Reducing Amyloid Plaque Burden via Ex Vivo Gene Delivery of an Aβ-Degrading Protease: A Novel Therapeutic Approach to Alzheimer Disease

Citation

Published Version
doi:10.1371/journal.pmed.0040262

Permanent link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:4724764

Terms of Use
This article was downloaded from Harvard University’s DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

Share Your Story
The Harvard community has made this article openly available. Please share how this access benefits you. Submit a story.

Accessibility
Reducing Amyloid Plaque Burden via Ex Vivo Gene Delivery of an Aβ-Degrading Protease: A Novel Therapeutic Approach to Alzheimer Disease

Matthew L. Hemming1, Michaela Patterson2, Casper Reske-Nielsen2, Ling Lin2, Ole Isacson2, Dennis J. Selkoe1*

1 Center for Neurologic Diseases, Brigham and Women’s Hospital and Harvard Medical School, Boston, Massachusetts, United States of America, 2 Neuroregeneration Laboratories, McLean Hospital and Harvard University Udall Parkinson’s Disease Research Center of Excellence, Belmont, Massachusetts, United States of America

Funding: This work was supported by National Institutes of Health Grants AG12749 (DJS) and NS39793 (OI), the Stern Foundation (OI), and the Anti-Aging Foundation (OI), and the Harvard Center for Neurodegeneration and Repair (MLH). DJS is a consultant to Elan. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Academic Editor: Sam Gandy, Farber Institute, United States of America


Received: March 19, 2007
Accepted: July 18, 2007
Published: August 28, 2007

Copyright: © 2007 Hemming et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abbreviations: Aβ, amyloid β-protein; ACE, angiotensin-converting enzyme; AD, Alzheimer disease; APP, β-amyloid precursor protein; CHO, Chinese hamster ovary; DAGNPNG, 3-dansyl-D-Ala-Gly-p-(nitro)-Phe-Gly; GFAP, glial fibrillary acidic protein; HEK, human embryonic kidney; NEP, neprilysin; SEM, standard error of the mean; sNEP, secreted neprilysin

* To whom correspondence should be addressed. E-mail: dselkoe@rics.bwh.harvard.edu

ABSTRACT

Background

Understanding the mechanisms of amyloid-β protein (Aβ) production and clearance in the brain has been essential to elucidating the etiology of Alzheimer disease (AD). Chronically decreasing brain Aβ levels is an emerging therapeutic approach for AD, but no such disease-modifying agents have achieved clinical validation. Certain proteases are responsible for the catabolism of brain Aβ in vivo, and some experimental evidence suggests they could be used as therapeutic tools to reduce Aβ levels in AD. The objective of this study was to determine if enhancing the clearance of Aβ in the brain by ex vivo gene delivery of an Aβ-degrading protease can reduce amyloid plaque burden.

Methods and Findings

We generated a secreted form of the Aβ-degrading protease neprilysin, which significantly lowers the levels of naturally secreted Aβ in cell culture. We then used an ex vivo gene delivery approach utilizing primary fibroblasts to introduce this soluble protease into the brains of β-amyloid precursor protein (APP) transgenic mice with advanced plaque deposition. Brain examination after cell implantation revealed robust clearance of plaques at the site of engraftment (72% reduction, p = 0.0269), as well as significant reductions in plaque burden in both the medial and lateral hippocampus distal to the implantation site (34% reduction, p = 0.0020; and 55% reduction, p = 0.0081, respectively).

Conclusions

Ex vivo gene delivery of an Aβ-degrading protease reduces amyloid plaque burden in transgenic mice expressing human APP. These results support the use of Aβ-degrading proteases as a means to therapeutically lower Aβ levels and encourage further exploration of ex vivo gene delivery for the treatment of Alzheimer disease.

The Editors’ Summary of this article follows the references.
Introduction

The pathologic hallmarks of Alzheimer disease (AD) are extracellular plaques of amyloid-β protein (Aβ) and intraneuronal neurofibrillary tangles of tau protein, both of which accumulate in brain regions mediating memory and cognition [1]. All known early-onset, inherited forms of AD arise from mutations in the β-amyloid precursor protein (APP), from which the Aβ peptide is generated, or the presenilins, which are the proteases that effect the cleavage of APP, which liberates Aβ. Both amyloid plaques and soluble Aβ oligomers are believed to have neurotoxic effects [2,3], and evidence suggests that Aβ neurotoxicity can promote neurofibrillary tangle formation [4,5]. Though approved treatments for AD ameliorate only the symptoms, potentially disease-modifying interventions under development focus on therapeutically lowering Aβ production [6] or enhancing Aβ clearance [7,8].

