



# Genetic and 'cultural' similarity in wild chimpanzees

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1 **Genetic similarity and ‘cultural’ similarity in wild chimpanzees**

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18

18 **Summary** Research interest in the importance of group-specific, socially transmitted  
19 behaviour (i.e., ‘cultures’ or ‘traditions’) in the lives of animals has increased in recent  
20 years, partly as a result of high profile reports of extensive variation in the behavioural  
21 repertoires of wild animals. The current dominant approach to identifying animal culture  
22 in the wild, which infers the existence of culture by eliminating ecological explanations  
23 for differences in the behavioural repertoires of groups, was first applied to chimpanzees.  
24 Some critics, however, argue that genetic differences between groups have not yet been  
25 ruled out as a factor contributing to differences in the behaviour of groups of  
26 chimpanzees and other animals. Here we examined this issue directly by combining  
27 genetic and behavioural data from nine groups of wild chimpanzees. We found that  
28 genetic differentiation among chimpanzees, at both the between-subspecies and within-  
29 subspecies level, cannot be ruled out as playing a major role in generating group  
30 differences in behaviour. We also found that behaviours involved in non-vocal  
31 communication may be more likely to represent true cultural variants, as the distribution  
32 of these types of behaviours fit the predictions of the genetics hypothesis less well than  
33 did other types of behaviours.  
34

34           The importance of group-specific, socially transmitted behaviour to the lives of  
35 non-human animals, and its relevance to the evolution of human culture, is a matter of  
36 considerable controversy (Galef and Heyes 1996; Boesch 2003; Laland and Janik 2006;  
37 Laland and Galef 2009). As our closest living relatives and the most intensively-studied  
38 non-human primate in the wild, chimpanzees have played a particularly important role in  
39 the study of animal culture (Goodall 1973; McGrew 1992; Boesch 1996; Boesch and  
40 Tomasello 1998). A landmark survey of geographic variation in chimpanzee behaviour  
41 found 39 behaviours, including various forms of tool use, grooming, and courtship  
42 behaviour, that were common in at least one chimpanzee group but absent in at least one  
43 other, and for which an ecological explanation for this absence was judged to be unlikely,  
44 (e.g., a lack of ‘termite fishing’ even though the appropriate termite species was present)  
45 (Whiten *et al.* 1999; Whiten *et al.* 2001). This so-called “method of exclusion”, which  
46 infers the existence of culture by eliminating ecological explanations for the patterning of  
47 between-group behavioural variation, has since been applied to several other primate and  
48 non-primate species, and is now the dominant approach used to identify animal culture in  
49 the wild (Rendell and Whitehead 2001; Hohmann and Fruth 2003; Perry *et al.* 2003; van  
50 Schaik *et al.* 2003).

51           In a pattern reminiscent of between-group variation in human behaviour, each  
52 chimpanzee group had its own unique combination of these putative cultural variants.  
53 This complex, mosaic pattern of between-group variation in behaviour was very different  
54 than that observed in prior studies of animal culture, where in most cases only one type of  
55 behaviour varied among groups (e.g., bird song dialect (Marler and Tamura 1964)).  
56 Despite various other species showing patterns of behavioural diversity that approach

57 levels shown in chimpanzees, for many, chimpanzees remain “the prime exemplar of  
58 animal culture” (Boesch 2003; McGrew 2004; Laland *et al.* 2009).

59         Research conducted in captivity shows that chimpanzees have the requisite social  
60 learning abilities to produce stable differences in group-specific behaviour (Whiten *et al.*  
61 2005; Horner *et al.* 2006; Bonnie *et al.* 2007; Whiten *et al.* 2007; Price *et al.* 2009). In  
62 one experiment (Whiten *et al.* 2005), for example, two chimpanzees were isolated from  
63 their respective groups and trained in alternative techniques to use a tool to gain access to  
64 food in an experimental apparatus. These two “demonstrators” were then re-introduced  
65 into their original groups, whose behaviour was then compared with each other and with  
66 that of a third group who had no such access to a trained demonstrator. Chimpanzees in  
67 the first two groups predominately performed the technique of their demonstrator, while  
68 chimpanzees in the third group did not learn to gain access to the food.

