Variable Cold-Induced Brown Adipose Tissue Response to Thyroid Hormone Status

The Harvard community has made this article openly available. Please share how this access benefits you. Your story matters.

<table>
<thead>
<tr>
<th>Citation</th>
<th>Gavrila, Alina, Per-Olof Hasselgren, Allison Glasgow, Ashley N. Doyle, Alice J. Lee, Peter Fox, Shiva Gautam, James V. Hennessey, Gerald M. Kolodny, and Aaron M. Cypess. 2017. “Variable Cold-Induced Brown Adipose Tissue Response to Thyroid Hormone Status.” Thyroid 27 (1) (January): 1–10. doi:10.1089/thy.2015.0646.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Published Version</td>
<td>doi:10.1089/thy.2015.0646</td>
</tr>
<tr>
<td>Citable Link</td>
<td><a href="http://nrs.harvard.edu/urn-3:HUL.InstRepos:32697818">http://nrs.harvard.edu/urn-3:HUL.InstRepos:32697818</a></td>
</tr>
<tr>
<td>Terms of Use</td>
<td>This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Open Access Policy Articles, as set forth at <a href="http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#OAP">http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#OAP</a></td>
</tr>
</tbody>
</table>

(Article begins on next page)
Variable Cold-induced Brown Adipose Tissue Response to Thyroid Hormone Status

Alina Gavrila, MD, MMSC¹; Per-Olof Hasselgren, MD, PhD²; Allison Glasgow³; Ashley N. Doyle³, Alice J. Lee³, Peter Fox⁵, Shiva Gautam, PhD¹, James V. Hennessey, MD¹; Gerald M. Kolodny, MD³; Aaron M. Cypess, MD, PhD, MMSC¹,⁵,⁶

¹Department of Medicine, ²Department of Surgery, ³Harvard Catalyst Clinical Research Center, ⁴Radiology, Beth Israel Deaconess Medical Center, Boston, MA, 02215; ⁵Section of Integrative Physiology and Metabolism, Research Division, Joslin Diabetes Center, Harvard Medical School, Boston, MA, 02215; ⁶Diabetes, Endocrinology, and Obesity Branch, National Institute of Diabetes and Digestive and Kidney Diseases, NIH, Bethesda, MD 20892, USA.

Email addresses: agavrila@bidmc.harvard.edu; phasselg@bidmc.harvard.edu; aglasgow02@gmail.com; andoyle2@gmail.com; alee16@bidmc.harvard.edu; pfox1986@gmail.com; sgautam@bidmc.harvard.edu; jhennes@bidmc.harvard.edu; gkolodny@bidmc.harvard.edu; aaron.cypess@nih.gov.

Running Title: Thyroid Hormone and Brown Adipose Tissue

Keywords: Brown adipose tissue, thyroid hormone, hypothyroidism, thyrotoxicosis, thermogenesis.
Abstract

**Background:** In addition to its role in adaptive thermogenesis, brown adipose tissue (BAT) may protect from weight gain, insulin resistance/diabetes and metabolic syndrome. Prior studies have shown contradictory results regarding the influence of thyroid hormone (TH) levels on BAT volume and activity. The aim of this pilot study was to gain further insights regarding the effect of TH treatment on BAT function in adult humans by evaluating the BAT mass and activity prospectively in six patients, first in the hypothyroid and then in the thyrotoxic phase.

**Methods:** The study subjects underwent 18F-FDG PET-CT scanning after cold exposure to measure BAT mass and activity while undergoing treatment for differentiated thyroid cancer, first while hypothyroid following thyroid hormone withdrawal at the time of the radioactive iodine treatment and then 3-6 months after starting TH suppressive treatment when they were iatrogenically thyrotoxic. We measured thermogenic and metabolic parameters in both phases.

**Results:** All study subjects had detectable BAT under cold stimulation in both the hypothyroid and thyrotoxic state. The majority but not all (4 out of 6) subjects showed an increase in detectable BAT volume and activity under cold stimulation between the hypothyroid and thyrotoxic phase (total BAT volume: 72.0 ± 21.0 vs. 87.7 ± 16.5 mL, P = 0.25; total BAT activity 158.1 ± 72.8 vs. 189.0 ± 55.5 SUV*g/mL, P = 0.34). Importantly, circulating T3 was a stronger predictor of energy expenditure changes compared to cold-induced BAT activity.

**Conclusions:** Iatrogenic hypothyroidism lasting 2-4 weeks does not prevent cold-induced BAT activation, while the use of TH to induce thyrotoxicosis does not consistently increase cold-induced BAT activity. It remains to be determined which physiological factors besides TH play a role in regulating BAT function.
Introduction

In the midst of worldwide obesity and diabetes epidemics, it is crucial to find new avenues to gain insight into the pathophysiology and treatment of these disorders. Brown adipose tissue (BAT) is of particular interest since in addition to its role in adaptive thermogenesis, it may also protect against weight gain, insulin resistance/diabetes and metabolic syndrome (1-3). Indeed, because of its very high metabolic activity, very little functional BAT can have a profound metabolic impact (4, 5).