The role of Aβ degradation in the clearance of the Aβ peptide is becoming more broadly understood and appreciated, with the proteases nephrilysin [9], insulin-degrading enzyme [10,11], endothelin-converting enzymes 1 and 2 [12], plasmin [13], and cathepsin B [14] all capable of regulating Aβ levels in vivo. Supporting a therapeutic function for Aβ-degrading proteases, both transgenic overexpression [15] and direct viral vector injection [14,16,17] of these enzymes have been shown to potently lower Aβ levels and plaque burden and reduce Aβ-associated neuropathology. The applicability of these approaches for AD treatment in humans is uncertain, and alternative methods of gene delivery merit exploration.

In ex vivo gene therapy, cells are taken from a patient, genetically modified in vitro, then implanted back into the patient to exert their new salutary effects. This approach has produced therapeutic improvements in experimental models of human diseases and conditions including hemophilia [18], retinal degeneration [19], cancer [20], spinal cord injury [21], myopathy [22], ischemia [23], Parkinson disease [24], Huntington disease [25], amyotrophic lateral sclerosis [26], and Alzheimer disease [27]. Several of these potential treatments have advanced to human trials [28–31], with encouraging outcomes for patients. In a recent Phase I clinical trial, autologous fibroblasts from AD patients were modified to produce nerve growth factor and implanted into the forebrain of human disease models [28]. These cells overexpressing empty vector, NEP, or sNEP and conditioned medium from APP-overexpressing CHO cells was combined with the V717F AD-causing mutation [37] were maintained in medium containing 200 µg/ml G418. Transfections were performed with Fugene 6 (Roche, http://www.roche.com), and stable cell lines were selected using 400 µg/ml hygromycin B. Cells were conditioned for Aβ measurements and protein analysis in Opti-MEM I (Invitrogen) for 16–18 h. For cell-free Aβ degradation assays, conditioned medium from APP-overexpressing CHO cells was combined with 10-fold concentrated conditioned medium from CHO cells overexpressing empty vector, NEP, or sNEP and incubated at 37 °C for 18 h.

Lentiviral vectors were generated by inserting the sNEP and GFP cDNAs into pCDH1 (System Biosciences, http://www.systembio.com/) and were cotransfected with the helper plasmids delta8.9 and VSV-G as previously described [38] into 293-FT cells (Invitrogen). Conditioned media were collected 50–60 h later, centrifuged at 500 g for 5 min to remove suspended cells, and stored at −80 °C. Viral multiplicity of infection was estimated by cellular fluorescence and Western blotting by transducing 293-FT cells with serial dilutions of viral supernatant. Cells were transduced for experiments at a multiplicity of infection of six to ten viral particles per cell with media containing 6 µg/ml polybrene.

Immunoblotting

Cells and tissue were lysed in 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1% NP-40, protease inhibitor cocktail (Roche), 2 mM 1,10-phenanthroline, and 5 mM EDTA, and the extracts centrifuged at 1,000 g for 10 min to remove nuclei. Protein concentrations were determined using a bicinchoninic acid-based assay (Pierce, http://www.perbiodirect.com/). Samples were then subjected to SDS-PAGE and Western blotting. We detected NEP with monoclonal antibody 56C6 (1:100 dilution; Abcam, http://www.abcam.com/); GFP with polyclonal antibody A11122 (1:1,000 dilution; Invitrogen); Aβ with mono-
Reactions were stopped by heating samples to 100°C using a Victor2 multilabel plate reader (excitation 342 nm; 400 lm min, then centrifuging at 5,000 g for 5 min to remove the supernatant). Western blots were probed with anti-mouse or anti-rabbit secondary antibodies (1:10,000 dilution) conjugated to Alexa Fluor 680 (Molecular Probes, http://probes.invitrogen.com/) or IRDye 800 (Rockland Immunochemicals, http://www.rockland-inc.com). Blots were detected using the Odyssey infrared imaging system (LI-COR, http://www.loric.com/).

**NEP Activity Assay**

NEP proteolytic activity was measured using the substrate 3-dansyl-D-Ala-Gly-p-(nitro)-Phe-Gly (DAGNPG; Sigma, http://www.sigma-aldrich.com) [39,40]. Cell lysate (100 μg) or the same relative amount of concentrated conditioned medium (prepared without protease inhibitors) was incubated with 50 μM DAGNPG and 1 μM captopril (to inhibit any ACE cleavage of DAGNPG) in a volume of 200 μl at 37°C. Reactions were stopped by heating samples to 100°C for 5 min, then centrifuging at 5,000 g for 5 min to remove the denatured protein. The whole supernatant was diluted into 400 μl of 50 mM Tris (pH 7.4) and fluorescence determined using a Victor2 multilabel plate reader (excitation 342 nm; emission 562 nm).

