



# Targeting the renin-angiotensin system to improve cancer treatment: Implications for immunotherapy

### Citation

Pinter, Matthias, and Rakesh K. Jain. 2018. "Targeting the renin-angiotensin system to improve cancer treatment: Implications for immunotherapy." Science translational medicine 9 (410): eaan5616. doi:10.1126/scitranslmed.aan5616. http://dx.doi.org/10.1126/scitranslmed.aan5616.

### **Published Version**

doi:10.1126/scitranslmed.aan5616

### Permanent link

http://nrs.harvard.edu/urn-3:HUL.InstRepos:37160084

## Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

# **Share Your Story**

The Harvard community has made this article openly available. Please share how this access benefits you. <u>Submit a story</u>.

<u>Accessibility</u>



# **HHS Public Access**

Author manuscript *Sci Transl Med.* Author manuscript; available in PMC 2018 May 01.

Published in final edited form as:

*Sci Transl Med.* 2017 October 04; 9(410): . doi:10.1126/scitranslmed.aan5616.

# Targeting the renin-angiotensin system to improve cancer treatment: Implications for immunotherapy

#### Matthias Pinter<sup>1,2</sup> and Rakesh K. Jain<sup>1,\*</sup>

<sup>1</sup>Edwin L. Steele Laboratories for Tumor Biology, Department of Radiation Oncology, Harvard Medical School and Massachusetts General Hospital, Boston, MA 02114, USA

<sup>2</sup>Division of Gastroenterology and Hepatology, Department of Internal Medicine III, Medical University of Vienna, Vienna, A-1090, Austria

#### Abstract

Renin-angiotensin system (RAS) inhibitors (RASi)—widely prescribed for the treatment of cardiovascular diseases— have considerable potential in oncology. The RAS plays a crucial role in cancer biology and affects tumor growth and dissemination directly and indirectly by remodeling the tumor microenvironment. We review clinical data on the benefit of RASi in primary and metastatic tumors and propose that, by activating immunostimulatory pathways, these inhibitors can enhance immunotherapy of cancer.

### INTRODUCTION

The circulating renin-angiotensin system (RAS) is mainly known for its pivotal role in maintaining cardiovascular homeostasis and fluid and electrolyte balance. In addition, a local RAS is expressed in many tissues and mainly acts at the cellular level, where it mediates cell proliferation, growth, and metabolism. The local RAS works synergistically and independently of the systemic RAS. Angiotensin II (AngII) is the main effector and maintains tissue homeostasis by exerting regulatory and counterregulatory effects through its different receptors. Alternative peptide-receptor axes also assist in maintaining this balance (1–7). Figure 1 provides an overview of the main components of the RAS. Dysregulation of the RAS, for example, by overexpression of certain RAS components [such as renin, Ang-converting enzyme (ACE), or AngII type 1 receptor (AT1R)], can be involved in the

SUPPLEMENTARY MATERIALS

www.sciencetranslationalmedicine.org/cgi/content/full/9/410/eaan5616/DC1

exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works. Distributed under a Creative Commons Attribution License 4.0 (CC BY).

<sup>\*</sup>Corresponding author. jain@steele.mgh.harvard.edu.

Author contributions: M.P. and R.K.J. performed the literature search, wrote the manuscript, and approved the final version of the manuscript.

**Competing interests:** M.P. received travel support from Bayer and speaking fees and consultant fees from Bayer and Bristol-Myers Squibb. R.K.J. received consultant fees from Ophthotech, Sun Pharma Advanced Research Corporation (SPARC), SynDevRx, and XTuit; owns equity in Enlight, Ophthotech, SynDevRx, and XTuit; and serves on the Board of Directors of XTuit and the Boards of Trustees of Tekla Healthcare Investors, Tekla Life Sciences Investors, Tekla Healthcare Opportunities Fund, and Tekla World Healthcare Fund. R.K.J. is an inventor on patent application (USSN 61/43 8,240 and USSN 61/643,487) submitted by MGH that covers the use of antihypertensive agents for cancer therapy.

pathophysiology and progression of a broad range of diseases, such as arterial hypertension, kidney disease, and other cardiovascular conditions (5, 8, 9).

The discoveries of captopril—the first orally active ACE inhibitor (ACEi)—in the mid-1970s (10) and losartan—the first orally active, selective AT1R blocker (ARB)—around a decade later (11) represent milestones in the history of the RAS. Numerous ACEis and ARBs have been developed since then. Now, ACEis and ARBs are the most common inhibitors of the RAS and are widely used in the management of several diseases, such as arterial hypertension, heart failure, myocardial infarction, and chronic kidney disease (12–15). Direct renin inhibitors (such as aliskiren) represent a third class of RAS-acting agents and have been added to the armamentarium more recently (16). A list of RAS inhibitors (RASi) approved by the U.S. Food and Drug Administration (FDA) is provided in table S1.

After being in clinical use for more than two decades in nonmalignant diseases, ACEi/ARBs have recently received considerable attention in oncology. A large-scale meta-analysis (17), published in 2010, found an increased overall occurrence of cancer in ARB users. However, two other meta-analyses published subsequently did not confirm these data (18, 19). The FDA also rebutted these findings with their own meta-analysis (20) and an integrated analysis of all 19 rodent carcinogenicity assays of ARBs (21). Thus, the data to date do not support an association between ACEi/ARB use and an increased cancer risk. However, they do not suggest a reduced occurrence of cancer either.

Of interest, an increasing number of preclinical studies support the involvement of RAS signaling in cancer development, growth, and progression (4). These data have led to investigations of the effects of RASi—both retrospectively and prospectively—in patients with different types of cancer. Interim analysis of a recent phase 2 trial—stemming from our preclinical findings (22)—showed encouraging R0 (microscopically margin-negative) resection rates in patients with locally advanced pancreatic ductal adenocarcinoma (PDAC) receiving neoadjuvant losartan plus chemoradiation (23). Moreover, our recent retrospective analysis indicated that RASi use is associated with improved survival of patients with nonmetastatic PDAC, presumably by stimulating the tumor's immune microenvironment, normalizing its extracellular matrix (ECM), and reducing the malignant potential of cancer cells (24).

In light of these emerging data, we discuss the role of the RAS in cancer biology with a special emphasis on tumor immunity. In addition, by carefully analyzing the studies with positive versus negative outcomes, we make a case for targeting the RAS to improve treatment of certain malignancies. Moreover, RASi may not only improve the outcome of immunotherapies but also reduce or even prevent adverse effects associated with these therapies.

# The Angll/AT1R axis shapes the tumor microenvironment and promotes an immunosuppressive milieu

Components of the RAS are expressed in various human cancers and cell lines (4). Overexpression of AT1R is typically associated with more aggressive tumor features (larger tumors, higher grade, and higher vascular density) and worse outcomes (25–29).

Moreover, RAS components are also expressed in many cell types of the tumor microenvironment, such as endothelial cells, fibroblasts, monocytes, macrophages, neutrophils, dendritic cells, and T cells (4, 30–34). RAS signaling in these cells can facilitate or hinder growth and dissemination and has been shown to affect cell proliferation, migration, invasion, metastasis, apoptosis, angiogenesis, cancer-associated inflammation, immunomodulation, and tumor fibrosis/desmoplasia (1, 4). Generally, the AngII/AT1R axis is considered to favor tumor growth, whereas AngII/AT2R and Ang(1–7)/MAS signaling have opposing effects (1, 4). However, there are also conflicting reports suggesting potential tumor type–specific differences (35–39).

The tumor-promoting actions of the ACE/AngII/AT1R axis, the main target of classical RASi, have been reviewed elsewhere (1, 4). In this section, we focus on its role in tumor immunity and propose RASi as an adjunct for immunotherapy. Immune checkpoint inhibitors have recently achieved compelling success in melanoma and other solid tumors (40). However, their efficacy is diminished by a major barrier—the immunosuppressive tumor microenvironment (41). Here, we review how AngII/AT1R signaling shapes the tumor immune microenvironment by modulating desmoplasia, vasculature, inflammation, and immune cells. We also discuss how RASi could alleviate immunosuppression and enhance the outcome of immunotherapy.

#### Tumor desmoplasia and solid stress

By regulating cancer-associated fibroblasts (CAFs) and profibrogenic pathways [such as transforming growth factor– $\beta$  (TGF- $\beta$ )], the RAS plays a key role in establishing a desmoplastic environment (22, 42), which affects the immune response in multiple ways (Fig. 2). CAFs can manipulate the immune system directly by inhibiting T and NK (natural killer) cell functions, promoting accumulation of suppressive cell types, and maintaining an inflammatory protumorigenic milieu (43). TGF- $\beta$  can also directly induce immune suppression by inhibiting the T cell response (44). Dense tumor fibrosis represents a physical barrier to T cell infiltration (45). It also compresses blood vessels by increasing solid stress (46, 47). The reduced tumor perfusion results in a hypoxic and acidic milieu, which promotes reprogramming of macrophages into an immunosuppressive phenotype, impairs tumor killing functions of immune cells, and up-regulates the expression of inhibitory immune checkpoint molecules, such as programmed death-ligand 1 (PD-L1), by immune, stromal, and tumor cells (Fig. 3) (46–51). Normalizing the desmoplastic milieu (for example, by targeting profibrotic pathways and CAFs) can improve the efficacy of immunotherapy (52–54).

Several studies have demonstrated that RASi can successfully normalize the fibrotic stroma. Co-injection of cancer cells with stromal cells increases tumor size and fibrosis, and treatment with ARBs attenuates these effects (55, 56). Losartan inhibits collagen I production by CAFs and reduces stromal collagen and hyaluronic acid (HA) in several desmoplastic tumor models by decreasing profibrotic signaling via TGF- $\beta$ , connective tissue growth factor, HA synthase 1 and 3, and endothelin-1 (22). Therefore, losartan reduces solid stress and improves vascular perfusion, resulting in decreased tumor hypoxia and improved distribution and efficacy of anticancer drugs and nanotherapeutics (22, 42). Similarly,

inhalation delivery of losartan and telmisartan reduces active TGF- $\beta$  and collagen I expression and increases the intratumoral distribution of nanoparticles (57, 58). Moreover, the cross-talk between tumor-associated neutrophils (TANs), adipocytes, and pancreatic stellate cells (PSCs) promotes tumor desmoplasia and pancreatic cancer growth in obesity (59). AT1R inhibition attenuates obesity-induced fibrosis and tumor progression and improves response to chemotherapy (CHT). The AT1R blockade also reduces TANs and regulatory T cells (T<sub>regs</sub>) and increases CD8<sup>+</sup> T cells through inhibition of PSC activation and subsequent reduction of interleukin-1 $\beta$  (IL-1 $\beta$ ) expression (59). In another orthotopic model of pancreatic cancer, inhibition of aberrant TGF- $\beta$  activity by losartan reduced collagen deposition and accumulation of T<sub>regs</sub> (60).

Collectively, these data support the idea that targeting AngII/AT1R signaling with RASi can effectively reduce tumor desmoplasia and thereby decrease solid stress, increase tumor perfusion, reduce hypoxia, enhance T cell infiltration and antitumor immunity, and improve delivery and efficacy of anticancer drugs. Thus, inhibiting the AngII/AT1R axis appears to be an attractive strategy, especially for highly desmoplastic tumors, such as PDAC and some subtypes of breast and lung cancer, and RASi may represent a promising combination partner for immunotherapy.

#### Angiogenesis and tumor vasculature

Considerable evidence suggests that AngII/AT1R signaling promotes VEGF-mediated angiogenesis in solid tumors. AT1R expression correlates with VEGF and VEGF receptor (VEGFR) expression and microvessel density (MVD) in different human tumors (26, 27, 29). In experimental studies, AngII promoted VEGF expression in tumor (61–63) and stromal cells (64). Treatment with either ACEi or ARB reduced VEGF expression and decreased MVD and neovascularization in vivo (65, 66).

VEGF also induces vascular hyperpermeability, one of the main characteristics of the abnormal tumor vasculature (46, 48). Tumor vessel leakiness promotes tumor hypoxia and acidosis by impairing tumor blood flow (Fig. 2) (48, 67). As mentioned above, hypoxia helps to create an immunosuppressive milieu (Fig. 3) and promotes tumor progression and dissemination (48, 68). Tumor vessel normalization can alleviate hypoxia, reprogram the immunosuppressive microenvironment, and improve the efficacy of immunotherapy in mice (68, 69). Glioblastoma patients who show enhanced tumor blood perfusion under antiangiogenic therapy have markedly prolonged survival compared to subjects who experience no change or a decrease in perfusion (70–72). RASi also reduces VEGF-mediated vascular leakiness in the dermis and retina of rodents (73, 74).

In an orthotopic model of PDAC, inhibition of aberrant TGF- $\beta$  signaling by losartan restored vessel diameter and permeability (60). In a retrospective study of glioblastoma patients receiving anticancer therapy, a concomitant treatment with matrix-depleting antihypertensive drugs improved vascular function as assessed by magnetic resonance imaging (75).