Studies using PET/CT scanning have shown metabolically active BAT in adult humans whose activity correlated inversely with body fat (5, 7, 8). Significant amounts of functional BAT have been detected in more than 95% of lean subjects in response to acute and chronic cold exposure in controlled prospective studies (5, 8, 9, 10). Since BAT activity is induced by cold exposure, BAT might play an important role in human thermogenesis and energy balance. This is in accordance with studies in small rodents, which showed that cold-exposure stimulates BAT activity by increasing sympathetic nerve activity and by affecting local thyroid hormone (TH) metabolism (11, 12). Cold exposure increases the expression and activity of tissue deiodinase 2 (DIO2), which stimulates the conversion of thyroxine (T4) to active triiodothyronine (T3), resulting in increased local T3 production from T4 and TH receptor saturation (13). Both catecholamines and local T3 act synergistically to stimulate uncoupling protein 1 (UCP-1) expression and through this thermogenesis in BAT. UCP-1 uncouples oxidative phosphorylation, thus representing the key protein in BAT thermogenesis (12). TH also has direct effects on oxidative phosphorylation and regulates the mitochondrial proliferation, differentiation and maturation through genomic and non-genomic mechanisms (14, 15). In addition, TH is known to play an important role in brown adipocyte differentiation (16, 17).
Thus, TH has been shown to have an important role in cold adaptation in rodents, and it may be important for temperature adaptation in humans (18). Understanding cold adaptation in adult humans is important, since factors that increase energy dissipation through facultative thermogenesis could potentially be used as anti-obesity agents (18, 19). If TH can influence BAT mass and activity and through this energy expenditure, then TH or its analogues might be useful in the treatment of obesity and the metabolic syndrome.

A few published human studies have shown contradictory results reporting increased BAT volume and activity in either hypothyroid or thyrotoxic subjects (20-23). To gain further insight regarding the effect of TH status on BAT in adult humans and whether it could be a feasible treatment strategy for activating BAT thermogenesis, we evaluated the BAT mass and activity prospectively in six patients who were undergoing treatment for thyroid cancer, first in the hypothyroid and then in the thyrotoxic phase. The study patients underwent thyroidectomy followed by TH withdrawal to induce hypothyroidism prior to the radioactive iodine (RAI) treatment and then started TH suppressive treatment with the goal of maintaining mild thyrotoxicosis long-term to prevent recurrence. The study patients underwent two PET-CT scans to measure BAT mass and activity, first while profoundly hypothyroid at the time of the RAI treatment and then three to six months after starting TH suppressive treatment when they were thyrotoxic. We compared the BAT mass and activity measured on these two scans. We also evaluated the relationship between BAT, TH status, and calorimetric and metabolic parameters.
Materials and Methods

Study Participants

Eight patients diagnosed with papillary thyroid cancer who underwent thyroidectomy and were scheduled to receive RAI treatment after TH withdrawal were recruited for the study. The study was approved by the Institutional Review Board of Beth Israel Deaconess Medical Center (BIDMC) and Joslin Diabetes Center, and patients provided written informed consent. Exclusion criteria included use of medications that can affect BAT, such as beta blockers, adrenergic agonists, benzodiazepines; use of medications that can affect thyroid function test interpretation, including estrogen, aspirin, dilantin; use of systemic, oral or intravenous corticosteroids or other medications known to cause insulin resistance in the previous 6 weeks. Patients with a history of any local or systemic infectious disease with fever or requiring antibiotic within four weeks of the PET/CT scans were not enrolled in the study.

Study Protocol (Figure 1)

The patients participated in a short screening visit to evaluate eligibility prior to the RAI treatment followed by two longer study visits when the whole-body BAT volume and activity were measured by 18F-FDG PET/CT under cold stimulation at 15-16°C, first in the hypothyroid state at the time of the RAI treatment and then in the thyrotoxic state 3-6 months after starting TH suppressive treatment. Six out of the eight patients who started the study completed all study visits and were included in the analysis. Prior to participating in the study, one patient underwent completion thyroidectomy for a thyroid nodule positive for papillary thyroid carcinoma on fine needle aspiration cytology, while five patients underwent hemithyroidectomy followed by completion thyroidectomy for thyroid nodules with fine needle aspiration showing
indeterminate cytology (2 patients), for a large thyroid nodule with benign cytology (1 patient),
for a large thyroid nodule with non-diagnostic cytology (1 patient), and for recurrent thyroid
cancer after undergoing lobectomy in the remote past (1 patient). Preparation for the
hypothyroid phase of the evaluation occurred at least 6 weeks postoperatively for 5 patients and
consisted of a 6 week L-thyroxine (LT4) withdrawal with a 4 week liothyronine bridging support
(12.5 mcg twice daily) for minimization of clinical symptoms followed by 2 weeks of complete
thyroid hormone withdrawal and a low iodine diet prior to the clinically indicated 131-I
administration. One study patient was already hypothyroid at the time of her initial endocrine
clinic visit with a TSH of 22 (reference range 0.27-4.2 µIU/mL); the levothyroxine was
discontinued at that time and the patient received RAI treatment 2 weeks later without the
liothyronine bridging support.

The patients were prepared for the study visits as previously described (24, 25). Baseline
vital signs and laboratory tests (thyroid function tests, insulin, HbA1c, and lipid panel) were
obtained. Baseline indirect calorimetry was performed during both the hypothyroid and
thyrotoxic phase after the patients were placed in a supine position and quiet environment for 30
minutes in the Clinical Research Center (CRC). The room temperature was maintained above
23°C throughout the stay in the CRC. Core body temperature was monitored during the study
visits.

