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#### ORIGINAL INVESTIGATION



# Genetic and phenotypic dissection of 1q43q44 microdeletion syndrome and neurodevelopmental phenotypes associated with mutations in *ZBTB18* and *HNRNPU*

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**Abstract** Subtelomeric 1q43q44 microdeletions cause a syndrome associating intellectual disability, microcephaly, seizures and anomalies of the corpus callosum. Despite several previous studies assessing genotype-phenotype correlations,

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the contribution of genes located in this region to the specific features of this syndrome remains uncertain. Among those, three genes, *AKT3*, *HNRNPU* and *ZBTB18* are highly expressed in the brain and point mutations in these genes have been recently identified in children with neurodevelopmental phenotypes. In this study, we report the clinical and molecular data from 17 patients with 1q43q44 microdeletions, four with

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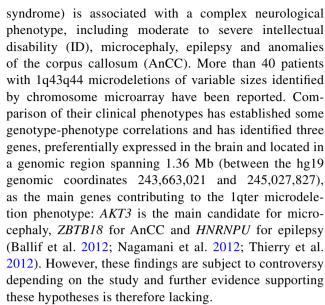


ZBTB18 mutations and seven with HNRNPU mutations, and review additional data from 37 previously published patients with 1q43q44 microdeletions. We compare clinical data of patients with 1q43q44 microdeletions with those of patients with point mutations in HNRNPU and ZBTB18 to assess the contribution of each gene as well as the possibility of epistasis between genes. Our study demonstrates that AKT3 haploinsufficiency is the main driver for microcephaly, whereas HNRNPU alteration mostly drives epilepsy and determines the degree of intellectual disability. ZBTB18 deletions or mutations are associated with variable corpus callosum anomalies with an incomplete penetrance. ZBTB18 may also contribute to microcephaly and HNRNPU to thin corpus callosum, but with a lower penetrance. Co-deletion of contiguous genes has additive effects. Our results confirm and refine the complex genotype-phenotype correlations existing in the 1qter microdeletion syndrome and define more precisely the neurodevelopmental phenotypes associated with genetic alterations of AKT3, ZBTB18 and HNRNPU in humans.

#### Introduction

Deletion of the subtelomeric region of the long arm of chromosome 1 (1q43q44 or 1qter microdeletion

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Interestingly, point mutations in *AKT3*, *ZBTB18* and *HNRNPU* have recently been identified by whole exome sequencing in patients with different neurodevelopmental phenotypes. *AKT3* encodes a serine/threonine protein kinase involved in the mammalian target of rapamycin (mTOR) signaling pathway. Gain-of-function point mutations or microduplications leading to abnormal AKT3 and

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mTOR activation, most of which are limited to somatic brain populations, cause a spectrum of disorders characterized by cerebral hemisphere overgrowth such as hemimegalencephaly (HME), megalencephaly-polymicrogyria-polydactyly-hydrocephalus (MPPH) and megalencephaly-capillary malformation (MCAP) (Lee et al. 2012; Mirzaa et al. 2013; Poduri et al. 2012; Riviere et al. 2012). Conversely,  $Akt3^{-/-}$  mice show a 20% reduction in brain size (Easton et al. 2005). ZBTB18 (also known as ZNF238 or RP58) encodes a C2H2-type zinc finger transcription factor negatively controlling the expression of genes involved in neuronal development, including cell division of progenitor cells and survival of postmitotic cortical neurons (Baubet et al. 2012; Heng et al. 2015; Xiang et al. 2012). Zbtb18-deficient mice show features reminiscent of the 1q43q44 microdeletion syndrome including microcephaly and agenesis of the corpus callosum (AgCC) (Xiang et al. 2012). Eight patients with de novo ZBTB18 mutations have been reported, including three with a normal corpus callosum (CC) (Cohen et al. 2016; de Munnik et al. 2014; Lopes et al. 2016; Rauch et al. 2012) and four with AnCC (Cohen et al. 2016). Finally, HNRNPU encodes the heterogeneous nuclear ribonucleoprotein (hnRNP) U, an abundant nucleoplasmic phosphoprotein able to bind pre-mRNA in vivo, possibly involved in pre-mRNA splicing (Roshon and Ruley 2005; Ye et al. 2015). Eighteen de novo and/or truncating mutations in HNRNPU mutations have been reported in ClinVar, Decipher and in different studies (Carvill et al. 2013; de Kovel et al. 2016; Epi4K Consortium et al. 2013; Hamdan et al. 2014; Monroe et al. 2016; Need et al. 2012; Zhu et al. 2015); however, since these mutations were reported each in separate studies and

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the phenotype of the patients was not described, a specific disorder related to HNRNPU mutations is not yet characterized. Although the description of these patients independently reinforced the previously proposed genotype-phenotype correlations, the dispersion of patients with point mutations in different studies and the absence of comparison with microdeletions did not permit to clearly address the clinical spectra associated with mutations in these genes, nor the possible epistatic or additive genetic interactions. The aim of this study was to describe in more details novel patients with HNRNPU and ZBTB18 point mutations identified by next generation sequencing and to compare their core phenotype with those of patients with 1q43q44 microdeletions to decipher the contribution of each gene to the 1 gter microdeletion syndrome. To this aim, we collected and compared the data of 17 patients with 1q43-q44 deletions, four patients with ZBTB18 mutations and seven with HNRNPU mutations.

#### Materials and methods

#### **Human subjects**

We independently identified two *ZBTB18* point mutations in unrelated patients with AnCC and one *HNRNPU* mutation in a patient with epileptic encephalopathy by, respectively, sequencing 423 genes associated with AnCC (callosome panel) in humans or mice (Mignot et al. 2016) or 4813 genes of the Trusight One panel (Illumina).

We then collected clinical and molecular data of patients with ZBTB18 and HNRNPU mutations or

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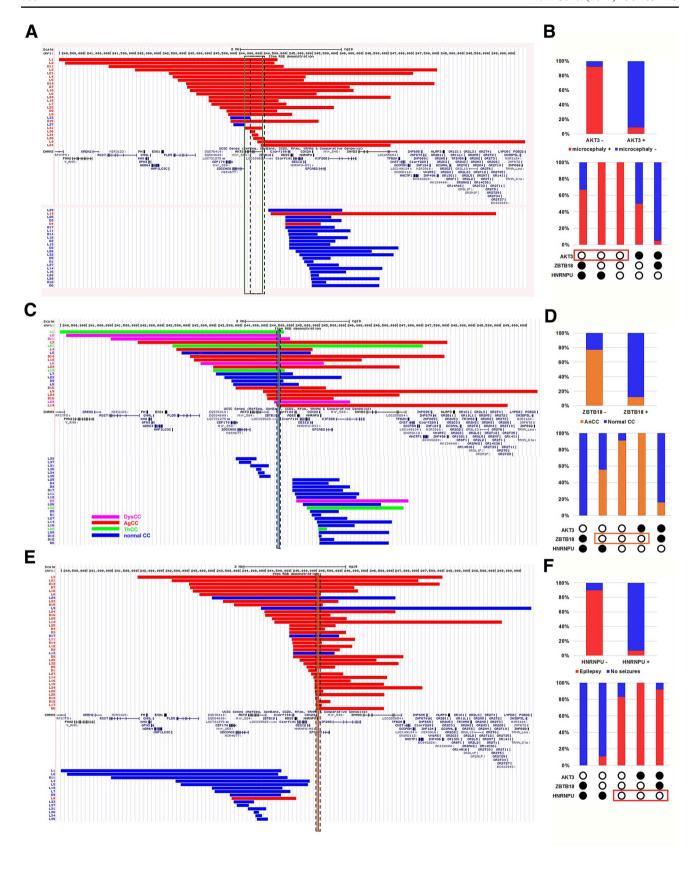


