Inhibition of SDF-1-induced migration of oncogene-driven myeloid leukemia by the L-RNA aptamer (Spiegelmer), NOX-A12, and potentiation of tyrosine kinase inhibition

The Harvard community has made this article openly available. Please share how this access benefits you. Your story matters

Citation

Published Version
doi:10.18632/oncotarget.22409

Citable link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:34868813

Terms of Use
This article was downloaded from Harvard University’s DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA
Inhibition of SDF-1-induced migration of oncogene-driven myeloid leukemia by the L-RNA aptamer (Spiegelmer), NOX-A12, and potentiation of tyrosine kinase inhibition

Ellen L. Weisberg¹,⁴, Martin Sattler¹,⁴, Abdel Kareem Azab¹,⁴, Dirk Eulberg³, Anna Kruschinski³, Paul W. Manley², Richard Stone¹,⁴ and James D. Griffin¹,⁴

¹Department of Medical Oncology, Dana-Farber Cancer Institute, Boston, MA 02215, USA
²Novartis Institutes of Biomedical Research, CH-4002 Basel, Switzerland
³Noxxon Pharma, Max-Dohrn-Strasse 8-10, 10589 Berlin, Germany
⁴Department of Medicine, Harvard Medical School, Boston, MA 02115, USA

Correspondence to: Ellen L. Weisberg, email: ellen_weisberg@dfci.harvard.edu
James D. Griffin, email: james_griffin@dfci.harvard.edu

Keywords: chronic myeloid leukemia; SDF-1; NOX-A12; nilotinib; drug resistance

Received: July 08, 2017  Accepted: October 25, 2017  Published: November 06, 2017

ABSTRACT

Resistance to targeted tyrosine kinase inhibitors (TKI) remains a challenge for the treatment of myeloid leukemias. Following treatment with TKIs, the bone marrow microenvironment has been found to harbor a small pool of surviving leukemic CD34+ progenitor cells. The long-term survival of these leukemic cells has been attributed, at least in part, to the protective effects of bone marrow stroma. We found that the NOX-A12 'Spiegelmer', an L-enantiomeric RNA oligonucleotide that inhibits SDF-1α, showed in vitro and in vivo activity against BCR-ABL- and FLT3-ITD-dependent leukemia cells. NOX-A12 was sufficient to suppress SDF-1-induced migration in vitro. The combination of NOX-A12 with TKIs reduced cell migration in the same in vitro model of SDF-1-induced chemotaxis to a greater extent than either drug alone, suggesting positive cooperativity as a result of the SDF-1 blocking function of NOX-A12 and cytotoxicity resulting from targeted oncogenic kinase inhibition. These results are consistent with our in vivo findings using a functional pre-clinical mouse model of chronic myeloid leukemia (CML), whereby we demonstrated the ability of NOX-A12, combined with the ABL kinase inhibitor, nilotinib, to reduce the leukemia burden in mice to a greater extent than either agent alone. Overall, the data support the idea of using SDF-1 inhibition in combination with targeted kinase inhibition to override drug resistance in oncogene-driven leukemia to significantly diminish or eradicate residual leukemic disease.

INTRODUCTION

CML is a malignancy of hematopoietic stem cells caused by the t(9;22) chromosome translocation product BCR-ABL [1]. Excessive proliferation and abnormal trafficking of transformed progenitor cells provides a survival advantage that is accompanied by premature release of these cells into the bloodstream of patients. This has been partially attributed to BCR-ABL-altered integrin expression or function affecting the communication between leukemic cells and the bone marrow stroma, as well as with important extracellular matrix proteins such as fibronectin [2–6]. Abnormal trafficking is also marked by the homing of malignant cells to bone marrow stroma and other tissue beds characterized by having high stromal cell content. This can lead to persistent minimal residual disease in CML patients undergoing therapy with targeted inhibitors such as imatinib and nilotinib, as stromal cells secrete growth factors and protect leukemia cells from targeted inhibitors [7]. Thus, in one study, CML recurred in more than 50% of patients who discontinued imatinib therapy after having achieved and maintained at least a
targeting CXCR4, including anti-CXCR4 monoclonal antibodies and peptide CXCR4 antagonists, for leukemia and lymphoma are currently under way [32].