69         However, although the social learning abilities of chimpanzees are sufficient to  
70 produce group differences in behaviour, this does not necessarily indicate that all or even  
71 most of the behavioural variants observed in the wild arise as a result of social learning,  
72 and thus represent true cultural variants (Galef 1990; Tomasello 1990; Boesch 1996;  
73 Galef 2009; Laland *et al.* 2009). Some critics argue that insufficient attention has been  
74 paid to the possibility that genetic differences are responsible for behavioural variation  
75 between chimpanzee groups (Galef 1990; Heyes 1994; Heyes and Galef 1996; Laland *et*  
76 *al.* 2006; Tennie *et al.* 2009). Recent research showing that individuals raised with no  
77 opportunity to acquire the behaviour through social learning nevertheless exhibit  
78 sophisticated abilities to use tools (crows) (Kenward *et al.* 2005) and process foods  
79 (gorillas) (Tennie *et al.* 2008) suggests that complex behaviours can at least partly be

80 under genetic control, and thus that group differences in the underlying genetic  
81 predispositions and abilities could plausibly contribute to group differences in behaviour.  
82 Indeed, almost one-third of the putative cultural variants in the original report (Whiten *et*  
83 *al.* 1999) of geographical variation in chimpanzee behaviour are found only in *Pan*  
84 *troglodytes verus*, the most genetically divergent of the four chimpanzee subspecies, and  
85 considered by some to represent a different species (Morin *et al.* 1994; Gagneux *et al.*  
86 1999).

87 In the only systematic investigation of this issue, Lycett and colleagues (Lycett *et*  
88 *al.* 2007; Lycett *et al.* in press) reasoned that if the patterning of intergroup variation in  
89 behaviour is primarily the product of genetic differences between subspecies, then a tree  
90 of the relationships between chimpanzee groups generated from a cladistic analysis of the  
91 39 putative cultural variants should have more phylogenetic structure when two  
92 subspecies are considered together than a tree generated from the East African subspecies  
93 alone. They failed to find more phylogenetic structure in the two-subspecies than the  
94 single-subspecies tree, and thus concluded that the patterning of between-group variation  
95 in chimpanzee behaviour fit a cultural explanation better than a genetic one.

96 While an important first step towards an explicit investigation of the role of  
97 genetic differentiation in generating intergroup variation in behaviour, this study has four  
98 major limitations. First, as acknowledged by Lycett and colleagues (Lycett *et al.* in  
99 press), at present there is no well-accepted statistical methodology for comparing the  
100 extent of phylogenetic structure in two different trees. Second, recent simulation studies  
101 show that measures of phylogenetic structure are very sensitive to rates of evolution  
102 (Nunn *et al.* in press). Even if the behavioural variants were genetically determined, then

103 a sufficiently high rate of evolution, which could be caused by selection, could produce a  
104 lower signal of phylogenetic structure in a tree with deeper phylogenetic separation  
105 (Nunn *et al.* in press). Third, the one-subspecies versus two-subspecies analysis  
106 presupposes that only genetic differentiation between subspecies, but not genetic  
107 differentiation among groups within a single subspecies, can affect behavioural variation  
108 among groups. Although the statement that “populations within East Africa cannot be  
109 distinguished” (Lycett *et al.* 2007) p. 17589 is technically correct in the sense that there is  
110 sharing of mitochondrial DNA (mtDNA) haplotypes across the subspecies range,  
111 quantitative analyses show that there is as much geographical structuring of genetic  
112 variation within the East African subspecies as within the entire human species (Goldberg  
113 and Ruvolo 1997). That the extent of genetic differentiation among chimpanzee groups  
114 belonging to the same subspecies is sufficient to affect between-group behavioural  
115 variation is suggested by the finding that genetically influenced variation in body size is  
116 implicated in differences between the qualities of vocalizations of two groups of  
117 chimpanzees belonging to the same East African subspecies (Mitani *et al.* 1999). Finally,  
118 the method employed by (Lycett *et al.* 2007; Lycett *et al.* in press) purports to test only  
119 whether genetics can be excluded as a cause for *overall* levels of similarity in the  
120 behavioural repertoires of chimpanzee groups; it offers no insight into how strongly the  
121 distribution of each of the individual behavioural variants follows (or does not follow)  
122 patterns of between-group genetic differentiation. Some authors have suggested that  
123 differences in non-vocal communicative behaviours are particularly unlikely to result  
124 from differences in underlying genetic predispositions, as this class of behaviours is less  
125 likely than other classes of behaviour (e.g., tool-use in a foraging context) to be

126 influenced by the differential operation of natural selection according to local ecological  
127 conditions (Boesch 1996; van Schaik 2009).