To maximize the BAT activity/FDG uptake and PET-CT scan BAT visualization, the
patients were then transported to a room set at 20°C and wore a surgeon's cooling vest (Polar
Products, Inc.) set to 15-16°C, as described previously (24, 25). Sixty minutes after wearing the
vest, a second set of vital signs and laboratory tests (thyroid function tests, glucose, insulin, free
fatty acids) were obtained and an intravenous bolus of 592 MBq (16 mCi) of 18F-FDG was
administered; this dose was calculated based on the sensitivity of the scanner available at the
time of the study. The patients remained in the semi-darkened quiet room for another 60 minutes
while wearing the cooling vest, and at this time, a second measure of indirect calorimetry was
performed. Following 120 minutes of cold exposure, the vest was removed, and BAT images
were acquired using a 4-MDCT scanner (Discovery/LightSpeed PET/CT, GE Healthcare)
following a previously described imaging protocol (24). BAT volume, metabolic activity and
maximum standardized uptake value (SUVmax) were measured in the cervical, supraclavicular,
and anterior thoracic depots from vertebral level C3 to T7 using the PET-CT Viewer shareware,
as described previously (24). The BAT metabolic activity represents the primary endpoint of
this study and is calculated in a three-step process. First, in each axial slice on CT, a voxel is
defined as containing adipose tissue if it has a HU density between -250 and -10. Second, the
amount of retained FDG in each voxel is quantified by measuring the mean amount of positron-
derived gamma radiation in the voxel compared to the injected dose per body mass (MBq/mL of
voxel)/ (MBq/g of body mass), giving the “meanSUV*g/mL”. Finally, the total-body metabolic
activity is calculated by taking the sum of the volume of BAT from each voxel
=“mL*meanSUV*g/mL.” As expected, according to the method used to quantify BAT
parameters, there was a strong positive correlation between the cold-induced BAT volume and
activity of the study subjects in the hypothyroid phase (rho = >0.99, P = 0.01) as well as
thyrotoxic phase (rho = 0.94, P = 0.005). SUVmax reflects the highest amount of retained FDG
anywhere within the tissue. We used this metric to compare BAT activity with more
homogeneous tissues such as liver and skeletal muscle. The SUVmax of the FDG uptake in the
right lobe of the liver and skeletal muscle (erector spinae) was also measured.
Physiological and Laboratory Parameters

Blood pressure and heart rate were measured using a SureSigns VS3 vital signs monitor (Philips Healthcare). Core body temperature was measured with a temporal artery thermometer (Exergen Co.). Indirect calorimetry measurements including the resting metabolic rate (RMR), energy expenditure (EE), respiratory quotient (RQ), and substrate utilization were calculated using Sensormedics VMAX Encore equipment (VIASYS Respiratory Care Inc.) as well as techniques well established in the laboratory and standardized values. Insulin and TSH were measured according to standard procedures at the Harvard Catalyst CRC (http://catalyst.harvard.edu/services/hccrc-lab/). Total T4, total T3, free T3, T3 uptake, free T4 index, thyroxin binding globulin (TBG), free fatty acids (FFAs), lipid panel and HbA1c were measured using standard procedures at the Laboratory Corporation of America. Glucose was measured at the BIDMC clinical laboratory.

Statistical Analysis

This pilot study was designed to evaluate whether the TH status had an effect on BAT mass and activity in adult humans, specifically whether treatment with TH increases BAT mass and activity as the subjects transitioned from hypothyroidism to thyrotoxicosis. We anticipated consenting and screening up to 10 patients, so that 6-8 patients would complete all segments of the study (completion rate of 80%).

Data were analyzed using Statistical Package for the Social Science (SPSS, IBM, Armonk, New York) and Statistical Analysis System (SAS, SAS Institute, Inc.). All P values presented are two-tailed, and values ≤ 0.05 were considered to indicate statistical significance, while values < 0.10 were considered to represent a trend to achieve significance. Changes in BAT volume,
activity and maximum standardized uptake value (SUVmax), as well as hormonal and metabolic changes in the study patients between the hypothyroid and thyrotoxic state were evaluated using the non-parametric Wilcoxon signed-rank test. Associations between BAT volume and activity, TH and metabolic parameters were evaluated by Spearman correlation.

Results

The baseline characteristics of the six patients who completed the study are summarized in Table 1. Five patients had a normal baseline TSH prior to being enrolled in the study. One patient who was enrolled at the time of her completion thyroidectomy, had a low baseline TSH of 0.19 (reference range 0.27-4.2 µIU/mL) on TH treatment prior to starting the study. All patients were hypothyroid after TH withdrawal at the time of the first study PET/CT scan. In 4 out of the 6 study patients, the final TSH was in the thyrotoxic range, while the fifth patient had a TSH in the low reference range at 0.38 µIU/mL 3-6 months after starting TH suppressive treatment at the time of the second PET/CT scan. However, the sixth patient had a final TSH of 5.46 µIU/mL. Therefore, although the mean was in the euthyroid range, all patients but one were thyrotoxic at the time of the second PET/CT scan.

Thyroid function tests and metabolic parameters. TSH was higher while total T4, free T4 index, free T3 and T3 uptake were lower in the hypothyroid compared to the thyrotoxic state (trend only for free T3) (Figure 2A-C, Table 2). There was no significant difference in the total T3 between the hypothyroid and thyrotoxic state. There was no significant change in any thyroid function test after 60 min of cold exposure during each state. TBG levels did not change between the hypothyroid and thyrotoxic states. The total and LDL cholesterol were higher in the
hypothyroid compared to the thyrotoxic state. There were no significant changes in fasting triglyceride, free fatty acids, glucose, insulin, and HbA1c between the hypothyroid and thyrotoxic state (Table 2).

