New Photosensitizers for Photodynamic Therapy

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Accessibility
New photosensitizers for photodynamic therapy

Heidi Abrahamse and Michael R. Hamblin

Abstract

Photodynamic therapy (PDT) was discovered more than 100 years ago, and has since become a well-studied therapy for cancer and various non-malignant diseases including infections. PDT uses photosensitizers (PSs, non-toxic dyes) that are activated by absorption of visible light to initially form the excited singlet state, followed by transition to the long-lived excited triplet state. This triplet state can undergo photochemical reactions in the presence of oxygen to form reactive oxygen species (including singlet oxygen) that can destroy cancer cells, pathogenic microbes and unwanted tissue. The dual-specificity of PDT relies on accumulation of the PS in diseased tissue and also on localized light delivery. Tetrapyrrole structures such as porphyrins, chlorins, bacteriochlorins and phthalocyanines with appropriate functionalization have been widely investigated in PDT, and several compounds have received clinical approval. Other molecular structures including the synthetic dyes classes as phenothiazinium, squaraine and BODIPY (boron-dipyrromethene), transition metal complexes, and natural products such as hypericin, riboflavin and curcumin have been investigated. Targeted PDT uses PSs conjugated to antibodies, peptides, proteins and other ligands with specific cellular receptors. Nanotechnology has made a significant contribution to PDT, giving rise to approaches such as nanoparticle delivery, fullerene-based PSs, titania photocatalysis, and the use of upconverting nanoparticles to increase light penetration into tissue. Future directions include photochemical internalization, genetically encoded protein PSs, theranostics, two-photon absorption PDT, and sonodynamic therapy using ultrasound.

Keywords

naturally occurring photosensitizers; photochemical mechanisms; photodynamic therapy; photosensitizers; synthetic dyes; tetrapyrroles

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INTRODUCTION

The concept of photodynamic therapy (PDT) dates back to 1900. A medical student, Oscar Raab, working in Munich, Germany [1], made an accidental discovery that micro-organisms such as paramecia that had been incubated with certain dyes could be killed when exposed to light, but not when they were kept in the dark. When it was subsequently discovered that oxygen in the air was also necessary for this light-mediated killing effect to occur, the term ‘photodynamic action’ was coined. It was not long after these discoveries that the first efforts were made to use this phenomenon as a cancer therapy, by painting dyes on to superficial skin tumours and then exposing them to light. However, the following two world wars and the impressive rise of the pharmaceutical industry in the 1950s and 1960s delayed the further exploration of PDT by over 60 years. The modern era of PDT started in the 1970s in the U.S.A., largely due to the efforts of Dr Thomas Dougherty working at Roswell Park Cancer Institute in Buffalo, NY. The first photosensitizer (PS) that was introduced by Dougherty and co-workers was a water-soluble mixture of porphyrins that was named ‘haematoporphyrin derivative’ (HpD), and a more purified preparation later became known as Photofrin. Although Photofrin is still the most frequently used PS throughout the world today, it has many acknowledged disadvantages including skin photosensitivity that can last for weeks or months and can be highly troubling for patients, and a relatively small absorbance peak at 630 nm making it somewhat inefficient in use, especially for bulky tumours where light penetration is problematic [2]. Since then medicinal chemists have attempted to synthesize and discover molecules that could act as improved PSs, and several hundred compounds have now been proposed as potentially useful to mediate PDT for tackling cancer, infections and many other diseases. In recent years PDT has returned to its earliest roots, and antimicrobial photodynamic inactivation (aPDI) has made something of a comeback. The structures of antimicrobial PSs have some features in common with anti-cancer PSs, but there are also major differences.

Photochemical mechanisms

The PS molecule is a singlet in its ground state because it has two electrons with opposite spins. Absorption of a photon of light with the appropriate quantum energy (wavelength) leads to the excitation of one electron into a higher-energy orbital (illustrated by the Jablonski diagram shown in Figure 1). This singlet excited-state PS is very unstable and loses its excess energy either as emission of light (fluorescence) or production heat (internal conversion). However, the excited singlet PS may undergo a process known as ‘intersystem crossing’ to form a more stable excited triplet state with parallel spins. The triplet-state PS molecule can decay back to the ground state (by emitting a phosphorescent photon) but this is a ‘forbidden process’ by the quantum selection rules, so the triplet state is much more stable than the singlet state having a lifetime of microseconds compared with only nanoseconds for the excited singlet. This long lifetime of the triplet state allows it sufficient time to transfer its energy by colliding with molecular oxygen (O₂), which is unique in being a molecular triplet in its ground state. This energy-transfer step leads to the formation of singlet oxygen (¹O₂) (and ground-state PS), and the reaction is referred to as a Type II photochemical process [3]. A Type I photochemical process can also occur whereby the excited-state PS undergoes electron transfer reactions that eventually forms reactive oxygen.
species (ROS). This mechanism may involve either acquisition or donation of an electron to form the radical cation or radical anion. The radical anion can react with oxygen to produce the superoxide radical anion ($\text{O}_2^{\cdot-}$). Dismutation or one-electron reduction of $\text{O}_2^{\cdot-}$ gives hydrogen peroxide ($\text{H}_2\text{O}_2$), which in turn can undergo another one-electron reduction to form the powerful oxidant hydroxyl radicals (HO$\cdot$). ROS generation via Type II chemistry is mechanistically much simpler than via Type I, and most PSs used for anti-cancer PDT are believed to operate via the Type II rather than the Type I mechanism.

