



# Comparison of Effects of Inhibitors of Viral and Cellular Protein Kinases on Human Cytomegalovirus Disruption of Nuclear Lamina and Nuclear Egress

## Citation

Sharma, M., and D. M. Coen. 2014. "Comparison of Effects of Inhibitors of Viral and Cellular Protein Kinases on Human Cytomegalovirus Disruption of Nuclear Lamina and Nuclear Egress." *Journal of Virology* 88 (18): 10982–85. <https://doi.org/10.1128/jvi.01391-14>.

## Permanent link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:41482940>

## Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA>

## Share Your Story

The Harvard community has made this article openly available.  
Please share how this access benefits you. [Submit a story](#).

[Accessibility](#)

# Comparison of Effects of Inhibitors of Viral and Cellular Protein Kinases on Human Cytomegalovirus Disruption of Nuclear Lamina and Nuclear Egress

Mayuri Sharma, Donald M. Coen

Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, Boston, Massachusetts, USA

**Human cytomegalovirus (HCMV) kinase UL97 is required for efficient nuclear lamina disruption during nuclear egress. However, cellular protein kinase C (PKC) has been implicated in this process in other systems. Comparing the effects of UL97 and cellular kinase inhibitors on HCMV nuclear egress confirms a role for UL97 in lamina disruption and nuclear egress. A pan-PKC inhibitor did not affect lamina disruption but did reduce the number of cytoplasmic capsids more than the number of nuclear capsids.**

Transit of herpesvirus capsids from the nucleus to the cytoplasm (nuclear egress) involves phosphorylation-driven disruption of the nuclear lamina underlying the inner nuclear membrane (reviewed in references 1 to 3). Lamina disruption in herpes simplex virus 1 (HSV-1) and murine cytomegalovirus correlates with the recruitment of cellular protein kinase C (PKC) isoforms by the viral nuclear egress complex (NEC) (4, 5). During HSV-1 infection, the PKC inhibitor bisindolylmaleimide 1 (Bim-1) reduced cytoplasmic capsid numbers with little effect on nuclear capsid numbers, suggesting a role for PKC in nuclear egress (6). Also, in a cellular process akin to herpesvirus nuclear egress, rearrangement of nuclear lamins requires an isoform of PKC (7). However, during human cytomegalovirus (HCMV) infection, the NEC recruits the viral kinase UL97, not PKC, to the nuclear rim (8). Moreover, UL97 is required for efficient lamin A/C phosphorylation and lamina disruption during nuclear egress (9–11). Nevertheless, a textbook view is that in HCMV nuclear egress, host PKC functions interchangeably with UL97 in the NEC for phosphorylation-driven disruption of the nuclear lamina (12). A role for PKC or other cellular kinases in these processes would be consistent with HCMV replication proceeding, albeit inefficiently, in the absence of UL97 (11, 13). Additionally, both UL97 and cellular cyclin-dependent kinase 1 (Cdk-1, which dissolves nuclear lamina during mitosis) phosphorylate lamin A/C residue Ser22, and Ser22 phosphorylation increases somewhat during HCMV infection in the absence of UL97 (9, 11).

To compare the roles of viral and cellular kinases during lamina disruption and nuclear egress, we utilized inhibitors of UL97, PKC, and Cdk-1 at concentrations that exert substantial effects in herpesvirus systems without major cytotoxicity (6, 14, 15) (see Fig. S1 at <https://coen.med.harvard.edu>), i.e., the UL97 inhibitor maribavir (MBV) (16) at 1  $\mu$ M; the PKC isoform  $\alpha$ ,  $\beta_1$ ,  $\beta_2$ ,  $\gamma$ ,  $\delta$ , and  $\epsilon$  inhibitor Bim-1 (17) at 10  $\mu$ M; and the Cdk-1, Cdk-2, and Cdk-5 inhibitor roscovitine (Rosc) (18) at 15  $\mu$ M (6, 14, 15). Neither Bim-1 nor Rosc inhibited UL97 autophosphorylation activity *in vitro* (see Fig. S2 at <https://coen.med.harvard.edu>). Each inhibitor or a vehicle control (0.1% dimethyl sulfoxide [DMSO]) was added to serum-fed (dividing) mock-infected or HCMV strain AD169-infected cells at 48 h postinfection (hpi) to limit the inhibition of steps prior to nuclear egress. At 72 hpi, we stained cells for lamin A/C and the viral DNA polymerase subunit UL44. Replication compartment formation (UL44 staining) had pro-