The impaired perfusion and hypoxic condition of tumors can be further aggravated by AngII-induced vasoconstriction and increased vascular resistance (Fig. 2) (76, 77). Our laboratory has shown that AngII transiently enhanced tumor blood flow and interstitial fluid

pressure by increasing the mean arterial blood pressure in different tumor types (78, 79). However, Thews and colleagues (80) found that AngII infusion decreased tumor perfusion and oxygenation in small subcutaneous sarcomas but increased both parameters in large tumors. They concluded that perfusion decreased due to vasoconstriction of preexisting functionally intact host vessels in small sarcomas, whereas the newly formed tumor vessels in large tumors did not seem to have this vasoresponsive capability, possibly due to lack of smooth muscle cells and/or angiotensin (AT) receptors (80).

Together, available data indicate that AngII/AT1R signaling impairs tumor blood supply through multiple mechanisms, such as desmoplasia-mediated vessel compression, VEGF-induced vessel leakiness and abnormal morphology, and AngII-mediated vasoconstriction of host vessels. The resulting tumor hypoxia aggravates immunosuppression and evasion. Although RASi can reduce VEGF-mediated angiogenesis and desmoplasia, additional studies are needed to ascertain whether RASis have the ability to normalize the tumor vasculature, similar to anti-VEGF agents (48).

#### Inflammation and immune cell modulation

The RAS promotes cancer-related inflammation and infiltration of tumor-promoting immune cells (1, 4, 81), both of which enhance the immunosuppressive micro-environment (41, 82). Here, we discuss how the RAS modulates the expression of inflammatory cytokines and orchestrates the recruitment of cancer-associated immune cells to the tumor microenvironment.

**Inflammatory cytokines**—A number of studies have shown that AngII/AT1R signaling can increase the production and release of several proinflammatory cytokines in both tumor and stromal cells (4). Fibroblasts represent a main target of the RAS and play a pivotal role in maintaining an inflammatory response. Cytokines released from tumor and stromal cells upon AT1R activation by AngII include TGF- $\beta$ , IL-1 $\alpha$ , IL-1 $\beta$ , IL-6,IL-8,MCP-1(monocyte chemoattractant protein–1), M-CSF, COX-2 (cyclooxygenase-2), and CRP (C-reactive protein) (Fig. 2) (4, 22, 42, 56, 59, 65, 83–87). Immunomodulatory cytokines (such as TGF- $\beta$ , IL-1 $\beta$ , MCP-1, IL-6, and IL-8) can up-regulate multiple—mostly immunosuppressive— pathways by modulating the differentiation and recruitment of both myeloid and lymphoid immune cell types (Fig. 2) (44, 82, 88–91). COX-2 suppresses antitumor immunity and contributes to resistance to immunotherapy, mainly through prostaglandin E<sub>2</sub> synthesis (92, 93). The role of tumor-derived CRP in tumor immunity is less clear, but it may impair dendritic cell function by reducing their migration activity (94).

Oxidative stress represents another aspect of cancer-related inflammation. Although reactive oxygen species (ROS) are involved in T cell activation (95, 96), exposure to ROS can reduce T cell fitness (90, 97, 98) and enhance the function of  $T_{regs}$  (99) and TAMs (100). TAMs typically show a polarized M2-like phenotype and contribute to immunosuppression, whereas M1-like macrophages are known to induce anti-tumor immunity (101). AngII/AT1R signaling induces ROS generation in tumor cells and stromal cells (4). In prostate cancer cells, AngII-mediated expression of oxidative stress–related proteins (such as

inducible nitric oxide synthase) and the generation of the ROS family member  $O_2^-$  radical are attenuated by the ARB candesartan (102).

**Immune cells**—Several studies have shown that RASi can reduce infiltration of TAMs. In human prostate cancer, high MCP-1 and macrophage infiltration are associated with more aggressive tumor features, and MCP-1 independently correlates with prostate-specific antigen recurrence (103). AngII/AT1R signaling promotes production and infiltration of TAMs in experimental tumor models; inhibition of AngII production or AT1R signaling down-regulates MCP-1, restrains tumor-induced TAM response, reduces tumor growth, and prolongs survival (34, 103–105).

AngII/AT1R signaling is also important for myeloid differentiation and functional maturation (106). ACE knockout mice show enhanced extramedullary myelopoiesis and increased numbers of cells with MDSC phenotype (32). In contrast, cultured bone marrow from ACE 10/10 mice, a mouse line overexpressing ACE in monocytic cells, demonstrates enhanced myeloid maturation and reduced MDSC production; macrophages from these mice have a more proinflammatory phenotype and more antitumor activity compared to those from wild-type mice (107). Similarly, tumor-bearing ACE 10/10 mice showed enhanced immune response, which ultimately resulted in a reduced tumor growth. Notably, ACEi reversed the beneficial effects on tumor growth, but AT1R blockade did not, suggesting that the effects of ACE overexpression were not dependent on AngII/AT1R signaling (108, 109).

Together, available data clearly demonstrate that AngII/AT1R signaling stimulates the expression of different cytokines and growth factors from tumor and stromal cells, which enhance cancer-related inflammation and promote an immunosuppressive microenvironment (Fig. 2). Beyond the tumor immune microenvironment, the AngII/AT1R axis is also crucial for the maturation and function of immunostimulatory myeloid cells, and ACE overexpression in monocytic cells enhances antitumor immunity, although the latter effect seems to be independent of the AngII/AT1R axis. These conflicting data highlight the complexity of the RAS in cancer immunity. However, because studies supporting a stimulatory role of RAS in tumor immunosuppression considerably outweigh opposing data, we propose that RASi can effectively reprogram the tumor microenvironment toward an immunostimulatory milieu and enhance the efficacy of immunotherapy.

#### RASi to reduce side effects of immunotherapy

As discussed above, RASi may increase the intratumoral delivery of T cells and immunotherapeutic agents by modulating tumor vasculature and desmoplasia. This may allow for reduction in the dose of immunotherapeutic agents without decreasing the therapeutic benefit and could ultimately result in a decreased number of severe (grades 3 and 4) immunotherapy-related adverse effects. These side effects can occur in more than 50% of patients, especially if certain checkpoint blockers are combined, and some can be even life-threatening (110, 111).

Obesity and associated chronic inflammation seem to play a critical role in inducing immunotherapy-associated toxicities (112, 113). Systemic stimulatory immunotherapy, such as aCD40/IL-2, can cause a cytokine storm, characterized by high tumor necrosis factor–a.

(TNF-α) and IL-6, resulting in multiorgan pathologies and lethality in obese but not in lean mice (112, 113). The TNF blockade ameliorates the observed toxicities in obese mice (113). Inhibition of the RAS can also ameliorate chronic inflammation, as shown by reduced serum concentrations of proinflammatory cytokines (TNF-α and IL-6) in patients with hypertension and diabetes (114–116). This represents another way that RASi may help to reduce or even prevent immunotherapy-induced toxicity.

#### RAS inhibition can improve treatment of certain tumors

The effect of RASi on the clinical outcome of patients with different tumor types has been extensively studied in recent years. Tables S2 and S3 provide an overview of the published prospective (117–126) and retrospective studies (24, 127–175), respectively. Here, we summarize the main conclusions based on the available data.

#### RASi usage in conjunction with CHT

Available clinical data suggest that RASi may potentiate the effect of certain systemic antitumor therapies. The use of RASi was associated with better outcomes in patients with different solid tumors who received platinum-based CHT (142, 143, 149, 165, 172). The gain in overall survival (OS; the length of time from either the date of diagnosis or the start of treatment that patients are still alive) ranged from ~3 months in advanced non–small cell lung cancer (NSCLC) to 5.7 months in advanced gastric cancer and even 11 months in metastatic colorectal cancer (CRC) (142, 149, 165, 172). In line with the clinical data, experimental studies showed that platinum-based CHT can increase VEGF production through up-regulation of AT1R expression. This seems to represent a mechanism for platinum resistance that can be successfully targeted by RASi (176, 177).

In addition, concomitant RASi treatment was associated with better survival in patients with metastatic renal cell carcinoma (RCC; gain in OS, 7 to 26 months) (137-140), metastatic CRC (gain in OS, ~11 months) (172), glioblastoma (175), and advanced hepatocellular carcinoma (HCC; gain in OS, ~5 months) (173) who received VEGF-targeted therapies. Because AngII/AT1R signaling promotes VEGF-mediated angiogenesis (4), RASi may potentiate the effect of anti-VEGF therapy. In a mouse model of Ehrlichs's ascites carcinoma, the ARB olmesartan augmented the anti-angiogenic effect of the tyrosine kinase inhibitor (TKI) sorafenib (178). RASi may also represent a strategy to inhibit rapid revascularization (179, 180) and regrowth of tumors (181, 182) after cessation of anti-VEGF therapy, which is often necessary due to treatment-related side effects, especially with VEGFR TKIs (183, 184). Notably, arterial hypertension is a common side effect of anti-VEGF therapy and can be associated with better survival outcomes (185). VEGF-targeted therapy-induced hypertension is often treated with RASi, which could represent a potential con-founder for the reported beneficial survival results associated with RASi use in patients who received anti-VEGF therapies. However, two points suggest otherwise: First, some studies reported the number of patients who received RASi either at baseline or after initiation of anti-VEGF therapy and showed that most of the patients were taking RASi already at baseline (137, 139). Second, McKay and colleagues (140) demonstrated that even in the subgroup of patients who developed anti-VEGF therapy-induced hypertension, RASi users had improved survival compared to nonusers.

Finally, two studies suggested a putative clinical benefit of RASi use in patients who received epidermal growth factor receptor (EGFR) TKIs (128, 143). This could be explained by the preclinical finding that AT1R signaling can regulate proliferation and migration of cancer cells through transactivation of the EGFR by metalloproteinase-dependent shedding of EGF ligands (4).

#### Tumor characteristics as determinants of RASi efficacy

RASi use was associated with better outcomes in multiple studies, whereas no association was found in others. This suggests that response to RASi treatment may also vary by tumor type and depend on certain tumor characteristics, as discussed below.

In breast cancer, only 2 of 13 studies shown in tables S2 and S3 reported beneficial effects of RASi use, whereas 3 studies found worse outcomes. A meta-analysis found no association of ACEi/ARB use with disease-free survival (DFS; the length of time after primary treatment for a cancer ends that the patient survives without any signs or symptoms of that cancer) or OS in breast cancer (186). The heterogeneity in terms of tumor stage, hormone receptor status, human epidermal growth factor receptor 2 overexpression, and (neo)adjuvant treatment regimen could have masked a potential benefit of RASi in certain subgroups and highlights the need for careful patient selection to obtain homogenous and comparable study cohorts.

The use of RASi was associated with better outcomes in patients with RCC, CRC, and HCC (tables S2 and S3). These tumors are well known to respond to anti-VEGF therapy (187–189). As discussed earlier, RASi may enhance the efficacy of VEGF-targeted therapies and thereby improve clinical outcome. However, in HCC (125, 126, 159, 164) and some CRC (167) and RCC (144) studies listed in tables S2 and S3, most of the patients were not treated with anti-VEGF treatment, suggesting that anti-VEGF–responsive tumors generally seem to be more sensitive to RASi.

RASi therapy had a clinical benefit in both slowly progressing cancers, such as prostate cancer, and highly aggressive tumor types, such as glioblastoma and pancreatic cancer (tables S2 and S3). A phase 2 study at the Massachusetts General Hospital (MGH) is currently investigating whether adding losartan to CHT (FOLFIRINOX), followed by chemoradiation, can convert locally advanced PDAC to resectable tumors (23). Preliminary results from this trial showed that R0 resection was achieved in 13 of 25 patients (52%), which is a major improvement compared to previously reported R0 resection rates obtained with neoadjuvant FOLFIRINOX and radiation in locally advanced PDAC (23 to 24%) (190, 191). The median OS was 33 months, with a 2-year survival rate of 65% for all patients and 83% for resected patients (23).

In addition, RASi use was effective in both early and advanced tumor stages. In some tumor types, the effect of RASi was investigated primarily in either early tumors (such as resected urinary tract cancer) (130, 147, 150, 151) or advanced stages (such as metastatic NSCLC) (142, 149). In RCC and CRC, positive outcomes were reported for both early (144, 167) and metastatic diseases (137–140, 172). Notably, in PDAC, a survival benefit in RASi users was

only shown for locally advanced/metastatic diseases treated with CHT (168–170) but not for resected early/locally advanced tumors (174).