An alternative to inhibiting the CXCR4 receptor is targeting its ligand, SDF-1, directly. SDF-1 is secreted and presented by stromal cells from bone marrow and other tissues by cell-surface-bound glycosaminoglycans (GAGs); this attracts leukemia cells and contributes to residual disease by conferring protection to the cytotoxic effect of small molecule inhibitors. NOX-A12 is an RNA oligonucleotide in L-configuration (or the mirror image of naturally occurring RNA) (Spiegelmer), which makes it unrecognizable by plasma nucleases and resistant to degradation, and which was originally designed to interfere with SDF-1 in the stromal microenvironment [33–34]. NOX-A12 has been shown to inhibit SDF-1-induced chemotaxis of chronic lymphocytic leukemia (CLL) cells and conversely augment migration of CLL cells beneath a bone marrow stromal cell (BMSC) layer by disrupting the BMSC-generated SDF-1 gradient [35]. It has also been found to reduce MM tumor burden in mice when combined with bortezomib [36], as well as to exhibit activity against glioblastoma in rat models [37] and potentiate anti-VEGF therapy against glioblastoma [38]. NOX-A12 was tested in healthy volunteers and found to be well-tolerated and to dose-dependently mobilize white blood cells and hematopoietic stem cells into the peripheral blood [39]. Thus far, encouraging results have been obtained in clinical trials testing NOX-A12 in combination with bendamustine and rituximab (BR) for the treatment of relapsed/refractory CLL patients, and testing of NOX-A12 in combination with bortezomib (Velcade) and dexamethasone (VD) in relapsed/refractory MM patients [40].

To date, to the best of our knowledge, therapeutic effects of NOX-A12 have not been explored in myeloid leukemia. Here, we show the ability of NOX-A12 to block SDF-1-induced migration of BCR-ABL-positive leukemia cells, as well as FLT3-ITD-positive leukemia cells. In addition, we demonstrate the combined effects of targeted kinase inhibition and NOX-A12 in vitro and in vivo.

RESULTS

NOX-A12 inhibits SDF-1-induced migration of BCR-ABL-expressing cells and potentiates the effects of ABL inhibition against BCR-ABL-positive cells in vitro

In transwell migration assays designed to test the ability of NOX-A12 to inhibit SDF-1-induced migration of BCR-ABL-expressing cells, we found that 50–100 nM NOX-A12 effectively inhibited the SDF-1-stimulated migration of BCR-ABL-expressing Ba/F3 and 32D cells and reduced migration to a level similar to unstimulated control cells (Figure 1A, 1D and Supplementary Figure 1A). Specifically, SDF-1-induced migration of
Ba/F3.p210 cells by 5–11.2 fold over control (Figure 1A, 1D), whereas in the presence of NOX-A12 (50–100 nM), SDF-1 only induced migration by 0.5–0.6 fold (Figure 1A, 1D). Similarly, SDF-1 induced migration of 32D.p210 cells by 23.1 fold, whereas in the presence of NOX-A12, SDF-1 only induced migration by 2 fold (Supplementary Figure 1A). Effects of SDF-1 observed in the transwell migration assay were confirmed to be due to effects on migration as opposed to increased growth of cells, as the proliferation of Ba/F3.p210 cells treated with SDF-1 for two days was found to be unchanged (Figure 1B). Specifically, in contrast to the 11.2-fold induction of Ba/F3.p210 cell migration by SDF-1 (Figure 1A), we observed a mere 1.1-fold difference between untreated and SDF-1-stimulated Ba/F3.p210 cells with respect to proliferation. In addition, we showed in a separate set of studies performed in parallel that 19 hours of stimulation of Ba/F3.p210 cells with SDF-1 led to a strong induction of cell migration in a transwell migration assay, whereas 19 hours of SDF-1 stimulation of Ba/F3.p210 cells did not lead to any measurable increase in cell proliferation (Supplementary Figure 1B–1C). We also tested (in parallel) the effects of NOX-A12 (50 nM) on proliferation as compared to migration, and we observed that while NOX-A12 strongly suppressed SDF-1-induced migration, it did not enhance or inhibit cell proliferation (Figure 1D–1E). These data are in support of the effects of NOX-A12 observed in the transwell migration assay being due to effects on migration as opposed to increased cell growth.