**Properties of ideal PSs**

Most of the PSs used in cancer therapy are based on the tetrapyrrole backbone, a structure similar to that contained in the protoporphyrin prosthetic group contained in haemoglobin. Depending on the precise structure, effective PSs can be synthesized with absorbance bands between 600 and 800 nm. Since the penetration of light into tissue increases with wavelength, agents with strong absorbance in the deep-red spectral region such as chlorins, bacteriochlorins and phthalocyanines tend to make much more efficient PSs although many other factors are also important.

A PS should ideally be a single pure compound to allow manufacturing under good manufacturing practice (GMP) conditions with quality control and low manufacturing costs, and leading to better stability in storage. It should have a strong absorption peak in the red to near-infrared spectral region (between 650 and 800 nm) because absorption of single photons with wavelengths longer than 800 nm does not provide enough energy to excite oxygen to its singlet state. PSs should possess a substantial triplet quantum yield leading to good production of ROS upon irradiation. It should have no dark toxicity and relatively rapid clearance from normal tissues, thereby minimizing the side effects of phototoxicity [4]. Although it used to be considered desirable for the interval between drug administration and irradiation (drug–light interval, DLI) to be as long as possible (up to 4 days), so that the PS was given sufficient time to clear from normal tissues, while remaining concentrated in tumours, many reports now suggest that the tumour response may be substantially better when light is delivered at a much shorter DLI (minutes or hours) when most of the PS is still present in the blood vessels, thus producing marked vascular damage [5]. Some reports have suggested that a pronounced inflammatory response resulting from necrotic cell death after PDT is important because it aids the immune-stimulating function of PDT, whereas other reports have suggested that PDT regimens that produce more apoptosis and less necrosis and inflammation are suitable for applications such as PDT of brain tumours where swelling is undesirable. Previous findings, however, show that certain PDT-induced apoptotic cell death mechanisms are also highly immunogenic and can stimulate antitumour immunity [6]. The light-mediated destruction of the PS (known as photobleaching) was thought to be undesirable, but some reports now suggest that this phenomenon may make light dosimetry during PDT less critical, as over-treatment is avoided when the remaining PS is destroyed during the illumination [7].

The discovery that 5-aminolaevulinic acid (ALA) could function as a biosynthetic precursor of the PS protoporphyrin IX [8] has led to many applications in which ALA or ALA-esters can be topically applied to tissues, or even administered orally. These ALA derivatives are
considered to be ‘pro-drugs’, needing to be metabolically converted into protoporphyrin IX in order to become active PSs. Successes in treatment of skin cancer and other dermatologic indications have been reported.

Tumour-targeting in PDT

The most effective PSs tend to be relatively hydrophobic compounds that rapidly diffuse into tumour cells and localize in intracellular membrane structures such as mitochondria and endoplasmic reticulum (ER). More polar compounds tend to be taken up by the active process of adsorptive or fluid phase endocytosis, and this process is slower than passive diffusion, necessitating a longer DLI. Many hypotheses have been proposed to account for the tumour-localizing properties that have long been observed in PDT [9]. These explanations include the occurrence of leaky and tortuous tumour blood vessels that are typical of the neovascularization process occurring in tumour angiogenesis, and together with the absence of lymphatic drainage in tumours are collectively known as the ‘enhanced permeability and retention (EPR) effect’ [10]. Some of the most effective PS compounds have been found preferentially to low-density lipoprotein (LDL) among various serum proteins, and it has been proposed that over-expressed LDL receptors that are sometimes found on tumour cells could be important in tumour localization [11].

Mechanisms of PDT-mediated cytotoxicity

The lifetime of singlet oxygen (\(^{1}\text{O}_2\)) is very short (~10–320 ns), limiting its diffusion to only approximately 10–55 nm in cells [12]. Thus, photodynamic damage is likely to occur very close to the intracellular location of the PS [13]. PDT can kill cells via the three main morphologies of cell death: apoptotic, necrotic and autophagy-associated cell death (Figure 2). It is thought that the subcellular localization of the PS in different organelles (mitochondria, lysosomes, endoplasmic reticulum, plasma membrane, etc.) plays a major role in the type of cell death mechanism that dominates, but other factors such as the overall PDT dose (PS concentration × light fluence) and DLI also play a role. Overall it is accepted that apoptosis is the principal modality of cell death when cells are treated with PDT in vitro.

Antimicrobial photoinactivation

It is somewhat ironic that the origins of PDT over 100 years ago were in the killing of micro-organisms, but throughout the period from the 1970s to 2010, cancer was the overwhelming most popular target disease in PDT research. Now with the inexorable rise of multi-drug-resistance among pathogenic microbes, a PDI has made something of a comeback.

The ideal PS structure is very different between anti-cancer drugs and antimicrobial drugs. Anti-cancer PSs tend to be lipophilic with little or no overall charge (either positive or negative). Antimicrobial PSs, on the other hand, should have pronounced cationic charges, and in many cases the more charges the better especially for targeting Gram-negative bacteria. Anti-cancer PSs are usually expected to have long wavelength (farred/near-infrared) absorption bands for good tissue penetration of the exciting light, whereas for
antimicrobial PSs, this property is much less important as infections that will be treated by PDT tend to be rather superficial in nature.