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**∢Fig. 1** Significant genotype–phenotype correlations gained from comparison of clinical data of patients with microdeletions. a, b Alignment of microdeletions found in patients with (red bars) and without (blue bars) microcephaly showed that deletions including AKT3 (upper panel) were mostly associated with microcephaly and those excluding AKT3 (lower panel) were mostly associated with normal OFC (a). The minimal critical region (vertical rectangle with dashed borders) overlapped the 5' region of AKT3. Diagrams in b show the percentages of patients with (red) or without (blue) microcephaly who had a microdeletion including (AKT3-) or excluding (AKT3+) AKT3 (upper panel), and comparison of the percentage of patients with (red) or without (blue) microcephaly who had a microdeletion encompassing only one of the three genes of interest, two genes or all three genes (lower panel, empty circles designate deleted genes, full circles are for non-deleted genes). c, d Alignment of the microdeletions found in patients with AnCC (AgCC red bars, DysCC pink bars, ThCC green bars) and patients without CC anomalies (blue bars) showed that deletions including ZBTB18 (upper panel) were mostly associated with all types of AnCC and those excluding ZBTB18 (lower panel) were mostly associated with normal CC. The minimal critical region (vertical rectangle with dashed borders) overlapped the entire coding sequence of ZBTB18. Diagrams in **d** show the percentages of patients with (orange) or without (blue) AnCC who had a microdeletion including (ZBTB18-) or excluding (ZBTB18+) ZBTB18 (upper panel), and comparison of the percentage of patients with (orange) or without (blue) AnCC who had a microdeletion encompassing only one of the three genes of interest, two genes or all three genes (lower panel, empty circles designate deleted genes, full circles are for non-deleted genes). e, f Alignment of the microdeletions found in patients with (red bars) and without (blue bars) epilepsy showed that deletions including HNRNPU (upper panel) were mostly associated with epilepsy and those excluding HNRNPU (lower panel) were mostly associated with no seizures. The minimal critical region (vertical rectangle with dashed borders) overlapped the entire coding sequence of HNRNPU and COX20. Diagrams in **f** show the percentages of patients with (red) or without (blue) epilepsy who had a microdeletion including (HNRNPU-) or excluding (HNRNPU+) HNRNPU (upper panel), and comparison of the percentage of patients with (red) or without (blue) epilepsy who had a microdeletion encompassing only one of the three genes of interest, two genes or all three genes (lower panel, empty circles designate deleted genes, full circles are for non-deleted genes)

1q43q44 microdeletions through members of the EUROEPINOMICS RES consortium, the French Achropuce network (http://www.renapa.univ-montp1.fr/), Genematcher (Sobreira et al. 2015) and Decipher (https:// decipher.sanger.ac.uk/) (Firth et al. 2009). This series includes previously reported patients with updated clinical data and previously reported genotypes with unreported clinical data: patients D2, D3, D4, D5 and D8 were reported as patients #7, #5, #3, #9 and #6, respectively, in (Thierry et al. 2012); patient H2 was reported as patient #15 in (Monroe et al. 2016); patient H3 was reported as patient 2012D06376 (de Kovel et al.); the mutation identified in patient H7 was present in Decipher (ID 268181—DDD-NIG268181); deletions present in Decipher correspond to patients D9 (ID 2762), D13 (ID 332095), D14 (ID 275142), D15 (ID 268383),

D16 (ID 253339) and D17 (ID 2926112). All other patients with deletions and point mutations are novel. In addition, we performed a review of the literature on 1q43q44 microdeletions and included microdeletions <6 Mb encompassing AKT3, ZBTB18 and/or HNRNPU with available breakpoints, excluding patients with other probably pathogenic chromosomic anomaly (numbered L1-L37: Ballif et al. 2012; Du et al. 2014; Gai et al. 2015; Gupta et al. 2014; Nagamani et al. 2012; Perlman et al. 2013; Thierry et al. 2012). Coordinates of the deletions reported in hg18 were converted into hg19/ GRCh37 with LiftOver (https://genome.ucsc.edu/cgibin/hgLiftOver). Clinical data were collected using a standardized questionnaire directly from the referring clinicians. Microcephaly was considered for patients with an occipitofrontal circumference (OFC) of at least -2.5 standard deviation (SD) below the mean. A radiologist, a neuropediatrician and a geneticist collegially ascertained brain MRI anomalies. AgCC designates the absence of one or all parts of the CC, DysCC is used for complete CC with an abnormal shape or abnormally small CC, ThCC is used for complete CC with insufficient thickness.

# Genotype-phenotype correlations, bioinformatics and statistics analyses

We retrieved the probability of loss-of-function (LoF) intolerance (pLI) calculated by the Exome Aggregation Consortium (ExAC) and the haploinsufficiency score (HI) established by Huang et al. (2010) for genes of the 1q43q44 region comprised between genomic positions 239,990,618 and 249,208,333 to determine genes intolerant to haploinsufficiency, contributing to 1qter deletion phenotypes (Table S1). The pLI calculates the probability that a gene is intolerant to LoF mutations, calculated from the difference between the number of LoF mutations observed in the 60,000 individuals present in ExAC and the theoretical number of expected LoF mutations in this gene in a population of same size if there was no selective constraint. Genes with a pLI ≥0.9 are considered to be significantly LoF intolerant. HI scores evaluate the probability that the gene is intolerant to haploinsufficiency, calculated from CNV data and integrating genomic, evolutionary and function properties of haploinsufficiency (Huang et al. 2010). High ranks (e.g. 0-10%) indicate a gene that is likely intolerant to haploinsufficient; low ranks (e.g. 90-100%) indicate a gene that likely tolerates haploinsufficiency.

Missense variants were assessed in silico for possible pathogenicity using Alamut Visual 2.7 (Biointeractive Software, France), PolyPhen-2 (http://genetics.bwh.harvard.edu/pph2) and SIFT (http://sift.bii.a-star.edu.sg).



We used UCSC (https://genome-euro.ucsc.edu) to align microdeletions on a schematic representation of the 1q chromosome. Alignments were performed using different colors explained in the figure legends. Minimal critical regions were defined as the smallest deleted region of overlap found in at least 95% (microcephaly and epilepsy) or 85% (AnCC) of patients harboring a given phenotypic trait. Frequencies were compared using the Fisher's exact test.

#### Results

#### Patients with 1q43-q44 deletion

To decipher the contribution of genes located in 1qter region to the corresponding microdeletion syndrome, we collected clinical data from 17 patients with 1q43q44 microdeletion (Table S2) and compiled them with those of 37 previously reported patients fulfilling our criteria (see Methods, Table S3). Altogether, the 54 deletions span a 10 Mb region comprising 83 genes, 39 of which encode clustered olfactive receptors (OR). Seven genes (RGS7, AKT3, ZBTB18, HNRNPU, KIF26B, CNST, AHCTF1) were predicted to be probably or possibly intolerant to haploinsufficiency among the 44 genes other than OR genes (Table S1). Among these genes, AKT3, ZBTB18 and HNRNPU are clearly those with the highest pLI scores and HI ranks as well as the highest expression in the brain (Table S1). We then decided to focus our study on these three genes that likely contribute to most clinical features of 1q43q44 microdeletion syndrome, as predicted from previous genotype-phenotype correlation studies. Specifically, 12 microdeletions encompassed AKT3, ZBTB18 and HNRNPU; six completely or partially included AKT3 but not ZBTB18 and HNRNPU; nine encompassed AKT3 and ZBTB18 but not HNRNPU, two ZBTB18 and HNRNPU but not AKT3, and 25 deleted HNRNPU but not AKT3 and ZBTB18 (Fig. S1).

Out of the 49/54 patients with available OFC, 26 had microcephaly (red bars in Fig. S2A); 47/54 patients had available brain imaging: 20 had AnCC, including ten with agenesis (AgCC, red bars in Fig.S2B), five with dysgenesis (DysCC, pink bars in Fig.S2B) and five with thin corpus callosum (ThCC, green bars in Fig. S2B). Finally, 36/54 patients had epilepsy (red bars in Fig. S2C).

The alignments of microdeletions found in patients with a known OFC and comparison of their gene content showed that microcephaly was present in all 20 patients with deletions encompassing both *AKT3* and *ZBTB18*, regardless of the presence of *HNRNPU* in the deletions (Fig. 1a, b). Conversely, 20/21 patients with a deletion encompassing *HNRNPU* but sparing *AKT3* and *ZBTB18* had normal OFC (versus 2/15 when *HNRNPU* 

not deleted, p = 0.00086, Fisher's exact test). AKT3 was included in 24/26 deletions identified in individuals with microcephaly, whereas only 2/23 patients with microdeletions sparing AKT3 had microcephaly  $(p = 1.46E^{-9})$ . The number of patients with microcephaly who had deletions including ZBTB18 (n = 21/22) and sparing ZBTB18 (5/27) was also significantly different ( $p = 4.25E^{-8}$ ). Four of the six deletions including coding sequences of AKT3 only and one of the two deletions encompassing ZBTB18 but not AKT3 were associated with microcephaly. The minimal critical region for microcephaly (g.243,778,438-g.244,125,269) mapped to a region encompassing the 5' upstream region and the five first exons of AKT3 (Figs. 1a, 2a). Altogether, these results indicate that (1) AKT3 is the main driver for microcephaly in the 1q43q44 region; (2) ZBTB18 haploinsufficiency may independently lead to microcephaly with a lower penetrance; and (3) co-deletion of AKT3 and ZBTB18, which are neighboring genes spaced from only ~200 Kb, may have an addictive effect, resulting in constant microcephaly.