128         Here we take a more direct approach to the question of whether genetics can be  
129 excluded as playing a role in generating intergroup variation in the behaviour of wild  
130 chimpanzees by measuring levels of genetic differentiation among nine groups of wild  
131 chimpanzees (three from the West African *P.t. verus* and six from the East African *P.t.*  
132 *schweinfurthii* subspecies), and comparing these results with patterns of between-group  
133 behavioural variation. Our study involved five steps. First, we sequenced most of the  
134 members of each of the nine groups at the first hypervariable region of the maternally  
135 inherited mtDNA and estimated levels of between-group genetic  
136 similarity/differentiation. Second, we converted the behavioural data of the original study  
137 of geographical variation in chimpanzee behaviour (Whiten *et al.* 1999) (as well as data  
138 from two newly assessed chimpanzee groups) into numerical codes reflecting how  
139 frequently each of the 39 behaviours occurred in each chimpanzee group. Third, we used  
140 these behavioural data to generate, for each of the 36 pairs of groups, (1) a measure of the  
141 overall level of behavioural similarity (*sensu* (Lycett *et al.* 2007; Lycett *et al.* in press)),  
142 and (2) a measure of the level of differentiation for each of the individual behavioural  
143 variants. Fourth, we examined how well patterns of between-group genetic variation  
144 matched patterns of between-group (1) overall behavioural similarity and (2)  
145 differentiation for each of the individual behavioural variants. And fifth, we tested  
146 whether the pattern of between-group genetic differentiation predicted patterns of  
147 between-group behavioural differentiation less well for behavioural variants involved in  
148 non-vocal communication than it did for other types of behavioural variants.



149

## 150 **Results and Discussion**

151 We found that the overall level of genetic differentiation among the 9 groups was  
152 substantial (AMOVA value = 0.49,  $p = 0.00001$ ), and that the majority of pairs of groups  
153 (28 / 36 = 77.8 %) were significantly genetically differentiated from one another (Table  
154 1). Of the 12 pairs of groups showing no significant genetic differentiation, three were  
155 located in the same block of continuous forest (Mahale K and M, Kibale Kanayawara and  
156 Ngogo, Tai North and South), and four involved pairs that contained the poorly  
157 genetically sampled Mahale K group.

158 When comparing the genetic and behavioural data, we found that levels of genetic  
159 similarity and levels of overall behavioural similarity between chimpanzee groups were  
160 strongly and significantly correlated (Pearson matrix permutation correlation,  $N = 26$   
161 pairs,  $R = 0.59$ ,  $p = 0.001$ ) (figure 1; table 1). The relationship between genetic similarity  
162 and overall behavioural similarity was less strong and failed to reach statistical  
163 significance when calculated for the much smaller data set of only the 6 groups belonging  
164 to the East African subspecies (Pearson matrix permutation correlation,  $N = 15$  pairs of  
165 groups,  $R = 0.35$ ,  $p = 0.18$ ). These results suggest that genetic differentiation cannot be  
166 excluded as an explanation for overall patterns of behavioural variation among  
167 chimpanzees, and may even play a role in generating behavioural variation among groups  
168 within a single subspecies.

169 The strong correlations between genetic and overall behavioural similarity,  
170 however, do not necessarily preclude social learning as contributing to the patterning of  
171 behavioural variation among chimpanzee groups. If emigrating females carry the

172 behavioural variants of their natal group to their new group, then groups that share more  
173 migrants are likely to be more behaviourally similar. High migration rates between  
174 groups could thus result in a positive correlation between genetic and behavioural  
175 similarity, even if behavioural variants were completely socially learned. It is thus only  
176 when patterns of behavioural and genetic similarity are discordant that inferences can be  
177 made about the role of social learning in generating geographical variation in behaviour.