In contrast, in our own retrospective analysis, RASi use was associated with longer OS in pancreatic cancer patients with resected primary tumors (median OS, 36.3 versus 19.3 months) and locally advanced tumors (median OS, 11.3 versus 9.3 months) but not in metastatic patients. To obtain mechanistic insights, we performed RNA sequencing expression profiling of prospectively collected cancer treatment-naïve pancreatic cancer samples (four lisinopril-treated patients versus four controls). Our data suggest that lisinopril, which was the most commonly used ACEi in our cohort, normalized the ECM, down-regulated genes involved in cancer progression (such as Wnt and Notch signaling), and up-regulated genes associated with the activity of T cells and antigen-presenting cells. In addition, we identified a predictive gene signature for RASi-mediated survival, which was validated in two publicly available cohorts (24). A recently published meta-analysis pooling data on different solid tumor types (192) showed that the use of ACEi or ARB was associated with improved DFS and OS. After pooling studies that were classified as early (I/II) or advanced (III/IV) stage-dominant, the association with DFS remained significant in both stages (P = 0.04 and P = 0.03, respectively); a positive association with OS was only observed in advanced tumor stage (192).

Finally, HCC usually develops in patients with underlying liver fibrosis/cirrhosis (193). The peritumoral liver tissue and the severity of liver dysfunction determine prognosis of HCC, and complications of cirrhosis (portal hypertension and variceal bleeding) are a common cause of death in patients with HCC (193). The AngII/AT1R axis plays a crucial role in the pathophysiology of liver cirrhosis (194), and RASi can improve both liver fibrosis (195) and portal hypertension (196). These effects, in addition to the direct antitumor effects of RASi, may also contribute to the improved outcome observed in HCC patients treated with RASi (125, 126, 159, 164, 173).

#### CONCLUSIONS

Preclinical studies have provided compelling evidence that the AngII/AT1R axis regulates almost all hallmarks of cancer. RASi can directly attenuate tumor growth and dissemination and improve the efficacy of systemic therapies by increasing drug delivery to the tumor tissue. The latter should help to reduce the dose of CHT and immunotherapy without decreasing the benefit and consequently decrease the anticancer therapy–induced side effects.

It is also clear that AngII/AT1R signaling contributes to the immunosuppressive tumor microenvironment in multiple ways. The immunosuppressive milieu is a major barrier for immunotherapy and may explain why immune checkpoint inhibitors have failed in some tumor types, such as PDAC, and have benefited only a fraction of patients in other indications where these agents are approved. Studies have shown that AT1R inhibition can decrease infiltration of immunosuppressive cell types and increase the number of effector T cells. This could also help to reduce the dose of immunotherapy without lowering drug efficacy, eventually resulting in a decreased number of severe immunotherapy-induced side

effects. Although not yet studied in the context of tumor immunity, the AngII/AT1R axis is also important for the maturation of immune effector cells.

Multiple clinical studies have also revealed that RASi may have beneficial effects in a broad range of malignancies. The gain in survival is tumor type– and stage-dependent and ranged from 3 months (advanced NSCLC) to more than 25 months (metastatic RCC) in retrospective studies. However, response to RASi treatment may not only vary with tumor types but also depend on certain tumor characteristics, cancer treatment, and RASi type and dosing. More precisely, RCC, HCC, PDAC, glioblastoma, urinary tract cancer, and NSCLC seem to belong to the responsive tumor types, whereas breast cancer is rather unresponsive to RASi. With respect to cancer treatment, RASi use was associated with better outcomes in patients with NSCLC, gastric cancer, and CRC who received platinum-based CHT and in those with RCC, HCC, and CRC treated with anti-VEGF therapy (for example, sunitinib). More data are needed for other tumor types, such as melanoma, thyroid cancer, head and neck cancer, and hematologic malignancies.

Because the clinical evidence largely came from retrospective studies and small prospective pilot trials, these findings should be considered as hypothesis-generating. However, given the large amount of preclinical and clinical data suggesting a beneficial effect of RASi in different cancer types, we propose that RASis have a great potential to become an adjunct within the oncological armamentarium. Ongoing trials testing whether RASi can improve the antitumor effect of certain anticancer treatments are listed in table S4.

#### Future perspectives and translational challenges

Advancing the promising strategy to reprogram the tumor micro-environment with RASi to enhance anticancer treatment will require a close interplay between basic and clinical research and addressing a number of outstanding questions. Preclinical research should combine immune checkpoint inhibitors or other immunotherapy approaches with RASi to confirm whether RASis have the potential to reprogram the immunosuppressive microenvironment and eventually render tumors more sensitive to immunotherapies. In addition, mechanistic studies should not only focus on effects of RASi on the tumor stroma but also investigate treatment-related changes within immune cell populations in the bone marrow and lymphoid organs. This will help to better understand the role of the RAS in cancer immunity.

Moreover, clinical pilot studies focusing on biological readouts—such as intratumoral ECM deposition, immune cell infiltration, and drug distribution—should be designed to confirm the available preclinical data and to pave the way for large randomized controlled efficacy trials. These studies should seek to identify those patients who may benefit most from concomitant RASi use. Such personalized approaches require a tight integration between measurements of various biomarkers—circulating (profibrotic molecules, immune cells, and chemokines), tissue (profibrotic molecules, collagen, and HA), and imaging (perfusion, oxygenation, and drug distribution)—and the treatment outcome (197). Assessing the intratumoral expression of the components of the RAS may also have the potential to predict response to RASi treatment.

Finally, the beneficial response of tumors to RASi is dose-dependent. For example, the collagen content of desmoplastic tumors decreases with an increasing dose of ARBs (42). However, increasing the dose can cause hypotension and other adverse effects. One potential solution to this challenge is to develop nanoformulations of RASi that will preferentially deliver RASi to the tumor microenvironment. Addressing these issues and challenges will unravel the complexity of RAS signaling and its role in different malignancies and enable development of new strategies to deliver RASi to tumors in safe doses with an even better outcome.

#### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

#### Acknowledgments

We thank Y. Boucher, I. Chen, A. Crane, M. Datta, M. Khandekar, H. Liu, M. Pittet, and K. Naxerova for helpful comments. We also apologize to all authors whose papers are not cited because of the limitations in number of references.

**Funding:** M.P. is supported by an Erwin-Schroedinger Fellowship by the Austrian Science Fund (project no. J 3747-B28). R.K.J. is supported by the National Cancer Institute (NCI; grants P01-CA080124, P50-CA165962, R01-CA129371, R01-CA208205, and U01-CA 224348), NCI Outstanding Investigator Award (R35-CA197743), the Lustgarten Foundation, the Ludwig Center at Harvard, the National Foundation for Cancer Research, and the Gates Foundation.

#### REFERENCES AND NOTES

- Ager EI, Neo J, Christophi C. The renin-angiotensin system and malignancy. Carcinogenesis. 2008; 29:1675–1684. [PubMed: 18632755]
- Bader M. Tissue renin-angiotensin-aldosterone systems: Targets for pharmacological therapy. Annu Rev Pharmacol Toxicol. 2010; 50:439–465. [PubMed: 20055710]
- de Gasparo M, Catt KJ, Inagami T, Wright JW, Unger T. International union of pharmacology. XXIII. The angiotensin II receptors. Pharmacol Rev. 2000; 52:415–472. [PubMed: 10977869]
- George AJ, Thomas WG, Hannan RD. The renin-angiotensin system and cancer: Old dog, new tricks. Nat Rev Cancer. 2010; 10:745–759. [PubMed: 20966920]
- Paul M, Poyan Mehr A, Kreutz R. Physiology of local renin-angiotensin systems. Physiol Rev. 2006; 86:747–803. [PubMed: 16816138]
- Qaradakhi T, Apostolopoulos V, Zulli A. Angiotensin (1–7) and alamandine: Similarities and differences. Pharmacol Res. 2016; 111:820–826. [PubMed: 27456244]
- 7. Rodrigues-Ferreira S, Nahmias C. G-protein coupled receptors of the renin-angiotensin system: New targets against breast cancer? Front Pharmacol. 2015; 6:24. [PubMed: 25741281]
- Kobori H, Nangaku M, Navar LG, Nishiyama A. The intrarenal renin-angiotensin system: From physiology to the pathobiology of hypertension and kidney disease. Pharmacol Rev. 2007; 59:251– 287. [PubMed: 17878513]
- Ferrario CM. Role of angiotensin II in cardiovascular disease—Therapeutic implications of more than a century of research. J Renin Angiotensin Aldosterone Syst. 2006; 7:3–14. [PubMed: 17083068]
- Cushman DW, Ondetti MA. History of the design of captopril and related inhibitors of angiotensin converting enzyme. Hypertension. 1991; 17:589–592. [PubMed: 2013486]
- Bhardwaj G. How the antihypertensive losartan was discovered. Expert Opin Drug Discov. 2006; 1:609–618. [PubMed: 23506070]
- 12. Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JGF, Coats AJS, Falk V, González-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT,

Pieske B, Riley JP, Rosano GMC, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P. Authors/ Task Force Members, 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC)Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. Eur Heart J. 2016; 37:2129–2200. [PubMed: 27206819]

- 13. James PA, Oparil S, Carter BL, Cushman WC, Dennison-Himmelfarb C, Handler J, Lackland DT, LeFevre ML, MacKenzie TD, Ogedegbe O, Smith SC Jr, Svetkey LP, Taler SJ, Townsend RR, Wright JT Jr, Narva AS, Ortiz E. 2014 evidence-based guideline for the management of high blood pressure in adults: Report from the panel members appointed to the Eighth Joint National Committee (JNC 8). JAMA. 2014; 311:507–520. [PubMed: 24352797]
- Levin A, Stevens PE. Summary of KDIGO 2012 CKD Guideline: Behind the scenes, need for guidance, and a framework for moving forward. Kidney Int. 2014; 85:49–61. [PubMed: 24284513]
- 15. O'Gara PT, Kushner FG, Ascheim DD, Casey DE Jr, Chung MK, de Lemos JA, Ettinger SM, Fang JC, Fesmire FM, Franklin BA, Granger CB, Krumholz HM, Linderbaum JA, Morrow DA, Newby LK, Ornato JP, Ou N, Radford MJ, Tamis-Holland JE, Tommaso CL, Tracy CM, Woo YJ, Zhao DX, Anderson JL, Jacobs AK, Halperin JL, Albert NM, Brindis RG, Creager MA, DeMets D, Guyton RA, Hochman JS, Kovacs RJ, Ohman EM, Stevenson WG, Yancy CW. American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines, 2013 ACCF/AHA guideline for the management of ST-elevation myocardial infarction: A report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. Circulation. 2013; 127:e362–e425. [PubMed: 23247304]
- Shafiq MM, Menon DV, Victor RG. Oral direct renin inhibition: Premise, promise, and potential limitations of a new antihypertensive drug. Am J Med. 2008; 121:265–271. [PubMed: 18374681]
- Sipahi I, Debanne SM, Rowland DY, Simon DI, Fang JC. Angiotensin-receptor blockade and risk of cancer: Meta-analysis of randomised controlled trials. Lancet Oncol. 2010; 11:627–636. [PubMed: 20542468]
- ARB Trialists Collaboration. Effects of telmisartan, irbesartan, valsartan, candesartan, and losartan on cancers in 15 trials enrolling 138,769 individuals. J Hypertens. 2011; 29:623–635. [PubMed: 21358417]
- Bangalore S, Kumar S, Kjeldsen SE, Makani H, Grossman E, Wetterslev J, Gupta AK, Sever PS, Gluud C, Messerli FH. Antihypertensive drugs and risk of cancer: Network meta-analyses and trial sequential analyses of 324,168 participants from randomised trials. Lancet Oncol. 2011; 12:65–82. [PubMed: 21123111]
- 20. FDA Drug Safety Communication. No increase in risk of cancer with certain blood pressure drugs —Angiotensin Receptor Blockers (ARBs). Jul 15. 2010 www.fda.gov/DrugS/DrugSafety/ ucm257516.htm
- Link WT, De Felice A. An FDA overview of rodent carcinogenicity studies of angiotensin II AT-1 receptor blockers: Pulmonary adenomas and carcinomas. Regul Toxicol Pharmacol. 2014; 70:555– 563. [PubMed: 25223563]
- 22. Chauhan VP, Martin JD, Liu H, Lacorre DA, Jain SR, Kozin SV, Stylianopoulos T, Mousa AS, Han X, Adstamongkonkul P, Popovi Z, Huang P, Bawendi MG, Boucher Y, Jain RK. Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. Nat Commun. 2013; 4:2516. [PubMed: 24084631]
- 23. Murphy JE, Wo JY-L, Ferrone C, Jiang W, Yeap BY, Blaszkowsky LS, Kwak EL, Allen JN, Clark JW, Faris JE, Zhu AX, Goyal L, Mamon HJ, Lillemoe KD, Ryan DP, DeLaney TF, Fernandez-del Castillo C, Boucher Y, Hong TS. TGF-B1 inhibition with losartan in combination with FOLFIRINOX (F-NOX) in locally advanced pancreatic cancer (LAPC): Preliminary feasibility and R0 resection rates from a prospective phase II study. J Clin Oncol. 2017; 35(suppl. 4S):386.
- 24. Liu H, Naxerova K, Pinter M, Incio J, Lee H, Shigeta K, Ho WW, Crain JA, Jacobson A, Michelakos T, Dias-Santos D, Zanconato A, Hong TS, Clark JW, Murphy JE, Ryan DP, Deshpande V, Lillemoe KD, Fernandez-del Castillo C, Downes M, Evans RM, Michaelson J, Ferrone CR, Boucher Y, Jain RK. Use of angiotensin system inhibitors is associated with immune activation and longer survival in non-metastatic pancreatic ductal adenocarcinoma. Clin Cancer Res. 2017