**Vital signs.** The systolic BP did not change significantly during the study. The diastolic BP increased after 60 min of cold exposure during both the hypothyroid and thyrotoxic states, the levels after cold exposure being higher in the hypothyroid compared to the thyrotoxic state. The HR decreased significantly after 60 min of cold exposure in the hypothyroid but not the thyrotoxic phase. There were no other significant changes in vital signs measured at room temperature and after 60 min of cold exposure during the hypothyroid as well as thyrotoxic state or between the two states (Table 2).

**Energy expenditure.** The baseline RMR of the study subjects measured at room temperature was higher in the thyrotoxic compared to the hypothyroid state. The RMR increased after 60 min of cold exposure during both the hypothyroid and thyrotoxic states. There was no difference between the RMR measured in the two states after cold exposure. There were no other significant changes in RMR or RQ measured at room temperature and after 60 min of cold exposure during the hypothyroid as well as thyrotoxic state or between the two states (Table 2). There was a positive correlation between the RQ measured at room temperature and after 60 min of cold exposure in the hypothyroid state (Spearman rho = 0.83, \( P = 0.04 \)). There was no correlation between the RMR measured at room temperature and after 60 min of cold exposure during each state.
Glucose uptake in BAT and other tissues. All patients had detectable BAT after acute cold exposure in both the hypothyroid and thyrotoxic state (Figure 3A-D, Table 3). Other tissues showed significant but small changes in glucose uptake. The SUVmax of the liver decreased (3.39 ± 0.27 vs. 3.03 ± 0.25 g/mL, \(P = 0.01\)), while the SUVmax of the skeletal muscle increased (0.62 ± 0.06 vs. 0.84 ± 0.10 g/mL, \(P = 0.02\)), when going from the hypothyroid to the thyrotoxic phase (Figure 4A-C).

The majority but not all (4 out of 6) patients showed an increase in detectable, acute, cold-induced BAT volume and activity between the hypothyroid and thyrotoxic states. The increases were not correlated with the seasonal trends in outdoor temperatures the patients experienced while they were taking thyroid hormone. The change in BAT volume, activity, and SUVmax between the two states were not significant. In the two patients who did not show an increase in BAT parameters between the hypothyroid and thyrotoxic state, the RMR also did not change after 60 min of cold exposure in both the hypothyroid and thyrotoxic state. However, there was a positive correlation between the BAT volume as well as BAT activity measured after cold exposure in the hypothyroid and thyrotoxic state (BAT volume: rho = 0.83, \(P = 0.04\); BAT activity: rho = 0.89, \(P = 0.02\)).

There was a negative correlation between the cold-induced BAT volume as well as activity and the subjects’ weight and BMI in both the hypothyroid and thyrotoxic state. There was no correlation between the BAT parameters and the subjects’ age and core body temperature. The baseline systolic BP measured in the hypothyroid state correlated strongly and positively with the BAT mass and activity measured in both states.
Inter-relationships among metabolic measures. Total T4, free T4 index, total T3 and free T3 correlated positively with the cold-induced BAT SUVmax in the hypothyroid state (all variables in hypothyroid state: rho = 0.88, \( P = 0.02 \)) (Figure 5A). Free T3 correlated negatively with the cold-induced BAT volume and activity in the thyrotoxic state (free T3 at room temperature/BAT volume: rho = -0.83, \( P = 0.04 \); free T3 at room temperature/BAT activity: rho = -0.77, \( P = 0.07 \); freeT3 after cold exposure/BAT volume: rho = -0.99, \( P = 0.01 \); free T3 after cold exposure/ BAT activity: rho = -0.99, \( P = 0.01 \)) (Figure 5B). Total T3 also correlated negatively with the cold-induced BAT volume and tended to correlate negatively with the BAT activity in the thyrotoxic state (total T3 at room temperature/BAT volume: rho = -0.83, \( P = 0.04 \); total T3 after cold exposure/BAT volume: rho = -0.83, \( P = 0.04 \); total T3 at room temperature/BAT activity: rho = -0.77, \( P = 0.07 \); total T3 after cold exposure/ BAT activity: rho = -0.77, \( P = 0.07 \)).

Several facets of thyroid status correlated with RMR. There was a significant inverse relationship between the baseline TSH and RMR in the hypothyroid state (rho = -0.89, \( P = 0.02 \)). In addition, there was a significant positive correlation between the change in RMR after 60 min of cold exposure and the change in total T3 as well as free T3 between the hypothyroid and thyrotoxic state (change in TT3 after 60 min of cold exposure: rho = 0.94, \( P = 0.005 \); change in FT3 after 60 min of cold exposure: rho = 0.94, \( P = 0.005 \)) (Figure 5C).