Importantly, NOX-A12 treatment combined with imatinib treatment of BCR-ABL-expressing cells resulted in reduced SDF-1-induced migration of cells as compared to NOX-A12 or imatinib alone, suggesting a positive combination effect of the two inhibitors in this in vitro assay when used together, likely due to direct inhibitory effects of NOX-A12 on SDF-1-induced cell migration coupled with cell cytotoxicity resulting from BCR-ABL kinase inhibition (Figure 1C). The difference between migration of control cells versus cells stimulated with SDF-1 was statistically significant \( (p < 0.001) \), and the differences between imatinib alone or NOX-A12 alone (each in the presence of SDF-1) and the combination of imatinib and NOX-A12 (in the presence of SDF-1) were statistically significant \( (p < 0.00005 \text{ and } p < 0.05, \text{ respectively}) \). The difference between migration for cells in the SDF-1 treatment group and migration for cells treated with imatinib+NOX-A12 in the presence of SDF-1 was significant \( (p < 0.0005) \), and there was no statistically significant difference between migration of control cells and cells treated with NOX-A12 in the absence of SDF-1 \( (p = 0.79) \).

The anti-SDF-1 effect of NOX-A12 was not limited to murine cells, as evidenced by its ability to reduce, however not abolish, SDF-1-induced migration of the human BCR-ABL-positive acute lymphoblastic leukemia (ALL) line, SUP-B15, and the human FLT3-ITD-positive line, MOLM14 (Figure 2). SUP-B15 cells, although not a myeloid line, is being used in our study as a representative human cell line model of Philadelphia chromosome-positive leukemia that is dependent on the BCR-ABL oncogene. Specifically, SDF-1 stimulation of SUP-B15 cells led to a 12.7-fold induction of migration of cells, whereas SDF-1 stimulation of SUP-B15 cells in the presence of NOX-A12 (10 nM) led to only a 4.7-fold increase in cell migration, and in the presence of NOX-A12 (100 nM) led to only a 2.8-fold increase in cell migration (Figure 2A).

**NOX-A12 inhibits SDF-1-induced migration of FLT3-ITD-expressing cells and potentiates the effects of FLT3 inhibition against mutant FLT3-positive cells**

As stromal cell-mediated chemoresistance has proven to be a challenge for TKI-treated leukemia, and TKIs, such as midostaurin, are now in clinical use for mutant FLT3-positive leukemia, it was of interest to explore the clinical and cooperative potential of NOX-A12 in the context of this disease. We first found that SDF-1 stimulation of MOLM14 cells led to a 4.96-fold induction of migration of cells, whereas SDF-1 stimulation of MOLM14 cells in the presence of NOX-A12 (100 nM) led to a 0.82-fold induction of migration of cells (Figure 2B). We next observed that NOX-A12 effectively inhibited the SDF-1-stimulated migration of FLT3-ITD-expressing MOLM13-luc+ cells in a transwell migration assay and reduced migration to a level similar to unstimulated control cells (Supplementary Figure 2A). Specifically, SDF-1 stimulation led to migration of 5,125 cells, as compared to 1,375 migrated cells in the control, whereas SDF-1 stimulation in the presence of NOX-A12 (25 nM) led to migration of only 1,500 cells (Supplementary Figure 2A). In addition, the combination of NOX-A12 (at 25 nM) and midostaurin led to slightly reduced SDF-1-induced migration of MOLM13-luc+ (813 cells) as compared to NOX-A12 alone (1,500 cells) or midostaurin alone (2,188 cells) in the transwell migration assay (Supplementary Figure 2A). A similar combination study was carried out for MOLM14 cells (Figure 2B). As expected, we observed robust SDF-1 induction of cell migration, which was strongly suppressed by 25 nM NOX-A12. The combination of midostaurin and NOX-A12 (at 25 nM) similarly led to slightly greater suppression of MOLM14 cell migration as compared to single agent-treated cells (Supplementary Figure 2B).