178         Our second goal was therefore to identify those behavioural variants whose  
179 geographical distribution did not match the predictions of the genetic hypothesis,  
180 suggesting they represent true socially learned cultural variants. We found that the  
181 individual behavioural variants varied continuously in the extent to which their  
182 distribution matched patterns of genetic differentiation (table 2). The combination of a  
183 relatively modest sample size ( $N = 6$  to 36 pairs of groups) and the stringent alpha level  
184 (e.g., in this case, a Bonferroni correction of  $0.05 / 24 = 0.002$ ) that is required to correct  
185 for multiple comparisons means that there is very low power to reject the null hypothesis  
186 that the distribution of each behavioural variant is predicted by genetic differentiation.  
187 Thus, it is not possible to make strong inferences about which of the individual  
188 behavioural variants can and which of the individual behavioural variants cannot be  
189 excluded as being explained by patterns of genetic differentiation.

190         However, we did find that, as a whole, the distribution of non-vocal  
191 communicative behavioural variants was less well predicted by patterns of genetic  
192 differentiation than was the distribution of the other types of behavioural variants (table  
193 2, figure 2). The mean of the Spearman rank correlation coefficients between behavioural  
194 differentiation and genetic differentiation was significantly higher among the

195 communicative behavioural variants ( $\bar{X} = 0.01$ , 95 % C.I. = 0.07) than among the non-  
196 communicative behavioural variants ( $\bar{X} = 0.22$ , 95 % C.I. = 0.11) (independent samples t  
197 test;  $t(29.3) = 3.234$ ,  $p = 0.003$ ).

198

## 199 **Conclusions**

200 Here we have shown that genetic differences cannot be excluded as playing a  
201 major role in structuring patterns of behavioural variation among chimpanzee groups,  
202 particularly for behaviours not involved in non-vocal communication. As others have  
203 noted, the method of exclusion may lead to an underestimation of the true number of  
204 cultural variants that exist in the wild (Boesch 1996; Laland *et al.* 2006; Laland *et al.*  
205 2009). Our results showing that chimpanzee groups separated by more than  $\approx 300$  km are  
206 likely to show significant genetic differentiation suggests that if the method of exclusion  
207 is strictly applied, then only behaviours that vary among chimpanzee groups that are very  
208 geographically close should be considered as cultural variants (e.g. (Mobius *et al.* 2008)).  
209 It has been proposed (Lefebvre 1995; Franz and Nunn 2009; Kendal *et al.* 2009) that  
210 mathematical methods to identify a statistical ‘signature’ of transmission of a behavioural  
211 variant through social learning are a promising way to investigate culture in the wild for  
212 animals, like chimpanzees, where definitive translocation experiments (e.g. (Helfman and  
213 Schultz 1984)) are impossible for logistic and ethical reasons. Unfortunately, while we  
214 often have a good understanding of the distribution of a behaviour among the individuals  
215 within a group, the diffusion processes that led to that distribution are rarely observed  
216 (Perry *et al.* 2003). Comparing patterns of behavioural variation with patterns of kinship  
217 among individuals within the same social group may be a more tractable way to

218 investigate animal culture in the wild (Krutzen *et al.* 2005), particularly in species like  
219 chimpanzees where the relatively weak influence of kinship on patterns of affiliation  
220 (Langergraber *et al.* 2007; Langergraber *et al.* 2009) means that evidence for social  
221 learning (i.e., animals that interact with each other more frequently are more likely to  
222 share the behavioural variant) is not confounded with evidence for genetic inheritance  
223 (i.e., close relatives are more likely to share the behavioural variant). In sum, diverse  
224 methodologies will be essential to resolving the long-standing debate of the relative  
225 contributions of genetics, social learning, and other factors in generating geographical  
226 variation in animal behaviour.