**TETRAPYRROLE STRUCTURES**

Tetrapyrrrole structures make up the largest group of PSs that have been employed for anti-cancer applications. Tetrapyrrrole backbones occur naturally in several important biomolecules such as haem, chlorophyll and bacteriochlorophyll. In fact, tetrapyrrroles have been termed the ‘pigments of life’ [14]. As the double-bonds are successively reduced when moving from backbones that are porphyrins to chlorins to bacteriochlorins the Q-band is substantially red-shifted and the height of the band also increases (Figure 3). Phthalocyanines also have a large band in the 670 nm region. Broadly speaking, tetrapyrrrole PSs (with the exception of bacteriochlorins) tend to produce predominantly Type II singlet oxygen, compared with the Type I ROS (such as hydroxyl radicals) that are often produced by PSs with other structures. The number of tetrapyrrrole compounds used as PSs in PDT is too high to even list them all, but in Table 1 we have tried to list the most important compounds and the newest examples.

**Porphyrins**

As mentioned above, HpD and Photofrin were the first PSs to receive regulatory approval, and these and similar preparations are still in widespread use around the world [4]. ALA-induced protoporphyrin IX is, of course, a porphyrin and this methodology is widely used around the world, mainly by dermatologists [15]. There have been some new porphyrin compounds proposed as PSs, but these have been tested mainly in the PDI field [16]. Although porphyrins are most efficiently activated at the Soret band (~400 nm), the presence of several Q-bands extending as far as the 630 nm region means that red light is most often used to activate porphyrins in vivo, although blue, green or white light has been used for more superficial applications.

**Chlorins**

Chlorins include several of the most important clinical PSs, namely m-tetrahydroxyphenylchlorin (Temoporfin or Foscan) [17], benzoporphyrin derivative (Verteporfin) [18] and Radachlorin (now Bremachlorin) [19]. Chlorin(e6) is derived from naturally occurring chlorophyll and it has been used on its own (formulated either as the trisodium salt known as photodithazine [20] or dissolved in polyvinylpyrrolidone [21]). The monoaspartyl derivative (laserphyrin, taloporphin sodium or LS11) [22] has been used in Japan. Other PSs that have advanced to clinical trials can also be classified as chlorins, including the pyrophaeophorbide derivative HPPH [23] and tin(II) etiopurpurin [24]. Red light between 650 and 700 nm is used to activate chlorins depending on the exact structure.

**Bacteriochlorins**

The bacteriochlorin group of compounds also contains clinically important PSs. The lead-containing bacteriophaeophorbide derivative known as TOOKAD [25] and its newer watersoluble derivative known as TOOKAD Soluble have been tested in clinical trials for prostate cancer [26]. The new bacteriochlorin derivative known as LUZ11 [27] recently entered
clinical trials for head and neck cancer in Portugal (Photodynamic Therapy With LUZ11 in Advanced Head and Neck Cancer, Clinical Trials.Gov NCT02070432). Other bacteriochlorin structures have been tested as PSs for both anti-cancer [28] and antimicrobial applications [29]. Near-infrared light between 700 and 800 nm is used to activate bacteriochlorins and this was shown to be particularly effective against pigmented tumours such as malignant melanoma [30].

**Phthalocyanines**

Phthalocyanines were some of the earliest compounds studied in the 1980s and 1990s. It should be noted that phthalocyanines are also synthetic dyes and could have also been included in the next section. There used to be a mixture of sulfonated chloroaluminium phthalocyanines known as CASPs that received considerable attention in the PDT field [31]. Unmodified zinc-phthalocyanine was tested in a liposomal formulation in both animal models and in clinical trials [32]. The silicon-substituted phthalocyanine PC4 has also been tested *in vivo* [33] and in clinical trials [34]. Cationic phthalocyanines such as RLP068 have been studied for antimicrobial applications, both *in vivo* [35] and in clinical trial for infected diabetic foot ulcers [36]. Phthalocyanines are activated by far-red light in the 670 nm range.

**SYNTHETIC DYES**

Structures that can be classified as synthetic dyes account for a sizeable fraction of the molecules that are being studied for a PDI and a selection are listed in Table 2.

**Phenothiazinium salts**

The two most popular phenothiazinium dyes (Methylene Blue [37] and Toluidine Blue [38]) have been widely studies for both antimicrobial applications [39] and less often for anti-cancer [40]. There are some new phenothiazinium dye structures that have been recently prepared mostly for antimicrobial use. PP904 was clinically tested for infected non-healing leg ulcers [41]. The benzophenothiazinium dye EtNBS has been studied for both antimicrobial [42] and anti-cancer [43] PDT.

**Rose Bengal**

Rose Bengal is a member of the xanthene class of fluorescent dyes that includes fluorescein (the most popular synthetic fluorophore). The introduction of heavy atoms into the rings, such as the halogen atoms bromine and iodine, increases the triplet yield of the molecule by facilitating intersystem crossing. Rose Bengal has a long history as a photoactive dye and has been explored for antimicrobial applications [44], tissue bonding applications [45] and anti-cancer applications [46].

**Squaraines**

The squaraine dye structure has a delocalized system of molecular orbitals providing good absorption in the visible range. The four-membered carbon ring is usually stabilized against nucleophilic attack by surrounding it with a rotaxane structure [47,48]. In a similar manner to xanthenes (see above), the introduction of iodine substituents into the ring increases the triplet yield by the heavy atom effect.
BODIPY dyes

The BODIPY (boron-dipyrrromethene) dye structure usually contains a BF$_2$ bridging unit, and constitutes a popular class of fluorophores [49]. Addition of heavy halogen atoms in the pyrrole rings increases the triplet yield and allows the molecules to function as PSs [50].