**NOX-A12 potentiates the anti-proliferative effects of ABL inhibition against BCR-ABL-positive cells in vivo**

The simultaneous demonstrated anti-SDF-1 influence of NOX-A12 coupled with cell killing by
TKI encouraged further studies exploring the ability of NOX-A12 to potentiate the effects of ABL inhibition in vivo, presumably by mobilizing BCR-ABL-positive leukemia cells out of bone marrow and into peripheral blood due to direct inhibition of SDF-1 and consequent blockade of its function in the stromal microenvironment as a chemoattractant. We utilized a mouse model of BCR-ABL-positive leukemia to examine the in vivo effects of SDF-1 inhibition by NOX-A12 (40 mg/kg every other day administered subcutaneously (sc)), combined with the ABL inhibitor, nilotinib, administered by oral gavage. In a pilot study designed to determine the optimal dose of nilotinib to use in combination with NOX-A12, mice treated for four days with 30mg/kg nilotinib showed strong and rapid loss of leukemia burden, whereas NOX-A12 alone did not inhibit in vivo leukemia growth (data not shown). As the effects of nilotinib were disproportionate to the effects of NOX-A12, the nilotinib dose was reduced...
by 2-fold and mice were treated orally for a total of 11 days with 15mg/kg nilotinib in the presence or absence of 40 mg/kg NOX-A12 every other day sc. Nilotinib-treated mice were observed to carry very low leukemia burden following 15 days total treatment (Supplementary Figure 3A, 3B) as compared to vehicle-treated mice and NOX-A12 only-treated mice. However, the combination of NOX-A12 and nilotinib led to further suppression of leukemia growth in 2 of 3 mice in the combination group as compared to nilotinib only-treated mice (Supplementary Figure 3C).

The results of the pilot study encouraged us to test nilotinib at a lower dose (7.5 mg/kg every day administered by oral gavage) alone and in combination with NOX-A12 in BCR-ABL-positive leukemia-bearing mice. As expected, NOX-A12 showed no single-agent

---

**Figure 2**: NOX-A12 inhibition of SDF-1-induced migration of BCR-ABL-expressing human SUP-B15 cells and FLT3-ITD-positive human MOLM14 cells. (A) Transwell migration assay (n = 1); error bars (standard deviation) represent intra-experimental variability, with multiple quantifications taken for one Transwell migration assay. SUP-B15 cells were stimulated with hSDF-1 (100 ng/mL) in the presence of NOX-A12 (1–100 nM). Data are presented as number of migrated cells. Transwell migration assay incubation time was 24 hours for SUP-B15. Data shown are mean +/- S.D. (B) Transwell migration assay (n = 1); error bars (standard deviation) represent intra-experimental variability, with multiple quantifications taken for one Transwell migration assay. MOLM14 cells were stimulated with hSDF-1 (50 ng/mL) in the presence of NOX-A12 (100 nM). Data are presented as fold migration/control, where control is normalized to a value of 1. Transwell migration assay incubation time was overnight. Data shown are mean +/- S.D.
activity, however in combination with nilotinib it led to reduced total leukemia burden assessed by total body bioluminescence (Figure 3A–3C). Spleens dissected from vehicle- and drug-treated mice were significantly smaller in the drug combination group than in vehicle-treated or single agent-treated mice (Figure 4). The size of spleens of NOX-A12-treated mice were significantly different from the spleen sizes of combination-treated mice ($p = 0.02564$), and the size of spleens of nilotinib-treated mice were significantly different from the spleen sizes of combination mice ($p = 0.03602$). Drugs, alone and combined, were generally well-tolerated by mice with no adverse toxicity to vital organs (data not shown). Since NOX-A12 exerted no efficacy as a single agent, these results demonstrate that NOX-A12, at a well-tolerated dose, acts cooperatively with nilotinib to suppress the growth of 32D.p210 leukemia.

**DISCUSSION**

The important role of stroma in leukemia cell trafficking is evident from the number of studies that have been carried out over the past couple of decades. Abnormal adhesion properties are believed to interfere with stroma:leukemia cell interaction and lead to increased leukemic cell proliferation and the untimely...
release of these cells out of patient bone marrow and into the peripheral blood. The converse homing of leukemic cells to bone marrow stroma dampens the efficacy of TKI therapy because of cytoprotection of leukemic cells and consequent development of minimal residual disease. New approaches to overriding stromal cell-associated chemoresistance, such as the use of agents that inhibit the CXCR4-SDF-1 axis, could potentially improve leukemia patient response and survival if developed as anticancer therapeutics. Of relevance in the case of CML, while there is a reduction of CXCR4 expression and signaling in BCR-ABL-positive cells, conversely treatment with imatinib or nilotinib increases CXCR4 surface expression on CML cells; this causes these cells to migrate to cytoprotective bone marrow stroma, rendering them resistant to TKI therapy [29, 41, 42].