Considering all types of AnCC, the alignment of deletions revealed that, contrary to microcephaly, AKT3 deletion was not significantly associated with AnCC. More precisely, 11/26 patients with AKT3 deletion versus 16/21 without AKT3 deletion had a normal CC (p = 0.06). Accordingly, all six patients with a microdeletion involving only AKT3 had a normal CC. The proportions of patients with AnCC who had deletions encompassing (17/32) or sparing HNRNPU (5/15) were also not significantly different (p = 0.23). In contrast, the number of patients with AnCC was significantly higher in cases of deletions containing ZBTB18 (17/22) compared with deletions sparing this gene (3/25,  $p = 6.84E^{-6}$ , Fig. 1c, d). Accordingly, the minimal critical region for AnCC overlaps ZBTB18 (Fig. 1c). When comparing patients with ZBTB18 deletions sparing HNRNPU versus deletions encompassing both genes, the proportions of AnCC as a whole were not significantly different (5/9 vs 12/13, respectively; Fig. S3). However, these proportions reached statistical significance when considering AgCC instead of AnCC (p = 0.01). These results suggest that (1) the main driver for AnCC, and more particularly AgCC, in the 1q region is ZBTB18, although with incomplete penetrance, (2) HNRNPU haploinsufficiency can be associated with ThCC, and (3) deletion of both ZBTB18 and HNRNPU, which are 792 kb distant, has an additive effect, resulting in more penetrant AnCC phenotype and more frequent AgCC.

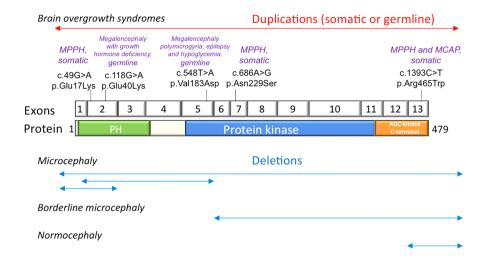
Deletions identified in epileptic patients showed a shift toward the telomeric extremity of the 1q region (Fig. S2). Of the 36 patients with epilepsy, 35 had a deletion including HNRNPU and only one had a deletion sparing HNRNPU ( $p = 1.28E^{-8}$ , Fig. 1e, f). The minimal critical



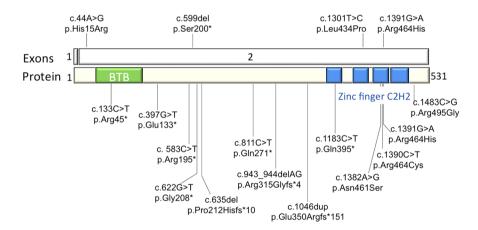
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Fig. 2 Summary of intragenic microdeletions and point mutations in AKT3, ZBTB18 and HNRNPU. a A schematic representation of the AKT3 gene and protein, location of point mutations (somatic) and duplications (somatic or germline) identified in patients with brain overgrowth syndromes (upper panel) and comparison of intragenic AKT3 microdeletions and their association with microcephaly (lower panel). b A schematic representation of the ZBTB18 gene and protein and location of pathogenic point mutations identified in patients with ID and/or AnCC, including this study (upper panel) and the literature (lower panel). c A schematic representation of the HNRNPU gene and protein and location of pathogenic point mutations identified in patients with ID and epilepsy, including this study (upper panel) and the literature (lower panel)

#### **A** AKT3 (NM\_005465.4)



#### **B** ZBTB18 (NM\_205758.2)



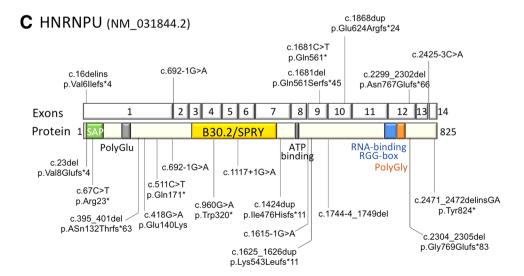




 Table 1
 Molecular and clinical characteristics of patients with HNRNPU mutations

Patient ID	H1	H2	Н3	H4	Н5	9Н	H7
Gender	F	F	F	M	M	M	F
Current age (years)	6.5	16.75	5.5	22	17	6.3	8
Genetic data							
Genomic position (hg19)	g.245027594delinsAAT	g.[245018776_245018779=/24501877 6_245018779del]	g.245019803dup	g.245017808G>T	g.245020092del	g.245020092G>A	g.245026033C>T
cDNA change (NM_031844.2)	c.16delinsATT	c.[2299_2302AACA=/2299_2302del]	c.1868dup	c.2425-3C>A	c.1681del	c.1681C>T	c.692-1G>A
Amino acid change	p.Val6Ilefs*4	p.Asn767Glufs*66 (mosaic)	p.Glu624Argfs*24	p.?	p.Gln561Serfs*45	p.Gln561* (mosaic)	p.?
Exon/intron	Exon 1	Exon 12	Exon 10	Intron 13	Exon 9	Exon 9	Intron 1
Inheritance Epilepsy	Paternal mosaicism	De novo	De novo	De novo	De novo	De novo	De novo
Epilepsy	Yes	No	Yes	Yes	Yes	Yes	Yes
Age of first seizure	2.5 months	NA	8 months	7 months (febrile sz)	4 months	1 febrile sz at 3 years then sz at 4.8 years	12 months
Type of seizures	Short tonic sz, unilateral clonic, eye blink-ing + atypical abs.	NA	GTCS, atonic	Abs., GTCS (febrile before age 3 years)	Atypical abs., cyanotic epi-sodes, noctumal tonic and GTCS	GTCS, abs.	NA
Triggering factors	Fever	NA	Fever	Fever, bright light, stress, tiredness	None	Fever	Fever
Max number of seizures	10/day	NA	1/months	>20/day	Tonic and abs.: multiple/day; GTCS: 5/months	24/months	1–2/months
Status epilepticus? Development	Status epilepticus? Yes (2 with fever) evelopment	NA	No	No	Only non-convulsive	No	No
Age of sitting	2 years	Delayed	NA	Delayed	NA	18 months	14 months
Age of walking	Never walked	Never walked (wheelchair bound)	2 years	Delayed	NA	2.5 years	3 years
First words	None	Delay	2.5 years	3 years	None	20 months	3 years
Current language abilities	Few bisyllable words	Delay (short sentences)	Delay	Single words	Absence of speech	4 years: 180 words, 2–3 word sen- tences; 6 years: regressed to 100- 150 single words	Words



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Moderate-severe ¥ Ā Ϋ́ (0)/54 (+2)/6105(-2), 21.9Moderate ID Hypotonia Normal Severe ID Thin CC Normal Normal autistic disorder tive functioning tional function-2.5 years, level normal (before of social emo-87 (+0.5)/100 ing 9 months, Level of cogni-Brain CT scan age 4 years) (>+3)/58.5(+0.5)/22ΝA Hypotonia, hyperlaxity of joints NA (-2)/NA (-0.5)/NA(-1)/2.5Deficient Normal Dilated ventricles (presumed aqueduct  $115 (-4)/42 (\gg +2.5)/56 (+1.5)/11.5$ Axial hypotonia, spastic diplegia stenosis), small splenium of CC Moderate/severe ID Enlarged lateral ventricles, complete CC 110(-1)/20(0)/47Global hypotonia (-3)/6.5Severe ID  $\Xi$ weight (kg)/OFC Clinical examination Global intellectual (cm) + SD/ageHeight (cm)/ Neurological exam data **3rain MRI** (years) Patient ID

 Table 1
 continued

WA not available or not applicable, sz seizures; GTCS generalized tonic-clonic seizures; abs. absences; ID intellectual deficiency; OFC occipitofrontal circumference; CC corpus callosum.\$ pre-

dicted to change splice acceptor site, potentially leading to a shorter protein

region for epilepsy was narrow and included HNRNPU and COX20, which is a gene that tolerates haploinsufficiency. Comparison of the number of epileptic patients who had AKT3 deleted (n=11/27) or spared (n=25/27) and the absence of seizures in patients with deletion restricted to this gene confirmed that AKT3 was not involved in epilepsy. The difference in the number of epileptic patients with or without ZBTB18 deletion was also not significant (13/23 vs 23/31, p=0.24). These results suggest that the loss of one HNRNPU allele is the primary cause of epilepsy.