gressed comparably across the infected samples (Fig. 1A). In vehicle-treated infected cells, lamin A/C staining exhibited a characteristic deformed shape, which is a marker of lamina disruption (9, 19, 20). There was a significant reduction in these nuclear deformities in MBV-treated infected cells (Fig. 1B), similar to when MBV is present throughout infection (9). However, MBV treatment did not significantly reduce the frequency of nuclear deformities in mock-infected cells (6% in both MBV-treated and vehicle-treated samples). Bim-1 or Rosc treatment did not result in significant differences from untreated HCMV-infected cells (Fig. 1B) or mock-infected cells (data not shown). These results confirm the importance of UL97 in lamina disruption during HCMV nuclear egress but provide no evidence of a role for PKC or Cdk-1 in this process.

In parallel, we measured viral titers at 96 hpi with MBV, Bim-1, and Rosc added at 48 hpi. All three inhibitors led to significant reductions in viral titers as follows: MBV, 10-fold; Bim-1, 100-fold; Rosc, 30-fold (Fig. 2, left). Thus, the lack of effect of Bim-1 or Rosc on lamina disruption was not due to a lack of activity. We assessed the effects of these compounds on viral protein expression (Fig. 2, right) as described previously (8, 11). MBV exerted little, if any, effect on the levels of the proteins assayed. Unexpectedly, while Bim-1 and Rosc did not reduce levels of UL44 and pp28, they did reduce levels of UL97 and UL50 2- to 5-fold, suggesting a role for PKC and Cdk in the expression of these proteins. Nevertheless, the reductions in UL97 and UL50 levels were substantially smaller than what one would expect to explain the effects of these compounds on viral titers, and they were not sufficient to affect lamina disruption (Fig. 1B).

We then compared nuclear egress in the presence or absence of kinase inhibitors from 48 to 96 hpi by using electron microscopy as described previously (11). MBV led to significant reductions (~10-fold) in the number of cytoplasmic capsids without de-