- Arrieta O, Pineda-Olvera B, Guevara-Salazar P, Hernández-Pedro N, Morales-Espinosa D, Cerón-Lizarraga TL, González-De la Rosa CH, Rembao D, Segura-Pacheco B, Sotelo J. Expression of AT1 and AT2 angiotensin receptors in astrocytomas is associated with poor prognosis. Br J Cancer. 2008; 99:160–166. [PubMed: 18594540]
- 26. Arrieta O, Villarreal-Garza C, Vizcaíno G, Pineda B, Hernández-Pedro N, Guevara-Salazar P, Wegman-Ostrosky T, Villanueva-Rodríguez G, Gamboa-Domínguez A. Association between AT1 and AT2 angiotensin II receptor expression with cell proliferation and angiogenesis in operable breast cancer. Tumour Biol. 2015; 36:5627–5634. [PubMed: 25682288]
- 27. Ino K, Shibata K, Kajiyama H, Yamamoto E, Nagasaka T, Nawa A, Nomura S, Kikkawa F. Angiotensin II type 1 receptor expression in ovarian cancer and its correlation with tumour angiogenesis and patient survival. Br J Cancer. 2006; 94:552–560. [PubMed: 16434990]
- Rocken C, Rohl FW, Diebler E, Lendeckel U, Pross M, Carl-McGrath S, Ebert MP. The angiotensin II/angiotensin II receptor system correlates with nodal spread in intestinal type gastric cancer. Cancer Epidemiol Biomarkers Prev. 2007; 16:1206–1212. [PubMed: 17548686]
- Shirotake S, Miyajima A, Kosaka T, Tanaka N, Maeda T, Kikuchi E, Oya M. Angiotensin II type 1 receptor expression and microvessel density in human bladder cancer. Urology. 2011; 77:1009.e19–1009.e25.
- Balyasnikova IV, Danilov SM, Muzykantov VR, Fisher AB. Modulation of angiotensin-converting enzyme in cultured human vascular endothelial cells. In Vitro Cell Dev Biol Anim. 1998; 34:545– 554. [PubMed: 9719414]
- 31. Danilov SM, Sadovnikova E, Scharenborg N, Balyasnikova IV, Svinareva DA, Semikina EL, Parovichnikova EN, Savchenko VG, Adema GJ. Angiotensin-converting enzyme (CD143) is abundantly expressed by dendritic cells and discriminates human monocyte-derived dendritic cells from acute myeloid leukemia-derived dendritic cells. Exp Hematol. 2003; 31:1301–1309. [PubMed: 14662338]
- 32. Lin C, Datta V, Okwan-Duodu D, Chen X, Fuchs S, Alsabeh R, Billet S, Bernstein KE, Shen XZ. Angiotensin-converting enzyme is required for normal myelopoiesis. FASEB J. 2011; 25:1145– 1155. [PubMed: 21148418]
- Shen XZ, Billet S, Lin C, Okwan-Duodu D, Chen X, Lukacher AE, Bernstein KE. The carboxypeptidase ACE shapes the MHC class I peptide repertoire. Nat Immunol. 2011; 12:1078– 1085. [PubMed: 21964607]
- 34. Cortez-Retamozo V, Etzrodt M, Newton A, Ryan R, Pucci F, Sio SW, Kuswanto W, Rauch PJ, Chudnovskiy A, Iwamoto Y, Kohler R, Marinelli B, Gorbatov R, Wojtkiewicz G, Panizzi P, Mino-Kenudson M, Forghani R, Figueiredo JL, Chen JW, Xavier R, Swirski FK, Nahrendorf M, Weissleder R, Pittet MJ. Angiotensin II drives the production of tumor-promoting macrophages. Immunity. 2013; 38:296–308. [PubMed: 23333075]
- 35. Azevedo H, Fujita A, Bando SY, Iamashita P, Moreira-Filho CA. Transcriptional network analysis reveals that AT1 and AT2 angiotensin II receptors are both involved in the regulation of genes essential for glioma progression. PLOS ONE. 2014; 9:e110934. [PubMed: 25365520]
- Li X, Zhang H, Soledad-Conrad V, Zhuang J, Uhal BD. Bleomycin-induced apoptosis of alveolar epithelial cells requires angiotensin synthesis de novo. Am J Physiol Lung Cell Mol Physiol. 2003; 284:L501–L507. [PubMed: 12573988]
- Nguyen L, Ager EI, Neo J, Christophi C. Regulation of colorectal cancer cell epithelial to mesenchymal transition by the renin angiotensin system. J Gastroenterol Hepatol. 2016; 31:1773– 1782. [PubMed: 26849969]
- Papp M, Li X, Zhuang J, Wang R, Uhal BD. Angiotensin receptor subtype AT<sub>1</sub> mediates alveolar epithelial cell apoptosis in response to ANG II. Am J Physiol Lung Cell Mol Physiol. 2002; 282:L713–L718. [PubMed: 11880296]
- Zheng S, Yang Y, Song R, Yang X, Liu H, Ma Q, Yang L, Meng R, Tao T, Wang S, He J. Ang-(1– 7) promotes the migration and invasion of human renal cell carcinoma cells via Mas-mediated AKT signaling pathway. Biochem Biophys Res Commun. 2015; 460:333–340. [PubMed: 25783053]
- 40. Smyth MJ, Ngiow SF, Ribas A, Teng MWL. Combination cancer immunotherapies tailored to the tumour microenvironment. Nat Rev Clin Oncol. 2016; 13:143–158. [PubMed: 26598942]

- Munn DH, Bronte V. Immune suppressive mechanisms in the tumor microenvironment. Curr Opin Immunol. 2016; 39:1–6. [PubMed: 26609943]
- 42. Diop-Frimpong B, Chauhan VP, Krane S, Boucher Y, Jain RK. Losartan inhibits collagen I synthesis and improves the distribution and efficacy of nanotherapeutics in tumors. Proc Natl Acad Sci USA. 2011; 108:2909–2914. [PubMed: 21282607]
- Ohlund D, Elyada E, Tuveson D. Fibroblast heterogeneity in the cancer wound. J Exp Med. 2014; 211:1503–1523. [PubMed: 25071162]
- 44. Li MO, Flavell RA. TGF-β: A master of all T cell trades. Cell. 2008; 134:392–404. [PubMed: 18692464]
- 45. Watt J, Kocher HM. The desmoplastic stroma of pancreatic cancer is a barrier to immune cell infiltration. OncoImmunology. 2013; 2:e26788. [PubMed: 24498555]
- Jain RK. Normalizing tumor microenvironment to treat cancer: Bench to bedside to biomarkers. J Clin Oncol. 2013; 31:2205–2218. [PubMed: 23669226]
- 47. Jain RK, Martin JD, Stylianopoulos T. The role of mechanical forces in tumor growth and therapy. Annu Rev Biomed Eng. 2014; 16:321–346. [PubMed: 25014786]
- Jain RK. Antiangiogenesis strategies revisited: From starving tumors to alleviating hypoxia. Cancer Cell. 2014; 26:605–622. [PubMed: 25517747]
- 49. Noman MZ, Hasmim M, Messai Y, Terry S, Kieda C, Janji B, Chouaib S. Hypoxia: A key player in antitumor immune response. A review in the theme: Cellular responses to hypoxia. Am J Physiol Cell Physiol. 2015; 309:C569–C579. [PubMed: 26310815]
- Palazón A, Aragonés J, Morales-Kastresana A, de Landázuri MO, Melero I. Molecular pathways: Hypoxia response in immune cells fighting or promoting cancer. Clin Cancer Res. 2012; 18:1207– 1213. [PubMed: 22205687]
- 51. Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet AL, Latreche S, Bergaya S, Benhamouda N, Tanchot C, Stockmann C, Combe P, Berger A, Zinzindohoue F, Yagita H, Tartour E, Taieb J, Terme M. VEGF-A modulates expression of inhibitory checkpoints on CD8<sup>+</sup> T cells in tumors. J Exp Med. 2015; 212:139–148. [PubMed: 25601652]
- 52. Chen Y, Ramjiawan RR, Reiberger T, Ng MR, Hato T, Huang Y, Ochiai H, Kitahara S, Unan EC, Reddy TP, Fan C, Huang P, Bardeesy N, Zhu AX, Jain RK, Duda DG. CXCR4 inhibition in tumor microenvironment facilitates anti-programmed death receptor-1 immunotherapy in sorafenib-treated hepatocellular carcinoma in mice. Hepatology. 2015; 61:1591–1602. [PubMed: 25529917]
- 53. Feig C, Jones JO, Kraman M, Wells RJB, Deonarine A, Chan DS, Connell CM, Roberts EW, Zhao Q, Caballero OL, Teichmann SA, Janowitz T, Jodrell DI, Tuveson DA, Fearon DT. Targeting CXCL12 from FAP-expressing carcinoma-associated fibroblasts synergizes with anti–PD-L1 immunotherapy in pancreatic cancer. Proc Natl Acad Sci USA. 2013; 110:20212–20217. [PubMed: 24277834]
- 54. Jiang H, Hegde S, Knolhoff BL, Zhu Y, Herndon JM, Meyer MA, Nywening TM, Hawkins WG, Shapiro IM, Weaver DT, Pachter JA, Wang-Gillam A, DeNardo DG. Targeting focal adhesion kinase renders pancreatic cancers responsive to checkpoint immunotherapy. Nat Med. 2016; 22:851–860. [PubMed: 27376576]
- 55. Masamune A, Hamada S, Kikuta K, Takikawa T, Miura S, Nakano E, Shimosegawa T. The angiotensin II type I receptor blocker olmesartan inhibits the growth of pancreatic cancer by targeting stellate cell activities in mice. Scand J Gastroenterol. 2013; 48:602–609. [PubMed: 23477656]
- 56. Okazaki M, Fushida S, Harada S, Tsukada T, Kinoshita J, Oyama K, Tajima H, Ninomiya I, Fujimura T, Ohta T. The angiotensin II type 1 receptor blocker candesartan suppresses proliferation and fibrosis in gastric cancer. Cancer Lett. 2014; 355:46–53. [PubMed: 25224569]
- Godugu C, Patel AR, Doddapaneni R, Marepally S, Jackson T, Singh M. Inhalation delivery of Telmisartan enhances intratumoral distribution of nanoparticles in lung cancer models. J Control Release. 2013; 172:86–95. [PubMed: 23838154]
- Patel K, Doddapaneni R, Chowdhury N, Boakye CHA, Behl G, Singh M. Tumor stromal disrupting agent enhances the anticancer efficacy of docetaxel loaded PEGylated liposomes in lung cancer. Nanomedicine. 2016; 11:1377–1392. [PubMed: 27171485]

