of nucleotide k in position j of the motif (θj) and the estimated proportion of nucleotide k in the background (θk), provided by our optimal motif, we scanned all upstream sequences to see if there were additional sites that matched our discovered motif, but were not strong enough to be detected by the motif discovery procedure. In each sequence, for each potential starting position i, we had a potential site Si = (r1, r2, ..., rw), for which we compute the following score:

\[\text{Strength} = \log \left( \frac{P(S_i | \text{motif})}{P(S_i | \text{background})} \right) = \log \left( \prod_{j=1}^{w} \frac{\theta_{j}}{\prod_{k=1}^{w} \theta_{k}} \right) \]  

We considered the site in each sequence with the largest Strength value to be the best candidate as an additional site. If a sequence already contained a site found by our motif-discovery procedure, we would expect that this same site would be the one with the largest Strength value. For any sequence that did not have an optimal site found by the motif-discovery procedure, this scanning procedure gave us new site predictions. However, for any new sites found by the scanning procedure, one must be cautious about the strength of these sites, since the procedure found sites in each sequence regardless of how well those sites matched our optimal motif. Therefore, we also calculated a p-value for each site by comparing the Strength value calculated for that site to the Strength value calculated for 10,000 random sequences. Only sites with low p-values were considered as potential sites. With the Bonferroni correction for multiple comparisons, we considered only sites with p-values less than 0.000183.

MDscan analysis of the SpoIIID-binding motif. We used the word-enumeration algorithm “MDscan” (Liu et al. 2002) to identify motifs in sequences most enriched by immunoprecipitation experiments. In this algorithm, it is assumed that the most enriched sequences have stronger motif signals than the remaining sequences. MDscan first identifies oligomers of width w (w-mers) in the top sequences, which are used as seed oligomers. Motif matrices are constructed for each seed oligomer using all similar segments from the top sequences. Segments are defined to be similar if they share at least m matched positions, with m determined so that the probability that a pair of randomly produced w-mers are m-matches is less than 0.15%. The resulting motif matrices are evaluated using the following semi-Bayesian scoring function:

\[\log(x_s) = w + \sum_{i=1}^{m} \log(p_i) + \frac{1}{x_s} \sum_{i=1}^{m} \log(p_i(s)) \]  

where ns is the number of segments aligned in the motif, p_i is the frequency of base j at motif position i, and p_i(s) is the probability of generating segment s from the background model. The top distinct highest-scoring motifs are defined as candidate motifs. These motifs are refined using the remaining sequences, by adding new w-mers to the matrix if the score is increased. The motifs are further refined by reexamining all segments of the motif matrix and removing segments if the motif score is increased.

We first ranked by enrichment ratio the 26 regions of the chromosome that were enriched by immunoprecipitation by a factor of 2 or greater. We used the top 20 regions as the top sequences, with the remaining six sequences used for refinement. The 26 regions were used as the background sequences, and we reported 30 candidate motifs. We first searched for motifs of width w = 8. In using alternative widths (w = 7, 9, and 10), and alternative definitions of top regions (15–25), the top reported motif was similar to that for width 8. As reported in Liu et al. (2002), MDscan is tolerant of different top sequence definitions (~3–20), and of moderate ranking errors.

Germination assays. Tests for germination using 2,3,5-triphenyltetrazolium chloride overlay were carried out as described in Nicholson and Selton (1990). Strains mutant for GerR (PE316) were compared to wild-type cells (PY79), and strains mutant for GerE (strain PE454) or CotE (strain RL322; Driks et al. 1994) were described as negative controls. All strains were sporulated in DSM. Heat activation was performed in a 65 °C oven for 3 h.

Measuring β-galactosidase activity. β-galactosidase activity assays were carried out as previously described (Miller 1972; Harwood and Cutting 1990).

Promoter mapping by 5′ RACE-PCR. The 5′ end of several σ^E-controlled mRNAs was determined by the RACE–PCR procedure (Frohman 1994; Price et al. 2001). Total RNA was extracted from strains PE454 (sigE^+, sigK^+), and PE455 (sigE^-, sigK^-) and analyzed as described by Eichenberger et al. (2003).

Measuring sporulation efficiency. Strains were grown to exhaustion in DSM for 30 h at 37 °C and assayed for heat resistance as previously described by van Ooij et al. (2004).

Supporting Information

Figure S1. DNAase I Footprinting of SpoIIID Binding to the Promoters of spoIID, spoIIE, and spoVE (A) Radioactive DNA fragments were incubated with no protein (left lane) or with 400 nM of SpoIIID protein (right lane) and then subjected to DNAase I footprinting. A chemical sequencing ladder was used as a marker (not shown). Protected regions are indicated by a bar.