Received 13 May 2014 Accepted 20 June 2014

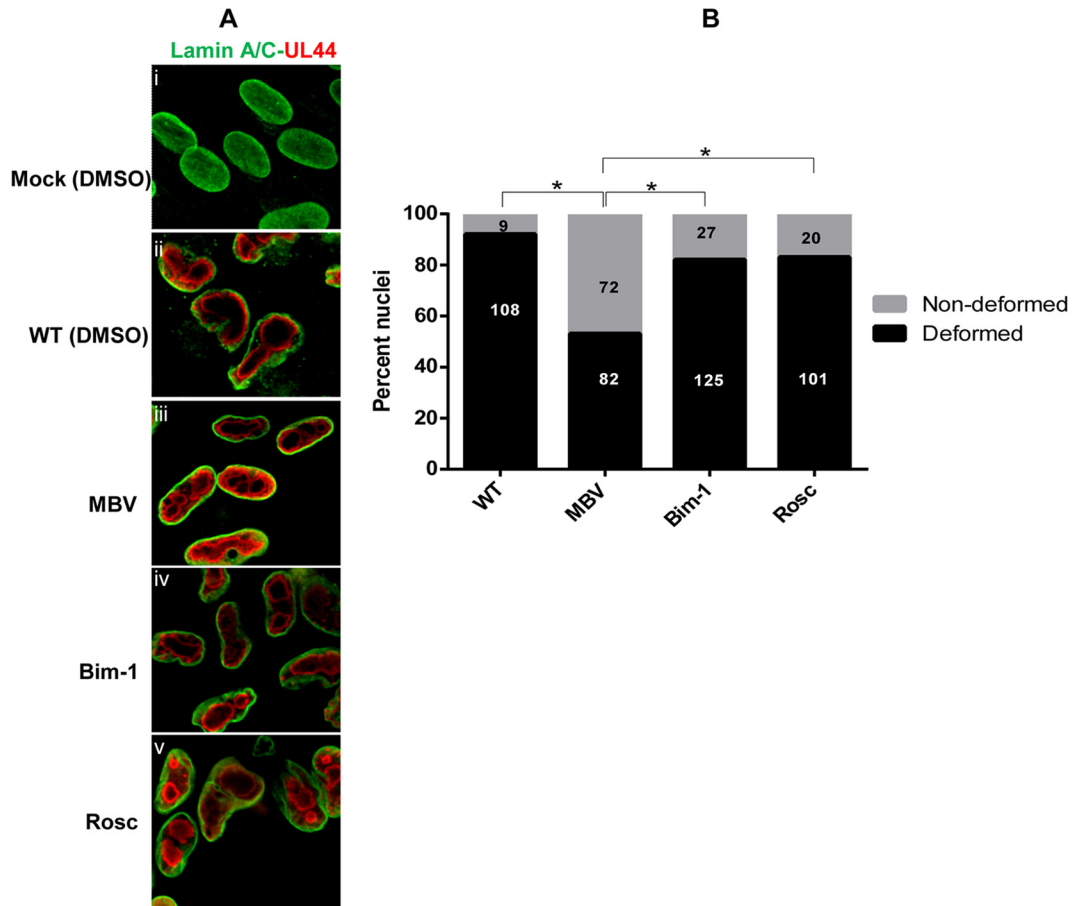
Published ahead of print 25 June 2014

Editor: R. M. Sandri-Goldin

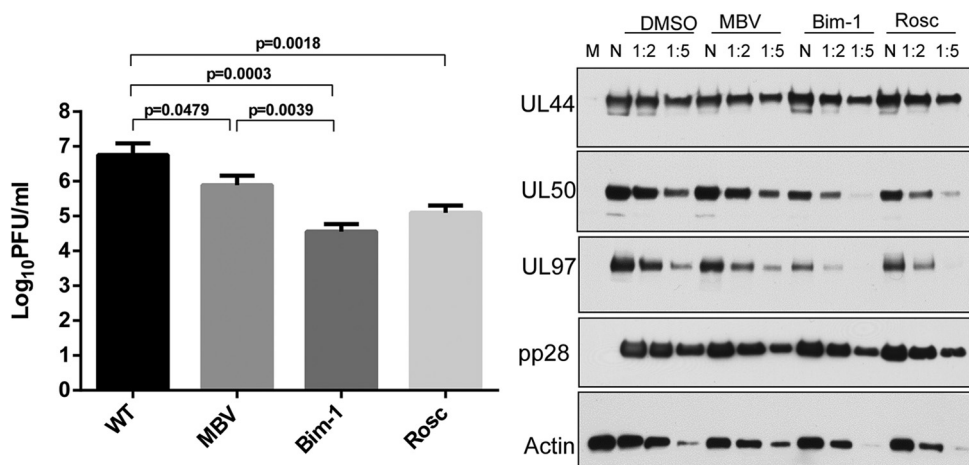
Address correspondence to Donald M. Coen, [don\\_coen@hms.harvard.edu](mailto:don_coen@hms.harvard.edu).

Copyright © 2014, American Society for Microbiology. All Rights Reserved.