NOX-A12 is an RNA oligonucleotide in L-configuration (Spiegelmer®) that binds and neutralizes SDF-1 thereby blocking its interaction with CXCR4 and CXCR7 and which also disrupts existing SDF-1 gradients by stripping the chemokine from GAGS [35]. By inhibiting SDF-1 with NOX-A12, the tumor microenvironment can be modulated in such a way as to mobilize leukemia cells to the peripheral blood where they are more susceptible to the cytotoxicity of anti-cancer agents. Additionally, as SDF-1 behaves as a survival factor for leukemic cells, blockade of SDF-1 could potentially reduce leukemia cell viability [40]. NOX-A12, because of its ability to block SDF-1 activity on both of its receptors, CXCR4 and CXCR7, provides additional benefit over a targeted CXCR4 receptor inhibitor because of its dual influence on CXCR7, which affects proliferation, migration, and metastasis of some cancers [43].

To the best of our knowledge, the activity of NOX-A12 alone and in combination with targeted therapies has not been explored for CML and AML. Here, we observed the ability of NOX-A12 to potentiate the effects of ABL inhibition against BCR-ABL-positive leukemia in vitro and in vivo. In transwell migration assays, NOX-A12 inhibited SDF-1-induced migration of BCR-ABL-expressing Ba/F3, 32D, and SUP-B15 (human Ph+ ALL) cells. NOX-A12 also potently inhibited SDF-

![Figure 4: NOX-A12 potentiation of effects of ABL inhibition against BCR-ABL-positive cells in vivo: Effects on spleen weight. Percent spleen/total weights for treatment groups. *NOX-A12 versus combination statistically significant (p = 0.025643). †Nilotinib versus combination statistically significant (p = 0.036021).](image-url)
1-induced migration of FLT3-ITD-expressing MOLM13 and MOLM14 cells.

Using a murine model of BCR-ABL-positive progressive disease, we found that NOX-A12 combined with nilotinib was more effective in reducing leukemia burden than either drug alone. In our models, NOX-A12 modestly enhanced the efficacy of nilotinib when nilotinib, as a single agent, was used at doses that led to substantial suppression of leukemia. However, a stronger positive combination effect was observed between NOX-A12 and a lower dose of nilotinib that was ineffective when administered as monotherapy. As such, combination treatment may have a particularly effective role in those patients not able to tolerate TKI treatment and for whom dose reduction of the TKI is needed.

Taken together, our data confirm the anti-SDF-1 activity of the ‘Spiegelmer’ and L-enantiomeric RNA oligonucleotide, NOX-A12. Our findings also suggest that NOX-A12 has the ability to potentiate the efficacy of TKIs against a subset of oncogene-driven leukemias.

MATERIALS AND METHODS

Cell lines and cell culture

BCR-ABL tyrosine kinase-dependent murine 32Dp210 cells were developed as described [44]. SUP-B15 cells were purchased from the American Type Culture Collection (ATCC) (Manassas, VA, USA). Ba/F3.p210 cells were obtained by transfecting the IL-3-dependent murine hematopoietic Ba/F3 cell line with a pGD vector containing p210BCR-ABL (B2A2) cDNA [45].

The human AML-derived, FLT3-ITD-expressing line, MOLM14, was provided to us by Dr. Scott Armstrong, Dana-Farber Cancer Institute (DFCI), Boston, MA. The human AML-derived, FLT3-ITD-expressing cell line, MOLM-13 (DSMZ (German Resource Centre for Biological Material), was engineered to express luciferase fused to neomycin phosphotransferase (pMMP-LucNeo) by transduction with a VSVG-pseudotyped retrovirus as previously described [46].

All cell lines used in this study were cultured with 5% CO₂ at 37°C, at a concentration of 2 x 10⁴ to 5 x 10⁴ in RPMI (Corning, Inc. Corning, NY) with 10% HyClone fetal bovine serum (FBS) (GE Healthcare Life Sciences, Pittsburgh, PA), and supplemented with 2% L-glutamine and 1% penicillin/streptomycin.