Among the seven patients with *HNRNPU* mutations, six had constitutive de novo mutations and one has a mosaic frameshift mutation (Table 1). All seven mutations (four frameshifts, one nonsense variant and two splice site mutations, Fig. 2b) theoretically introduced a premature termination codon in the protein sequence, a mutation spectrum compatible with *HNRNPU* haploinsufficiency as the main consequence.

All six patients with constitutive de novo *HNRNPU* mutations had early-onset epilepsy (Table 1, Table S4). Seizure onset ranged from age 2.5 months to 4 years, and was within or at the first year of life in 5/6 patients. Fever was a factor triggering seizures in five patients. Seizures types included tonic—clonic, tonic, unilateral clonic or atypical absences occurring one to 20 times a day. Two patients experienced *status epilepticus*.

Early developmental delay was observed in all patients, including the individual with the mosaic mutation. The severity of ID ranged from moderate to severe. None of the seven patients was able to make sentences, four of them spoke single words at a time and one never acquired any word. Two patients older than 6 years never learned to walk independently and three others walked after the age of 30 months. Microcephaly was noted in one patient and global hypotonia in three. Among the five patients who underwent brain MRI, one displayed a small splenium of the CC and one had a globally thin CC (Fig. 3).

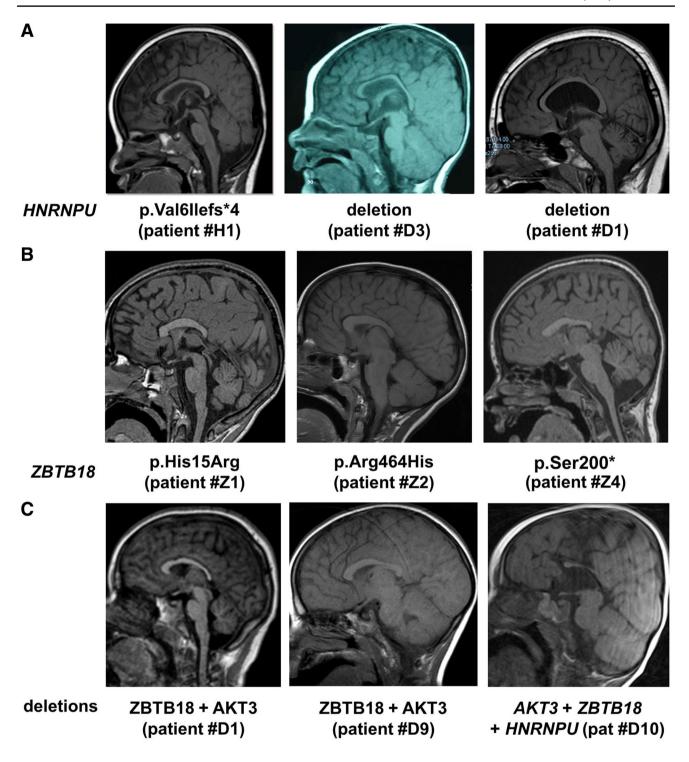
#### **Patients with ZBTB18 mutations**

Mutations identified in *ZBTB18* included three missense variants altering highly conserved amino acids of the protein (Fig. S4). Two of them are located in zinc finger domains in which missense mutations tend to cluster (Fig. 2c). The remaining was a nonsense mutation. Analysis of the parents showed that all variants occurred de novo.

All four patients with *ZBTB18* mutations had ID ranging from mild to severe (Table 2). All had developmental delay with walking achieved by age 24 months (range



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**Fig. 3** Aspects of the corpus callosum (CC) on MRI in patients with *ZBTB18* and *HNRNPU* mutations and deletions. **a** Normal CC in a patient with *HNRNPU* mutation (*left*) and ThCC in two patients with *HNRNPU* deletions (*middle* and *right*). **b** Partial AgCC (*left*)

and *right*) and short DysCC (*middle*) in three patients with *ZBTB18* mutations. **c** ThCC (*left*) and normal CC (*middle*) in two patients with *AKT3* + *ZBTB18* deletions. Partial AgCC in a patient with a deletion encompassing *AKT3* + *ZBTB18* + *HNRNPU* (*right*)



Table 2 Molecular and clinical characteristics of patients with ZBTB18 mutations

	į	51		174
Fatient ID	71	77	73	7.4
General data				
Ethnic origin	Caucasian	Caucasian	Caucasian	Caucasian
Gender	M	Т	M	Ĺ
Age at last examination (years)	14	12	23	12
Genetic data				
Variant position (hg19)	g.244217120A>G	g.244218467G>A	g.244218377T>C	g.244217675del
cDNA change (NM_205768.2)	c.44A > G	c.1391G > A	c.1301T > C	c.599del
Amino acid change	p.His15Arg	p.Arg464His	p.Leu434Pro	p.Ser200*
Exon no	2	2	2	2
Inheritance	De novo	De novo	De novo	De novo
Epilepsy				
Epilepsy	No	No	Yes	Yes
Age of first seizure	NA	NA	9 years	8 months
EEG	NA	5 years: few spikes	NA	Normal, then sz recorded, originating from left occipital lobe; later, spikes and occasional SW in the right
Development				centro-parietal region
Age of sitting	NA	NA	10 months	<9 months
Age of walking	27 months	22 months		24 months
First words	delayed	3 years	NA	24 months
Current language abilities	11 years: 100 words; 12–14 years: short sentences	12 years: reads syllables	16 years: few words	Short sentences, pronunciation dif- ficulties, good comprehension
Use of hands	Purposeful	Purposeful	Flapping of hands	Purposeful
Other	Oral dyspraxia, appropriate behavior	Hyperactive, tick disorder, obsessive-compulsive behavior (10 years)	NA	Severe behavioral problems: opposition, intolerance to frustration, psychic rigidity, temper tantrums, hyperphagia
Global developmental level	Moderate ID with prominent speech delay	Mild-moderate ID (WISC-IV 6 years: VIQ 58, PIQ 49)	Moderate/severe ID	Moderate ID
Clinical examination				
Height (SD)/weight (SD)/OFC (SD)/age in years	172 cm (+1)/44 kg (-0.25)/54.5 cm (0)/14	142 cm $(-1.75)/29.5$ kg $(-1.5)/50$ cm $(-2.25)/12$	192 cm (>+3)/NA/54.5 cm (-1)/23	155.5 cm (+1)/54.9 kg (+2)/54 cm (+0.5)/12.5
Neurological examination	Infantile hypotonia	Normal	Normal	Normal
Brain MRI	Partial agenesis of CC	Short and dysgenetic CC	NA	Thin CC with hypoplastic splenium, mild enlargement of cerebellar interfolial spaces, wide Virchow-Robin spaces, diffuse hypomyelination



22–27 months). Language abilities were highly variable, with two 12-year-old patients speaking short sentences, one individual being able to read syllables at the same age, and the most severely affected adult patient speaking only a few words. Two patients experienced seizures from the age of 8 months and 9 years. Spells were tonic–clonic seizures and head turning with cyanosis. Epilepsy in the patient with the earliest onset rapidly responded to valproate therapy. The OFC was normal in three patients and borderline in another. AnCC was observed in the three patients who underwent brain MRI, with two having a partial AgCC and the other short CC (size <3rd percentile) classified as a DysCC (Fig. 3).

## Developmental delay and intellectual disability in patients with deletions and mutations

Except one, all patients with 1q43q44 microdeletions or point mutations in *ZBTB18* and *HNRNPU* had developmental delay or ID with a wide range of severity (Tables 1, 2, S2 and S3). The only exception was a patient with a borderline intellectual quotient and a deletion limited to *AKT3* (#L31) inherited from his father who was reported to have an intelligence within the normal range (Gai et al. 2015).