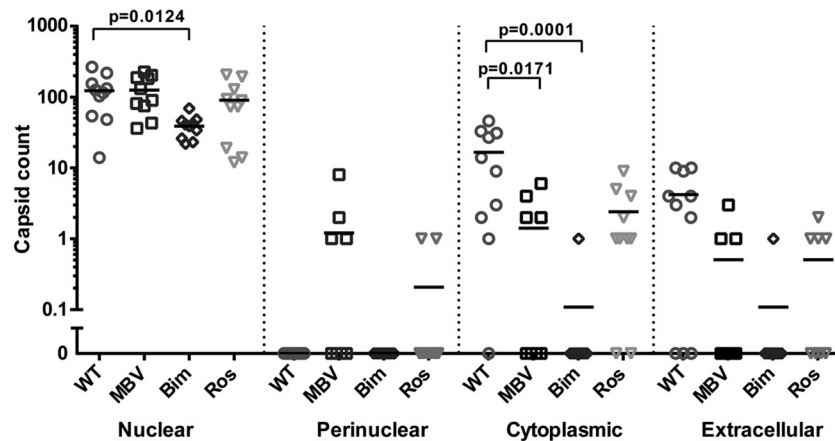
doi:10.1128/JVI.01391-14



**FIG 1** Effects of kinase inhibitors on nuclear lamina morphology. (A) Human foreskin fibroblasts were mock infected or infected with wild-type (WT) HCMV AD169rv (multiplicity of infection = 1). At 48 hpi, cells were treated with DMSO or with the viral or cellular kinase inhibitor MBV, Bim-1, or Rosc. Cells were fixed and stained for lamin A/C (green) and UL44 (red) at 72 hpi. Images were acquired by confocal microscopy and are presented as median planes from Z-stacks. (B) Mock-infected or virus-infected cells from the confocal microscopy images ( $n = 117$  to 154 per condition) were assessed for nuclear lamina deformities and analyzed for significance with Fisher's exact tests. For a family-wise type I error rate of 0.05 in a set of six comparisons, a result can be considered significant only when the  $P$  value is  $<0.0085$ . \*,  $P < 0.0001$ . No asterisk indicates no significant difference.



**FIG 2** (Left) Effects of viral or cellular kinase inhibitors on HCMV replication as determined by plaque assays with supernatants from cells infected with wild-type (WT) HCMV AD169rv (multiplicity of infection = 1) in the absence or presence of MBV, Bim-1, or Rosc (added at 48 hpi) at 96 hpi. Mean log titers (with error bars displaying standard errors of the means) from three independent experiments were assessed for statistically significant differences by one-way analysis of variance, followed by Sidak's multiple-comparison tests (five comparisons). The  $P$  values obtained are shown. No label indicates no significant difference. (Right) Effects of viral or cellular kinase inhibitors on viral protein expression. Lysates were obtained from mock-infected (lane M) or wild-type HCMV-infected cells (in the absence or presence of the kinase inhibitors shown at the top) from a parallel setup at 96 hpi. The undiluted lysates (lanes N) or serial dilutions (2-fold [1:2] or 5-fold [1:5]) were separated by SDS-PAGE, which was followed by Western blotting with antibodies against UL44, UL50, UL97, and pp28, as well as a loading control ( $\beta$ -actin), as indicated to the left.



**FIG 3** Effects of viral or cellular kinase inhibitors on HCMV nuclear egress. Human foreskin fibroblasts infected with wild-type (WT) HCMV AD169rv (multiplicity of infection = 1) in the absence (WT) or presence of MBV, Bim-1, or Rosc (added at 48 hpi) were fixed for electron microscopy at 96 hpi. Viral capsids in the nucleus, perinuclear space, or cytoplasm or outside the cell (extracellular) were counted in 10 electron microscopy sections that each represented a whole cell. Statistical analysis for each location was performed by using Kruskal-Wallis tests with Dunn's multiple-comparison posttests. The *P* value in each case is shown. No label indicates no significant difference.

creasing the number of nuclear capsids (Fig. 3), consistent with the importance of UL97 for nuclear egress (10, 11). Bim-1 led to significant reductions (~100-fold) in cytoplasmic capsid numbers, similar to results obtained with Bim-1 in the HSV-1 system (6), but also significant reductions (3-fold) in nuclear capsid numbers (Fig. 3). Rosc treatment did not cause any significant alterations in capsid numbers consistent with a role for Cdk after nuclear egress (21, 22).