Phenalenones

The parent compound phenalenone was often used as a reference standard for generation of $^1$O$_2$ [51]. Maisch and co-workers have recently synthesized a quaternized cationic derivative of phenalenone as an antimicrobial PS [52,53].

Transition metal compounds

Transition metal co-ordination compounds are a relatively new class of PS. Ruthenium(II) polypyridyl complexes are perhaps the most studied [54], although other ruthenium ligands [55], rhodium [56] and cyclometalated iridium [57] complexes have also been studied. Rhodium compounds are often in the form of Rh(II)–Rh(II) bridged dimer compounds [58]. There is also evidence that some luminescent platinum(II) and gold(III) compounds can act as PSs [59].

NATURAL PRODUCTS

The idea to use a naturally occurring compound as a PS inherently contains possible contradictions. If plants have evolved over millennia to grow in sunlight, then surely they cannot contain a highly active PS molecule or they would have burned up in the sun? Nevertheless, there are several natural product isolates that have been extensively explored as PSs and some of these are listed in Table 3.

Hypericin

Hypericin is a perylenequinone isolated from St John’s wort, a long-known medicinal plant [60]. Hypericin is a hydrophobic molecule that requires formulation in a drug-delivery vehicle (liposomes, micelles, nanoparticles) [61]. The peak absorption band of hypericin is 600 nm, so orange light is used. Hypericin has been found to efficiently localize in the endoplasmic reticulum and to cause ER stress after light application, that can produce ‘damage associated molecular patterns’ that efficiently activate the immune system [62].

Hypocrellin

Hypocrellins A and B are another group of perylenequinone pigments which have been isolated from the parasitic fungi Hypocrella bambuase sacc and Shiraia bambusicola P. Heen found in China and other parts of Asia including Sri Lanka [63]. These compounds have been used in PDT and can also benefit from encapsulation in liposomes [64].

Riboflavin

Riboflavin (vitamin B$_2$) has been explored as an antimicrobial PS. It has been tested for antimicrobial [65] and blood product sterilization [66] applications, and also as a photoactivated cross-linker for corneal stiffening [67]. Riboflavin has two peaks in the UVA
(360 nm) and blue (440 nm) regions. Maisch et al. [68] have synthesized a cationic version of riboflavin designed as an antimicrobial PS.

**Curcumin**

Curcumin is a relative newcomer in the PDT field, although it has been known as a spice and medicinal compound for centuries [69]. Again curcumin is a very hydrophobic molecule and requires some kind of formulation vehicle to allow it to be used as a PS [70]. Curcumin is activated by blue light [71]. Curcumin has found most applications as an antimicrobial PS in dentistry to eradicate oral pathogens [72].

**TARGETED PDT**

There have been a large number of targeting studies in which PSs are covalently attached to various molecules that have some affinity for neoplasia or to receptors expressed on specific tumours [73]. The intention is to rely on the ability of the targeting vehicle to control localization so that the PS can be chosen based on its photochemical properties rather than on its tumour targeting properties, which are often unimpressive. These targeting vehicles include monoclonal antibodies [74], antibody fragments [75], peptides [76], proteins such as transferrin [77], epidermal growth factor [77] and insulin [78], LDL [79], various carbohydrates [80], somatostatin [81], folic acid [82] and many others. Table 4 lists some of these targeting ligand–PS conjugates.

**NANOTECHNOLOGY**

The nanotechnology revolution has had a major impact on PDT as it also has had in other areas of biomedicine [83]. The fact that most effective PSs tend to be insoluble, hydrophobic molecules with a high propensity to aggregate means that encapsulation in nano-drug carriers may make a big difference to their performance [84]. Moreover many other nanostructures such as plasmonic gold nanoparticles, mesoporous silica nanoparticles, carbon nanotubes, graphene and upconversion nanoparticles have found uses in PDT [85]. There is another group of nanostructures where the actual nanoparticle itself acts as the PS absorbing light and producing ROS, as in the case of fullerenes [86], titanium dioxide [84] and some types of quantum dots [87].

**Nanoparticle PS delivery**

There has been an astonishing variety of nanoparticles used to solubilize, encapsulate and deliver PSs to both tumours [88] and microbial cells [89]. As mentioned above, the planar conjugated structures of PSs that are needed to absorb the light means the molecules tend to be hydrophobic and prone to aggregation. Liposomes, micelles, nanoemulsions can all be constructed out of lipids or amphiphilic polymers that self-assemble into delivery vehicles for PSs [90]. These nanovehicles have many advantages, the most important of which are providing a big increase in photochemical efficiency, and the ability to localize in tumours after IV injection due to the EPR effect [91].
Fullerenes

Fullerenes are closed-cage all-carbon nanostructures composed of sp2 hybridized carbon atoms (C_{60}, C_{70}, C_{84}, etc.) and with a large molar absorption coefficient and high triplet yields. In organic solvents or hydrophobic environments, fullerenes are very efficient in producing photoexcited $^{1}\text{O}_2$, whereas in aqueous environments, fullerenes switch the photochemical mechanism to Type I producing HO$^\bullet$ [92]. Pristine fullerenes are insoluble in water, but the cage can be readily functionalized with polar groups such as carboxylic acids and quaternary amino groups that improve solubility and biological compatibility. Fullerenes have been used to mediate PDT of cancer cells [93] and microbial cells [94] \textit{in vitro}, to treat tumours [95] and infections [96] \textit{in vivo}, and there is even one case report of a clinical application.