Since formal evaluations of cognitive functioning were unavailable for most other patients, we used the postural and language milestones to evaluate their developmental level. Walking abilities were available for 22 patients with 1q43q44 deletions older than 2 years. Four of them were unable to walk and 18 walked independently at a mean age of 35 months (median age 24 months). Among ambulatory patients, (1) three had microdeletions encompassing ZBTB18 and AKT3 but not HNRNPU and walked at a mean age of 18 months (range 17-19 months, median age 18 months), (2) 13 with a deletion including HNRNPU but not AKT3 and ZBTB18 walked at a mean age of 32.6 months (range 21–59 months, median age 24 months), and (3) two had a large deletion encompassing the three genes and walked at a mean age of 6.5 years (range 5-7 years). Among non-ambulatory patients, one carried a HNRNPU deletion and the three others had a deletion of the three genes. These data suggest that the loss of one HNRNPU allele has a more deleterious effect on walking acquisition than the loss of ATK3/ZBTB18 genes but larger deletions including all three genes have even more severe consequences. Walking abilities in patients with HNRNPU point mutations were similar to those with deletions of the gene. Patients with ZBTB18 mutations globally walked later than those with deletions including AKT3 and ZBTB18 although this should be confirmed on larger patient series.

Language abilities were available for 15 patients with 1q43q44 deletions older than 4 years of our series only.

Three patients did not speak any word, six had acquired a few words, six were able to speak short (n = 5) or full (n = 1) sentences. None of the four patients with a deletion of all three genes made sentences, whereas 2/2 patients with deletions including AKT3 and ZBTB18 but not HNRNPU, and 4/10 patients with HNRNPU deletions did. Thus, patients with HNRNPU point mutations globally had more severe speech impairments than patients with deletions including HNRNPU, who had more variable language abilities. Yet, language abilities were more preserved in patients with ZBTB18 point mutations and individuals with deletions including ZBTB18 but sparing HNRNPU, since 6/7 could speak sentences.

#### **Discussion**

The association of ID, microcephaly, AnCC and epilepsy characterizes the full neurodevelopmental phenotype of the 1q43q44 microdeletion syndrome. The recent identification of point mutations in *ZBTB18* and *HNRNPU* in patients with neurodevelopmental disability can help decipher genotype/phenotype correlations. In this study, we confirm that *AK3*, *ZBTB18* and *HNRNPU* are the main genes contributing to the phenotype of the 1q43q44 microdeletion syndrome, with each gene driving a specific feature although genetic interactions between these genes also exist.

Microcephaly has been reported in about half of the patients with 1q43q44 deletions. Previous genotype-phenotype correlation studies suggested that microcephaly is mainly associated with AKT3 haploinsufficiency (Ballif et al. 2012; Gai et al. 2015; Nagamani et al. 2012; Thierry et al. 2012). Our data, analyzing the alignment of 49 deletions from patients with known OFC confirmed this correlation. The occurrence of microcephaly in patients with microdeletions restricted to AKT3 (Ballif et al. 2012; Gai et al. 2015; Nagamani et al. 2012) narrows the minimal critical region to this single gene. This observation is consistent with a mirror phenotype consisting in macrocephaly in individuals with duplications encompassing AKT3 and segmental hypertrophy (in the form of hemimegalencephaly or syndromic megalencephaly) in individuals with missense mutations leading to increased mTOR signaling that are usually—but not always—limited to mosaic brain tissues (Fig. 2a) (Conti et al. 2015; Lee et al. 2012; Poduri et al. 2012; Riviere et al. 2012; Wang et al. 2013; Nellist et al. 2015; Takagi et al. 2017). It also confirms that dosage of AKT3 is crucial for controlling brain size during development. The observation of two patients and a healthy father with microdeletions limited to AKT3 and normal brain size (Gai et al. 2015) suggests that AKT3-related microcephaly is not fully penetrant. Alternatively, the fact that AKT3 deletions in



patients with normal OFC alter the 3' end of the gene could suggest that the region of AKT3 critical for microcephaly encompasses at least the first 5 exons but not the 3' coding part of the gene (Fig. 3a). This hypothesis is compatible with the description of three AKT3 isoforms encoding two distinct proteins differing in their 3' exons. No patient with constitutive point mutation leading to LoF of AKT3 has been reported so far, so we were unable to compare the phenotype of patients with point mutations and microdeletions; but our data suggest that point mutations resulting in LoF of AKT3, especially if located in the 5' exons common to the two known AKT3 isoforms, would result in microcephaly with or without ID.

The preponderant role of AKT3 in microcephaly does not exclude minor involvement of other genetic determinants. This is exemplified by the observation that: (1) three deletions sparing AKT3 (one encompassing ZBTB18 and HNRNPU and two HNRNPU but not ZBTB18) were also associated with microcephaly, and (2) all patients with deletion comprising AKT3 but extending to and including ZBTB18 and/or HNRNPU had microcephaly. The OFC is known for nine patients with ZBTB18 mutations [from the literature (Table S6) and our series] and three have microcephaly. This observation, combined with the microcephaly phenotype described in  $Zbtb18^{-/-}$ mice, suggests that microcephaly can also be associated with ZBTB18 mutations and deletions with a lower penetrance. In contrast, data from patients with HNRNPU mutations (Tables 1, S5) shows that heterozygous loss of HNRNPU is rarely associated with microcephaly. Therefore, we suggest that: (1) AKT3 haploinsufficiency is sufficient to cause microcephaly with high but incomplete penetrance, (2) the heterozygous loss of ZBTB18 may cause microcephaly with a lower penetrance, and (3) other regions located more distally (including ZBTB18) may contribute to microcephaly in addition to AKT3 deletion.

Previous genotype-phenotype correlation studies determined ZBTB18 to be the main candidate gene for AnCC in the 1q43q44 deletion syndrome (Ballif et al. 2012; Nagamani et al. 2012; Thierry et al. 2012). This hypothesis was supported by the analysis of deletion alignments and by the absence of CC in mice lacking both copies of RP58, the murine homologue of ZBTB18 (Xiang et al. 2012). However, the first three patients with heterozygous ZBTB18 mutations were reported to have a normal CC (de Munnik et al. 2014; Lopes et al. 2016; Rauch et al. 2012). This unexpected result has been challenged by the recent report of AnCC in four patients with ZBTB18 de novo mutations (Cohen et al. 2016). Similarly, three of our four patients with ZBTB18 mutations had AnCC. Genotype-phenotype correlations in our series of patients with 1q43q44 deletions confirm that *ZBTB18* is the main gene driving AnCC in the 1qter region. However, 5/22 patients with *ZBTB18* haploinsufficiency and 4/11 with *ZBTB18* point mutations had normal CC, indicating that like for many other genes previously associated with AgCC in humans, the AnCC related to *ZBTB18* is not a fully penetrant trait. Interestingly, most *ZBTB18* point mutations are truncating or located in the functional zinc-finger domain of the ZBTB18 protein. This mutation spectrum suggests that missense mutations could also lead to a LoF of *ZBTB18*, although this has to be confirmed by functional studies.

Considering different categories of AnCC, it appeared that: (1) no patient with 1q43q44 deletion sparing ZBTB18 had AgCC, (2) three patients carrying HNRNPU deletions sparing ZBTB18 and two with HNRNPU point mutations had ThCC (one had DysCC), and (3) patients with ZBTB18 point mutations had either partial AgCC, DysCC or ThCC. Thus, ThCC is the main category of AnCC observed when HNRNPU is deleted and is possibly related to insufficient myelination of crossing axons rather than indicating malformation of the CC. We conclude that ZBTB18 haploinsufficiency predisposes to different types of AnCC, particularly partial AgCC, while HNRNPU anomalies are more specifically associated with ThCC. Furthermore, AgCC is significantly more frequent in patients with microdeletions comprising ZBTB18 extending towards the telomeric end of the 1q region, i.e., encompassing both ZBTB18 and HNRNPU, compared to those encompassing ZBTB18 but sparing HNRNPU. This suggests that the loss of genetic determinant(s) in 3' of the ZBTB18 coding sequence or that the co-deletion of ZBTB18 and HNRNPU has an additive effect resulting in AgCC. Moreover, the AgCC observed in patients with point mutations or small deletions altering mainly ZBTB18 is partial and characterized by a small splenium with the absence of beak (patients #Z1 and #Z4 and #2, #3 and #5 in Cohen et al. 2016). However, complete or subtotal AgCC have been reported in patients with larger 1q deletions not included in our study (Boland et al. 2007; Caliebe et al. 2010; Hemming et al. 2016; Zaki et al. 2012), suggesting that one or more proximal genes on chromosome 1 could also lead or predispose to AgCC.