We also scored for each of the three capsid forms (A, B, and C) (23) in the nucleus by analyzing the data by using Kruskal-Wallis tests with Dunn's multiple-comparison posttests (Table 1). MBV led to no more than modest reductions in the A and C forms (2.5- and 2-fold, respectively), which were not significant ( $P = 0.4314$  and  $0.7291$ ). Bim-1 led to reductions in all three forms of capsids, with a drastic (32-fold) and significant ( $P = 0.0007$ ) effect on C capsids. Since C capsids contain viral DNA, their reduction likely makes a major contribution to the severe effect of Bim-1 on viral titers. Rosc did not significantly affect the numbers of any of the nuclear capsid forms.

In summary, our results show that HCMV UL97 and PKC are not interchangeable. UL97 is important for lamina disruption and nuclear egress. PKC appears to be important for capsid formation and accumulation in both the nucleus and the cytoplasm. As Bim-1 led to a more drastic reduction in cytoplasmic capsid numbers than in nuclear capsid numbers, PKC may also be important for nuclear egress. If so, PKC could act indirectly by promoting

capsid formation and expression of UL97 and UL50. Alternatively, PKC may act directly during nuclear egress, but if it does, its role is evidently not disruption of the nuclear lamina. This is consistent with HCMV not inducing changes in the staining pattern of lamin B (8) (see Fig. S3A and B at <https://coen.med.harvard.edu>), which is an important substrate of PKC (24, 25). Thus, functions for PKC in HCMV-infected cells differ from those attributed to this kinase during nuclear egress in other systems (4, 5, 7).

#### ACKNOWLEDGMENTS

We thank Jean Pesola for help with statistical analysis and Adrian Wilkie for helpful comments. We are also grateful for the assistance of the staff and the availability of equipment at the Nikon Imaging Center at Harvard Medical School during the acquisition and analysis of immunofluorescence data, the Electron Microscopy Core Facility at Harvard Medical School for acquisition and analysis of electron microscopy data, and the ICCB-Longwood Screening Facility at Harvard Medical School for acquisition of cell viability data.

This work was supported by NIH grant R01 AI026077 to D.M.C.

#### REFERENCES

- Mettenleiter TC, Klupp BG, Granzow H. 2006. Herpesvirus assembly: a tale of two membranes. *Curr. Opin. Microbiol.* 9:423–429. <http://dx.doi.org/10.1016/j.mib.2006.06.013>.
- Mettenleiter TC and Minson T. 2006. Egress of alphaherpesviruses. *J. Virol.* 80:1610–1612. <http://dx.doi.org/10.1128/JVI.80.3.1610-1612.2006>.
- Mettenleiter TC. 2002. Herpesvirus assembly and egress. *J. Virol.* 76:1537–1547. <http://dx.doi.org/10.1128/JVI.76.4.1537-1547.2002>.
- Park R, Baines JD. 2006. Herpes simplex virus type 1 infection induces activation and recruitment of protein kinase C to the nuclear membrane and increased phosphorylation of lamin B. *J. Virol.* 80:494–504. <http://dx.doi.org/10.1128/JVI.80.1.494-504.2006>.
- Muranyi W, Haas J, Wagner M, Krohne G, Koszinowski UH. 2002. Cytomegalovirus recruitment of cellular kinases to dissolve the nuclear lamina. *Science* 297:854–857. <http://dx.doi.org/10.1126/science.1071506>.
- Leach NR, Roller RJ. 2010. Significance of host cell kinases in herpes simplex virus type 1 egress and lamin-associated protein disassembly from the nuclear lamina. *Virology* 406:127–137. <http://dx.doi.org/10.1016/j.virol.2010.07.002>.
- Speese SD, Ashley J, Jokhi V, Nunnari J, Barria R, Li Y, Ataman B, Koon A, Chang YT, Li Q, Moore MJ, Budnik V. 2012. Nuclear envelope budding enables large ribonucleoprotein particle export during synaptic