For exploratory analyses, we looked at the relationships between detectable cold-induced BAT metabolism and other metabolic parameters. Serum TG levels correlated negatively with BAT mass and activity in each state (hypothyroid state, BAT volume and activity: rho = -0.89, \( P = 0.02 \); hyperthyroid state, BAT volume: rho = -0.83, \( P = 0.04 \) and BAT activity: rho = -0.94, \( P = 0.005 \), and HbA1c correlated negatively with BAT volume and activity in the hypothyroid state (rho = -0.90, \( P = 0.015 \)). Total and LDL cholesterol correlated negatively with BAT
activity (rho = -0.83, P = 0.04), while HDL cholesterol correlated positively with BAT mass and activity (BAT volume: rho = +0.90, P = 0.015, BAT activity: rho = +0.84, P = 0.04) in the thyrotoxic state.
Discussion

Given the central role that thyroid hormone plays in regulating whole-body energy expenditure and BAT function, we designed this pilot study to evaluate how promising treatment with TH would be toward increasing BAT thermogenesis. However, because of the numerous adverse effects of TH supplementation in healthy humans, a novel study population is required in order to evaluate the full capacity of dosing with TH. We therefore chose as our model a series of patients with thyroid cancer because their standard course of treatment leads to iatrogenic hypothyroidism followed by TH treatment to the thyrotoxic state. All six study patients demonstrated detectable BAT under cold stimulation in both the hypothyroid and thyrotoxic state. The majority but not all (4 out of 6) patients showed an increase in BAT volume and activity under cold stimulation between the hypothyroid and thyrotoxic phase. In addition, there was a positive correlation between the cold-induced BAT volume as well as BAT activity in the hypothyroid and thyrotoxic phase for each patient.

It is important to consider that all study patients showed detectable cold-activated BAT in the hypothyroid state, even at TSH levels higher than 100 µIU/mL, which indicates that even low levels of TH are adequate for BAT activation in adult humans. In our study, serum T3 levels correlated negatively with cold-induced BAT mass and activity in the thyrotoxic state in adult humans. These apparently counterintuitive findings suggest that circulating T3 may be a marker of thyroid function without direct effects on BAT. Studies in rodents have shown that the BAT type 2 deiodinase (DIO2) is stimulated by the sympathetic nervous system (SNS) and inhibited by its substrate T4 (11, 12, 26). Excessive circulating T4 levels could result in inhibition of the DIO2 and proportionally lower local T3 concentrations and UCP1 activation in hyperthyroid
subjects (12), while low circulating T4 levels could activate DIO2, increase the local T3 production and activate the UCP1 in hypothyroid patients (27, 28). In addition, we need to take into consideration that the SNS and TH act synergistically to respond and adjust the body to different environmental factors, such as cold exposure. Studies in small rodents have shown that cold-exposure stimulates BAT activity by increasing central sympathetic nerve activity and by affecting local TH metabolism (11, 12). Both catecholamines and local T3 act synergistically to stimulate uncoupling protein 1 (UCP-1) expression and through this thermogenesis in BAT. In hypothyroidism, there is an increase, while in thyrotoxicosis there is a decrease in the central sympathetic outflow, as measured by plasma and urinary norepinephrine (NE) levels as well as NE turnover in different tissues such as BAT. However, there is also a reduced/exaggerated response to catecholamines in different tissues, because of decreased/increased beta-adrenergic receptor density as well as postreceptor mechanisms, which usually overrides the effect of TH on the central SNS outflow and explains the clinical manifestations in hypothyroidism and thyrotoxicosis, respectively (12, 34). Thus, the regulatory mechanisms involved in regulating BAT are complex and one of these mechanisms may be predominant, which could explain the different BAT responses to cold exposure noted in different patients.

We did not find a correlation between the change in thyroid function or the change in RMR and the change in cold-induced BAT volume and activity between the hypothyroid and thyrotoxic state. This may be explained by the fact that TH influences both the basal/obligatory and facultative thermogenesis through different mechanisms (11, 29). A treatment that would selectively stimulate facultative thermogenesis could be beneficial for weight loss and improvement in metabolic parameters without the side effects of elevated circulating TH levels.
Our study showed that circulating T3 is a stronger predictor of the change in energy expenditure compared to cold-induced BAT activity. However, the response to TH treatment led to opposite responses among two of the principal thermogenic organs: glucose uptake in the liver was lower, but skeletal muscle uptake was higher. Therefore, it cannot be assumed that all tissues respond the same to chronic treatment with TH. Our findings also support the hypothesis that TH contributes significantly to cold-induced thermogenesis by affecting multiple organs including skeletal muscle in addition to BAT (30, 31, 21, 22). In our study, there was a more consistent increase in the skeletal muscle activity as compared to the BAT activity between the hypothyroid and thyrotoxic state. This may be explained by the fact that the skeletal muscle activity is mainly regulated by thyroid hormone, while the synergistic activity with the SNS may be less important in this tissue (32). In addition, the difference between the two tissues could be potentially explained by the fact that our study patients took high-dose levothyroxine at the time of the second PET-CT scan, which can decrease DIO2 activity; this enzyme has more important effects in BAT than skeletal muscle (33).

The few published human studies looking at the relationship between thyroid status and BAT function have shown contradictory results (20-23). Skarulis et al. reported detectable BAT measured by PET-CT in a hypothyroid patient and an increase in BAT parameters when the patient became thyrotoxic upon resuming TH treatment as part of follow-up for thyroid cancer (20). In contrast, a severely hypothyroid child with Hashimoto’s thyroiditis was found with a significant supraclavicular BAT volume measured by MRI and BAT activity measured by infrared thermal imaging, which both decreased after two months of TH replacement (23).
Lahesmaa et al. reported a three-fold increase in glucose uptake measured by FDG-PET in ten hyperthyroid patients compared to euthyroid patients (21), while Zhang et al. reported no detectable BAT in ten patients with newly diagnosed Graves’ hyperthyroidism before starting treatment (22). Additional human studies are needed to gain insight into the factors that moderate the effect of TH on BAT volume and activity. It may be that TH directly stimulates BAT, but the general increase in the resting metabolic rate may obviate the need for additional BAT in some patients. The end result of these two simultaneous but opposite effects of TH on BAT may explain why some patients increase their BAT while others have less. In addition, as described above, the complex interaction between TH and sympathetic nervous system in regulating BAT may play a role.