Two thirds of patients with 1q43q44 deletions have epilepsy. The minimal critical region for epilepsy included *HNRNPU* and *COX20*. *COX20* encodes a protein contributing to the assembly of mitochondrial cytochrome C oxidase and has been involved in a recessive disease with healthy heterozygous carriers (Doss et al. 2014; Szklarczyk et al. 2013). Thus, heterozygous deletions of *COX20* are unlikely to be responsible for the epilepsy phenotype. *HNRNPU* is the main gene accounting for seizures since: (1) epilepsy is present in 90% of patients with deletions comprising *HNRNPU* and absent in 14/15 patients with deletions sparing *HNRNPU*, and (2) all patients with constitutive *HNRNPU* mutations



have epilepsy. The only epileptic patient with a deletion sparing *HNRNPU* had a deletion encompassing *ZBTB18*. Given that 3/11 patients with *ZBTB18* mutations, including two of our series, had seizures; epilepsy may also be a minor phenotypic trait in some patients with *ZBTB18* haploinsufficiency. The pro-epileptogenic effect of *ZBTB18* alteration could be masked by the strong penetrance of *HNRNPU*-related epilepsy in patients with loss of both genes.

The mean age at seizure onset in patients with HNRNPU point mutations from both our series and the literature (n=9) was 13.5 months (median 8.5 months), versus 12.3 months (median 12 months) in individuals with deletions encompassing HNRNPU but sparing ZBTB18 (n=13) and 12.9 months (median 12 months) in patients with deletions encompassing both HNRNPU and ZBTB18 (n=7). These observations suggest that the age at seizure onset is independent of the size of the deletion, and that the loss of one HNRNPU allele is the strongest factor determining the age at seizure onset.

To date, available data did not reveal specific epileptic features in patients with *HNRNPU* mutations or deletions. Tonic–clonic seizures and atypical absences are the most frequently reported seizure types. Seizures occur with variable frequencies, are frequently triggered by fever at the onset of the disease and are pharmacoresistant in some patients. Diffuse or focal slow-waves or a slow background activity are recurrently reported on EEG recordings together with various epileptiform features.

ID is reported in almost all patients carrying 1q43q44 deletions, but its severity is frequently unmentioned. The only individuals without ID had a deletion limited to the whole *AKT3* gene (Gai et al. 2015). The cognitive abilities of other patients with microdeletions limited to *AKT3* are unknown but at least two of them were reported to have ID (Nagamani et al. 2012). All patients with *ZBTB18* mutations known to date have ID with variable degrees of severity, except one with "overall cognitive ability in the low average range" (Cohen et al. 2016). In contrast, no patients with *HNRNPU* mutation or with *HNRNPU* deletion and normal development have been reported.

Patients with ZBTB18 mutations from the literature walked at a mean age of 28 months (n=8) and 5/8 of them aged 3–37 years were not able to speak sentences or to associate several words. Thus, the impression of a relatively preserved development in patients with ZBTB18 mutations from our series should probably be attenuated. These differences are apparently unrelated to the nature (missense *versus* truncating) of the mutation but likely to the small sample sizes. An overview of acquired developmental milestones in these series of patients shows that those with HNRNPU deletions or mutations have a globally more severe postural and speech delay that those with AKT3/ZBTB18 deletions and those with ZBTB18 mutations.

Because the most severe group of patients have deletions encompassing *HNRNPU*, *AKT3* and *ZBTB18*, co-deletions of these three genes could have an additive detrimental effect on neurodevelopment.

In conclusion, the complete neurodevelopmental phenotype of the 1q43q44 microdeletion syndrome is the consequence of the deletion of three main genes spanning 1.36 Mb. Our data confirm that AKT3 is the main gene driving microcephaly, ZBTB18 defect is responsible for AnCC and HNRNPU is the main gene accounting for epilepsy. These correlations can be summarized as follows: (1) AKT3 deletion causes microcephaly with incomplete penetrance but ZBTB18 and HNRNPU deletions may also be involved with a weaker effect; (2) epilepsy and the loss of one HNRNPU allele are strongly associated; and (3) AgCC, is dependent on the loss of ZBTB18 allele but is also influenced by the alteration of neighboring genes. Neurodevelopmental impairment in patients with ZBTB18 LoF is more variable and less severe than that with HNRNPU LoF. Additional studies are required to investigate factors controlling this phenotypic variability in more details, including in particular the possibility of modifiers variants located on the trans allele.

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#### Compliance with ethical standards

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Conflict of interest The authors declare no conflict of interest.

**Ethical approval** The study received approval from local ethical standards committees on human experimentation.



**Informed consent** Informed written consent was obtained from each individual or their parents or legal representatives before blood sampling.

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#### References

- Ballif BC, Rosenfeld JA, Traylor R, Theisen A, Bader PI, Ladda RL, Sell SL, Steinraths M, Surti U, McGuire M, Williams S, Farrell SA, Filiano J, Schnur RE, Coffey LB, Tervo RC, Stroud T, Marble M, Netzloff M, Hanson K, Aylsworth AS, Bamforth JS, Babu D, Niyazov DM, Ravnan JB, Schultz RA, Lamb AN, Torchia BS, Bejjani BA, Shaffer LG (2012) High-resolution array CGH defines critical regions and candidate genes for microcephaly, abnormalities of the corpus callosum, and seizure phenotypes in patients with microdeletions of 1q43q44. Hum Genet 131:145–156. doi:10.1007/s00439-011-1073-y
- Baubet V, Xiang C, Molczan A, Roccograndi L, Melamed S, Dahmane N (2012) Rp58 is essential for the growth and patterning of the cerebellum and for glutamatergic and GABAergic neuron development. Development 139:1903–1909. doi:10.1242/dev.075606
- Boland E, Clayton-Smith J, Woo VG, McKee S, Manson FD, Medne L, Zackai E, Swanson EA, Fitzpatrick D, Millen KJ, Sherr EH, Dobyns WB, Black GC (2007) Mapping of deletion and translocation breakpoints in 1q44 implicates the serine/ threonine kinase AKT3 in postnatal microcephaly and agenesis of the corpus callosum. Am J Hum Genet 81:292–303. doi:10.1086/519999
- Caliebe A, Kroes HY, van der Smagt JJ, Martin-Subero JI, Tonnies H, van't Slot R, Nievelstein RA, Muhle H, Stephani U, Alfke K, Stefanova I, Hellenbroich Y, Gillessen-Kaesbach G, Hochstenbach R, Siebert R, Poot M (2010) Four patients with speech delay, seizures and variable corpus callosum thickness sharing a 0.440 Mb deletion in region 1q44 containing the HNRPU gene. Eur J Med Genet 53:179–185. doi:10.1016/j.eimg.2010.04.001
- Carvill GL, Heavin SB, Yendle SC, McMahon JM, O'Roak BJ, Cook J, Khan A, Dorschner MO, Weaver M, Calvert S, Malone S, Wallace G, Stanley T, Bye AM, Bleasel A, Howell KB, Kivity S, Mackay MT, Rodriguez-Casero V, Webster R, Korczyn A, Afawi Z, Zelnick N, Lerman-Sagie T, Lev D, Moller RS, Gill D, Andrade DM, Freeman JL, Sadleir LG, Shendure J, Berkovic SF, Scheffer IE, Mefford HC (2013) Targeted resequencing in epileptic encephalopathies identifies de novo mutations in CHD2 and SYNGAP1. Nat Genet 45:825–830. doi:10.1038/ng.2646
- Cohen JS, Srivastava S, Farwell Hagman KD, Shinde DN, Huether R, Darcy D, Wallerstein R, Houge G, Berland S, Monaghan KG, Poretti A, Wilson AL, Chung WK, Fatemi A (2016) Further evidence that de novo missense and truncating variants in ZBTB18 cause intellectual disability with variable features. Clin Genet. doi:10.1111/cge.12861
- Conti V, Pantaleo M, Barba C, Baroni G, Mei D, Buccoliero AM, Giglio S, Giordano F, Baek ST, Gleeson JG, Guerrini R (2015) Focal dysplasia of the cerebral cortex and infantile spasms associated with somatic 1q21.1-q44 duplication including the AKT3 gene. Clin Genet 88:241–247. doi:10.1111/cge.12476