**TABLE 1** Nuclear capsid forms in the presence or absence of kinase inhibitors<sup>a</sup>

Condition	No. of whole-cell sections scored	No. (%) of capsids of form:		
		A	B	C
Vehicle	10	67 (5.4)	880 (71.3)	288 (23.3)
MBV	10	27 (2.2)	1,079 (86)	148 (11.8)
Bim-1	10	11 (2.8)	370 (94.8)	9 (2.4)
Rosc	10	84 (9.2)	644 (70)	185 (20.8)

<sup>a</sup> Statistical analysis for each comparison was performed by using Kruskal-Wallis tests with Dunn's multiple-comparison posttests. The *P* values are provided in the text.

- Wnt signaling. *Cell* 149:832–846. <http://dx.doi.org/10.1016/j.cell.2012.03.032>.
8. Sharma M, Kamil JP, Coughlin M, Reim NI, Coen DM. 2014. Human cytomegalovirus UL50 and UL53 recruit viral protein kinase UL97, not protein kinase C, for disruption of nuclear lamina and nuclear egress in infected cells. *J. Virol.* 88:249–262. <http://dx.doi.org/10.1128/JVI.02358-13>.
  9. Hamirally S, Kamil JP, Ndassa-Colday YM, Lin AJ, Jahng WJ, Baek MC, Noton S, Silva LA, Simpson-Holley M, Knipe DM, Golan DE, Marto JA, Coen DM. 2009. Viral mimicry of Cdc2/cyclin-dependent kinase 1 mediates disruption of nuclear lamina during human cytomegalovirus nuclear egress. *PLoS Pathog.* 5(1):e1000275. <http://dx.doi.org/10.1371/journal.ppat.1000275>.
  10. Krosky PM, Baek MC, Coen DM. 2003. The human cytomegalovirus UL97 protein kinase, an antiviral drug target, is required at the stage of nuclear egress. *J. Virol.* 77:905–914. <http://dx.doi.org/10.1128/JVI.77.2.905-914.2003>.
  11. Reim NI, Kamil JP, Wang D, Lin A, Sharma M, Ericsson M, Pesola JM, Golan DE, Coen DM. 2013. Inactivation of retinoblastoma protein does not overcome the requirement for human cytomegalovirus UL97 in lamina disruption and nuclear egress. *J. Virol.* 87:5019–5027. <http://dx.doi.org/10.1128/JVI.00007-13>.
  12. Mocarski ES, Shenk JT, Griffith P, Pass RF. 2013. Cytomegaloviruses, p 1960–2014. In Knipe DM, Howley PM (ed), *Fields virology*, 6th ed, vol 1. Lippincott Williams & Wilkins, Philadelphia, PA.
  13. Kamil JP, Hume AJ, Jurak I, Munger K, Kalejta RF, Coen DM. 2009. Human papillomavirus 16 E7 inactivator of retinoblastoma family proteins complements human cytomegalovirus lacking UL97 protein kinase. *Proc. Natl. Acad. Sci. U. S. A.* 106:16823–16828. <http://dx.doi.org/10.1073/pnas.0901521106>.
  14. Williams SL, Hartline CB, Kushner NL, Harden EA, Bidanset DJ, Drach JC, Townsend LB, Underwood MR, Biron KK, Kern ER. 2003. In vitro activities of benzimidazole D- and L-ribonucleosides against herpesviruses. *Antimicrob. Agents Chemother.* 47:2186–2192. <http://dx.doi.org/10.1128/AAC.47.7.2186-2192.2003>.