227

## 228 **Materials and Methods**

229

230 We examined mitochondrial (mtDNA) variation (354 bases of the first  
231 hypervariable region) in nine groups of chimpanzees (numbers of adolescent and adult  
232 individuals sequenced / number of adolescent and adult group members present at time of  
233 sampling in brackets): the Bossou group (12/12) of Guinea, the North (8/11) and South  
234 (16/26) groups of Taï National Park, Cote d'Ivoire, the Sonso group (28/28) of Budongo  
235 Forest Reserve, Uganda, the Ngogo (89/89) and Kanyawara (20/20) groups of Kibale  
236 National Park, Uganda, the M (32/32) and K (3/Unknown) groups of Mahale Mountains  
237 National Park, Tanzania, and the Kasekela (38/38) group of Gombe Stream National  
238 Park, Tanzania. mtDNA sequences for five of the groups were previously published:  
239 Bossou (Shimada *et al.* 2009), Sonso (Langergraber *et al.* 2007), Gombe (Liu *et al.*  
240 2008), Ngogo (Langergraber *et al.* 2007), Kanyawara (Langergraber *et al.* 2007). We

241 generated new mtDNA sequences for the Taï North and South groups, the Mahale M and  
242 K groups, and the Gombe Kasekela group, following previously described procedures  
243 [24, 25]. The Genbank (<http://www.ncbi.nlm.nih.gov>) accession numbers for the  
244 chimpanzee mtDNA sequences used in this study are EU077270-EU077418,  
245 XXXXXXXX-XXXXXXX, and XXXXXXXX-XXXXXXX.

246 AMOVA and pairwise  $F_{ST}$  values were calculated using Arlequin 3.1 (Excoffier *et*  
247 *al.* 2005), with genetic distances between haplotypes measured using the number of  
248 nucleotide differences, and statistical significance was assessed by comparison with  
249 genetic differentiation calculated on random data sets where mtDNA haplotypes were  
250 permuted among groups. Although mtDNA is unlikely to directly code for any of the  
251 behavioural variants considered here, it is an excellent marker for determining overall  
252 levels of genetic similarity among groups, particularly in species such as chimpanzees  
253 where females rather than males move between groups. This is supported by the strong  
254 correlation between mtDNA genetic distance and geographical distance among the nine  
255 chimpanzee groups included in this study (Pearson correlation matrix permutation,  $N =$   
256 36 pairs of groups,  $R = 0.96$ ,  $p = 0.00005$ ).

257 For six of the chimpanzee groups in this study (Bossou, Sonso, Kanyara, Mahale  
258 M and K, Kasekela), we used the behavioural codes as described in the original paper on  
259 geographical variation in chimpanzee behaviour (Whiten *et al.* 1999; Whiten *et al.* 2001).  
260 For Taï North we changed three codes from the original paper, as one of us (C.B., the  
261 director of this field site) decided that these new codes were more accurate: ‘Hand-clasp’  
262 was changed from habitual to present, ‘Knuckle-knock’ was changed from customary to  
263 habitual, and ‘Branch-slap’ was changed from customary to absent. We repeated our

264 analyses of the relationship between overall behavioural and genetic similarity using the  
265 original codes for Tai North and found no qualitative changes in our results (Pearson  
266 correlation matrix permutation,  $N = 36$ ,  $R = 0.61$ ,  $p = 0.001$ ). Tai South group, newly  
267 coded for this study by C.B., had the same codes as Tai North, with the following two  
268 exceptions: ‘Bee-probe’ was coded as present and ‘Marrow-pick’ as habitual. Ngogo,  
269 newly coded by J.M. and K.L. for this study, had the same codes as Kanyawara, with the  
270 following two exceptions: ‘Seat-vegetation’ was coded as habitual, and ‘Leaf-clip,  
271 mouth’ was coded as customary. The behavioural variant ‘Branch-din’ was excluded  
272 from all analyses because it was absent in all nine of the chimpanzee groups included in  
273 this study. Thus, our study examined only 38 of the 39 putative cultural variants  
274 described in the original study on geographical variation in chimpanzee behaviour  
275 (Whiten *et al.* 1999; Whiten *et al.* 2001). We made no attempt to include behaviours now  
276 known or suspected to vary between chimpanzee groups that were not included in this  
277 original study (e.g., ‘social scratch’ (Nishida *et al.* 2004)).