- de Kovel CG, Brilstra EH, van Kempen MJ, Van't Slot R, Nijman IJ, Afawi Z, De Jonghe P, Djemie T, Guerrini R, Hardies K, Helbig I, Hendrickx R, Kanaan M, Kramer U, Lehesjoki AE, Lemke JR, Marini C, Mei D, Moller RS, Pendziwiat M, Stamberger H, Suls A, Weckhuysen S, Euro ERESC, Koeleman BP (2016) Targeted sequencing of 351 candidate genes for epileptic encephalopathy in a large cohort of patients. Mol Genet Genom Med 4:568–580. doi:10.1002/mgg3.235
- de Munnik SA, Garcia-Minaur S, Hoischen A, van Bon BW, Boycott KM, Schoots J, Hoefsloot LH, Knoers NV, Bongers EM, Brunner HG (2014) A de novo non-sense mutation in ZBTB18 in a patient with features of the 1q43q44 microdeletion syndrome. Eur J Hum Genet 22:844–846. doi:10.1038/ejhg.2013.249
- Doss S, Lohmann K, Seibler P, Arns B, Klopstock T, Zuhlke C, Freimann K, Winkler S, Lohnau T, Drungowski M, Nurnberg P, Wiegers K, Lohmann E, Naz S, Kasten M, Bohner G, Ramirez A, Endres M, Klein C (2014) Recessive dystonia-ataxia syndrome in a Turkish family caused by a COX20 (FAM36A) mutation. J Neurol 261:207–212. doi:10.1007/s00415-013-7177-7
- Du X, An Y, Yu L, Liu R, Qin Y, Guo X, Sun D, Zhou S, Wu B, Jiang YH, Wang Y (2014) A genomic copy number variant analysis implicates the MBD5 and HNRNPU genes in Chinese children with infantile spasms and expands the clinical spectrum of 2q23.1 deletion. BMC Med Genet 15:62. doi:10.1186/1471-2350-15-62
- Easton RM, Cho H, Roovers K, Shineman DW, Mizrahi M, Forman MS, Lee VM, Szabolcs M, de Jong R, Oltersdorf T, Ludwig T, Efstratiadis A, Birnbaum MJ (2005) Role for Akt3/protein kinase Bgamma in attainment of normal brain size. Mol Cell Biol 25:1869–1878. doi:10.1128/MCB.25.5.1869-1878.2005
- Epi4K consortium, Epilepsy Phenome/Genome P, Allen AS, Berkovic SF, Cossette P, Delanty N, Dlugos D, Eichler EE, Epstein MP, Glauser T, Goldstein DB, Han Y, Heinzen EL, Hitomi Y, Howell KB, Johnson MR, Kuzniecky R, Lowenstein DH, Lu YF, Madou MR, Marson AG, Mefford HC, Esmaeeli Nieh S, O'Brien TJ, Ottman R, Petrovski S, Poduri A, Ruzzo EK, Scheffer IE, Sherr EH, Yuskaitis CJ, Abou-Khalil B, Alldredge BK, Bautista JF, Berkovic SF, Boro A, Cascino GD, Consalvo D, Crumrine P, Devinsky O, Dlugos D, Epstein MP, Fiol M, Fountain NB, French J, Friedman D, Geller EB, Glauser T, Glynn S, Haut SR, Hayward J, Helmers SL, Joshi S, Kanner A, Kirsch HE, Knowlton RC, Kossoff EH, Kuperman R, Kuzniecky R, Lowenstein DH, McGuire SM, Motika PV, Novotny EJ, Ottman R, Paolicchi JM, Parent JM, Park K, Poduri A, Scheffer IE, Shellhaas RA, Sherr EH, Shih JJ, Singh R, Sirven J, Smith MC, Sullivan J, Lin Thio L, Venkat A, Vining EP, Von Allmen GK, Weisenberg JL, Widdess-Walsh P, Winawer MR (2013) De novo mutations in epileptic encephalopathies. Nature 501:217-221. doi:10.1038/nature12439
- Firth HV, Richards SM, Bevan AP, Clayton S, Corpas M, Rajan D, Van Vooren S, Moreau Y, Pettett RM, Carter NP (2009) DECI-PHER: database of chromosomal imbalance and phenotype in humans using ensembl resources. Am J Hum Genet 84:524–533. doi:10.1016/j.ajhg.2009.03.010
- Gai D, Haan E, Scholar M, Nicholl J, Yu S (2015) Phenotypes of AKT3 deletion: a case report and literature review. Am J Med Genet A 167A:174–179. doi:10.1002/ajmg.a.36710
- Gupta R, Agarwal M, Boqqula VR, Phadke RV, Phadke SR (2014) Hemiconvulsion-hemiplegia-epilepsy syndrome with 1q44 microdeletion: causal or chance association. Am J Med Genet A 164A:186–189. doi:10.1002/ajmg.a.36198
- Hamdan FF, Srour M, Capo-Chichi JM, Daoud H, Nassif C, Patry
   L, Massicotte C, Ambalavanan A, Spiegelman D, Diallo O,
   Henrion E, Dionne-Laporte A, Fougerat A, Pshezhetsky AV,



Venkateswaran S, Rouleau GA, Michaud JL (2014) De novo mutations in moderate or severe intellectual disability. PLoS Genet 10:e1004772. doi:10.1371/journal.pgen.1004772

- Hemming IA, Forrest AR, Shipman P, Woodward KJ, Walsh P, Ravine DG, Heng JI (2016) Reinforcing the association between distal 1q CNVs and structural brain disorder: a case of a complex 1q43-q44 CNV and a review of the literature. Am J Med Genet B Neuropsychiatr Genet 171B:458–467. doi:10.1002/ajmg.b.32427
- Heng JI, Qu Z, Ohtaka-Maruyama C, Okado H, Kasai M, Castro D, Guillemot F, Tan SS (2015) The zinc finger transcription factor RP58 negatively regulates Rnd2 for the control of neuronal migration during cerebral cortical development. Cereb Cortex 25:806–816. doi:10.1093/cercor/bht277
- Huang N, Lee I, Marcotte EM, Hurles ME (2010) Characterising and predicting haploinsufficiency in the human genome. PLoS Genet 6:e1001154. doi:10.1371/journal.pgen.1001154
- Lee JH, Huynh M, Silhavy JL, Kim S, Dixon-Salazar T, Heiberg A, Scott E, Bafna V, Hill KJ, Collazo A, Funari V, Russ C, Gabriel SB, Mathern GW, Gleeson JG (2012) De novo somatic mutations in components of the PI3K-AKT3-mTOR pathway cause hemimegalencephaly. Nat Genet 44:941–945. doi:10.1038/ng.2329
- Lopes F, Barbosa M, Ameur A, Soares G, de Sa J, Dias AI, Oliveira G, Cabral P, Temudo T, Calado E, Cruz IF, Vieira JP, Oliveira R, Esteves S, Sauer S, Jonasson I, Syvanen AC, Gyllensten U, Pinto D, Maciel P (2016) Identification of novel genetic causes of Rett syndrome-like phenotypes. J Med Genet 53:190–199. doi:10.1136/jmedgenet-2015-103568
- Mignot C, Moutard ML, Rastetter A, Boutaud L, Heide S, Billette T, Doummar D, Garel C, Afenjar A, Jacquette A, Lacombe D, Verloes A, Bole-Feysot C, Nitschke P, Masson C, Faudet A, Lesne F, Bienvenu T, Alby C, Attie-Bitach T, Depienne C, Nava C, Heron D (2016) ARID1B mutations are the major genetic cause of corpus callosum anomalies in patients with intellectual disability. Brain. doi:10.1093/brain/aww181
- Mirzaa GM, Riviere JB, Dobyns WB (2013) Megalencephaly syndromes and activating mutations in the PI3 K-AKT pathway: MPPH and MCAP. Am J Med Genet C Semin Med Genet 163C:122–130. doi:10.1002/ajmg.c.31361
- Monroe GR, Frederix GW, Savelberg SM, de Vries TI, Duran KJ, van der Smagt JJ, Terhal PA, van Hasselt PM, Kroes HY, Verhoeven-Duif NM, Nijman IJ, Carbo EC, van Gassen KL, Knoers NV, Hovels AM, van Haelst MM, Visser G, van Haaften G (2016) Effectiveness of whole-exome sequencing and costs of the traditional diagnostic trajectory in children with intellectual disability. Genet Med 18:949–956. doi:10.1038/gim.2015.200
- Nagamani SC, Erez A, Bay C, Pettigrew A, Lalani SR, Herman K, Graham BH, Nowaczyk MJ, Proud M, Craigen WJ, Hopkins B, Kozel B, Plunkett K, Hixson P, Stankiewicz P, Patel A, Cheung SW (2012) Delineation of a deletion region critical for corpus callosal abnormalities in chromosome 1q43-q44. Eur J Hum Genet 20:176–179. doi:10.1038/ejhg.2011.171
- Need AC, Shashi V, Hitomi Y, Schoch K, Shianna KV, McDonald MT, Meisler MH, Goldstein DB (2012) Clinical application of exome sequencing in undiagnosed genetic conditions. J Med Genet 49:353–361. doi:10.1136/jmedgenet-2012-100819
- Nellist M, Schot R, Hoogeveen-Westerveld M, Neuteboom RF, van der Louw EJ, Lequin MH, Bindels-de Heus K, Sibbles BJ, de Coo R, Brooks A, Mancini GM (2015) Germline activating AKT3 mutation associated with megalencephaly, polymicrogyria, epilepsy and hypoglycemia. Mol Genet Metab 114(3):467–473. doi:10.1016/j.ymgme.2014.11.018
- Perlman SJ, Kulkarni S, Manwaring L, Shinawi M (2013) Haploinsufficiency of ZNF238 is associated with corpus callosum abnormalities in 1q44 deletions. Am J Med Genet A 161A:711–716. doi:10.1002/ajmg.a.35779