Human cell lines were authenticated through cell line short tandem repeat (STR) profiling (DDC Medical, Fairfield, OH and Molecular Diagnostics Laboratory, Dana-Farber Cancer Institute). All cell lines tested matched > 80% with lines listed in the ATCC or DSMZ Cell Line Bank STR. SUP-B15 cells were newly purchased from ATCC at the time of this study. All cell lines were confirmed to be virus- and mycoplasma-free.

Chemicals

Midostaurin (Rydapt) and nilotinib (Tasigna) were synthesized by Novartis Pharma AG, Basel, Switzerland and dissolved in DMSO to obtain a 10 mM stock solution. Serial dilutions were then made, to obtain final dilutions for cellular assays with a final concentration of DMSO not exceeding 0.1%. Stock solutions of the RNA nucleic acid, NOX-A12/olaptesed pegol (Noxxon, Germany), were prepared in 5% glucose in water at a 5 mg/mL concentration and stored as aliquots, light-protected, at -20°C.

Proliferation studies

The trypan blue exclusion assay has been previously described [47] and was used for quantification of cells prior to seeding for CellTiter-Glo Luminescent Cell Viability assays (Promega, Madison, WI). These assays were used for cell growth studies and carried out according to manufacturer’s instructions. Cell viability is reported as percentage of control (untreated) cells, and error bars represent the standard deviation for each data point.

Transwell migration assay

For murine Ba/F3.p210 and 32D.p210 cells, the lower chamber of a Transwell plate (6.5mm Transwell® with 5.0µm Pore Polycarbonate Membrane Insert, Sterile (Product #3421), Corning Costar Corporation, Cambridge, MA) was filled with 600 µL of migration (starvation) media (RPMI 1640/PBS+0.01% bovine serum albumin (BSA)), followed by placement of inserts into the wells. For human SUP-B15, MOLM13 and MOLM14 cells, the lower chamber of a Transwell plate (6.5mm Transwell® with 8.0µm Pore Polycarbonate Membrane Insert, Sterile (Product #3422), Corning Costar Corporation, Cambridge, MA) was filled with 600 µL of migration (starvation) media (IMDM/PBS+0.05% BSA) followed by placement of inserts into the wells. Cells were counted by Trypan Blue exclusion with a hemocytometer, and resuspended in migration media at a concentration of 2 x 10⁶ cells/mL. 100 µL of this cell suspension was transferred to the upper chamber (for a total of 2 x 10⁶ cells). The media in the lower chamber contained either no SDF-1 (for control samples) or 100 ng/mL mouse SDF-1 (for Ba/F3.p210 and 32D.p210 cells) or 50–100 ng/mL human SDF-1 (for MOLM13, MOLM14, or SUP-B15 cells). NOX-A12 and kinase inhibitors were included in the assay and added to the top and bottom chambers to avoid gradient formation at the indicated concentrations. After the indicated times, cells in the lower compartment were resuspended and counted using Trypan Blue exclusion. The spontaneous Transwell migration of cells was expressed as the total number of cells that migrated in the presence of SDF-1.
versus SDF-1+NOX-A12. Shown are mean+/− S.D. for each sample.

**Bioluminescent BCR-ABL model of CML**

32D.p210 cells were transduced with a retrovirus encoding firefly luciferase (MSCV-Luc), and selected with G418 at a concentration of 1 mg/mL to produce the 32D.p210-luciferase (luc+) cell-line. Bioluminescence imaging was carried out as previously described [48]. Briefly, virus- and Mycoplasma-free cells were washed in Hank’s Balanced Salt Solution (HBSS; Mediatech, Inc.,VA), resuspended in PBS for injection, and administered via IV tail vein injection (250 µL, 1 × 10^6 cells) into 15 female NOD.Cg-PrkdcscidIl2rgtm1WjlSzJ mice (or NOD scid gamma (NSG™)) (6 weeks of age) (cat # 005557), purchased from The Jackson Laboratory (Bar Harbor, ME). Mice were imaged 3 days later to establish baseline bioluminescence and were randomized into the following four treatment groups: Vehicles (administered NMP/PEG solution), NOX-A12-only (40 mg/kg sc every other day), nilotinib (7.5 mg/kg oral gavage every day), or a combination. Mice were continuously administered treatments until morbidity, at which time spleens were dissected and weighed and reported as a percentage of total mouse weight.