**ELISA**

ELISAs for Aβ were performed as previously described [39,41] with few modifications. 96-well ELISA plates (Costar, http://www.corning.com/) were coated with 3.5 μg/ml of the capture antibody. Aβ was measured by capturing with monoclonal antibody 266, specific to residues 13–28 of Aβ. Captured Aβ was detected with biotinylated monoclonal antibody 3D6, specific to residues 1–5 of the Aβ N terminus (all ELISA antibodies were gifts of Elan Pharmaceuticals, http://www.elan.com). ELISAs were developed by incubating the Aβ-bound biotinylated 3D6 with avidin-horseradish peroxidase (Vector Labs, http://www.vectorlabs.com) followed by a 1 h incubation with QuantaBlu fluorogenic peroxidase substrate (Pierce), and the resulting fluorescence (excitation 340 nm; emission 400 nm) was measured. Plate washing after the antibody and enzyme binding steps was performed three times for 1 min with Tris-buffered saline, 0.05% Tween 20.

**Primary Fibroblast Generation and Transduction**

Primary fibroblasts were generated from young wild-type littermates of J20 transgenic mice that expressed human APP. Skin biopsies were swabbed with 70% ethanol to sterilize the tissue, and then were minced with sterile scalpels in 0.25% trypsin-EDTA. Minced tissue was incubated in trypsin-EDTA at 37°C for 15 min and triturated to further disrupt the tissue. Cells remaining in suspension were plated in growth medium and allowed to expand to 90% confluency. Fibroblasts were then passaged and transduced with two serial applications of either sNEP (treatment cells) or GFP (control cells) lentiviral vectors to attain strong transgene expression in all cells. Fibroblasts were passaged a maximum of five times before injections. Confluent cultures of transduced fibroblasts were maintained to observe cell longevity and transgene expression. After 12 mo in culture, the cells remained metabolically active and continued to express the introduced transgene.

**Mice and Surgical Procedures**

The J20 line of APP<sup>Swe</sup> heterozygous transgenic mice (C57BL/6 x DBA2 background) [42] were aged to approximately 22–23 mo before surgery to allow for substantial plaque deposition. For surgical procedures, mice received intraperitoneal injections of a preanesthetic mixture (a 1:1 solution of atropine sulfate and acepromazine maleate), followed by a 2:1 solution of ketamine and xylazine. Mice received a unilateral hippocampal injection of 500,000 fibroblasts expressing sNEP or GFP to allow for comparison to the contralateral side. The stereotaxic coordinates for graft placement were calculated from bregma: AP −3.3 mm; L −3.3 mm; DV −3.12 mm and −1.9 mm. Cells, in a total volume of 4 μl, were injected at two dorsoventral positions to better distribute the graft. Brains were harvested 28 d after surgery and fixed in 10% formalin for subsequent immunohistochemical analysis. All animal procedures were approved by the Harvard Standing Committee for Animal Use and by the Animal Care and Use Committee at McLean Hospital (Belmont, Massachusetts, United States).

**Immunohistochemistry and Image Analysis**

Sagittal brain sections of 18 μm thickness were deparaffinized and hydrated through a series of graded alcohol steps and washed in phosphate-buffered saline. Endogenous peroxidase activity was quenched with 0.6% hydrogen peroxide in methanol for 15 min. Sections were blocked in serum for 25 min, then incubated with the anti-Aβ antibody R1282 (1:1,000 dilution) [39], anti-NEP antibody 56C6 (1:100), anti-GFP antibody A11122 (1:100), or anti-glial fibrillary acidic protein (GFAP) antibody (1:1,000; Dako, http://www.dako.com/) for 2 h. After washing, an anti-rabbit or antimouse biotinylated secondary antibody was applied for 30 min, washed, and developed using the avidin/biotin/HRP method (ABC Elite Kit, Vector Labs) and DAB chromogenic reaction (Liquid DAB, BioGenex, http://www.biogenex.com). Images were captured and quantified from approximately 20 stained sections per brain of each mouse using a 2× or 5× objective. Thioflavin-S staining was performed by incubating tissue sections for 8 min in 1% thioflavin-S, followed by washing in 80% and 95% ethanol, then water. The brain area covered by Aβ, thioflavin plaques, and GFAP staining was determined in a blinded fashion using IPLab Spectrum 3.1 Image Analyzer software (Signal Analytics, http://www.scanalytics.com/) and NIH ImageJ software [43]. The ratio between the immunohistochemical and thioflavin staining on the ipsilateral versus contralateral hemispheres was calculated for comparisons.

**Statistical Analysis**

The data were analyzed using a one-way analysis of variance and Tukey post-hoc comparison or a two-tailed Student t-test, where appropriate. Calculated comparisons of p < 0.05 were considered significant. All reported values represent the means ± standard error of the mean (SEM).