Titanium dioxide

It has long been known that titanium dioxide (TiO$_2$) or titania acts as a large band gap semiconductor. When excited with UVA light, an electron is excited from the valence band into the conductance band leaving behind a positively charged hole. The electron can produce superoxide from oxygen, whereas the hole can produce hydroxyl radicals from water. These ROS have been used in the process of photocatalysis which is used to kill micro-organisms and degrade organic pollutants [97]. In recent times TiO$_2$ nanoparticles have been used as PDT agents often as composites or hybrids [98–100].

Quantum dots

Although many researchers have prepared conjugates between quantum dots and various different PSs to carry out PDT [101–103], a recent study showed that graphene quantum dots could mediate PDT on their own without any added PS [87].

Upconversion nanoparticles

Upconversion is a process in which the sequential absorption of two or more photons leads to the emission of light at shorter wavelength than the excitation wavelength. The mechanism relies on absorption of two consecutive photons, where second near-infrared photon is absorbed by a ‘virtual excited state’ to give the actual excited state that can then emit a much shorter wavelength photon. Lanthanide-doped nanoparticles emerged in the late 1990s and the optical transitions from compounds containing yttrium, ytterbium and erbium were found to be suitable for photon upconversion. In contrast with two-photon absorption (2PA) which needs a very high peak power density provided by femtosecond pulsed lasers, photon upconversion can operate with reasonable efficiency using a CW laser (often at 980 nm) [104]. Near-infrared wavelengths are preferred for their greater tissue penetration, and the shorter wavelengths emitted are able to excite more different PSs, that have been attached to the lanthanide-doped nanoparticles. Recently a NaYbF$_4$:Nd@NaGdF$_4$:Yb/Er@NaGdF$_4$ core–shell–shell nanoparticle loaded with chlorin(e6) and folic acid for targeting was reported that could be excited with 808 nm laser [105].

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FUTURE DIRECTIONS

Photochemical internalization (PCI)

The PCI is a new technique in which a highly amphiphilic PS such as aluminium phthalocyanine adjacent disulfonate or Amphinex (meso-tetraphenylchlorin adjacent disulfonate) [106] is administered together with a cytotoxic molecule such as bleomycin [107] or a ribosome-inactivating protein [108]. The cytotoxic molecule is taken up by endocytosis into endosomes which also have the PS in their membranes. When light is delivered, the endosomes are broken open, releasing the cytotoxic molecule into the cytoplasm where its toxicity is much higher. This approach has advanced through in vitro studies [109], to in vivo [110], and into clinical trials for head and neck cancer [111]. PCI can also be used for non-viral delivery of DNA and other nucleic acid-based therapeutics as a form of gene therapy [112].

Genetically encoded proteins

It was previously discovered that some modified fluorescent proteins based on the fundamental chromophore structure present in green fluorescent protein (GFP) can not only emit fluorescence, but also produce ROS upon excitation with light [113]. The most well known of these photodynamically active fluorescent proteins is called KillerRed [114]. The intracellular location of the expressed protein can be controlled by additional genetic elements attached to the DNA sequence for the KillerRed molecule. For instance KillerRed has been targeted to cell membranes [115], to lysosomes [116], to mitochondria [117] and to the nucleus [118]. KillerRed has been used as an optogenetic tool to allow the light-mediated inactivation of specific groups of neurons in Caenorhabditis elegans [119] in transgenic zebrafish [120], in Xenopus laevis embryos [121] and in the mouse retina [122]. Another genetically encoded PS is the flavoprotein ‘miniSOG’ [123].

Theranostics

The term ‘theranostics’ was coined as a combination of diagnostics and therapeutics [124], but should really combine more elements than simply an imaging agent that allows a tumour to be visualized (often by fluorescence), and a therapeutic agent (often photodynamic) that allows a tumour to be destroyed [125]. Theranostic agents could in addition be designed to act as ‘smart drug-delivery vehicles’ that could be activated by a change in the environment of the tumour or the infection such as lower pH [126], redox, enzyme activation [127], elevated temperature or magnetic fields [128]. Furthermore theranostic agents could also be designed to incorporate a method to monitor the effectiveness of treatment, possibly in real-time, and in the days following treatment. Porphysomes (self-assembled nanostructures from lipid-conjugated porphyrins) are an interesting example of a theranostic PDT agent [129].

Two-photon excitation

Another way to use long wavelength near-infrared light that penetrates deeply into tissue to excite PSs with short wavelength absorptions (in addition to upconversion nanoparticles) is 2-PA. Here two separate near-infrared photons have to arrive at the PS molecule virtually simultaneously, and if they are absorbed at the same time they will be equivalent to a single
photon of half the wavelength [130]. However, the need for simultaneous absorption of both photons means that an extremely high peak power density (in the order of GW/cm²) is required. Moreover, the efficiency of 2-PA PDT depends strongly on what is known as the 2-PA cross-section of the PS molecule, which is measured in Goeppert-Mayer units (1 GM = 10⁻⁵⁰ cm⁴·s·molecule⁻¹·photon⁻¹). Most regular PSs have a 2-PA cross-section between 1 and 100, but specially designed PSs can have values as high as 33 000 [131]. 2-PA PDT has been tested in vivo when the femtosecond laser beam was delivered through the whole body of the mouse to reach the tumour on the other side [132].