  15. Sanchez V, McElroy AK, Yen J, Tamrakar S, Clark CL, Schwartz RA, Spector DH. 2004. Cyclin-dependent kinase activity is required at early times for accurate processing and accumulation of the human cytomegalovirus UL122-123 and UL37 immediate-early transcripts and at later times for virus production. *J. Virol.* 78:11219–11232. <http://dx.doi.org/10.1128/JVI.78.20.11219-11232.2004>.
  16. Biron KK, Harvey RJ, Chamberlain SC, Good SS, Smith AA, III, Davis MG, Talarico CL, Miller WH, Ferris R, Dornsife RE, Stanat SC, Drach JC, Townsend LB, Kozzalka GW. 2002. Potent and selective inhibition of human cytomegalovirus replication by 1263W94, a benzimidazole L-riboside with a unique mode of action. *Antimicrob. Agents Chemother.* 46:2365–2372. <http://dx.doi.org/10.1128/AAC.46.8.2365-2372.2002>.
  17. Toullec D, Pianetti P, Coste H, Bellevergue P, Grand-Perret T, Ajakane M, Baudet V, Boissin P, Boursier E, Loriolle F, Duhamel L, Charon D, Kirilovsky J. 1991. The bisindolylmaleimide GF 109203X is a potent and selective inhibitor of protein kinase C. *J. Biol. Chem.* 266:15771–15781.
  18. Meijer L, Borgne A, Mulner O, Chong JP, Blow JJ, Inagaki N, Inagaki M, Delcros JG, Moulinoux JP. 1997. Biochemical and cellular effects of roscovitine, a potent and selective inhibitor of the cyclin-dependent kinases cdc2, cdk2 and cdk5. *Eur. J. Biochem.* 243:527–536. <http://dx.doi.org/10.1111/j.1432-1033.1997.t01-2-00527.x>.
  19. Buser C, Walther P, Mertens T, Michel D. 2007. Cytomegalovirus primary envelopment occurs at large infoldings of the inner nuclear membrane. *J. Virol.* 81:3042–3048. <http://dx.doi.org/10.1128/JVI.01564-06>.
  20. Camozzi D, Pignatelli S, Valvo C, Lattanzi G, Capanni C, Dal Monte P, Landini MP. 2008. Remodelling of the nuclear lamina during human cytomegalovirus infection: role of the viral proteins pUL50 and pUL53. *J. Gen. Virol.* 89:731–740. <http://dx.doi.org/10.1099/vir.0.83377-0>.
  21. Sanchez V, McElroy AK, Spector DH. 2003. Mechanisms governing maintenance of Cdk1/cyclin B1 kinase activity in cells infected with human cytomegalovirus. *J. Virol.* 77:13214–13224. <http://dx.doi.org/10.1128/JVI.77.24.13214-13224.2003>.
  22. Sanchez V, Spector DH. 2006. Cyclin-dependent kinase activity is required for efficient expression and posttranslational modification of human cytomegalovirus proteins and for production of extracellular particles. *J. Virol.* 80:5886–5896. <http://dx.doi.org/10.1128/JVI.02656-05>.
  23. Gibson W. 1996. Structure and assembly of the virion. *Intervirology* 39:350–354.
  24. Collas P, Thompson L, Fields AP, Poccia DL, Courvalin JC. 1997. Protein kinase C-mediated interphase lamin B phosphorylation and solubilization. *J. Biol. Chem.* 272:21274–21280. <http://dx.doi.org/10.1074/jbc.272.34.21274>.
  25. Hocevar BA, Burns DJ, Fields AP. 1993. Identification of protein kinase C (PKC) phosphorylation sites on human lamin B. Potential role of PKC in nuclear lamina structural dynamics. *J. Biol. Chem.* 268:7545–7552.