We recruited thyroid cancer patients for this pilot study. This represents an excellent study model for BAT evaluation since treatment for thyroid cancer involves several steps including thyroidectomy, followed by induction of short-term hypothyroidism prior to the radioactive iodine treatment and then long-term suppressive TH treatment to prevent recurrence. Therefore, we can evaluate the changes in BAT mass and activity between the hypothyroid and thyrotoxic state in this patient population.

This is a pilot study designed with the goal to obtain preliminary data in a small but physiologically informative group of patients with thyroid cancer whose clinical course involves treatment with high-dose levothyroxine to change their thyroid status from severe hypothyroidism to mild thyrotoxicosis. The signal from this cohort would be a best-case scenario for any studies in healthy subjects or patients with milder hypothyroidism. Our data
show that based on the mean change in BAT metabolic activity, we would have to study more than a total of 50 pairs of subjects to demonstrate a significant effect of thyroid hormone treatment on detectable BAT. Therefore, this pilot study has achieved one of its essential goals and demonstrated that it is not very feasible to use levothyroxine treatment in most populations as a way of increasing BAT mass or metabolic activity.

As known from prior animal as well as human studies, chronic cold exposure is probably the strongest determinant of BAT volume and activity (9, 10). Based on our study design, we cannot easily distinguish the effects of chronic cold from the chronic administration of high doses of thyroid hormone. However, the change in BAT activity was not correlated with the season of thyroid hormone treatment, so the effects of cold exposure were not likely to have substantially confounded the modest effect we saw from chronic exposure to high doses of thyroid hormone.

In our study, we measured cold-induced BAT, since significant amounts of functional BAT have been detected in more than 95% of lean subjects in response to acute cold exposure (5, 8), while significantly fewer subjects showed detectable BAT via PET-CT scanning when studied at room temperature. With the current study design we were able to evaluate the effect of TH on cold-induced BAT by studying the same patients after cold exposure in both the hypothyroid and thyrotoxic state. Therefore, we can infer that the change in the BAT mass and activity between the two phases is related to the thyroid hormone status change.

In conclusion, in this pilot study we found that all 6 study patients had detectable BAT after 2 hours of cold exposure in both the hypothyroid and thyrotoxic state, and the majority but not all (4 out of 6) patients showed an increase in cold-induced BAT volume and activity between the hypothyroid and thyrotoxic state. Therefore, iatrogenic hypothyroidism lasting 2-4 weeks does
not prevent cold-induced BAT activation. At the same time, the use of TH to induce thyrotoxicosis does not consistently increase cold-induced BAT activity. Although the use of levothyroxine is not feasible, using a BAT-specific TH analog may be still feasible to increase BAT thermogenesis as a treatment for obesity and metabolic disease. Further studies are needed to evaluate this and also determine which physiological factors besides TH are involved in stimulating BAT growth and thermogenesis.
Acknowledgments: This work was supported by the 2012 Harvard Catalyst Clinical Research Center (HCCRC) Resources and Services Funding Award; the 2013 Harvard Catalyst Clinical Research Center (HCCRC) Laboratory Support and Genotyping Award; National Institutes of Health (NIH) grants K23 DK081604, the Intramural Research Program of the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK); and the Clinical Translational Science Awards UL1RR025758, UL1TR000170 and UL1TR001102 to Harvard University and its affiliated academic health care centers from the National Center for Research Resources (NCRR) and the National Center for Advancing Translational Science and financial contributions from Harvard University and its affiliated academic healthcare centers. The content is solely the responsibility of the authors and does not necessarily represent the official views of Harvard Catalyst, Harvard University and its affiliated academic healthcare centers, or the National Institutes of Health.

We thank the Beth Israel Deaconess Medical Center (BIDMC) Clinical Research Center nursing team, Bionutrition Core, and nuclear medicine technologists for the excellent support they provided; Michelle Beck for supervising the study development and progression; Anthony Hollenberg for reviewing the manuscript; Matthew R Palmer for reviewing the PET-CT study protocol; Lauren Weiner for her assistance in coordinating the clinical studies; and our volunteers for their commitment to the studies.

Disclosures: The authors have nothing to disclose.
Corresponding author:

Alina Gavrila, M.D., M.M.Sc.
Division of Endocrinology and Metabolism
Beth Israel Deaconess Medical Center
330 Brookline Avenue GZ6
Boston, MA 02215
Phone: 617-667-9344
Fax: 617-667-7060
agavrila@bidmc.harvard.edu
**References:**


[16]. Obregon MJ 2008 Thyroid hormone and adipocyte differentiation. Thyroid 18(2):185-95.


Figure Legends

Figure 1. Study Protocol.

Figure 2. Changes in thyroid function in individual subjects measured at room temperature and after 60 min of cold exposure (15-16°C) in the hypothyroid and then thyrotoxic state. (A) Changes in serum TSH. (B) Changes in serum free T4. (C) Changes in serum free T3.