- Poduri A, Evrony GD, Cai X, Elhosary PC, Beroukhim R, Lehtinen MK, Hills LB, Heinzen EL, Hill A, Hill RS, Barry BJ, Bourgeois BF, Riviello JJ, Barkovich AJ, Black PM, Ligon KL, Walsh CA (2012) Somatic activation of AKT3 causes hemispheric developmental brain malformations. Neuron 74:41–48. doi:10.1016/j.neuron.2012.03.010
- Rauch A, Wieczorek D, Graf E, Wieland T, Endele S, Schwarzmayr T, Albrecht B, Bartholdi D, Beygo J, Di Donato N, Dufke A, Cremer K, Hempel M, Horn D, Hoyer J, Joset P, Ropke A, Moog U, Riess A, Thiel CT, Tzschach A, Wiesener A, Wohlleber E, Zweier C, Ekici AB, Zink AM, Rump A, Meisinger C, Grallert H, Sticht H, Schenck A, Engels H, Rappold G, Schrock E, Wieacker P, Riess O, Meitinger T, Reis A, Strom TM (2012) Range of genetic mutations associated with severe non-syndromic sporadic intellectual disability: an exome sequencing study. Lancet 380:1674–1682. doi:10.1016/S0140-6736(12)61480-9
- Riviere JB, Mirzaa GM, O'Roak BJ, Beddaoui M, Alcantara D, Conway RL, St-Onge J, Schwartzentruber JA, Gripp KW, Nikkel SM, Worthylake T, Sullivan CT, Ward TR, Butler HE, Kramer NA, Albrecht B, Armour CM, Armstrong L, Caluseriu O, Cytrynbaum C, Drolet BA, Innes AM, Lauzon JL, Lin AE, Mancini GM, Meschino WS, Reggin JD, Saggar AK, Lerman-Sagie T, Uyanik G, Weksberg R, Zirn B, Beaulieu CL, Finding of Rare Disease Genes Canada C, Majewski J, Bulman DE, O'Driscoll M, Shendure J, Graham JM Jr, Boycott KM, Dobyns WB (2012) De novo germline and postzygotic mutations in AKT3, PIK3R2 and PIK3CA cause a spectrum of related megalencephaly syndromes. Nat Genet 44:934–940. doi:10.1038/ng.2331
- Roshon MJ, Ruley HE (2005) Hypomorphic mutation in hnRNP U results in post-implantation lethality. Transgenic Res 14:179–192
- Sobreira N, Schiettecatte F, Valle D, Hamosh A (2015) GeneMatcher: a matching tool for connecting investigators with an interest in the same gene. Hum Mutat 36:928–930. doi:10.1002/ humu.22844
- Szklarczyk R, Wanschers BF, Nijtmans LG, Rodenburg RJ, Zschocke J, Dikow N, van den Brand MA, Hendriks-Franssen MG, Gilissen C, Veltman JA, Nooteboom M, Koopman WJ, Willems PH, Smeitink JA, Huynen MA, van den Heuvel LP (2013) A mutation in the FAM36A gene, the human ortholog of COX20, impairs cytochrome c oxidase assembly and is associated with ataxia and muscle hypotonia. Hum Mol Genet 22:656–667. doi:10.1093/hmg/dds473
- Takagi M, Dobashi K, Nagahara K, Kato M, Nishimura G, Fukuzawa R, Narumi S, Hasegawa T (2017) A novel de novo germline mutation Glu40Lys in AKT3 causes megalencephaly with growth hormone deficiency. Am J Med Genet A. doi:10.1002/ajmg.a.38099 (Epub ahead of print)
- Thierry G, Beneteau C, Pichon O, Flori E, Isidor B, Popelard F, Delrue MA, Duboscq-Bidot L, Thuresson AC, van Bon BW, Cailley D, Rooryck C, Paubel A, Metay C, Dusser A, Pasquier L, Beri M, Bonnet C, Jaillard S, Dubourg C, Tou B, Quere MP, Soussi-Zander C, Toutain A, Lacombe D, Arveiler B, de Vries BB, Jonveaux P, David A, Le Caignec C (2012) Molecular characterization of 1q44 microdeletion in 11 patients reveals three candidate genes for intellectual disability and seizures. Am J Med Genet A 158A:1633–1640. doi:10.1002/ajmg.a.35423
- Wang D, Zeesman S, Tarnopolsky MA, Nowaczyk MJ (2013) Duplication of AKT3 as a cause of macrocephaly in duplication 1q43q44. Am J Med Genet A 161A:2016–2019. doi:10.1002/ajmg.a.35999
- Xiang C, Baubet V, Pal S, Holderbaum L, Tatard V, Jiang P, Davuluri RV, Dahmane N (2012) RP58/ZNF238 directly modulates proneurogenic gene levels and is required for neuronal differentiation and brain expansion. Cell Death Differ 19:692–702. doi:10.1038/cdd.2011.144



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- Ye J, Beetz N, O'Keeffe S, Tapia JC, Macpherson L, Chen WV, Bassel-Duby R, Olson EN, Maniatis T (2015) hnRNP U protein is required for normal pre-mRNA splicing and postnatal heart development and function. Proc Natl Acad Sci USA 112:E3020–E3029. doi:10.1073/pnas.1508461112
- Zaki MS, Gillessen-Kaesbach G, Vater I, Caliebe A, Siebert R, Kamel AK, Mohamed AM, Mazen I (2012) Bladder exstrophy and extreme genital anomaly in a patient with pure terminal 1q deletion: expansion of phenotypic spectrum. Eur J Med Genet 55:43–48. doi:10.1016/j.ejmg.2011.09.003
- Zhu X, Petrovski S, Xie P, Ruzzo EK, Lu YF, McSweeney KM, Ben-Zeev B, Nissenkorn A, Anikster Y, Oz-Levi D, Dhindsa RS, Hitomi Y, Schoch K, Spillmann RC, Heimer G, Marek-Yagel D, Tzadok M, Han Y, Worley G, Goldstein J, Jiang YH, Lancet D, Pras E, Shashi V, McHale D, Need AC, Goldstein DB (2015) Whole-exome sequencing in undiagnosed genetic diseases: interpreting 119 trios. Genet Med 17:774–781. doi:10.1038/gim.2014.191

#### Urls/Resources

BIOBASE HGMD Professional: http://www.biobase-international.com/product/hgmd

Exome Variant Server: http://evs.gs.washington.edu/EVS/

ExAC Browser (Beta) | Exome Aggregation Consortium: http://exac.broadinstitute.org/

GeneMatcher: https://genematcher.org/

NCBI Pubmed: http://www.ncbi.nlm.nih.gov/pubmed

Uniprot: http://www.uniprot.org/