278         We converted the behavioural codes into numbers as follows: (3) ‘customary’, the  
279 behaviour occurs in all or most able-bodied members of at least one age-sex class (such  
280 as adult males); (2) ‘habitual’, the behaviour is not customary but has occurred repeatedly  
281 in several individuals, consistent with some degree of social transmission; (1) ‘present’,  
282 the behaviour is neither customary nor habitual but is clearly identified; (0) ‘absent’, the  
283 behaviour has not been recorded and no ecological explanation is apparent; (?)  
284 ‘ecological explanation’, absence is explicable because of a local ecological feature; and  
285 (?) ‘unknown’, the behaviour has not been recorded, but this may be due to inadequacy of  
286 relevant observational opportunities. Our numerical coding system differs from that used

287 in the study of Lycett and colleagues (Lycett *et al.* 2007; Lycett *et al.* in press), in that  
288 these authors did not distinguish between behavioural variants that were absent and those  
289 that were absent due to an ecological reason, coding both types as 0.

290 We then used these data to calculate Pearson correlations between each of the 36  
291 pairs of chimpanzee groups, giving a measure of overall behavioural similarity that could  
292 potentially range from -1 to 1. We compared this matrix of overall behavioural similarity  
293 to the matrix of genetic similarity ( $1 - F_{ST}$ ) using a Pearson matrix correlation test. For the  
294 analysis of the relationship between genetic differentiation and the distribution of the  
295 individual behavioural variants, we created a matrix consisting of the absolute value of  
296 the difference in the numerical code between each of the communities, such that  
297 communities that were similar for the behavioural variant had a score of 0 (i.e., both had  
298 the same code), and communities that were very different for the behavioural variant had  
299 a score of 3 (i.e., ‘habitual’ in one group and ‘absent’ in the other). We then calculated a  
300 Spearman rank correlation matrix permutation test between each of these 38 matrices and  
301 a matrix of genetic differentiation ( $F_{ST}$ ). The independent samples t-test we used to  
302 compare the average of the Spearman rank correlations between behavioural  
303 differentiation and genetic differentiation of the communicative and non-communicative  
304 behaviours accounted for unequal variances between these two groups (Levene’s test for  
305 equality of variance,  $F = 5.159$ ,  $p = 0.03$ ). This independent samples t-test was computed  
306 on the 34 behavioural variants where at least 15 pairs of groups did not have a missing  
307 value (i.e., ‘ecological explanation’ or ‘unknown’).

308 All matrix permutation tests used 10,000 permutations.

309

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317

### 318 **Table and Figure Captions**

319

320 Figure 1. The relationship between levels of overall behavioural similarity and levels of  
321 genetic similarity in 36 pairs of wild chimpanzee groups.

322

323 Table 1. Genetic similarity (below diagonal) and overall behavioural similarity (above  
324 diagonal) in 36 pairs of chimpanzee groups. Pairs where genetic differentiation is  
325 statistically significantly different from 0 are in listed in bold.

326

327 Table 2. Spearman rank correlations between behavioural distance and genetic distance  
328 for the individual behavioural variants. Also shown are the classifications of behavioural  
329 variants as non-vocal communicative or not. Behavioural variants are listed in descending  
330 order of strength of negative correlation with genetic distance.

331



332 Figure 2. Mean of the correlation coefficients between genetic and behavioural  
333 differentiation for non-vocal communicative and other types of behavioural variants.  
334 Error bars represent 95 % confidence intervals.

335

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475 Table 1

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477

	Bossou	Tai North	Tai South	Kibale Ngogo	Kibale Kanyawara	Budongo Sonso	Gombe Kasekela	Mahale K	Mahale M
Bossou		-0.02	0.03	-0.10	-0.16	0.24	-0.19	-0.16	0.22
Tai North	0.98		0.96	0.02	-0.04	-0.11	-0.08	-0.02	-0.14
Tai South	0.99	1.06		0.11	0.05	-0.05	-0.01	-0.02	-0.08
Kibale Ngogo	<b>0.29</b>	<b>0.28</b>	<b>0.29</b>		0.98	0.23	0.10	0.36	0.40
Kibale Kanyawara	<b>0.31</b>	<b>0.29</b>	<b>0.32</b>	0.98		0.20	0.18	0.35	0.41
Budongo Sonso	<b>0.26</b>	<b>0.24</b>	<b>0.28</b>	<b>0.90</b>	<b>0.85</b>		-0.20	-0.02	0.19
Gombe Kasekela	<b>0.32</b>	<b>0.30</b>	<b>0.32</b>	<b>0.89</b>	<b>0.85</b>	<b>0.88</b>		0.12	0.13
Mahale K	<b>0.44</b>	<b>0.45</b>	<b>0.44</b>	0.94	0.84	<b>0.77</b>	0.93		0.58
Mahale M	<b>0.24</b>	<b>0.22</b>	<b>0.26</b>	<b>0.77</b>	<b>0.65</b>	<b>0.63</b>	<b>0.75</b>	1.18	