**Drug formulations for in vivo studies**

Solutions of nilotinib (Novartis Pharma AG, Basel) were prepared just prior to administration, by dissolving 100 mg in 1.0 mL of NMP to give a clear solution and diluting with 9.0 mL PEG300. Working solutions of NOX-A12 were prepared by adding ca. 50% of target volume (e.g. 5 mL to 100 mg container) of 5% glucose in water and dissolving the lyophilisate under aseptic conditions. Incubation for at least 30 minutes at 37°C was required to completely dissolve the lyophilisate with occasional vortexing. The solution was rinsed two times with ca. 20% of the target volume of solution medium (e.g. 2 mL) and the rinsed solution was transferred to a preparation container. Solution medium was added to target volume (e.g. 10 mL). Aliquots prepared at 10 mg/mL were stored, light protected, as stock solutions at −20°C and thawed prior to use.

**Statistical analysis**

Results were reported as the mean ± standard deviation. Samples were compared by the Student’s t-test, and results were considered significantly different for p values less than 0.05.

**Abbreviations**

tyrosine kinase inhibitors (TKI); chronic myeloid leukemia (CML); stromal-cell derived factor-1 (SDF-1 or CXCL-12); human SDF-1 (hSDF-1), mouse SDF-1 (mSDF-1); acute myeloid leukemia (AML); multiple myeloma (MM); glycosaminoglycans (GAGs); chronic lymphocytic leukemia (CLL); acute lymphoblastic leukemia (ALL); bone marrow stromal cell (BMSC); bendamustine and rituximab (BR); bortezomib (Velcade) and dexamethasone (VD); luciferase fused to neomycin phosphotransferase (pMMP-LucNeo); fetal bovine serum (FBS); bovine serum albumin (BSA); short tandem repeat (STR); American Type Culture Collection (ATCC); retrovirus encoding firefly luciferase (MSCV-Luc); luciferase (luc+); Hank’s Balanced Salt Solution (HBSS); subcutaneously (sc).

**Author contributions**

Ellen L. Weisberg is responsible for generation of research findings reported in paper (design/performance of experiments, including transwell migration assays and in vivo studies), integrity and analysis of the data, writing of the manuscript. Martin Sattler offered valuable scientific feedback and helped with conception of research reported in paper. Abdel Kareem Azab offered valuable scientific feedback and helped with conception of research reported in paper. Dirk Eulberg offered valuable scientific feedback, provided NOX-A12, and helped with conception of research reported in paper. Anna Kruschinski offered valuable scientific feedback, provided NOX-A12, and helped with conception of research reported in paper. Paul W. Manley offered valuable scientific feedback, provided nilotinib, imatinib, and midostaurin, and helped with conception of research reported in paper. Richard Stone offered valuable scientific feedback and helped with conception of research reported in paper. James D. Griffin offered valuable scientific feedback and helped with conception of research reported in paper.

**ACKNOWLEDGMENTS**

We would like to acknowledge Catherine Sypher, Daisy Moreno, and the Animal Resources Facility at DFCI staff for their contributions to the in vivo studies presented in this manuscript.

**CONFLICTS OF INTEREST**

D.E. and A.K. are employees of Noxxon Pharmaceuticals and have a financial interest with Noxxon Pharmaceuticals.

**FUNDING**

This work was funded by the NIH grant P01CA66996. P.W.M. is an employee of Novartis Pharma AG, J.D.G. and P.M. have a financial interest with Novartis Pharma AG.
REFERENCES


31. Weisberg E, Azab AK, Manley PW, Kung AL, Christie AL, Bronson R, Gobrial IM, Griffin JD. Inhibition of CXCR4 in CML cells disrupts their interaction with the bone marrow microenvironment and sensitizes them to nilotinib. Leukemia. 2012; 26:985–990.


47. Weisberg E, Boulton C, Kelly LM, Manley P, Fabbro D, Meyer T, Gilliland DG, Griffin JD. Inhibition of mutant FLT3 receptors in leukemia cells by the small molecule