**Results**

**NEP Expression and Generation of sNEP**

Several studies point to NEP as one of the prominent Aβ-degrading proteases within the brain [9,15,44,45]. To determine the tissue distribution of NEP in the brain and several...
peripheral tissues, we dissected cortex, hippocampus, cerebellum, and the basal ganglia/brainstem from the murine brain and compared protein expression levels to those in the liver, lung, and kidney. As positive and negative controls, lysates from CHO cells transfected with NEP or empty vector were examined in parallel. Equal amounts of protein from each sample were examined by quantitative fluorescent Western blotting. Because transfected NEP may run as a

**Figure 1.** Tissue Distribution of NEP and Generation of Secreted NEP

(A) Identical amounts of protein from transfected CHO cells and the indicated mouse tissues were probed by NEP immunoblot. CHO cells were transiently transfected with empty vector (CHO-control) or human NEP (CHO-NEP). The indicated peripheral tissues and brain regions (Bg/Bs, basal ganglia/brainstem; Cb, cerebellum; Ctx, cortex; Hip, hippocampus) were homogenized and probed by Western blot for NEP. The lower blot is a longer exposure of the brain samples, demonstrating low levels of NEP in the brain. Each sample was either left untreated (−) or deglycosylated with PNGase F (+) to remove N-linked sugars.

(B) Schematic representation of wild-type NEP and sNEP proteins. The sNEP construct was generated by replacing the NEP transmembrane (TM) domain and cytosolic N terminus with a signal peptide (SP) ending at the luminal residue 52 of NEP. Cleavage of the signal peptide produces a soluble, secreted form of NEP.

(C) NEP immunoblot of cellular lysates from CHO cells transfected with empty vector, NEP, or sNEP constructs.

(D) NEP activity assay in which the fluorogenic NEP substrate DAGNPG was incubated with lysates from CHO cells stably transfected with the indicated constructs. NEP enzymatic activity was inhibited by phosphoramidon.

(E) NEP immunoblot of conditioned medium from CHO cells transfected with the indicated constructs.

(F) NEP activity assay on conditioned media from cells transfected with the indicated constructs. NEP activity assays of conditioned media (F) were performed using the same relative amounts of material as for the lysates (D).

Immunoblots are representative of at least three experiments; NEP activity assays report the mean ± SEM of six experiments.

doi:10.1371/journal.pmed.0040262.g001
CHO cells overexpressing APP were cocultured with CHO cells stably expressing either empty vector, NEP, or sNEP constructs. Equal numbers of cells were seeded, grown to confluence, and media were conditioned for 18 h. Conditioned media were analyzed for total Aβ levels by ELISA. Each condition is shown in duplicate.

To determine if the sNEP construct is able to degrade naturally secreted Aβ to a similar extent as wild-type NEP, we cocultured CHO cells stably expressing APP at a 1:1 ratio with CHO cells expressing either empty vector as control, wild-type NEP, or sNEP. Cells were allowed to grow to confluency. Media were conditioned for Aβ catabolism, with the NEP and sNEP conditions reducing Aβ levels in the media to 74% and 55% of control, respectively (*p < 0.01 and ***p < 0.001). Concentrating the conditioned media from these cells for Western blot analysis revealed a strong signal for sNEP, consistent with proper sNEP expression and secretion (Figure 1E). Quantifying NEP activity in conditioned media (using the same relative amount of material used for cell lysates in Figure 1D) demonstrated robust NEP activity in the sNEP samples alone, and this was again completely inhibited by phosphoramidon (Figure 1F). These data demonstrate that the sNEP construct is processed and secreted by cells as expected and that sNEP is enzymatically active against a prototypical NEP substrate.

sNEP Reduces Aβ Levels in Cell Culture

To determine if the sNEP construct is able to degrade naturally secreted Aβ to a similar extent as wild-type NEP, we cocultured CHO cells stably expressing APP at a 1:1 ratio with CHO cells expressing either empty vector as control, wild-type NEP, or sNEP. Cells were allowed to grow to confluency, and media were conditioned for Aβ ELISA determinations. Both wild-type NEP and sNEP were capable of promoting Aβ catabolism, with the NEP and sNEP conditions reducing Aβ levels in the media to 74% and 55% of control, respectively (p < 0.01 for NEP and p < 0.001 for sNEP) (Figure 2A). These reductions in Aβ levels were not due to differences in the proliferation rates of the cell lines, as there was equivalent APP expression in each cocultured condition, indicating a similar number of APP expressing cells at confluency.

In order to clarify the location of NEP- and sNEP-mediated Aβ degradation, we mixed conditioned media from the APP-overexpressing cell line with concentrated media from cells

contrast to lung and kidney, brain levels of NEP are very low and detectable only upon longer blot exposure (Figure 1A, lower blot). Thus, although NEP has been characterized as an essential protease for Aβ catabolism, its protein expression levels are relatively low in the brain compared to peripheral tissues that express NEP. Moreover, since cerebral NEP expression decreases with age and in AD [33,34], elevating NEP levels could be neuroprotective by enhancing Aβ catabolism.