- 59. Incio J, Liu H, Suboj P, Chin SM, Chen IX, Pinter M, Ng MR, Nia HT, Grahovac J, Kao S, Babykutty S, Huang Y, Jung K, Rahbari NN, Han X, Chauhan VP, Martin JD, Kahn J, Huang P, Desphande V, Michaelson J, Michelakos TP, Ferrone CR, Soares R, Boucher Y, Fukumura D, Jain RK. Obesity-induced inflammation and desmoplasia promote pancreatic cancer progression and resistance to chemotherapy. Cancer Discov. 2016; 6:852–869. [PubMed: 27246539]
- 60. Arnold SA, Rivera LB, Carbon JG, Toombs JE, Chang CL, Bradshaw AD, Brekken RA. Losartan slows pancreatic tumor progression and extends survival of SPARC-null mice by abrogating aberrant TGFβ activation. PLOS ONE. 2012; 7:e31384. [PubMed: 22348081]
- 61. Anandanadesan R, Gong Q, Chipitsyna G, Witkiewicz A, Yeo CJ, Arafat HA. Angiotensin II induces vascular endothelial growth factor in pancreatic cancer cells through an angiotensin II type 1 receptor and ERK1/2 signaling. J Gastrointest Surg. 2008; 12:57–66. [PubMed: 18026817]
- Ji Y, Wang Z, Li Z, Li K, Le X, Zhang T. Angiotensin II induces angiogenic factors production partly via AT1/JAK2/STAT3/SOCS3 signaling pathway in MHCC97H cells. Cell Physiol Biochem. 2012; 29:863–874. [PubMed: 22613986]
- 63. Kosaka T, Miyajima A, Shirotake S, Kikuchi E, Hasegawa M, Mikami S, Oya M. Ets-1 and hypoxia inducible factor-1alpha inhibition by angiotensin II type-1 receptor blockade in hormonerefractory prostate cancer. Prostate. 2010; 70:162–169. [PubMed: 19760626]
- 64. Fujita M, Hayashi I, Yamashina S, Fukamizu A, Itoman M, Majima M. Angiotensin type 1a receptor signaling-dependent induction of vascular endothelial growth factor in stroma is relevant to tumor-associated angiogenesis and tumor growth. Carcinogenesis. 2005; 26:271–279. [PubMed: 15637093]
- 65. Kosugi M, Miyajima A, Kikuchi E, Horiguchi Y, Murai M. Angiotensin II type 1 receptor antagonist candesartan as an angiogenic inhibitor in a xenograft model of bladder cancer. Clin Cancer Res. 2006; 12:2888–2893. [PubMed: 16675585]
- 66. Yoshiji H, Kuriyama S, Kawata M, Yoshii J, Ikenaka Y, Noguchi R, Nakatani T, Tsujinoue H, Fukui H. The angiotensin-I–converting enzyme inhibitor perindopril suppresses tumor growth and angiogenesis: Possible role of the vascular endothelial growth factor. Clin Cancer Res. 2001; 7:1073–1078. [PubMed: 11309359]
- Jain RK. Determinants of tumor blood flow: A review. Cancer Res. 1988; 48:2641–2658. [PubMed: 3282647]
- Huang Y, Goel S, Duda DG, Fukumura D, Jain RK. Vascular normalization as an emerging strategy to enhance cancer immunotherapy. Cancer Res. 2013; 73:2943–2948. [PubMed: 23440426]
- 69. Huang Y, Yuan J, Righi E, Kamoun WS, Ancukiewicz M, Nezivar J, Santosuosso M, Martin JD, Martin MR, Vianello F, Leblanc P, Munn LL, Huang P, Duda DG, Fukumura D, Jain RK, Poznansky MC. Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. Proc Natl Acad Sci USA. 2012; 109:17561–17566. [PubMed: 23045683]
- 70. Batchelor TT, Gerstner ER, Emblem KE, Duda DG, Kalpathy-Cramer J, Snuderl M, Ancukiewicz M, Polaskova P, Pinho MC, Jennings D, Plotkin SR, Chi AS, Eichler AF, Dietrich J, Hochberg FH, Lu-Emerson C, Iafrate AJ, Ivy SP, Rosen BR, Loeffler JS, Wen PY, Sorensen AG, Jain RK. Improved tumor oxygenation and survival in glioblastoma patients who show increased blood perfusion after cediranib and chemoradiation. Proc Natl Acad Sci USA. 2013; 110:19059–19064. [PubMed: 24190997]
- 71. Emblem KE, Mouridsen K, Bjornerud A, Farrar CT, Jennings D, Borra RJH, Wen PY, Ivy P, Batchelor TT, Rosen BR, Jain RK, Sorensen AG. Vessel architectural imaging identifies cancer patient responders to anti-angiogenic therapy. Nat Med. 2013; 19:1178–1183. [PubMed: 23955713]
- 72. Sorensen AG, Emblem KE, Polaskova P, Jennings D, Kim H, Ancukiewicz M, Wang M, Wen PY, Ivy P, Batchelor TT, Jain RK. Increased survival of glioblastoma patients who respond to antiangiogenic therapy with elevated blood perfusion. Cancer Res. 2012; 72:402–407. [PubMed: 22127927]
- 73. Gilbert RE, Kelly DJ, Cox AJ, Wilkinson-Berka JL, Rumble JR, Osicka T, Panagiotopoulos S, Lee V, Hendrich EC, Jerums G, Cooper ME. Angiotensin converting enzyme inhibition reduces retinal

overexpression of vascular endothelial growth factor and hyperpermeability in experimental diabetes. Diabetologia. 2000; 43:1360–1367. [PubMed: 11126403]

- 74. Sano H, Hosokawa K, Kidoya H, Takakura N. Negative regulation of VEGF-induced vascular leakage by blockade of angiotensin II type 1 receptor. Arterioscler Thromb Vasc Biol. 2006; 26:2673–2680. [PubMed: 16973968]
- 75. Emblem KE, Gerstner ER, Sorensen G, Rosen BR, Wen PY, Batchelor TT, Jain RK. Abstract 3975: Matrix-depleting anti-hypertensives decompress tumor blood vessels and improve perfusion in patients with glioblastoma receiving anti-angiogenic therapy. Cancer Res. 2016; 76(suppl. 14): 3975.
- 76. Bell KM, Prise VE, Shaffi KM, Chaplin DJ, Tozer GM. A comparative study of tumour-blood-flow modification in two rat-tumour systems using endothelin-1 and angiotensin II: Influence of tumour size on angiotensin-II response. Int J Cancer. 1996; 67:730–738. [PubMed: 8782666]
- 77. Tozer GM, Shaffi KM. The response of tumour vasculature to angiotensin II revealed by its systemic and local administration to 'tissue-isolated' tumours. Br J Cancer. 1995; 72:595–600. [PubMed: 7669567]
- Zlotecki RA, Baxter LT, Boucher Y, Jain RK. Pharmacologic modification of tumor blood flow and interstitial fluid pressure in a human tumor xenograft: Network analysis and mechanistic interpretation. Microvasc Res. 1995; 50:429–443. [PubMed: 8583955]
- Zlotecki RA, Boucher Y, Lee I, Baxter LT, Jain RK. Effect of angiotensin II induced hypertension on tumor blood flow and interstitial fluid pressure. Cancer Res. 1993; 53:2466–2468. [PubMed: 8495405]
- Thews O, Kelleher DK, Vaupel P. Disparate responses of tumour vessels to angiotensin II: Tumour volume-dependent effects on perfusion and oxygenation. Br J Cancer. 2000; 83:225–231. [PubMed: 10901375]
- Okwan-Duodu D, Landry J, Shen XZ, Diaz R. Angiotensin-converting enzyme and the tumor microenvironment: Mechanisms beyond angiogenesis. Am J Physiol Regul Integr Comp Physiol. 2013; 305:R205–R215. [PubMed: 23739345]
- Balkwill FR, Mantovani A. Cancer-related inflammation: Common themes and therapeutic opportunities. Semin Cancer Biol. 2012; 22:33–40. [PubMed: 22210179]
- Ji Y, Wang Z, Li Z, Zhang A, Jin Y, Chen H, Le X. Angiotensin II enhances proliferation and inflammation through AT1/PKC/NF-κB signaling pathway in hepatocellular carcinoma cells. Cell Physiol Biochem. 2016; 39:13–32. [PubMed: 27322819]
- 84. Uemura H, Ishiguro H, Nagashima Y, Sasaki T, Nakaigawa N, Hasumi H, Kato S, Kubota Y. Antiproliferative activity of angiotensin II receptor blocker through cross-talk between stromal and epithelial prostate cancer cells. Mol Cancer Ther. 2005; 4:1699–1709. [PubMed: 16275991]
- Matsuzuka T, Miller K, Pickel L, Doi C, Ayuzawa R, Tamura M. The synergistic induction of cyclooxygenase-2 in lung fibroblasts by angiotensin II and pro-inflammatory cytokines. Mol Cell Biochem. 2009; 320:163–171. [PubMed: 18827978]
- Pham H, Chong B, Vincenti R, Slice LW. Ang II and EGF synergistically induce COX-2 expression via CREB in intestinal epithelial cells. J Cell Physiol. 2008; 214:96–109. [PubMed: 17559081]
- Slice LW, Chiu T, Rozengurt E. Angiotensin II and epidermal growth factor induce cyclooxygenase-2 expression in intestinal epithelial cells through small GTPases using distinct signaling pathways. J Biol Chem. 2005; 280:1582–1593. [PubMed: 15525649]
- David JM, Dominguez C, Hamilton DH, Palena C. The IL-8/IL-8R axis: A double agent in tumor immune resistance. Vaccines. 2016; 4:E22. [PubMed: 27348007]
- Martin F, Apetoh L, Ghiringhelli F. Controversies on the role of Th17 in cancer: A TGF-β– dependent immunosuppressive activity? Trends Mol Med. 2012; 18:742–749. [PubMed: 23083809]
- Ugel S, De Sanctis F, Mandruzzato S, Bronte V. Tumor-induced myeloid deviation: When myeloid-derived suppressor cells meet tumor-associated macrophages. J Clin Invest. 2015; 125:3365–3376. [PubMed: 26325033]
- 91. Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, Lichtor T, Decker WK, Whelan RL, Kumara HMCS, Signori E, Honoki K, Georgakilas AG, Amin A, Helferich WG, Boosani CS,

Guha G, Ciriolo MR, Chen S, Mohammed SI, Azmi AS, Keith WN, Bilsland A, Bhakta D, Halicka D, Fujii H, Aquilano K, Ashraf SS, Nowsheen S, Yang X, Choi BK, Kwon BS. Immune evasion in cancer: Mechanistic basis and therapeutic strategies. Semin Cancer Biol. 2015; 35(Suppl):S185–S198. [PubMed: 25818339]

- Brown JR, DuBois RN. COX-2: A molecular target for colorectal cancer prevention. J Clin Oncol. 2005; 23:2840–2855. [PubMed: 15837998]
- 93. Zelenay S, van der Veen AG, Böttcher JP, Snelgrove KJ, Rogers N, Acton SE, Chakravarty P, Girotti MR, Marais R, Quezada SA, Sahai E, Reis e Sousa C. Cyclooxygenase-dependent tumor growth through evasion of immunity. Cell. 2015; 162:1257–1270. [PubMed: 26343581]
- Frenzel H, Pries R, Brocks CP, Jabs WJ, Wittkopf N, Wollenberg B. Decreased migration of myeloid dendritic cells through increased levels of C-reactive protein. Anticancer Res. 2007; 27:4111–4115. [PubMed: 18225580]
- 95. Jackson SH, Devadas S, Kwon J, Pinto LA, Williams MS. T cells express a phagocyte-type NADPH oxidase that is activated after T cell receptor stimulation. Nat Immunol. 2004; 5:818–827. [PubMed: 15258578]
- 96. Sena LA, Li S, Jairaman A, Prakriya M, Ezponda T, Hildeman DA, Wang CR, Schumacker PT, Licht JD, Perlman H, Bryce PJ, Chandel NS. Mitochondria are required for antigen-specific T cell activation through reactive oxygen species signaling. Immunity. 2013; 38:225–236. [PubMed: 23415911]
- 97. Gringhuis SI, Leow A, Papendrecht-van der Voort EAM, Remans PHJ, Breedveld FC, Verweij CL. Displacement of linker for activation of T cells from the plasma membrane due to redox balance alterations results in hyporesponsiveness of synovial fluid T lymphocytes in rheumatoid arthritis. J Immunol. 2000; 164:2170–2179. [PubMed: 10657671]
- Lahdenpohja N, Savinainen K, Hurme M. Pre-exposure to oxidative stress decreases the nuclear factor-κB–dependent transcription in T lymphocytes. J Immunol. 1998; 160:1354–1358. [PubMed: 9570554]
- 99. Kim HR, Lee A, Choi EJ, Hong MP, Kie JH, Lim W, Lee HK, Moon BI, Seoh JY. Reactive oxygen species prevent imiquimod-induced psoriatic dermatitis through enhancing regulatory T cell function. PLOS ONE. 2014; 9:e91146. [PubMed: 24608112]
- 100. Lin X, Zheng W, Liu J, Zhang Y, Qin H, Wu H, Xue B, Lu Y, Shen P. Oxidative stress in malignant melanoma enhances tumor necrosis factor-α secretion of tumor-associated macrophages that promote cancer cell invasion. Antioxid Redox Signal. 2013; 19:1337–1355. [PubMed: 23373752]
- 101. Sica A, Schioppa T, Mantovani A, Allavena P. Tumour-associated macrophages are a distinct M2 polarised population promoting tumour progression: Potential targets of anti-cancer therapy. Eur J Cancer. 2006; 42:717–727. [PubMed: 16520032]
- 102. Uemura H, Ishiguro H, Ishiguro Y, Hoshino K, Takahashi S, Kubota Y. Angiotensin II induces oxidative stress in prostate cancer. Mol Cancer Res. 2008; 6:250–258. [PubMed: 18314486]
- 103. Shirotake S, Miyajima A, Kosaka T, Tanaka N, Kikuchi E, Mikami S, Okada Y, Oya M. Regulation of monocyte chemoattractant protein-1 through angiotensin II type 1 receptor in prostate cancer. Am J Pathol. 2012; 180:1008–1016. [PubMed: 22226738]
- 104. Egami K, Murohara T, Shimada T, Sasaki K-I, Shintani S, Sugaya T, Ishii M, Akagi T, Ikeda H, Matsuishi T, Imaizumi T. Role of host angiotensin II type 1 receptor in tumor angiogenesis and growth. J Clin Invest. 2003; 112:67–75. [PubMed: 12840060]
- 105. Chehl N, Gong Q, Chipitsyna G, Aziz T, Yeo CJ, Arafat HA. Angiotensin II regulates the expression of monocyte chemoattractant protein-1 in pancreatic cancer cells. J Gastrointest Surg. 2009; 13:2189–2200. [PubMed: 19816747]
- 106. Shen XZ, Bernstein KE. The peptide network regulated by angiotensin converting enzyme (ACE) in hematopoiesis. Cell Cycle. 2011; 10:1363–1369. [PubMed: 21441775]
- 107. Shen XZ, Okwan-Duodu D, Blackwell WL, Ong FS, Janjulia T, Bernstein EA, Fuchs S, Alkan S, Bernstein KE. Myeloid expression of angiotensin-converting enzyme facilitates myeloid maturation and inhibits the development of myeloid-derived suppressor cells. Lab Invest. 2014; 94:536–544. [PubMed: 24614194]