Figure 3. Changes in BAT volume, activity and SUVmax in individual subjects measured after 120 min of cold exposure (15-16°C) in the hypothyroid and then thyrotoxic state. (A) Brown fat FDG uptake in a study subject (53 years old female, BMI 19.6 kg/m²) illustrated using coronal representations of PET in the hypothyroid state (TSH 70.90 µIU/mL) and then thyrotoxic state (TSH 0.03 µIU/mL), the green arrows pointing to the right and left supraclavicular BAT depots. (B) Changes in BAT volume. (C) Changes in BAT activity. (D) Changes in BAT maximum standardized uptake value (SUVmax).

Figure 4. Changes in 18F-FDG uptake between the hypothyroid and hyperthyroid state measured in different tissues in individual study subjects. (A) BAT. (B) Liver. (C) Skeletal muscle.

Figure 5. Correlations of circulating T3 with BAT and with RMR. (A) Negative correlation between FT3 and BAT in the hypothyroid state. (B) Positive correlation between FT3 and BAT
in the hyperthyroid state. (C) Positive correlation between the change in RMR and the change in
FT3 between the hypothyroid and hyperthyroid state.
<table>
<thead>
<tr>
<th></th>
<th>Mean ± SEM</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td>1 M / 5 F</td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>43.5 ± 4.7</td>
<td>22-53</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>75.3 ± 5.7</td>
<td>60.6-98.8</td>
</tr>
<tr>
<td>Body Mass Index (kg/m²)</td>
<td>25.45 ± 2.5</td>
<td>19.6-35.85</td>
</tr>
<tr>
<td>Baseline TSH (µIU/mL)</td>
<td>1.36 ± 0.66</td>
<td>0.19-2.09</td>
</tr>
<tr>
<td>Thyroid Cancer TNM Stage</td>
<td>1-IVA</td>
<td></td>
</tr>
</tbody>
</table>
Table 2. Changes in the subject characteristics measured at room temperature and after cold exposure (15-16°C) in the hypothyroid and then thyrotoxic state.

<table>
<thead>
<tr>
<th>Characteristic (units)</th>
<th>Hypothyroid Room Temperature</th>
<th>Cold Exposure</th>
<th>Thyrotoxic Room Temperature</th>
<th>Cold Exposure</th>
</tr>
</thead>
<tbody>
<tr>
<td>Temperature (°C)</td>
<td>36.7 ± 0.1</td>
<td>36.2 ± 0.2</td>
<td>36.5 ± 0.1</td>
<td>36.5 ± 0.2</td>
</tr>
<tr>
<td>Systolic BP (mmHg)</td>
<td>118.3 ± 4.0</td>
<td>127.6 ± 3.2</td>
<td>118.5 ± 4.3</td>
<td>126.2 ± 5.8</td>
</tr>
<tr>
<td>Diastolic BP (mmHg)</td>
<td>75.5 ± 3.5</td>
<td>84.0 ± 2.9(^a)</td>
<td>71.2 ± 2.5</td>
<td>78.7 ± 2.5(^{bd})</td>
</tr>
<tr>
<td>Heart Rate (bpm)</td>
<td>66.8 ± 3.5</td>
<td>60.0 ± 1.5(^a)</td>
<td>64.5 ± 4.4</td>
<td>65.2 ± 5.7</td>
</tr>
<tr>
<td>Free Fatty Acids (mEq/L)</td>
<td>--</td>
<td>0.63 ± 0.09</td>
<td>--</td>
<td>0.62 ± 0.07</td>
</tr>
<tr>
<td>Triglycerides (mg/dL)</td>
<td>99.8 ± 28.4</td>
<td>--</td>
<td>84.2 ± 20.0</td>
<td>--</td>
</tr>
<tr>
<td>Total Cholesterol (mg/dL)</td>
<td>279.7 ± 46.3</td>
<td>--</td>
<td>201.3 ± 24.6(^c)</td>
<td>--</td>
</tr>
<tr>
<td>LDL Cholesterol (mg/dL)</td>
<td>173.2 ± 39.6</td>
<td>--</td>
<td>110.7 ± 23.3(^c)</td>
<td>--</td>
</tr>
<tr>
<td>HDL Cholesterol (mg/dL)</td>
<td>86.5 ± 6.8</td>
<td>--</td>
<td>74.2 ± 6.6</td>
<td>--</td>
</tr>
<tr>
<td>Glucose (mg/dL)</td>
<td>--</td>
<td>81.5 ± 4.8</td>
<td>--</td>
<td>84.7 ± 3.8</td>
</tr>
<tr>
<td>Insulin (µU/mL)</td>
<td>6.5 ± 2.0</td>
<td>5.6 ± 1.9</td>
<td>6.4 ± 2.5</td>
<td>5.4 ± 1.6</td>
</tr>
<tr>
<td>Hemoglobin A1c (%)</td>
<td>5.6 ± 0.2</td>
<td>--</td>
<td>5.6 ± 0.03</td>
<td>--</td>
</tr>
<tr>
<td>TSH (µIU/mL)</td>
<td>60.7 ± 14.9</td>
<td>60.8 ± 15.0</td>
<td>1.2 ± 1.1(^c)</td>
<td>1.0 ± 0.9(^d)</td>
</tr>
<tr>
<td>Total T4 (µg/dL)</td>
<td>2.7 ± 1.4</td>
<td>2.9 ± 1.5</td>
<td>10.6 ± 0.8(^c)</td>
<td>11.0 ± 0.9(^d)</td>
</tr>
<tr>
<td>Free T4 index</td>
<td>0.77 ± 0.4</td>
<td>0.81 ± 0.43</td>
<td>3.8 ± 0.3(^c)</td>
<td>3.9 ± 0.25(^d)</td>
</tr>
<tr>
<td>Total T3 (mg/dL)</td>
<td>49.7 ± 24.3</td>
<td>53.3 ± 26.1</td>
<td>102.2 ± 14.5</td>
<td>106.0 ± 12.3</td>
</tr>
<tr>
<td>Free T3 (pg/mL)</td>
<td>1.2 ± 0.7</td>
<td>1.3 ± 0.8</td>
<td>3.2 ± 0.3(^c)</td>
<td>3.2 ± 0.2</td>
</tr>
<tr>
<td>T3 Uptake (%)</td>
<td>25.8 ± 1.7</td>
<td>25.7 ± 1.5</td>
<td>36.2 ± 1.6(^c)</td>
<td>35.3 ± 1.5(^d)</td>
</tr>
<tr>
<td>Thyroxin Binding Globulin (µg/mL)</td>
<td>--</td>
<td>18.2 ± 2.7</td>
<td>--</td>
<td>14.2 ± 1.6</td>
</tr>
<tr>
<td>Resting Metabolic Rate (Kcal/day)</td>
<td>1299.0 ± 57.7</td>
<td>1497.2 ± 106.7(^a)</td>
<td>1470.0 ± 81.9(^c)</td>
<td>1612.8 ± 46.9(^b)</td>
</tr>
</tbody>
</table>