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480 Table 2

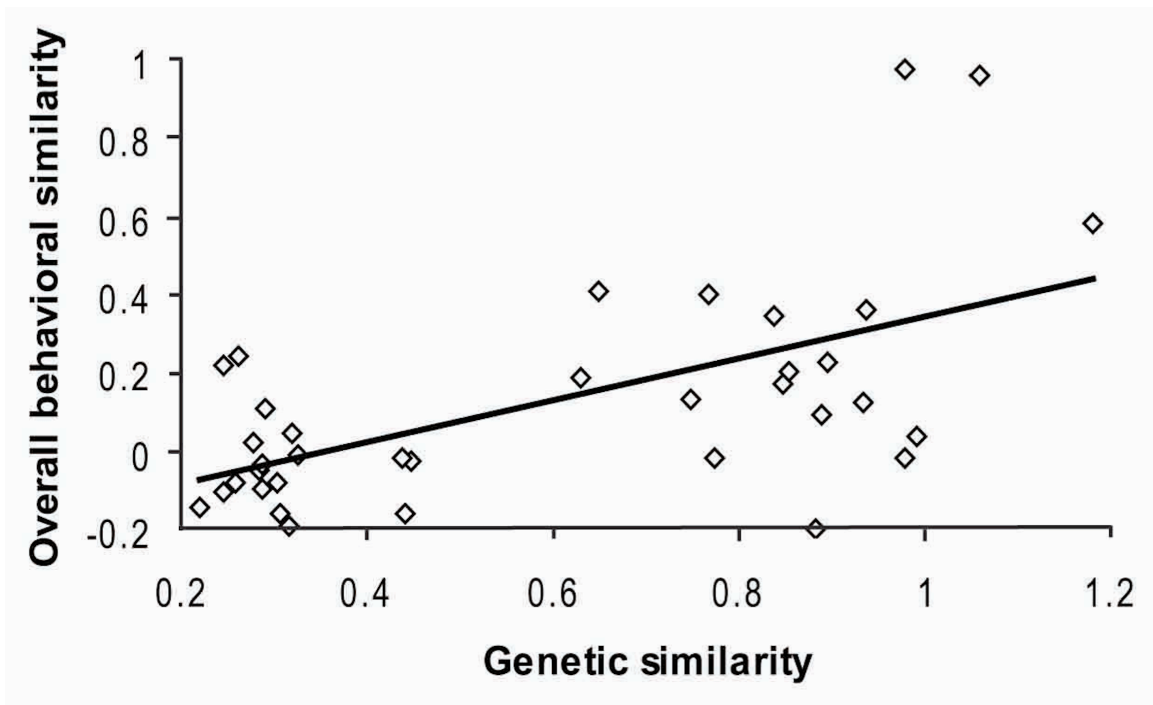
behavioral variant	N	spearman's r	p	non-vocal communicative?
Leaf-squash (squash ecto-parasite on leaf)	28	-0.21	0.26	no
Self-tickle (tickle self using objects)	36	-0.14	0.56	no
Leaf-clip, fingers (rip single leaf with fingers)	36	-0.11	0.48	yes
Leaf-clip, mouth (rip parts off leaf, with mouth)	36	-0.09	0.61	yes
Ant-dip-wipe (manually wipe ants off wand)	36	-0.07	0.63	no
Leaf-inspect (inspect ecto-parasite on hand)	28	-0.03	0.89	no
Leaf-dab (leaf dabbed on wound, examined)	36	-0.03	0.82	no
Leaf-strip (rip leaves off stem, as threat)	36	-0.02	0.88	yes
Fly-whisk (leafy stick used to fan flies)	21	0.00	0.99	no
Food-pound onto other (e.g., stone)	21	0.00	0.99	no
Stem pull-through (pull stems noisily)	36	0.00	0.99	yes
Expel/stir stick expels or stirs insects)	21	0.02	0.93	no
Branch-slap (slap branch, for attention)	36	0.03	0.82	yes
Ant-fish (probe used to extract ants)	36	0.03	0.78	no
Aimed-throw (throw object directionally)	36	0.06	0.60	no
Shrub-bend (squash stems underfoot)	36	0.07	0.63	yes
Lever open (stick used to enlarge entrance)	36	0.10	0.58	no
Fluid-dip (use of probe to extract fluids)	36	0.14	0.31	no
Bee-probe (disable bees, flick with probe)	36	0.15	0.35	no
Knuckle-knock (knock to attract attention)	36	0.16	0.28	yes
Hand-clasp (clasp arms overhead, groom)	36	0.18	0.20	no
Club (strike forcefully with stick)	36	0.24	0.10	no
Leaf-napkin (leaves used to clean body)	36	0.25	0.10	no
Nut-hammer, wood hammer on stone anvil	15	0.28	0.24	no
Nut-hammer, other (e.g., on ground)	15	0.28	0.24	no
Food-pound onto wood (smash food)	21	0.35	0.07	no
Index-hit (squash ecto-parasite on arm)	36	0.35	0.05	no
Marrow-pick (pick bone marrow out)	36	0.39	0.04	no
Rain dance (slow display at start of rain)	36	0.39	0.03	no
Seat-vegetation (large leaves as set)	36	0.49	0.02	no
Leaf-groom (intense 'grooming' of leaves)	36	0.72	0.01	no
Ant-dip-single (one handed dip stick on ants)	36	0.84	0.00	no
Nut-hammer, stone hammer on stone anvil	15	0.85	0.02	no