- 108. Shen XZ, Li P, Weiss D, Fuchs S, Xiao HD, Adams JA, Williams IR, Capecchi MR, Taylor WR, Bernstein KE. Mice with enhanced macrophage angiotensin-converting enzyme are resistant to melanoma. Am J Pathol. 2007; 170:2122–2134. [PubMed: 17525278]
- 109. Shen XZ, Lukacher AE, Billet S, Williams IR, Bernstein KE. Expression of angiotensinconverting enzyme changes major histocompatibility complex class I peptide presentation by modifying C termini of peptide precursors. J Biol Chem. 2008; 283:9957–9965. [PubMed: 18252713]
- 110. Hofmann L, Forschner A, Loquai C, Goldinger SM, Zimmer L, Ugurel S, Schmidgen MI, Gutzmer R, Utikal JS, Göppner D, Hassel JC, Meier F, Tietze JK, Thomas I, Weishaupt C, Leverkus M, Wahl R, Dietrich U, Garbe C, Kirchberger MC, Eigentler T, Berking C, Gesierich A, Krackhardt AM, Schadendorf D, Schuler G, Dummer R, Heinzerling LM. Cutaneous, gastrointestinal, hepatic, endocrine, and renal side-effects of anti–PD-1 therapy. Eur J Cancer. 2016; 60:190–209. [PubMed: 27085692]
- 111. Postow MA, Chesney J, Pavlick AC, Robert C, Grossmann K, McDermott D, Linette GP, Meyer N, Giguere JK, Agarwala SS, Shaheen M, Ernstoff MS, Minor D, Salama AK, Taylor M, Ott PA, Rollin LM, Horak C, Gagnier P, Wolchok JD, Hodi FS. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. N Engl J Med. 2015; 372:2006–2017. [PubMed: 25891304]
- 112. Bouchlaka MN, Sckisel GD, Chen M, Mirsoian A, Zamora AE, Maverakis E, Wilkins DEC, Alderson KL, Hsiao HH, Weiss JM, Monjazeb AM, Hesdorffer C, Ferrucci L, Longo DL, Blazar BR, Wiltrout RH, Redelman D, Taub DD, Murphy WJ. Aging predisposes to acute inflammatory induced pathology after tumor immunotherapy. J Exp Med. 2013; 210:2223–2237. [PubMed: 24081947]
- 113. Mirsoian A, Bouchlaka MN, Sckisel GD, Chen M, Pai CCS, Maverakis E, Spencer RG, Fishbein KW, Siddiqui S, Monjazeb AM, Martin B, Maudsley S, Hesdorffer C, Ferrucci L, Longo DL, Blazar BR, Wiltrout RH, Taub DD, Murphy WJ. Adiposity induces lethal cytokine storm after systemic administration of stimulatory immunotherapy regimens in aged mice. J Exp Med. 2014; 211:2373–2383. [PubMed: 25366964]
- 114. Fliser D, Buchholz K, Haller H, EUropean Trial on Olmesartan and Pravastatin in Inflammation and Atherosclerosis (EUTOPIA) Investigators. Antiinflammatory effects of angiotensin II subtype 1 receptor blockade in hypertensive patients with microinflammation. Circulation. 2004; 110:1103–1107. [PubMed: 15313950]
- 115. Manabe S, Okura T, Watanabe S, Fukuoka T, Higaki J. Effects of angiotensin II receptor blockade with valsartan on pro-inflammatory cytokines in patients with essential hypertension. J Cardiovasc Pharmacol. 2005; 46:735–739. [PubMed: 16306795]
- 116. Pavlatou MG, Mastorakos G, Margeli A, Kouskouni E, Tentolouris N, Katsilambros N, Chrousos GP, Papassotiriou I. Angiotensin blockade in diabetic patients decreases insulin resistance-associated low-grade inflammation. Eur J Clin Invest. 2011; 41:652–658. [PubMed: 21175613]
- 117. Holmes MD, Hankinson SE, Feskanich D, Chen WY. Beta blockers and angiotensin-converting enzyme inhibitors' purported benefit on breast cancer survival may be explained by aspirin use. Breast Cancer Res Treat. 2013; 139:507–513. [PubMed: 23649190]
- 118. Jones PH, Christodoulos K, Dobbs N, Thavasu P, Balkwill F, Blann AD, Caine GJ, Kumar S, Kakkar AJ, Gompertz N, Talbot DC, Ganesan TS, Harris AL. Combination antiangiogenesis therapy with marimastat, captopril and fragmin in patients with advanced cancer. Br J Cancer. 2004; 91:30–36. [PubMed: 15162145]
- 119. Nakai Y, Isayama H, Ijichi H, Sasaki T, Kogure H, Yagioka H, Miyabayashi K, Mizuno S, Yamamoto K, Mouri D, Kawakubo K, Yamamoto N, Hirano K, Sasahira N, Tateishi K, Tada M, Koike K. Phase I trial of gemcitabine and candesartan combination therapy in normotensive patients with advanced pancreatic cancer: GECA1. Cancer Sci. 2012; 103:1489–1492. [PubMed: 22515232]
- 120. Nakai Y, Isayama H, Ijichi H, Sasaki T, Takahara N, Ito Y, Matsubara S, Uchino R, Yagioka H, Arizumi T, Hamada T, Miyabayashi K, Mizuno S, Yamamoto K, Kogure H, Yamamoto N, Hirano K, Sasahira N, Tateishi K, Tada M, Koike K. A multicenter phase II trial of gemcitabine and candesartan combination therapy in patients with advanced pancreatic cancer: GECA2. Invest New Drugs. 2013; 31:1294–1299. [PubMed: 23690239]

- 121. Ronquist G, Frithz G, Wang YH, Lindeborg T. Captopril may reduce biochemical (prostatespecific antigen) failure following radical prostatectomy for clinically localized prostate cancer. Scand J Urol Nephrol. 2009; 43:32–36. [PubMed: 18932051]
- 122. Sørensen GV, Ganz PA, Cole SW, Pedersen LA, Sørensen HT, Cronin-Fenton DP, Garne JP, Christiansen PM, Lash TL, Ahern TP. Use of β-blockers, angiotensin-converting enzyme inhibitors, angiotensin II receptor blockers, and risk of breast cancer recurrence: A Danish nationwide prospective cohort study. J Clin Oncol. 2013; 31:2265–2272. [PubMed: 23650417]
- 123. Tatokoro M, Fujii Y, Kawakami S, Saito K, Koga F, Matsuoka Y, Iimura Y, Masuda H, Kihara K. Phase-II trial of combination treatment of interferon-α, cimetidine, cyclooxygenase-2 inhibitor and renin-angiotensin-system inhibitor (I-CCA therapy) for advanced renal cell carcinoma. Cancer Sci. 2011; 102:137–143. [PubMed: 20973869]
- 124. Uemura H, Hasumi H, Kawahara T, Sugiura S, Miyoshi Y, Nakaigawa N, Teranishi J-I, Noguchi K, Ishiguro H, Kubota Y. Pilot study of angiotensin II receptor blocker in advanced hormone-refractory prostate cancer. Int J Clin Oncol. 2005; 10:405–410. [PubMed: 16369744]
- 125. Yoshiji H, Noguchi R, Ikenaka Y, Kaji K, Aihara Y, Yamazaki M, Yamao J, Toyohara M, Mitoro A, Sawai M, Yoshida M, Morioka C, Fujimoto M, Uemura M, Fukui H. Combination of branched-chain amino acids and angiotensin-converting enzyme inhibitor suppresses the cumulative recurrence of hepatocellular carcinoma: A randomized control trial. Oncol Rep. 2011; 26:1547–1553. [PubMed: 21874260]
- 126. Yoshiji H, Noguchi R, Toyohara M, Ikenaka Y, Kitade M, Kaji K, Yamazaki M, Yamao J, Mitoro A, Sawai M, Yoshida M, Fujimoto M, Tsujimoto T, Kawaratani H, Uemura M, Fukui H. Combination of vitamin K<sub>2</sub> and angiotensin-converting enzyme inhibitor ameliorates cumulative recurrence of hepatocellular carcinoma. J Hepatol. 2009; 51:315–321. [PubMed: 19501932]
- 127. Alashkham A, Paterson C, Windsor P, Struthers A, Rauchhaus P, Nabi G. The incidence and risk of biochemical recurrence following radical radiotherapy for prostate cancer in men on angiotensin-converting enzyme inhibitors (ACEIs) or angiotensin receptor blockers (ARBs). Clin Genitourin Cancer. 2016; 14:398–405. [PubMed: 27053500]
- 128. Aydiner A, Ciftci R, Sen F. Renin-Angiotensin system blockers may prolong survival of metastatic non–small cell lung cancer patients receiving erlotinib. Medicine. 2015; 94:e887. [PubMed: 26039117]
- 129. Babacan T, Balakan O, Kuzan TY, Sarici F, Koca E, Kertmen N, Petekkaya I, Altundag K. The effect of renin-angiotensin-system inhibition on survival and recurrence of N3+ breast cancer patients. J BUON. 2015; 20:50–56. [PubMed: 25778296]
- 130. Blute ML Jr, Rushmer TJ, Shi F, Fuller BJ, Abel EJ, Jarrard DF, Downs TM. Renin-angiotensin inhibitors decrease recurrence after transurethral resection of bladder tumor in patients with nonmuscle invasive bladder cancer. J Urol. 2015; 194:1214–1219. [PubMed: 26173101]
- 131. Botteri E, Munzone E, Rotmensz N, Cipolla C, De Giorgi V, Santillo B, Zanelotti A, Adamoli L, Colleoni M, Viale G, Goldhirsch A, Gandini S. Therapeutic effect of β-blockers in triple-negative breast cancer postmenopausal women. Breast Cancer Res Treat. 2013; 140:567–575. [PubMed: 23912960]
- 132. Boudreau DM, Yu O, Chubak J, Wirtz HS, Bowles EJA, Fujii M, Buist DSM. Comparative safety of cardiovascular medication use and breast cancer outcomes among women with early stage breast cancer. Breast Cancer Res Treat. 2014; 144:405–416. [PubMed: 24557337]
- 133. Chae YK, Brown EN, Lei X, Melhem-Bertrandt A, Giordano SH, Litton GN, Hortobagyi JK, Gonzalez-Angulo AM, Chavez-MacGregor M. Use of ACE inhibitors and angiotensin receptor blockers and primary breast cancer outcomes. J Cancer. 2013; 4:549–556. [PubMed: 23983819]
- 134. Chae YK, Valsecchi ME, Kim J, Bianchi AL, Khemasuwan D, Desai A, Tester W. Reduced risk of breast cancer recurrence in patients using ACE inhibitors, ARBs, and/or statins. Cancer Invest. 2011; 29:585–593. [PubMed: 21936625]
- 135. Ganz PA, Habel LA, Weltzien EK, Caan BJ, Cole SW. Examining the influence of beta blockers and ACE inhibitors on the risk for breast cancer recurrence: Results from the LACE cohort. Breast Cancer Res Treat. 2011; 129:549–556. [PubMed: 21479924]
- 136. Goldvaser H, Rizel S, Hendler D, Neiman V, Shepshelovich D, Shochat T, Sulkes A, Brenner B, Yerushalmi R. The association between angiotensin receptor blocker usage and breast cancer characteristics. Oncology. 2016; 91:217–223. [PubMed: 27544756]