In exploring potential therapeutic approaches to reducing brain Aβ levels by elevating NEP expression, we evaluated a secreted form of NEP, sNEP. A secreted Aβ-degrading protease would possess the advantage of diffusion through the interstitial fluid, where it may clear Aβ at sites of extracellular deposition. To generate this sNEP construct, we used an overlapping PCR method to fuse the 29-amino acid signal peptide from ACE to the luminal domain of NEP beginning at residue 52 (Figure 1B). This construct, upon being translated and inserted into the endoplasmic reticulum membrane, has the signal peptide removed to liberate a soluble, secreted form of NEP.

Transfecting cells with empty vector, NEP, or the sNEP constructs and examining cell lysates by Western blotting revealed a robust immunoreactive NEP band in the wild-type NEP condition and a weaker, incompletely glycosylated, lower molecular weight signal in the sNEP lane (Figure 1C). Measuring NEP enzymatic activity in cell lysates demonstrated NEP activity only in the wild-type NEP condition, and this was inhibitable to baseline signal by the metalloprotease inhibitor phosphoramidon (Figure 1D). The incompletely glycosylated intracellular sNEP showed no enzymatic activity, consistent with reports that glycosylation is required for NEP activity [46]. Concentrating the conditioned media from these cells for Western blot analysis revealed a strong signal for sNEP, consistent with proper sNEP expression and secretion (Figure 1E). Quantifying NEP activity in conditioned media (using the same relative amount of material used for cell lysates in Figure 1D) demonstrated robust NEP activity in the sNEP samples alone, and this was again completely inhibited by phosphoramidon (Figure 1F). These data demonstrate that the sNEP construct is processed and secreted by cells as expected and that sNEP is enzymatically active against a prototypical NEP substrate.

sNEP Reduces Aβ Levels in Cell Culture

To determine if the sNEP construct is able to degrade naturally secreted Aβ to a similar extent as wild-type NEP, we cocultured CHO cells stably expressing APP at a 1:1 ratio with CHO cells expressing either empty vector as control, wild-type NEP, or sNEP. Cells were allowed to grow to confluency, and media were conditioned for Aβ ELISA determinations. Both wild-type NEP and sNEP were capable of promoting Aβ catabolism, with the NEP and sNEP conditions reducing Aβ levels in the media to 74% and 55% of control, respectively (p < 0.01 for NEP and p < 0.001 for sNEP) (Figure 2A). These reductions in Aβ levels were not due to differences in the proliferation rates of the cell lines, as there was equivalent APP expression in each cocultured condition (Figure 2B), indicating a similar number of APP expressing cells at confluency.

In order to clarify the location of NEP- and sNEP-mediated Aβ degradation, we mixed conditioned media from the APP-overexpressing cell line with concentrated media from cells

broad band due to its varying degrees of glycosylation, all samples were assessed in the absence or presence of PNGase F to remove N-linked glycosylation and reduce NEP immunoreactivity to a single signal. Mouse lung and kidney express high levels of NEP that comigrates with transfected human NEP, whereas the liver lacks NEP expression (Figure 1A). In
expressing empty vector, wild-type NEP, or sNEP. Only the sNEP-conditioned media exhibited an increased capacity to degrade Aβ compared to control (p < 0.001) (Figure 2C), elevating the amount of Aβ degraded 2-fold. Taken together, these data demonstrate that wild-type NEP degrades Aβ principally while anchored to the cell membrane, whereas sNEP is efficiently secreted into the extracellular environment to mediate Aβ catabolism.

**Generation of Viral and Cell-Based Aβ-Degrading Vectors**

To pursue an ex vivo gene delivery approach, we generated lentiviral vectors able to efficiently transduce most cell types. Retroviral vectors have been routinely utilized to genetically enhance cells ex vivo prior to engraftment for the experimental treatment of human diseases [47–49]. We inserted the cDNA of sNEP or GFP (as control) into a lentiviral vector driven by the cytomegalovirus promotor (Figure 3A). The resulting vectors were able to transduce all cultured cell types tested (HeLa, SH-SY5Y, SK-N-SH, HEK, CHO, and primary immortalized fibroblasts), and produced high levels of GFP or sNEP expression. To assess the ability of the sNEP lentiviral vector to reduce Aβ levels in APP overexpressing cell lines, we transduced HEK cells stably expressing a familial AD mutant form of APP695 and CHO cells stably expressing a familial AD mutant form of APP751 with the sNEP or GFP vectors. Examining cell lysates and conditioned media by Western blot showed strong transgene expression, with HEK cells attaining higher sNEP and GFP production than did CHO cells (Figure 3B, top three blot pairs). Expression of sNEP did not change APP levels in these cells compared with GFP transduction (Figure 3B, fourth blot pair); however, sNEP expression resulted in a marked decrease in Aβ monomer in the conditioned media (Figure 3B, fifth blot pair).
ELISA demonstrated significant reductions in Aβ due to sNEP expression in both HEK and CHO APP-overexpressing cell lines ($p < 0.001$) (Figure 3C).