481

482

482 Figure 1.

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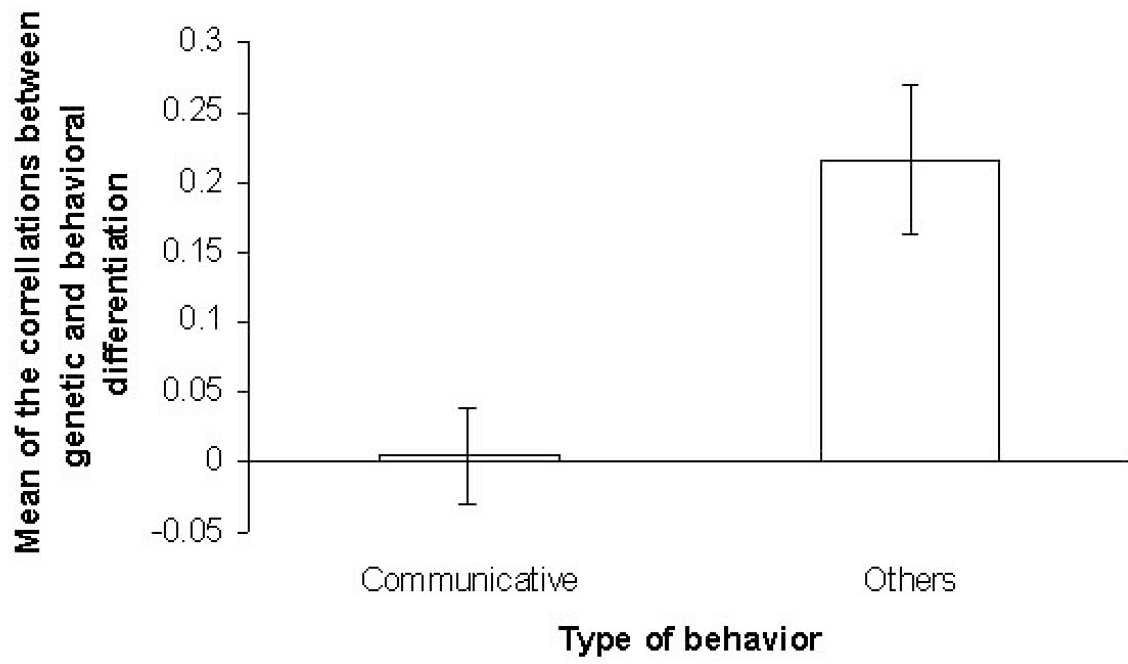
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486 Figure 2

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