Sonodynamic therapy (SDT)
The ability to activate some kinds of PS molecule with ultrasound energy instead of light has been known for some years [133]. The ultrasound employed is usually between 1 and 2 MHz delivered at power densities between 0.5 and 10 W/cm². In fact, although many sonosensitizers are the same types of molecule as those used as PSs, this is not always the case, and some chemotherapeutic drugs have been shown to be potentiated by ultrasound [134]. The main proposed mechanisms of actions are: (a) generation of flashes of light (sonoluminescence) from collapse of cavitation bubbles that activates the PS to produce singlet oxygen; (b) generation of free radicals which initiate chain peroxidation of membrane lipids via peroxyl and/or alkoxyl radicals; (c) physical destabilization of the cell membrane by the sonosensitizer thereby rendering the cell more susceptible to ultrasound induced shear forces; (d) ultrasound-enhanced drug transport across the cell membrane (sonoporation). The main advantage claimed for SDT is much deeper tissue penetration by ultrasound compared with light. However, it must be mentioned that SDT is still considered controversial due to its use in alternative medicine clinics.

CONCLUSIONS
PDT is a highly multidisciplinary field that involves chemists, physicists, biologists, engineers and physicians. Chemists, of course, are constantly seeking to design, synthesize, purify and characterize new compounds that can be used as PSs. Many significant advances have been made in PS design during the last 20 years, and second-, third- and even fourth-generation PSs have been described. Drug delivery and formulation requirements have increasingly been recognized as being of high importance, as many old-style PSs suffered from sub-optimal formulation. The advent of the nanotechnology revolution has had a big impact on PDT, and is expected to continue to influence the field. New tumour-targeting modalities are being reported frequently, and it is expected that these will be utilized to deliver PSs, as the need for activation by spatially confined light application provides yet another level of selectivity. Many commentators agree that the antimicrobial applications of PDT are now growing faster than the traditional anti-cancer applications. Moreover, it is also possible that other medical applications will emerge, as happened in the past for choroidal neovascularization.

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Abbreviations

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<tr>
<th>Abbreviation</th>
<th>Description</th>
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<tr>
<td>ALA</td>
<td>5-aminolaevulinic acid</td>
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<td>aPDI</td>
<td>antimicrobial photodynamic inactivation</td>
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<td>BODIPY</td>
<td>boron-dipyrromethene</td>
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<tr>
<td>DLI</td>
<td>drug–light interval</td>
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<td>EPR</td>
<td>enhanced permeability and retention</td>
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<td>ER</td>
<td>endoplasmic reticulum</td>
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<td>HpD</td>
<td>haematoporphyrin derivative</td>
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<td>LDL</td>
<td>low-density lipoprotein</td>
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<td>2PA</td>
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<td>reactive oxygen species</td>
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<td>SDT</td>
<td>sonodynamic therapy</td>
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Figure 1. Jablonski diagram
When light ($hv$) is absorbed by the PS, the electron moves from a non-excited low-energy singlet state into a high-energy singlet state. This excited state can lose energy by emitting a photon (fluorescence) or by internal conversion (non-radiative decay). The process known as intersystem crossing involves flipping of the spin of the high-energy electron, leading to a long-lived excited triplet state. In the presence of molecular oxygen, superoxide and hydroxyl radicals are formed in Type I reactions and singlet oxygen in a Type II reaction. These ROS can damage most types of biomolecules (amino acids, lipids, nucleic acids).
**Figure 2. Cell death mechanisms**
The subcellular localization of the PS in different organelles (mitochondria, lysosomes, endoplasmic reticulum, plasma membrane, etc.) plays a major role in the type of cell death mechanism that dominates, but other factors such as the overall PDT dose (PS concentration × light fluence) and DLI also play a role.
Figure 3. Structure and absorption spectra of tetrapyrrole photosensitizers
Tetrapyrrole absorption spectrum showing porphyrins, chlorins and bacteriochlorins.
Table 1