Fibroblasts were chosen as the optimal cell type for use in ex vivo gene delivery for a number of reasons. Fibroblasts survive grafting into several organs, including the brain; they survive and can sustain transgene expression for years [28]; they do not form tumors or migrate from the graft site [47]; they cause no detectible toxicity to the host tissue [50]; and they can be noninvasively extracted from patients via skin biopsy. For this study, fibroblasts were prepared from the wild-type littermates of the J20 APP transgenic mice. Cells were dissociated from skin biopsies and plated in growth medium for brief expansion in culture. Following their first passage in culture, the cells were transduced with either the sNEP or the GFP lentiviral vector. The fibroblasts transduced with GFP produce a strong native fluorescent signal (Figure 4A), while cells transduced with sNEP robustly secrete the sNEP protein into the extracellular medium (Figure 4B). To minimize the possibility of in vitro cell transformation, the fibroblasts used in the study were not passaged more the five times before engraftment into the experimental transgenic mice.

**sNEP Cell Grafting Reduces Plaque Burden Locally and Distal to the Implantation Site**

A cohort of J20 APP transgenic mice was aged to achieve significant plaque deposition [42] for use in these studies. To each of these mice, 500,000 cells expressing sNEP or GFP were stereotaxically injected unilaterally into the hippocampus (stereotaxic coordinates: AP $-3.3$ mm; L $-3.3$ mm; DV $-3.12$ and $-1.9$ mm), with the un.injected contralateral hippocampus serving as a control. Cells were injected at two dorsoventral positions to better distribute the graft. The brains of the mice were harvested and immunohistochemically assayed 28 d after surgery. Mice implanted with sNEP grafts exhibited NEP immunoreactivity at the graft site, as well as some diffuse NEP signal in nearby areas of the hippocampus that extended approximately 800 μm from the grafted cells (Figure 5A, right photomicrograph), consistent with sNEP secretion. Endogenous NEP immunoreactivity is localized to the cerebral blood vessels and the choroid plexus (Figure 5A, left photomicrograph). GFP fibroblasts exhibit GFP staining restricted to the grafted cells (Figure 5B, right).

To determine the extent of Aβ clearance from the brain due to sNEP delivery, we stained sagittal sections for Aβ immunoreactivity from the medial to the lateral hippocampus. At the graft site, the ipsilateral hippocampus was largely cleared of plaques near the sNEP-producing fibroblasts (Figure 5E, right). There was some reduction in plaque load at the GFP graft (Figure 5F, right), which was likely due to the displacement of plaque-laden tissue by the grafted GFP-expressing fibroblasts. The ipsilateral hippocampus medial to the graft site showed significant reductions in Aβ immunoreactivity in the sNEP-injected mice (compared to
Figure 5. Reduction in Hippocampal Plaque Burden following sNEP Cell Engraftment

Aged J20 APP transgenic mice were implanted with fibroblasts expressing either sNEP (n = 5) (A, C, E, and G) or GFP (n = 3) (B, D, F, and H). Cells were stereotaxically placed into the right (ipsilateral, right) hippocampus, with the uninjected left (contralateral, left) hippocampus serving as control. Brains were harvested for analysis 28 d after surgery. Grafted cells were immunoreactive for either NEP (A) or GFP (B), with superior and inferior aspects of the graft, respectively, indicated by arrows. Images (C–H) show staining for plaque burden at sites medial to the graft (C and D), at the site of the graft (E and F), and at sites lateral to the graft (G and H). These fields were quantified by calculating the ratio of the area covered by plaques on the ipsilateral versus the contralateral side; quantification was done in a blinded fashion. This ratio was determined separately for the sNEP and GFP groups and compared at the hippocampus medial to the graft (I), the graft site (J), and the hippocampus lateral to the graft (K). Data represent the ratio's mean ± SEM, compared to GFP control: ** $p = 0.0020$ for the medial hippocampus (I); * $p = 0.0269$ for the graft site (J); and ** $p = 0.0081$ for the lateral hippocampus (K).

doi:10.1371/journal.pmed.0040262.g005
the uninjected contralateral hippocampus), whereas no effect was observed in the GFP-injected mice (Figure 5C versus 5D, respectively). Similarly, in the lateral hippocampus distal to the graft, we observed significant reductions in plaque burden due to sNEP expression (Figure 5G versus 5H). Changes in plaque burden were measured in blinded fashion with automated image analysis, and the data expressed as a ratio of the area covered by Aβ plaque on the grafted ipsilateral versus the control contralateral side. Comparing the ipsilateral/contralateral ratios of the sNEP and GFP groups, significant reductions in plaque burden were observed in the ipsilateral hippocampus from mice engrafted with sNEP-secreting fibroblasts medial to the graft site (34% reduction, \( p < 0.01 \)), within the graft site (72% reduction, \( p < 0.05 \)), and lateral to the graft site (55% reduction, \( p < 0.01 \)) (Figure 5I–5K). Thus, ex vivo gene delivery of a soluble, secreted form of human NEP resulted in substantial and significant reductions in Aβ plaque burden both in the vicinity of the graft and adjacent to the graft site.