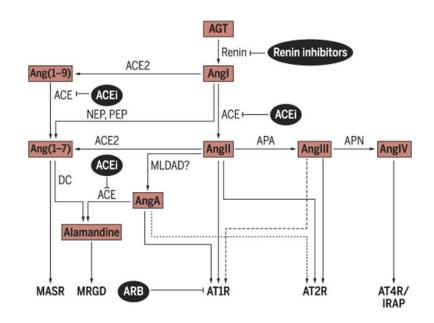
- 137. Izzedine H, Derosa L, Le Teuff G, Albiges L, Escudier B. Hypertension and angiotensin system inhibitors: Impact on outcome in sunitinib-treated patients for metastatic renal cell carcinoma. Ann Oncol. 2015; 26:1128–1133. [PubMed: 25795198]
- 138. Keizman D, Gottfried M, Ish-Shalom M, Maimon N, Peer A, Neumann A, Hammers H, Eisenberger MA, Sinibaldi V, Pili R, Hayat H, Kovel S, Sella A, Boursi B, Weitzen R, Mermershtain W, Rouvinov K, Berger R, Carducci MA. Active smoking may negatively affect response rate, progression-free survival, and overall survival of patients with metastatic renal cell carcinoma treated with sunitinib. Oncologist. 2014; 19:51–60. [PubMed: 24309979]
- 139. Keizman D, Huang P, Eisenberger MA, Pili R, Kim JJ, Antonarakis ES, Hammers H, Carducci MA. Angiotensin system inhibitors and outcome of sunitinib treatment in patients with metastatic renal cell carcinoma: A retrospective examination. Eur J Cancer. 2011; 47:1955–1961. [PubMed: 21600760]
- 140. McKay RR, Rodriguez GE, Lin X, Kaymakcalan MD, Hamnvik OPR, Sabbisetti VS, Bhatt RS, Simantov R, Choueiri TK. Angiotensin system inhibitors and survival outcomes in patients with metastatic renal cell carcinoma. Clin Cancer Res. 2015; 21:2471–2479. [PubMed: 25724518]
- 141. Melhem-Bertrandt A, Chavez-MacGregor M, Lei X, Brown EN, Lee RT, Meric-Bernstam F, Sood AK, Conzen SD, Hortobagyi GN, Gonzalez-Angulo AM. Beta-blocker use is associated with improved relapse-free survival in patients with triple-negative breast cancer. J Clin Oncol. 2011; 29:2645–2652. [PubMed: 21632501]
- 142. Menter AR, Carroll NM, Sakoda LC, Delate T, Hornbrook MC, Jain RK, Kushi LH, Quinn VP, Ritzwoller DP. Effect of angiotensin system inhibitors on survival in patients receiving chemotherapy for advanced non–small-cell lung cancer. Clin Lung Cancer. 2017; 18:189–197. [PubMed: 27637408]
- 143. Miao L, Chen W, Zhou L, Wan H, Gao B, Feng Y. Impact of angiotensin I-converting enzyme inhibitors and angiotensin II type-1 receptor blockers on survival of patients with NSCLC. Sci Rep. 2016; 6:21359. [PubMed: 26883083]
- 144. Miyajima A, Yazawa S, Kosaka T, Tanaka N, Shirotake S, Mizuno R, Kikuchi E, Oya M. Prognostic impact of renin–angiotensin system blockade on renal cell carcinoma after surgery. Ann Surg Oncol. 2015; 22:3751–3759. [PubMed: 25691280]
- 145. Sendur MAN, Aksoy S, Yaman S, Ozdemir NY, Zengin N, Altundag K. Efficacy of angiotensinreceptor blockers on demographic and clinico-pathological characteristics of breast cancer. Breast. 2012; 21:419–420. [PubMed: 22326438]
- 146. Sorich MJ, Kichenadasse G, Rowland A, Woodman RJ, Mangoni AA. Angiotensin system inhibitors and survival in patients with metastatic renal cell carcinoma treated with VEGFtargeted therapy: A pooled secondary analysis of clinical trials. Int J Cancer. 2016; 138:2293– 2299. [PubMed: 26685869]
- 147. Tanaka N, Miyajima A, Kikuchi E, Matsumoto K, Hagiwara M, Ide H, Kosaka T, Masuda T, Nakamura S, Oya M. Prognostic impact of renin-angiotensin system blockade in localised uppertract urothelial carcinoma. Br J Cancer. 2012; 106:290–296. [PubMed: 22187036]
- 148. Wang H, Liao Z, Zhuang Y, Liu Y, Levy LB, Xu T, Yusuf SW, Gomez DR. Incidental receipt of cardiac medications and survival outcomes among patients with stage III non–small-cell lung cancer after definitive radiotherapy. Clin Lung Cancer. 2015; 16:128–136. [PubMed: 25450873]
- 149. Wilop S, von Hobe S, Crysandt M, Esser A, Osieka R, Jost E. Impact of angiotensin I converting enzyme inhibitors and angiotensin II type 1 receptor blockers on survival in patients with advanced non–small-cell lung cancer undergoing first-line platinum-based chemotherapy. J Cancer Res Clin Oncol. 2009; 135:1429–1435. [PubMed: 19399518]
- 150. Yoshida T, Kinoshita H, Fukui K, Matsuzaki T, Yoshida K, Mishima T, Yanishi M, Komai Y, Sugi M, Inoue T, Murota T, Matsuda T. Prognostic impact of renin-angiotensin inhibitors in patients with bladder cancer undergoing radical cystectomy. Ann Surg Oncol. 2017; 24:823–831. [PubMed: 27730369]
- Yuge K, Miyajima A, Tanaka N, Shirotake S, Kosaka T, Kikuchi E, Oya M. Prognostic value of renin-angiotensin system blockade in non-muscle-invasive bladder cancer. Ann Surg Oncol. 2012; 19:3987–3993. [PubMed: 22872290]
- 152. Buchler T, Krejci M, Svobodnik A, Adam Z, Minarik J, Bacovsky J, Scudla V, Mayer J, Vorlicek J, Hajek R. Outcome of patients with multiple myeloma and hypertension treated with

angiotensin-I–converting enzyme inhibitors during high-dose chemotherapy. Hematol J. 2005; 5:559–564. [PubMed: 15692600]

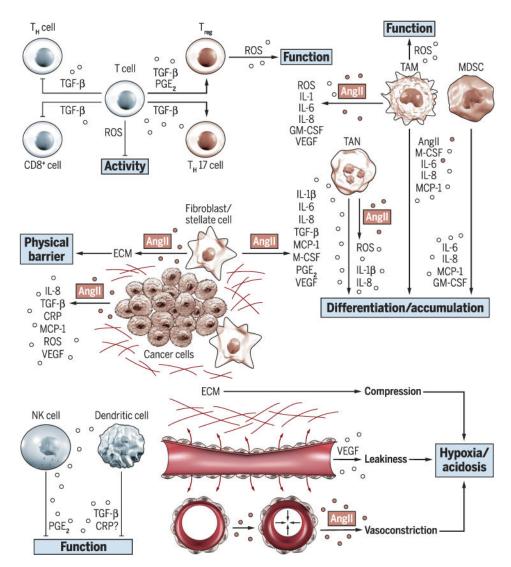
- 153. Cardwell CR, Mc Menamin UC, Hicks BM, Hughes C, Cantwell MM, Murray LJ. Drugs affecting the renin-angiotensin system and survival from cancer: A population based study of breast, colorectal and prostate cancer patient cohorts. BMC Med. 2014; 12:28. [PubMed: 24521426]
- 154. Carpentier AF, Ferrari D, Bailon O, Ursu R, Banissi C, Dubessy AL, Belin C, Levy C. Steroidsparing effects of angiotensin-II inhibitors in glioblastoma patients. Eur J Neurol. 2012; 19:1337– 1342. [PubMed: 22650322]
- 155. Chae YK, Dimou A, Pierce S, Kantarjian H, Andreeff M. The effect of calcium channel blockers on the outcome of acute myeloid leukemia. Leuk Lymphoma. 2014; 55:2822–2829. [PubMed: 24628293]
- 156. Chen YH, Huang CH, Lu HI, Chen CH, Huang WT, Hsieh MJ, Rau KM, Chang AYW, Lin WC, Li SH. Prognostic impact of renin-angiotensin system blockade in esophageal squamous cell carcinoma. J Renin Angiotensin Aldosterone Syst. 2015; 16:1185–1192. [PubMed: 24961505]
- 157. De Giorgi V, Gandini S, Grazzini M, Benemei S, Marchionni N, Geppetti P. Effect of β-blockers and other antihypertensive drugs on the risk of melanoma recurrence and death. Mayo Clin Proc. 2013; 88:1196–1203. [PubMed: 24182700]
- 158. Engineer DR, Burney BO, Hayes TG, Garcia JM. Exposure to ACEI/ARB and β-blockers is associated with improved survival and decreased tumor progression and hospitalizations in patients with advanced colon cancer. Transl Oncol. 2013; 6:539–545. [PubMed: 24151534]
- 159. Facciorusso A, Del Prete V, Crucinio N, Muscatiello N, Carr BI, Di Leo A, Barone M. Angiotensin receptor blockers improve survival outcomes after radiofrequency ablation in hepatocarcinoma patients. J Gastroenterol Hepatol. 2015; 30:1643–1650. [PubMed: 25974743]
- 160. He LR, Qiao W, Liao ZX, Komaki R, Ho L, Hofstetter WL, Lin SH. Impact of comorbidities and use of common medications on cancer and non-cancer specific survival in esophageal carcinoma. BMC Cancer. 2015; 15:1095. [PubMed: 25777421]
- 161. Heinzerling JH, Anthony T, Livingston EH, Huerta S. Predictors of distant metastasis and mortality in patients with stage II colorectal cancer. Am Surg. 2007; 73:230–238. [PubMed: 17375777]
- 162. Holmes S, Griffith EJ, Musto G, Minuk GY. Antihypertensive medications and survival in patients with cancer: A population-based retrospective cohort study. Cancer Epidemiol. 2013; 37:881–885. [PubMed: 24075077]
- 163. Januel E, Ursu R, Alkhafaji A, Marantidou A, Doridam J, Belin C, Levy-Piedbois C, Carpentier AF. Impact of renin-angiotensin system blockade on clinical outcome in glioblastoma. Eur J Neurol. 2015; 22:1304–1309. [PubMed: 26053493]
- 164. Kaibori M, Ishizaki M, Matsui K, Kitade H, Matsui Y, Kwon AH. Evaluation of metabolic factors on the prognosis of patients undergoing resection of hepatocellular carcinoma. J Gastroenterol Hepatol. 2011; 26:536–543. [PubMed: 21332549]
- 165. Kim ST, Park KH, Oh SC, Seo JH, Kim JS, Shin SW, Kim YH. How does inhibition of the reninangiotensin system affect the prognosis of advanced gastric cancer patients receiving platinumbased chemotherapy? Oncology. 2012; 83:354–360. [PubMed: 23052034]
- 166. Kourilsky A, Bertrand G, Ursu R, Doridam J, Barlog C, Faillot T, Mandonnet E, Belin C, Levy C, Carpentier AF. Impact of angiotensin-II receptor blockers on vasogenic edema in glioblastoma patients. J Neurol. 2016; 263:524–530. [PubMed: 26754004]
- 167. Morris ZS, Saha S, Magnuson WJ, Morris BA, Borkenhagen JF, Ching A, Hirose G, McMurry V, Francis DM, Harari PM, Chappell R, Tsuji S, Ritter MA. Increased tumor response to neoadjuvant therapy among rectal cancer patients taking angiotensin-converting enzyme inhibitors or angiotensin receptor blockers. Cancer. 2016; 122:2487–2495. [PubMed: 27203227]
- 168. Nakai Y, Isayama H, Ijichi H, Sasaki T, Sasahira N, Hirano K, Kogure H, Kawakubo K, Yagioka H, Yashima Y, Mizuno S, Yamamoto K, Arizumi T, Togawa O, Matsubara S, Tsujino T, Tateishi K, Tada M, Omata M, Koike K. Inhibition of renin-angiotensin system affects prognosis of advanced pancreatic cancer receiving gemcitabine. Br J Cancer. 2010; 103:1644–1648. [PubMed: 20978506]