PSs based on tetrapyrrole backbone

<table>
<thead>
<tr>
<th>Class</th>
<th>Name</th>
<th>Structure</th>
<th>$\lambda_{\text{max}}$</th>
<th>Application</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Porphyrin</td>
<td>Photofrin (NB actual structure of Photofrin is a complex mixture of ester- and ether-linked dimers and oligomers)</td>
<td><img src="image1" alt="Structure of Photofrin" /></td>
<td>630 nm</td>
<td>Cancer, in <em>vitro</em>, in <em>vivo</em>, clinical</td>
<td>[135]</td>
</tr>
<tr>
<td>Porphyrin</td>
<td>ALA-induced protoporphyrin IX</td>
<td><img src="image2" alt="Structure of ALA-induced protoporphyrin IX" /></td>
<td>635 nm</td>
<td>Cancer, in <em>vitro</em>, in <em>vivo</em>, clinical</td>
<td>[136]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
</tr>
<tr>
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</tr>
<tr>
<td>Porphyrin</td>
<td>5,10,15,20-Tetrakis(1-methylpyridinium-4-yl) porphyrin tosylate</td>
<td><img src="image1.png" alt="Structure 1" /></td>
<td></td>
<td>Cancer, antimicrobial, in vitro, in vivo</td>
<td>[137]</td>
</tr>
<tr>
<td>Porphyrin</td>
<td>XF70</td>
<td><img src="image2.png" alt="Structure 2" /></td>
<td></td>
<td>Antimicrobial, in vitro, in vivo</td>
<td>[138]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
</tr>
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<td>----------------------------------------------------------------------</td>
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</tr>
<tr>
<td>Chlorin</td>
<td>Foscan, m-tetrahydroxyphenylchlorin</td>
<td><img src="image" alt="Foscan" /></td>
<td>652 nm</td>
<td>Cancer, in vivo, in vivo, clinical</td>
<td>[139]</td>
</tr>
<tr>
<td>Chlorin</td>
<td>Verteporfin, benzoporphyrin derivative mono acid ring A</td>
<td><img src="image" alt="Verteporfin" /></td>
<td>690 nm</td>
<td>Cancer, ophthalmology, in vivo, in vivo, clinical</td>
<td>[18]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
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</tr>
<tr>
<td>Chlorin</td>
<td>Chlorin(e6)</td>
<td></td>
<td>660 nm</td>
<td>Cancer, <em>in vivo</em>, <em>in vivo</em>, clinical</td>
<td>[19]</td>
</tr>
<tr>
<td>Chlorin</td>
<td>Monoaspartyl chlorin(e6), talaporfin sodium</td>
<td></td>
<td>660 nm</td>
<td>Cancer, cardiology, <em>in vivo</em>, <em>in vivo</em>, clinical</td>
<td>[140]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
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<tr>
<td>Chlorin</td>
<td>HPPH</td>
<td><img src="image" alt="HPPH Structure" /></td>
<td>660 nm</td>
<td>Cancer, in vitro, in vivo, clinical</td>
<td>[141]</td>
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<tr>
<td>Bacteriochlorin</td>
<td>TOOKAD Soluble, WST-11</td>
<td><img src="image" alt="Bacteriochlorin Structure" /></td>
<td>753 nm</td>
<td>Cancer, in vitro, in vivo, clinical</td>
<td>[142]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
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<tr>
<td>Bacteriochlorin</td>
<td>LUZ11</td>
<td><img src="image" alt="Structure LUZ11" /></td>
<td>748 nm</td>
<td>Cancer, <em>in vitro, in vivo</em>, clinical</td>
<td>[27]</td>
</tr>
<tr>
<td>Bacteriochlorin</td>
<td>BC19</td>
<td><img src="image" alt="Structure BC19" /></td>
<td>732 nm</td>
<td>Cancer, <em>in vitro, in vivo</em></td>
<td>[28]</td>
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<tr>
<td>Bacteriochlorin</td>
<td>BC21</td>
<td><img src="image" alt="Structure BC21" /></td>
<td>732 nm</td>
<td>Antimicrobial, <em>in vitro</em></td>
<td>[143]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
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</tr>
<tr>
<td>Phthalocyanine</td>
<td>Liposomal ZnPC</td>
<td><img src="image" alt="Liposomal ZnPC" /></td>
<td>670 nm</td>
<td>Cancer, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[144]</td>
</tr>
<tr>
<td>Phthalocyanine</td>
<td>Chloroaluminium sulfonated phthalocyanine (CASP)</td>
<td><img src="image" alt="Chloroaluminium sulfonated phthalocyanine" /></td>
<td>670 nm</td>
<td>Cancer, <em>in vitro</em>, <em>in vivo</em></td>
<td>[145]</td>
</tr>
<tr>
<td>Phthalocyanine</td>
<td>Silicon phthalocyanine (PC4)</td>
<td><img src="image" alt="Silicon phthalocyanine" /></td>
<td>675 nm</td>
<td>Cancer, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[33]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application, in vivo, in vitro</td>
<td>Reference</td>
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<tr>
<td>Phthalocyanine</td>
<td>RLP068</td>
<td><img src="image" alt="Phthalocyanine Structure" /></td>
<td>690 nm</td>
<td>Antimicrobial, in vitro, in vivo</td>
<td>[146]</td>
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Table 2

PSs classed as synthetic dyes

<table>
<thead>
<tr>
<th>Class</th>
<th>Name</th>
<th>Structure</th>
<th>$\lambda_{\text{max}}$</th>
<th>Application</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phenothiazinium salt</td>
<td>Methylene Blue</td>
<td><img src="image1.png" alt="Structure" /></td>
<td>660 nm</td>
<td>Cancer, antimicrobial, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[37]</td>
</tr>
<tr>
<td>Phenothiazinium salt</td>
<td>Toluidine Blue O</td>
<td><img src="image2.png" alt="Structure" /></td>
<td>630 nm</td>
<td>Antimicrobial, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[147]</td>
</tr>
<tr>
<td>Phenothiazinium salt</td>
<td>PP904</td>
<td><img src="image3.png" alt="Structure" /></td>
<td>630 nm</td>
<td>Antimicrobial, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[41]</td>
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<tr>
<td>Benzophenothiazinium salt</td>
<td>EtNBS</td>
<td><img src="image4.png" alt="Structure" /></td>
<td>670 nm</td>
<td>Cancer, antimicrobial, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[148]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
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</tr>
<tr>
<td>Halogenated xanthene</td>
<td>Rose Bengal</td>
<td><img src="image" alt="Rose Bengal Structure" /></td>
<td>540 nm</td>
<td>Cancer, antimicrobial, tissue bonding, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[149]</td>
</tr>
<tr>
<td>Squaraine</td>
<td>ASQI</td>
<td><img src="image" alt="ASQI Structure" /></td>
<td>610 nm</td>
<td>Antimicrobial, <em>in vitro</em></td>
<td>[150]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
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</tr>
<tr>
<td>BODIPY</td>
<td>Zinc(II)-dipicolylamine di-ido-BODIPY</td>
<td><img src="image" alt="Structure of Zinc(II)-dipicolylamine di-ido-BODIPY" /></td>
<td>540 nm</td>
<td>Antimicrobial, <em>in vitro</em></td>
<td>[151]</td>
</tr>
<tr>
<td>BODIPY</td>
<td>DIMPy-BODIPY</td>
<td><img src="image" alt="Structure of DIMPy-BODIPY" /></td>
<td>530 nm</td>
<td>Antimicrobial, <em>in vitro</em></td>
<td>[152]</td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
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<tr>
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</tr>
<tr>
<td>Phenalenone</td>
<td>Cationic</td>
<td><img src="image1" alt="Structure" /></td>
<td>380 nm</td>
<td>Antimicrobial, <em>in vitro</em></td>
<td>[52]</td>
</tr>
<tr>
<td>Transition metal complex</td>
<td>Ruthenium</td>
<td><img src="image2" alt="Structure" /></td>
<td>450 nm</td>
<td>Antimicrobial, <em>in vitro</em></td>
<td>[153]</td>
</tr>
<tr>
<td>Transition metal complex</td>
<td>Rhodium</td>
<td><img src="image3" alt="Structure" /></td>
<td>450 nm</td>
<td>Cancer, antimicrobial, <em>in vitro</em></td>
<td>[154]</td>
</tr>
<tr>
<td>Transition metal complex</td>
<td>Iridium</td>
<td><img src="image4" alt="Structure" /></td>
<td>600 nm</td>
<td>Cancer, antimicrobial, <em>in vitro</em></td>
<td>[155]</td>
</tr>
</tbody>
</table>
Table 3