**Reduction in Thioflavin-Positive Plaques, but Not Astrocytosis, Following sNEP Cell Engraftment**

To determine if the engrafted sNEP was able to clear fibrillar plaques, we examined the number of thioflavin-positive plaques in the ipsilateral and contralateral hippocampus. Sagittal sections throughout the hippocampus were stained with thioflavin-S, and the ratio between the number of plaques on the ipsilateral versus contralateral hemispheres were calculated for sNEP and GFP groups for the medial hippocampus (E), the graft site (F), and the lateral hippocampus (G). Data represent the mean ± SEM, compared to GFP control: \( *p = 0.0432 \) for the medial hippocampus (E), and \( *p = 0.0337 \) for the graft site (F). The difference at the lateral hippocampus was not significant.

---

**Figure 6. Reduction in Hippocampal Thioflavin Staining Following sNEP Cell Engraftment**

Brain sections from mice engrafted with cells expressing sNEP or GFP were stained with thioflavin-S to quantify fibrillar plaques. Photomicrographs (A–D) show thioflavin staining at sites medial to the graft (A and B) and at the site of the graft (C and D). Thioflavin-positive plaques were counted, and the ratios of plaques on the ipsilateral versus contralateral hemispheres were calculated for sNEP and GFP groups for the medial hippocampus (E), the graft site (F), and the lateral hippocampus (G). Data represent the mean ± SEM, compared to GFP control: \( *p = 0.0432 \) for the medial hippocampus (E), and \( *p = 0.0337 \) for the graft site (F). The difference at the lateral hippocampus was not significant.

---

\[ \text{doi:10.1371/journal.pmed.0040262.g006} \]
compared to control (Figure 6E–6G). The lateral hippocampus had few or no thioflavin-positive plaques in either condition and was thus not a reliable reporter of plaque clearance. These data demonstrate that sNEP delivery promotes the clearance of fibrillar, thioflavin-positive plaques from the brain of APP transgenic mice.

The activation of astrocyte growth (astrocytosis) is a marker for brain inflammation, and astrocyte hypertrophy and number are increased in AD and may play a role in disease pathogenesis [1]. Transgenic expression of neprilysin has been shown previously to reduce astrocytosis over the lifetime of an APP-transgenic mouse [15], presumably due to reductions in Aβ accumulation. However, because we used a surgical approach to exogenously deliver a soluble form of NEP that clears existing plaques, there could be an inflammatory response associated with plaque removal or with the soluble protease itself. To examine this possibility, we stained sNEP and GFP brains for the astrocyte GFAP. At the site of cell engraftment, few astrocytes were found to have infiltrated the graft, although there was a modest increase in GFAP staining along the edge of the graft and along the needle track (Figure 7A and 7B). However, comparing overall hippocampal GFAP staining on the ipsilateral versus contralateral hemispheres showed that there was no change in astrocytosis due to cell engraftment or sNEP expression in any region of the hippocampus (Figure 7C–7E). Thus, sNEP cell delivery did not promote astrocytosis when measured at 28 d postimplantation. Future experiments examining longer time points of sNEP delivery could show that brain inflammation decreases after Aβ clearance.

**Discussion**

Various approaches to lowering Aβ levels within the brain have come to represent the central focus in AD therapeutic development. In this study, we describe the use of ex vivo gene therapy as a viable approach to reducing Aβ levels utilizing a modified Aβ-degrading protease. Replacing the transmembrane domain of the normally membrane-anchored NEP with a signal peptide resulted in the robust secretion of an enzymatically active protease. When syngenic fibroblasts modified to produce sNEP were engrafted into aged APP transgenic mice, the majority of the plaque burden surrounding the graft site was proteolytically cleared. Distal to the graft, in both the medial and lateral hippocampus, significant reductions in plaque burden were also observed as a result of sNEP gene delivery. These results demonstrate the utility of locally elevating the expression of a secreted Aβ-degrading protease as a potential therapeutic approach for AD.

The finding that sNEP was able to reduce plaque burden distal to the graft site may be explained by the diffusion of the sNEP enzyme through the parenchyma, by diffusion of soluble forms of Aβ toward the graft (which may thus be serving as an Aβ sink), or both of these mechanisms. Because
of the low sensitivity of available anti-NEP antibodies, we were unable to detect sNEP diffusion farther than ~800 μm from the graft site, nor could we detect the very low (Figure 1A) endogenous NEP levels expressed by neural tissue [51].

Potential side effects of sNEP expression may arise from the degradation of NEP substrates other than Aβ present in the brain, such as enkephalins. Though studies explicitly searching for such side effects were not performed here, no adverse phenotype was reported in NEP transgenic mice that were protected from Aβ-induced premature death [15], and in this study we observed no enhanced astrocitosis in response to sNEP expression.