(B) Position of SpoIIID-binding sites. The nucleotide sequence upstream of the transcriptional start site (+1) is shown for spoIID, spoIIE, spoVE-P1, and spoVE-P2. The boundaries of the region protected from DNAase I digestion by SpoIIID are indicated by bars.

The bold letters identify the sequences within the protected regions that match with the SpoIIID consensus sequence.

Found at DOI: 10.1371/journal.pbio.0020328.sg001 (1.72 MB PPT).

Figure S2. Mapping of Transcription Start Sites by 5′ RACE–PCR The underlined uppercase bold letters identify the 5′ ends of mRNAs from σ^E-controlled promoters by RACE-PCR. Also indicated are the corresponding −35 and −10 regions (upstream promoters in bold); the ribosome-binding site (double underlining), and the translation start site (upstream letters). RNA collected from strain PE454 (sigE^+, sigK^+) and strain PE455 (sigE^-, sigK^-) was used for the determination of transcription start sites. In four cases indicated with an asterisk (sigE^+, yedO, yedC, and yedA), the transcription start site was identified for strains PE454 and PE455, which is interpreted as evidence that the promoters for these three transcription units are recognized both by σ^E and σ^E'. In all of the other
cases, a transcription start site was obtained only with RNA collected from strain PE54.

Found at: DOI: 10.1371/journal.pbio.0020328.s002 (22 KB DOC).

Table S1. Mother-Cell Gene Expression

Found at: DOI: 10.1371/journal.pbio.0020328.s001 (1.5 MB XLS).

Table S2. Effect of SpoIIIID and Ger on the expression of genes in the σC regulon.

Found at: DOI: 10.1371/journal.pbio.0020328.s002 (351 KB XLS).

Table S3. ChiP-on-chip data for SpoIIIID.

Found at: DOI: 10.1371/journal.pbio.0020328.s003 (352 KB XLS).

Table S4. Effect of GerE on the expression of genes in the σC regulon.

Found at: DOI: 10.1371/journal.pbio.0020328.s004 (238 KB XLS).

Accession Numbers

The Swiss-Prot (http://www.ebi.ac.uk/swissprot) accession numbers for the gene products discussed in this paper are spsG (BSU06790), spoVK (BSU11690), spoIVCB (BSU25760), treA (BSU11680), spoIID (BSU18470), proJ (BSU02240), spoIIGA (BSU21280), ydhT (BSU12330), ymF (BSU12550), yktU (BSU14250), ykV (BSU14320), ykb (BSU14330), ybdB (BSU14400), ykd (BSU13570), ykv (BSU13850), ylbf (BSU15030), yvdOGerK (BSU15090), yngE (BSU18120), yngF (BSU18220), yngG (BSU18230), yngH (BSU18240), yngI (BSU18250), yofB (BSU18560), yofD (BSU19810), yofA (BSU22240), yofP (BSU21670), yofT (BSU25120), yofV (BSU24440), yofD (BSU26680), yofB (BSU26820), yofC (BSU40210), yofD (BSU40290), ydGD (BSU40170), ydHD (BSU40160), ydfB (BSU40140), and ydfD (BSU40140).

Microarray data were deposited in the Gene Expression Omnibus database under the accession number GSE1620, where they are accessible at http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE1620.

Acknowledgments

We are grateful to Uri Alon, Harley McAdams, Lee Kroos, Patrick Piggot, and Patrick Stragier for critical comments of the manuscript. We also thank Uri Alon for helpful discussions and Patrick Stragier for strains. We are indebted to Eduardo Gonzalez-Pastor for his contribution in setting up the tools for the generation and utilization of B. subtilis DNA microarrays in the laboratory. We thank Claire Ball and Paul Grossi at the Bauer Center for Genomics Education for help in the preparation of the DNA microarrays and with the Rosetta Resolver analysis, respectively. We also thank the members of the Losick laboratory for advice.

This work was supported by National Institutes of Health grant GM18458 to RL and in part by a Grant-in-Aid for Scientific Research on Priority Areas (genome biology) from the Ministry of Education, Science, Sports, and Culture of Japan to TS. PE was supported by a Merck Core Educational Support Program and the Swiss National Science Foundation. STJ, EMC, and JSL were supported by National Science Foundation grants DMS-0244638 to JSL.

Conflicts of interest.

The authors declare that no conflicts of interest exist.

Author contributions.

PE, MF, STJ, EMC, DZR, TS, JSL, and RL conceived and designed the experiments. PE, MF, DZR, STW, CF, and KH performed the experiments. PE, MF, STJ, EMC, DZR, and JSL analyzed the data. PE, MF, STJ, EMC, DZR, and JSL contributed reagents/materials/analysis tools. PE, MF, STJ, EMC, JSL, and RL wrote the paper.

References