- 169. Nakai Y, Isayama H, Sasaki T, Mizuno S, Sasahira N, Kogure H, Kawakubo K, Yamamoto N, Hirano K, Ijichi H, Tateishi K, Tada M, Koike K. Clinical outcomes of chemotherapy for diabetic and nondiabetic patients with pancreatic cancer: Better prognosis with statin use in diabetic patients. Pancreas. 2013; 42:202–208. [PubMed: 23000889]
- 170. Nakai Y, Isayama H, Sasaki T, Takahara N, Saito K, Ishigaki K, Hamada T, Mizuno S, Miyabayashi K, Yamamoto K, Mohri D, Kogure H, Yamamoto N, Ijichi H, Tateishi K, Tada M, Koike K. The inhibition of renin-angiotensin system in advanced pancreatic cancer: An exploratory analysis in 349 patients. J Cancer Res Clin Oncol. 2015; 141:933–939. [PubMed: 25398651]
- 171. Nakai Y, Isayama H, Sasaki T, Takahara N, Saito K, Takeda T, Umefune G, Saito T, Takagi K, Watanabe T, Hamada T, Uchino R, Mizuno S, Yamamoto K, Kogure H, Matsubara S, Yamamoto N, Ijichi H, Tateishi K, Tada M, Koike K. No survival benefit from the inhibition of reninangiotensin system in biliary tract cancer. Anticancer Res. 2016; 36:4965–4970. [PubMed: 27630357]
- 172. Osumi H, Matsusaka S, Wakatsuki T, Suenaga M, Shinozaki E, Mizunuma N. Angiotensin II type-1 receptor blockers enhance the effects of bevacizumab-based chemotherapy in metastatic colorectal cancer patients. Mol Clin Oncol. 2015; 3:1295–1300. [PubMed: 26807236]
- 173. Pinter M, Weinmann A, Wörns MA, Hucke F, Bota S, Marquardt JU, Duda DG, Jain RK, Galle PR, Trauner M, Peck-Radosavljevic M, Sieghart W. Use of inhibitors of the renin–angiotensin system is associated with longer survival in patients with hepatocellular carcinoma. United European Gastroenterol J. 2017
- 174. Tingle SJ, Moir JA, White SA. Role of anti-stromal polypharmacy in increasing survival after pancreaticoduodenectomy for pancreatic ductal adenocarcinoma. World J Gastrointest Pathophysiol. 2015; 6:235–242. [PubMed: 26600982]
- 175. Levin VA, Chan J, Datta M, Yee JL, Jain RK. Effect of angiotensin system inhibitors on survival in newly diagnosed glioma patients and recurrent glioblastoma patients receiving chemotherapy and/or bevacizumab. J Neurooncol. 2017; 134:325–330. [PubMed: 28631191]
- 176. Tanaka N, Miyajima A, Kosaka T, Shirotake S, Hasegawa M, Kikuchi E, Oya M. Cisdichlorodiammineplatinum upregulates angiotensin II type 1 receptors through reactive oxygen species generation and enhances VEGF production in bladder cancer. Mol Cancer Ther. 2010; 9:2982–2992. [PubMed: 20978160]
- 177. Tanaka N, Miyajima A, Kosaka T, Miyazaki Y, Shirotake S, Shirakawa H, Kikuchi E, Oya M. Acquired platinum resistance enhances tumour angiogenesis through angiotensin II type 1 receptor in bladder cancer. Br J Cancer. 2011; 105:1331–1337. [PubMed: 21970881]
- 178. Abd-Alhaseeb MM, Zaitone SA, Abou-El-Ela SH, Moustafa YM. Olmesartan potentiates the anti-angiogenic effect of sorafenib in mice bearing Ehrlich's ascites carcinoma: Role of angiotensin (1–7). PLOS ONE. 2014; 9:e85891. [PubMed: 24465768]
- 179. Mancuso MR, Davis R, Norberg SM, O'Brien S, Sennino B, Nakahara T, Yao VJ, Inai T, Brooks P, Freimark B, Shalinsky DR, Hu-Lowe DD, McDonald DM. Rapid vascular regrowth in tumors after reversal of VEGF inhibition. J Clin Invest. 2006; 116:2610–2621. [PubMed: 17016557]
- 180. Griffioen AW, Mans LA, de Graaf AMA, Nowak-Sliwinska P, de Hoog CLMM, de Jong TAM, Vyth-Dreese FA, van Beijnum JR, Bex A, Jonasch E. Rapid angiogenesis onset after discontinuation of sunitinib treatment of renal cell carcinoma patients. Clin Cancer Res. 2012; 18:3961–3971. [PubMed: 22573349]
- 181. Fox WD, Higgins B, Maiese KM, Drobnjak M, Cordon-Cardo C, Scher HI, Agus DB. Antibody to vascular endothelial growth factor slows growth of an androgen-independent xenograft model of prostate cancer. Clin Cancer Res. 2002; 8:3226–3231. [PubMed: 12374693]
- 182. Powles T, Blank C, Chowdhury S, Horenblas S, Peters J, Shamash J, Sarwar N, Boleti E, Sahdev A, O'Brien T, Berney D, Beltran L, Nathan P, Haanen J, Bex A. The outcome of patients treated with sunitinib prior to planned nephrectomy in metastatic clear cell renal cancer. Eur Urol. 2011; 60:448–454. [PubMed: 21612860]
- 183. Rini BI, Bellmunt J, Clancy J, Wang K, Niethammer AG, Hariharan S, Escudier B. Randomized phase III trial of temsirolimus and bevacizumab versus interferon alfa and bevacizumab in metastatic renal cell carcinoma: INTORACT trial. J Clin Oncol. 2014; 32:752–759. [PubMed: 24297945]

- 184. Haas NB, Manola J, Uzzo RG, Flaherty KT, Wood CG, Kane C, Jewett M, Dutcher JP, Atkins MB, Pins M, Wilding G, Cella D, Wagner L, Matin S, Kuzel TM, Sexton WJ, Wong YN, Choueiri TK, Pili R, Puzanov I, Kohli M, Stadler W, Carducci M, Coomes R, DiPaola RS. Adjuvant sunitinib or sorafenib for high-risk, non-metastatic renal-cell carcinoma (ECOG-ACRIN E2805): A double-blind, placebo-controlled, randomised, phase 3 trial. Lancet. 2016; 387:2008–2016. [PubMed: 26969090]
- 185. Ravaud A, Schmidinger M. Clinical biomarkers of response in advanced renal cell carcinoma. Ann Oncol. 2013; 24:2935–2942. [PubMed: 23925998]
- 186. Raimondi S, Botteri E, Munzone E, Cipolla C, Rotmensz N, DeCensi A, Gandini S. Use of betablockers, angiotensin-converting enzyme inhibitors and angiotensin receptor blockers and breast cancer survival: Systematic review and meta-analysis. Int J Cancer. 2016; 139:212–219. [PubMed: 26916107]
- 187. European Association For The Study Of The Liver. European Organisation For Research And Treatment Of Cancer, EASL-EORTC clinical practice guidelines: Management of hepatocellular carcinoma. J Hepatol. 2012; 56:908–943. [PubMed: 22424438]
- Fakih MG. Metastatic colorectal cancer: Current state and future directions. J Clin Oncol. 2015; 33:1809–1824. [PubMed: 25918280]
- 189. Escudier B, Porta C, Schmidinger M, Rioux-Leclercq N, Bex A, Khoo V, Gruenvald V, Horwich A, ESMO Guidelines Committee. Renal cell carcinoma: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. Ann Oncol. 2016; 27(suppl. 5):v58–v68. [PubMed: 27664262]
- 190. Faris JE, Blaszkowsky LS, McDermott S, Guimaraes AR, Szymonifka J, Huynh MA, Ferrone CR, Wargo JA, Allen JN, Dias LE, Kwak EL, Lillemoe KD, Thayer SP, Murphy JE, Zhu AX, Sahani DV, Wo JY, Clark JW, Fernandez-del Castillo C, Ryan DP, Hong TS. FOLFIRINOX in locally advanced pancreatic cancer: The Massachusetts General Hospital Cancer Center experience. Oncologist. 2013; 18:543–548. [PubMed: 23657686]
- 191. Mellon EA, Hoffe SE, Springett GM, Frakes JM, Strom TJ, Hodul PJ, Malafa MP, Chuong MD, Shridhar R. Long-term outcomes of induction chemotherapy and neoadjuvant stereotactic body radiotherapy for borderline resectable and locally advanced pancreatic adenocarcinoma. Acta Oncol. 2015; 54:979–985. [PubMed: 25734581]
- 192. Song T, Choi CH, Kim MK, Kim ML, Yun BS, Seong SJ. The effect of angiotensin system inhibitors (angiotensin-converting enzyme inhibitors or angiotensin receptor blockers) on cancer recurrence and survival: A meta-analysis. Eur J Cancer Prev. 2017; 26:78–85. [PubMed: 27158979]
- 193. Pinter M, Trauner M, Peck-Radosavljevic M, Sieghart W. Cancer and liver cirrhosis: Implications on prognosis and management. ESMO Open. 2016; 1:e000042. [PubMed: 27843598]
- 194. Yoshiji H, Kuriyama S, Yoshii J, Ikenaka Y, Noguchi R, Nakatani T, Tsujinoue H, Fukui H. Angiotensin-II type 1 receptor interaction is a major regulator for liver fibrosis development in rats. Hepatology. 2001; 34:745–750. [PubMed: 11584371]
- 195. Jonsson JR, Clouston AD, Ando Y, Kelemen LI, Horn MJ, Adamson MD, Purdie DM, Powell EE. Angiotensin-converting enzyme inhibition attenuates the progression of rat hepatic fibrosis. Gastroenterology. 2001; 121:148–155. [PubMed: 11438504]
- 196. Tandon P, Abraldes JG, Berzigotti A, Garcia-Pagan JC, Bosch J. Renin-angiotensin-aldosterone inhibitors in the reduction of portal pressure: A systematic review and meta-analysis. J Hepatol. 2010; 53:273–282. [PubMed: 20570385]
- 197. Jain RK. Lessons from multidisciplinary translational trials on anti-angiogenic therapy of cancer. Nat Rev Cancer. 2008; 8:309–316. [PubMed: 18337733]



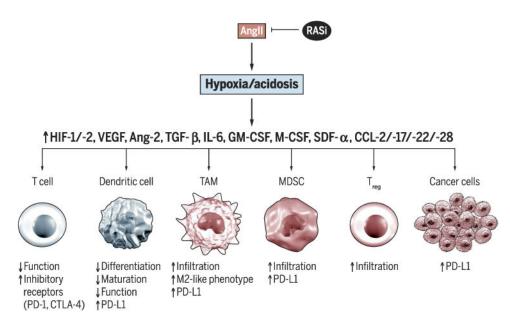
**Fig. 1. The RAS is a complex system whose bioactive peptides signal through different receptors** Angiotensinogen (AGT), generated and released into circulation by the liver, is hydrolyzed by renin, a product of the kidneys' juxtaglomerular cells, to form AngI. AngI is then hydrolyzed by ACE, predominantly expressed by endothelial cells in the vascular territory of the lungs, to form the biologically active AngII. In addition to AngII, other truncated bioactive peptides have been identified, such as AngIII, AngIV, Ang(1–7), Ang(1–9), AngA, and alamandine. AngII interacts with two seven-transmembrane receptors, AT1R and AT2R, both of which also mediate the effects of AngA. Ang(1–7) mainly acts via the MAS receptor (MASR), and alamandine binds and signals through MRGD (MAS-related G protein– coupled receptor D). IRAP (insulin-regulated membrane aminopeptidase; also known as AT4R) is a binding site for AngIV (1–7). APA, aminopeptidase A; APN, aminopeptidase N; DC, decarboxylase; MLDAD, mononuclear leukocyte-derived aspartate DC; NEP, neutral endopeptidase; PEP, prolyendopeptidase.



# Fig. 2. The AngII/AT1R axis regulates the tumor stroma and contributes to an immunosuppressive micro-environment

AngII/AT1R signaling can increase production and release of several proinflammatory cytokines in both tumor and stromal cells. Immunomodulatory cytokines regulate a myriad of immunosuppressive immune responses by modulating differentiation, recruitment, and function of both myeloid and lymphoid immune cell types (4, 43, 44). More precisely, these cytokines suppress the differentiation and function of immunostimulatory cell types [for example,  $T_H$  (T helper) and CD8<sup>+</sup> cells, NK cells, and dendritic cells] and activate recruitment and function of tumor-promoting cell types [such as  $T_{regs}$ ,  $T_H17$  cells, TANs, TAMs (tumor-associated macrophages), and MDSCs (myeloid-derived suppressor cells)]. Fibroblasts are a major source of cytokines and also play a key role in establishing a desmoplastic stroma by production and deposition of ECM. The dense tumor fibrosis represents a physical barrier to immune cell infiltration (45) and compresses blood vessels by increasing tissue stiffness and solid stress. The reduced tumor perfusion results in a hypoxic and acidic milieu, which further promotes immunosuppression (46–48). Vascular endothelial growth factor (VEGF)–induced vascular leakiness (48) and AngII-mediated

vasoconstriction (76, 77, 80) further impair tumor perfusion and aggravate hypoxia. GM-CSF, granulocyte-macrophage colony-stimulating factor. PGE<sub>2</sub>, prostaglandin E<sub>2</sub>.



#### Fig. 3. Tumor hypoxia and acidosis promote immunosuppression

AngII/AT1R-mediated effects on tumor vas-culature (shown in Fig. 2) can impair tumor perfusion and oxygenation, resulting in hypoxia and acidosis within the tumor stroma. The resulting up-regulation of various cytokines, growth factors, and transcription factors [including HIF (hypoxia-inducible factor), VEGF, and TGF- $\beta$ ] enhances an immunosuppressive microenvironment, characterized by impaired T and dendritic cell function, accumulation of immunosuppressive cell types (M2-like macrophages, MDSCs, and T<sub>regs</sub>), and increased expression of inhibitory immune checkpoint molecules such as PD-L1 in tumor and immune cell types (48–50, 68). Ang-2, angiopoietin-2; CCL, CC chemokine ligand; CTLA-4, cytotoxic T lymphocyte–associated protein 4; SDF, stromal cell–derived factor.