A major obstacle to translating the ex vivo methods used in this study into a human disease therapy is the larger dimensions of the human brain, which might require an elevation in Aβ-degrading activity throughout a much broader area. However, our experiment involved just a 28 d exposure to the sNEP-secreting cells, and we found these primary cells to maintain transgene expression in vitro for over a year. One approach to the problem of optimal diffusion of the gene product is to implant the cells where they may have the best access to circulating Aβ, such as in the cerebrospinal fluid or the ventricular wall. Several studies have reported the successful use of encapsulated cell implants that allow for diffusion of therapeutic factors into biological fluids, including cerebrospinal fluid [29,30]. By analogy to the peripheral sink hypothesis for Aβ immunotherapy [52], it may also be feasible and attractive to implant genetically modified cells into peripheral compartments (e.g., the peritoneal cavity or skeletal muscle), where they may substantially increase the Aβ-degrading capacity of the periphery and thus increase the rate of efflux of Aβ across the blood–brain barrier. Moreover, some of the secreted sNEP provided by such a peripheral implant may reach the brain and cerebrospinal fluid to help decrease brain Aβ levels directly. The existence of a subset of hematopoietic cells capable of infiltrating the brain and surrounding plaques has been described [53,54]. Though these cells may not efficiently degrade Aβ themselves, if they could be modified to express an Aβ-degrading protease in a manner similar to the approach we report here, they may provide an anatomically targeted means for local Aβ elimination.

In summary, our findings demonstrate that ex vivo gene delivery of an Aβ-degrading protease rapidly lowers hippocampal Aβ plaque burden in a mouse model of Alzheimer disease. Ex vivo gene delivery and other strategies to elevate Aβ-degrading activity in Alzheimer patients warrant further investigation as potential therapeutic approaches to treat this common and devastating disease.

Acknowledgments

We thank Lennart Mucke for the J20 line of APP transgenic mice, Elan Pharmaceuticals for providing the Aβ ELISA antibodies, and Kelly Dakin for critical review and discussion of the manuscript.

Author contributions. MLH, OL, and DJS designed the study. MLH, MP, CRN, and LL conducted surgical experiments. MLH collected and analyzed the data. MLH and DJS interpreted the data and drafted the manuscript. All authors have reviewed and approved the manuscript.

References

Why Was This Study Done? The researchers here wanted to investigate whether they could use this approach to deliver to the brain one of the proteases that breaks down amyloid plaques in mouse models of Alzheimer disease. The US National Institutes of Health National Institute of Aging has a page of links on Alzheimer disease available online

What Did the Researchers Do and Find? The researchers used a mouse model of Alzheimer disease in which, as the mice age, they develop plaques in the brain made of the same protein found in human Alzheimer disease. The researchers took a particular type of cell from the mice called a fibroblast, used a virus called a lentivirus to insert into the cells a protease called neprilysin that can degrade Aβ, and then injected these altered cells into the brains of the mice. They then compared results of these experiments with what happened when cells with a protease were injected. In addition, before this approach could be used in humans more work would need to be done, including showing that the plaques cause. In addition, before this approach could be used in humans more work would need to be done, including showing that the plaques cause. 

What Do These Findings Mean? These results suggest that this ex vivo gene therapy approach to Alzheimer disease is worth considering further. However, despite showing that the plaques were cleared the researchers did not show whether or not there was any effect on the behavior of the mice, i.e., whether there was any effect on the symptoms that the plaques cause. In addition, before this approach could be used in humans more work would need to be done, including showing that the lentivirus and the protease used were safe in humans, and the injection could be scaled up to the much larger human brain.

Additional Information. Please access these Web sites via the online version of this summary at http://dx.doi.org/10.1371/journal.pmed.0040262. 

- The slides from the original cases that Alois Alzheimer described are available online.
- Current research is discussed by the Alzheimer Research Forum
- News and information for researchers, doctors, and patients, including a 24-hour help line, are available from the Alzheimer’s Association
- Medline Plus, the health information site for patients from the US National Library of Medicine, has a page of links on Alzheimer disease
- The US National Institutes of Health National Institute of Aging has a fact sheet on Alzheimer disease, which is also available in Spanish
- The UK’s National Health Service online information site has an information page on Alzheimer disease

Editors’ Summary 

Background. Alzheimer disease is the most common form of dementia and is increasingly common as people age. It may affect up to half of the population in people over 85 years of age. The classic pathological features of Alzheimer disease in the brain were described 100 years ago by a German neuropathologist, Alois Alzheimer. They consist of plaques, which are made up of a protein called amyloid-β protein (shortened to Aβ) and tangles of another protein, called tau. These proteins accumulate especially in areas of the brain where memory and thought are processed and are believed to be toxic to neurons. There are a number of inherited forms of Alzheimer disease which are caused by mutations either in the precursor protein responsible for the production of Aβ from APP or enhancing clearance of the plaques.