Naturally occurring compounds as PSs

<table>
<thead>
<tr>
<th>Class</th>
<th>Name</th>
<th>Structure</th>
<th>$\lambda_{\text{max}}$</th>
<th>Application</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Perylenequinone</td>
<td>Hypericin</td>
<td>570 nm</td>
<td>Cancer, antimicrobial, \textit{in vitro}, \textit{in vivo}</td>
<td>[60]</td>
<td></td>
</tr>
<tr>
<td>Perylenequinone</td>
<td>Hypocrellin</td>
<td>470 nm</td>
<td>Cancer, antimicrobial, \textit{in vitro}, \textit{in vivo}, clinical</td>
<td>[63]</td>
<td></td>
</tr>
<tr>
<td>Class</td>
<td>Name</td>
<td>Structure</td>
<td>$\lambda_{\text{max}}$</td>
<td>Application</td>
<td>Reference</td>
</tr>
<tr>
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</tr>
<tr>
<td>Flavin</td>
<td>Cationic riboflavin</td>
<td><img src="image1.png" alt="Structure" /></td>
<td>UVA/440 nm</td>
<td>Antimicrobial, <em>in vitro</em></td>
<td>[68]</td>
</tr>
<tr>
<td>Curcuminoid</td>
<td>Curcumin</td>
<td><img src="image2.png" alt="Structure" /></td>
<td>420 nm</td>
<td>Antimicrobial, <em>in vitro</em>, <em>in vivo</em>, clinical</td>
<td>[156]</td>
</tr>
</tbody>
</table>
### Table 4

**Targeting ligands in PDT**

<table>
<thead>
<tr>
<th>Class</th>
<th>Ligand</th>
<th>Target</th>
<th>PS</th>
<th>Application</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Monoclonal antibody</td>
<td>OC125</td>
<td>Ovarian cancer</td>
<td>Chlorin(e6)</td>
<td>Cancer, <em>in vitro</em>, <em>in vivo</em></td>
<td>[157]</td>
</tr>
<tr>
<td>Peptide</td>
<td>Octreotide</td>
<td>Somatostatin receptor</td>
<td>Chlorin(e6)</td>
<td>Leukaemia, <em>in vitro</em></td>
<td>[81]</td>
</tr>
<tr>
<td>Peptide</td>
<td>RGD tripeptide</td>
<td>αvβ3 integrin</td>
<td>Phaeophorbide a</td>
<td>Cancer, <em>in vitro</em></td>
<td>[76]</td>
</tr>
<tr>
<td>Lipoprotein</td>
<td>Low-density lipoprotein</td>
<td>LDL receptor</td>
<td>Tetra-t-butyl silicon phthalocyanine</td>
<td>Cancer, atherosclerosis, <em>in vitro</em>, <em>in vivo</em></td>
<td>[158]</td>
</tr>
<tr>
<td>Serum protein</td>
<td>Transferrin</td>
<td>Transferrin receptor</td>
<td>Hematoporphyrin</td>
<td>Cancer, <em>in vitro</em></td>
<td>[77]</td>
</tr>
<tr>
<td>Serum protein</td>
<td>Modified albumin</td>
<td>Scavenger receptor</td>
<td>Chlorin(e6)</td>
<td>Atherosclerosis, <em>in vitro</em>, <em>in vivo</em></td>
<td>[159]</td>
</tr>
<tr>
<td>Vitamin</td>
<td>Folic acid</td>
<td>Folate receptor</td>
<td>Phaeophorbide a</td>
<td>Cancer, <em>in vitro</em></td>
<td>[76]</td>
</tr>
<tr>
<td>Steroid</td>
<td>Oestradiol</td>
<td>Steroid receptor</td>
<td>Phaeophorbide a</td>
<td>Breast cancer, <em>in vitro</em></td>
<td>[160]</td>
</tr>
<tr>
<td>Carbohydrate</td>
<td>Mannose</td>
<td>Mannose receptor</td>
<td>Meso-tetraphenylporphyrin</td>
<td>Cancer, <em>in vitro</em></td>
<td>[161]</td>
</tr>
</tbody>
</table>