Values represent mean ± SEM. Vitals, laboratory tests and indirect calorimetry were measured after 60 min of cold exposure, while BAT activity, volume and SUVmax were measured after 120 min of cold exposure at 15-16°C.

\( a = P \leq 0.05, \) hypothyroid state room temperature vs. hypothyroid state after cold exposure.

\( b = P \leq 0.05, \) thyrotoxic state room temperature vs. thyrotoxic state after cold exposure.

\( c = P \leq 0.05, \) hypothyroid state room temperature vs. thyrotoxic state room temperature.

\( d = P \leq 0.05, \) hypothyroid state after cold exposure vs. thyrotoxic state after cold exposure.
<table>
<thead>
<tr>
<th></th>
<th>Mean ± SEM</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>BAT Volume (mL)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hypothyroid</td>
<td>72.0 ± 21.0</td>
<td>20 - 150</td>
</tr>
<tr>
<td>Thyrotoxic</td>
<td>87.7 ± 16.5</td>
<td>23.5 - 136.9</td>
</tr>
<tr>
<td>Change</td>
<td>15.7 ± 13.9</td>
<td>-39.8 - 55.3</td>
</tr>
<tr>
<td>% Change</td>
<td>53.35 ± 35.9%</td>
<td>-26.5 - 203.4%</td>
</tr>
<tr>
<td><em><em>BAT Activity (mL x mean SUV</em> g/mL)</em>*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hypothyroid</td>
<td>158.1 ± 72.8</td>
<td>23.1 - 495.8</td>
</tr>
<tr>
<td>Thyrotoxic</td>
<td>189.0 ± 55.5</td>
<td>27.85 - 398.8</td>
</tr>
<tr>
<td>Change</td>
<td>30.9 ± 31.5</td>
<td>-97 – 125.85</td>
</tr>
<tr>
<td>% Change</td>
<td>74.35 ± 44.5%</td>
<td>-23.8 - 218.2%</td>
</tr>
<tr>
<td><strong>BAT SUVmax (g/mL)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hypothyroid</td>
<td>7.3 ± 2.6</td>
<td>2.0 - 19.5</td>
</tr>
<tr>
<td>Thyrotoxic</td>
<td>7.9 ± 2.95</td>
<td>2.1 - 22</td>
</tr>
<tr>
<td>Change</td>
<td>0.6 ± 0.95</td>
<td>-2.15 - 3.9</td>
</tr>
<tr>
<td>% Change</td>
<td>25.6 ± 35.1%</td>
<td>-42.8 – 195%</td>
</tr>
</tbody>
</table>

% Change from the baseline hypothyroid value.
Figure 3

A

TSH 70.90  TSH 0.03

88 days

B

Hypothyroid

Thyotoxic

C

D

190x254mm (96 x 96 DPI)
Figure 4

A

Maximal BAT Activity (max SUV/gm)

P = 0.6

B

Maximal Liver Activity (max SUV/gm)

P = 0.01

C

Maximal Skeletal Muscle Activity (max SUV/gm)

P = 0.02

190x254mm (96 x 96 DPI)
Figure 5

A

\[ \rho = 0.88 \quad \text{P} = 0.02 \]

\[ \text{BAT} \ \text{S7/	ext{mL}} \]

\[ \text{FT3 (pg/mL)} \]

Hypothyroid State

B

\[ \rho = -0.83 \quad \text{P} = 0.04 \]

\[ \text{BAT Volume (mL)} \]

\[ \text{FT3 (pg/mL)} \]

Thyrotoxic State

C

\[ \rho = 0.94 \quad \text{P} = 0.005 \]

\[ \text{Change in RMR (kcal/day)} \]

\[ \text{Change in FT3 (pg/mL)} \]

Hypothyroid to Hyperthyroid State